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FATTY ACID PROFILE OF FOUR ANTHEMIS SPECIES GROWING IN İZMİR, TURKEY

ABSTRACT

In this study, the fatty acid compositions of Anthemis aciphylla var. aciphylla, Anthemis pseudocotula, Anthemis macrotis and Anthemis coelopoda var. bourgaei growing in İzmir, Turkey were determined. The fatty acid composition was analyzed using gas chromatography. Generally, C 4:0 butyric acid, C 18:2 linoleic acid and C 6:0 caproic acid were found to be the major fatty acids in all species.

Keywords: Anthemis, Fatty Acid Composition, Butyric Acid,

Linoleic Acid, Caproic Acid

izmir, türkiye'de yetişen dört *anthemis* türünün yağ asidi profili

ÖZ

Bu çalışmada, İzmir, Türkiye'de yetişen Anthemis aciphylla var. Aciphylla, Anthemis pseudocotula, Anthemis macrotis ve Anthemis coelopoda var. bourgaei bitkilerinin yağ asidi içeriği tespit edilmiştir. Yağ asidi içeriği, gaz kromatografisi kullanılarak tespit edilmiştir. Genel olarak tüm türlerde majör yağ asitleri olarak C 4:0 butirik asit, C 18:2 linoleik asit ve C 6:0 kaproik asit bulunmuştur. Anahtar Kelimeler: Anthemis, Yağ Asidi İçeriği, Butirik Asit, Linoleik Asit, Kaproik Asit

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1. INTRODUCTION

The genus Anthemis L., in the family Asteraceae, is represented in the Flora of Turkey by 51 species, 29 of which are endemic to Turkey¹. Generally Anthemis species are known as "papatya" and "yavşan" in the west part of Turkey². These species are extensively used in Turkish folk medicine for the treatment of some diseases like gastrointestinal disorders, hemorrhoid, stomachache, abdominal pain, hepatic diseases and cough²⁻⁶. Anthemis species usually contain some chemical compounds such as sesquiterpene lactones, flavonoids and polyacetylenes⁷⁻¹¹. Also they have wide range of biological activities such as antioxidant^{12-16,19}, antimicrobial¹⁷⁻¹⁹, antiprotozoal²⁰ and antiinflammatory²¹ activities.

2. RESEARCH SIGNIFICANCE

To the best of our knowledge, no previous work has been reported on the fatty acid compositions of four Anthemis species from Turkey except the antioxidant activity study on Anthemis pseudocotula from Konya, Turkey¹⁹. The aim of the present study is to evaluate the fatty acid compositions of Anthemis aciphylla var. aciphylla Boiss., Anthemis pseudocotula Boiss., Anthemis macrotis (Rech.f.) Oberpr. & Vogt and Anthemis coelopoda var. bourgaei Boiss. growing in Izmir, Turkey.

3. MATERIALS AND METHODS

3.1. Plant Materials

The species Anthemis aciphylla var. aciphylla, Anthemis pseudocotula, Anthemis macrotis and Anthemis coelopoda var. bourgaei were collected from Izmir. The plants were identified by one of authors (B. Kivcak) of Ege University. The voucher specimens (herbarium numbers; 1365 for Anthemis aciphylla var. aciphylla, 1333 for Anthemis pseudocotula , 1368 for Anthemis macrotis and 1332 for Anthemis coelopoda var. bourgaei have been deposited at the Herbarium of the Ege University, Faculty of Pharmacy, Department of Pharmacognosy.

3.2. Oil Extraction

The dried and powdered aerial parts of the plant material (40 g) have been extracted by petroleum ether (400 ml) for 6 h at 60°C by Soxhlet extractor. The solvent was evaporated by a rotary evaporator²². The obtained oil was esterified to determine the fatty acid composition. The extraction yields were found of Anthemis aciphylla var. aciphylla, Anthemis pseudocotula, Anthemis macrotis and Anthemis coelopoda var. bourgaei were 0.96%, 1.08%, 2.34% and 1.51%, respectively.

3.3. Preparation of Fatty Acid Methyl Esters (FAMEs)

The fatty acids were esterified into methyl esters by saponification with methanol (50%) containing 5% sodium hydroxide at 100°C for 10 min and transesterified with 14% (v/v) boron trifluoride (BF₃) in methanol 100°C for 5 min²³.

3.4. Fatty Acid Analysis

Fatty acid methyl esters (FAMEs) were analyzed on a HP (Hewlett Packard) Agilent 6890 N model gas chromatograph (GC), equipped with a flame ionization detector (FID) and fitted to a Supelco SP-2380 Fased Silica capillary column (60m, 0.25mm i.d. and 0.2µm). Injector and



detector temperatures were set at 250° C and 260° C, respectively. The oven was programmed at an initial temperature of 140° C and an initial time of 5 min. Thereafter the temperature was increased up to 240° C at a rate of 3° C min⁻¹. The total run time was 41.33 min. Helium was used as the carrier gas (1 ml min⁻¹). Identification of fatty acids was carried out by comparing sample FAME peaks from samples with standarts. The results were expressed as FID response area in the relative percentages. Each reported result is given as the average value of three GC analyses. The results are offered as means \pm S.D.

4. RESULTS AND DISCUSSION

The fatty acid compositions of studied Anthemis species are given in Table 1. GC analysis revealed that the major fatty acids of A. aciphylla var. aciphylla, A. pseudocotula, A. macrotis and A. coelopoda var. bourgaei. were C 4:0 (butyric acid) (73.72%, 58.64%, 72.09% and 68.74%), C 18:2 (linoleic acid) (18.42%, 20.56%, 17.02% and 28.92%) and C 6:0 (caproic acid) (7.68%, 7.76%, 5.13% and 6.61%). The principal fatty acid in our Anthemis species investigated was C 4:0 (butyric acid), in the saturated form of fatty acids (SFAs). Also C 18:2 (linoleic acid) was the major PUFAs (polyunsaturated form of fatty acids).

Table 1. Facty acta compositions of Antinemis species (a				(0)
Fatty Acids	A. aciphylla var. aciphylla	A. pseudocotula	A. macrotis	A. coelopoda var. bourgaei
C 4:0(Butyric acid)	73.72ª	58.64	72.09	68.74
C 6:0 Caproic acid)	7.68	7.76	5.13	6.61
C 10:0(Capric acid)	0.02	0.03	0.01	0.03
C 12:0(Lauric acid)	0.04	0.02	0.03	0.02
C14:0(Mrystic acid)	0.17	0.21	0.13	0.19
C16:0(Palmitic acid)	2.55	1.96	2.03	2.37
C17:0(Heptadecanoic acid)	0.05	0.09	0.11	0.17
C18:0(Stearic acid)	0.79	0.85	1.02	1.11
C23:0(Tricosanoic acid)	0.05	0.07	0.02	0.12
$\sum SFA^{b}$	85.07	69.63	80.57	79.36
C16:1ω7(Palmitoleic acid)	0.09	0.17	0.07	0.11
C 18:1 ω9(Oleic acid)	3.75	4.58	2.25	6.82
∑MUFA ^b	3.84	4.75	2.32	6.93
C 18:2 ω 6(Linoleic acid)	18.42	20.56	17.02	28.92
C20:5n3(Eicosapentanoic acid)	0.79	1.47	2.02	1.52
∑PUFA ^b	19.21	22.03	19.04	30.44
^a Automatic of three lete analyzed b_{CEA} . Caturated fatty acide				

Table 1. Fatty acid compositions of Anthemis species (%)

^a Average of three lots analysed MUFA: Monounsaturated fatty acids ^bSFA: Saturated fatty acids, PUFA: Polyunsaturated fatty acids

In our earlier study, we found that butyric acid (36.13%), arachidic acid (18.80%) and linoleic acid (22.22%) were the major fatty acids of Anthemis widemanniana oil extract²⁴ . recently, cerotic and palmitic acids were claimed to be the major MUFAs in the oil of Anthemis tinctoria var. tinctoria and A. austriaca ²⁵. In a previous study, palmitic acid and α -linolenic acid were reported to be the main constituents of A. triumfetti¹¹. The species have a high content of butyric acid. Butyric acid is found in colon as well as other parts of the digestive tract. This acid has the ability to induce HIV reactivation²⁶. Butyric acid a short chain fatty acid and butyrate



derivatives producing bacteria are promising probiotic treatment for gastrointestinal tract diseases such as inflammatory bowel disease, Chron's disease and ulcerative colitis²⁷. PUFAs ranged from 19.04 to 30.44% while linoleic acid content ranged from 17.02 to 28.92%. Linoleic acid cannot be synthesized by the human body²⁸. Linoleic acid with protective effect against heart diseade has been shown to play arole in the development of the brain and retina²⁹. In conclusion, this is the first report on the fatty acid compositions of Anthemis aciphylla var. aciphylla, Anthemis pseudocotula, Anthemis macrotis and Anthemis coelopoda var. bourgaei.

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REFERENCES

- 1. Davis, P.H., (1984). Flora of Turkey and East Aegean Islands, Edinburgh University Press, Edinburgh.
- Baytop, T., (1984). Therapy with Medicinal Plants in Turkey (past and present), Publication of Istanbul University (first edition), Istanbul.
- Gürhan, G. ve Ezer, N., (2004). Halk Arasında Hemoroit Tedavisinde Kullanılan Bitkiler-I. Hacettepe University Journal of Faculty of Pharmacy, Volume:2, pp:37-60.
- Ugurlu, E. and Secmen, O., (2008). Medicinal Plants Popularly Used in the Villages of Yunt Mountain (Manisa- Turkey). Fitoterapia, Volume: 79, pp:126-131.
- 5. Honda, G., Yeşilada, E., and Tabata, M., (1996). Traditional Medicine in Turkey. VI. Folk Medicine in West Anatolia: Afyon, Kütahya, Denizli, Muğla, Aydın Provinces. Journal of Ethnopharmacology, Volume:5, pp:75-86.
- Kultur, S., (2007). Medicinal Plants Used in Kırklareli Provience (Turkey). Journal of Ethnopharmacology, Volume:111, pp: 341-364.
- Konstantinopoulou, M., Karioti, A., and Skaltsas, S., (2003). Sesquiterpene Lactones from Anthemis altissima and Their Antihelicobacter pylori Activity. Natural Prododucts, Volume:66, pp:699-702.
- Vuckovic, I., Vujusic, L., and Milosavljevic, S., (2005). Phytochemical Investigation of Anthemis cotula L. Serbian Chemical Society, Volume:71, pp:127-133.
- 9. Christensen, L.P., (1992). Acethylenes and Related Compounds in Anthemidae. Phytochemistry, Volume:31, pp:47-51.
- Gonenc, T., Aryropoulou, C., Erdogan, T., Gousiadou, C., Juergenliemk, G., Kıvcak, B., and Skaltsa, H., (2011). Chemical Constituents from Anthemis wiedemanniana Fisch. & Mey. Biochemical Systematics and Ecology, Volume:39, pp:51-55.
- Pavlovic, P., Kovacevic, N., Couladis, M., and Tzakou, O., (2006). Phenolic Constituents of Anthemis triumfetti (L.) DC. Biochemical Systematics and Ecology Volume:34, pp:449-452.
- 12. Bandoniene, D., Pukalskas, A., Venskutonis, P.R., and Gruzdiene, D., (2000). Preliminary Screening of Antioxidant Activity of Some Plant Extracts in Rapeseed Oil. Food Research International, Volume 33, pp: 785-799.



- 13. Povilaityte, V. and Venskutonis, P.R., (2000). Investigation of Antioxidative Activity of Purple Peri, Moldavian Dragonhead and Roman Chamomile Extracts in Rapeseed Oil. Jounal of American Oil Chemical Society, Volume:77, pp:951-956.
- 14. Tawaka, K., Alali, F.Q., Gharaibeh, M., Mohammad, M. and El-Elimat, T., (2007). Antioxidant Activity and Total Phenolic Content of Selected Jordanian Plant Species. Food Chemistry, Volume:104, pp:1372-1378.
- 15. Djeridane, A., Yousfi, M., Nadjemi, B., Boutassouna, D., Stocker, P., and Vidal, N., (2006). Antioxidant Activity of Some Algerian Medicinal Plants Extracts Containing Phenolic Compounds. Food Chemitry, Volume:97, pp:654-660.
- 16. Albayrak, S. and Aksoy, A., (2013). Evaluation of Antioxidant and Antimicrobial Activities of Two Endemic Anthemis Species in Turkey. J Food Biocem. Volume: 37, pp:639-645.
- 17. Qarenghi, M.V., Tereschuk, M.L., Baigori, M.D., and Abdala, L.R., (2000). Antimicrobial Activity of Flowers from Anthemis cotula. Fitoterapia, Volume:71, pp:710-712.
- 18. Buruk, K., Sokmen, A., Aydın, F., and Erturk, M., (2006). Antimicrobial Activity of Some Endemic Plants Growing in the Eastern Black Sea Region, Turkey, Fitoterapia, Volume:77, pp:388-391.
- 19. Orhan, D.D., Ozcelik, B., Hosbas, S., and Vural, M., (2012). Assessment of Antioxidant, Antibacterial, Antimycobacterial, and Antifungal Activities of Some Plants Used as Folk Remedies in Turkey Against Dermatophytes and Yeastlike Fungi. Turkish Journal of Biology, Volume: 36, pp:672-686.
- 20. Karloti, A., Skaltsa, H., and Linden, A., (2007). Anthecularin: A Novel Sesquiterpene Lactone from Anthemis auriculata with Antiprotozoal Activity. Journal of Organic Chemistry, Volume:72, pp:8103-8106.
- 21. Baltacı, S., Kolatan, H. E., Yilmaz, O., and Kivcak, B., (2011). Antiinflammatory Activity of Anthemis aciphylla var. aciphylla Boiss. Turkish Journal of Biology, Volume:35, pp:757-762.
- 22. IUPAC, (1979). Standartds Methods for Analysis of Oils, Fats and Derivatives. 6th ed. Pergamon Press, Oxford, pp: 59-66.
- 23. Yıldırım, N., Sunar, S., Agar, G., Bozari, S., and Aksakal, O., (2009). Biochemical and Molecular Characterization of Some *Centaurea* species Growing in the Eastern Anatolia Region of Turkey. Biochemical Genetics, Volume:47, pp:850-859.
- 24. Gönenç, T.M., Akkol, E.K., Süntar, İ., Erdoğan, T.F., and Kıvçak, B., (2014). Fatty Acid Composition and Preclinical Researchees on Anthemis widemanniana Fisch. & Mey.: Discovery of a New Anti-inflammatory Agent. Pharmacognosy Magazine, Volume:10, Issue:37, pp:1-8.
- 25. Orhan, İ., Deliorman-Orhan D., and Özçelik, B., (2009). Antiviral Activity and Cytotoxicity of the Lipophilic Extracts of Various Edible Plants and Their Fatty Acids. Food Chemistry, Volume:115, pp:701-705.
- 26. Baskett, R.C. and Hentges, D.J., (1973). Shigella flexneri Inhibition by Acetic Acid. Infection and Immunity, Volume:8, pp:91-97.
- 27. Van Immerseel, F., Boyen, F., Gantois, I., Timbermont, L., Bohoz, F., Pasmons, F., Haesebrouck, F., and Ducatelle, R., (2005). Supplementation of Coated Butyric Acid in the Feed Reduces Colonization and Shedding of *Salmonella* in Poultry, Poultry Science, Volume:84, Issue:12, pp:1851-1856.



- 28. Gao, Z., Yin, J., Zhang, J., Ward, R.E., Martin, R.J., and Lefevre, R.J., (2009). Butyrate Improves Insulin Sensitivity and Increase Energy Expenditure in Mice. Diabetes, Volume:58, pp:1509-1517.
- 29. Brown, J.E., (2005). A Critical Review of Methods Used to Estimate Linoleic Acid 6-desaturation Exvivo and Invivo, European Journal of Lipid Science Technology, Volume:107, pp:119-134.
- 30. Swanson, D., Block, R., and Mousa, S.A., (2012). Omega-3 Fatty Acids EPA and DHA: Health Benefits throughout Life, Advances Nutrition, Volume:3, Issue:1, pp:1-7.