



THE POSSIBLE RELATIONSHIPS BETWEEN SOME GENE POLYMORPHISMS AND SJÖGREN'S SYNDROME

SJÖGREN SENDROMU İLE BAZI GEN POLİMORFİZMLERİ ARASINDAKİ OLASI BAĞLANTILAR

Ulkü TERZİ¹ , İlker ATEŞ^{1*} 

¹Ankara University, Faculty of Pharmacy, Department of Toxicology, 06560, Ankara, Türkiye

ABSTRACT

Objective: Sjögren's syndrome is a complex and widespread autoimmune disease whose pathogenesis is not fully elucidated and environmental and genetic factors affect the development of the disease. In order to reveal the effect of genetic contribution, studies have been conducted on the genes previously shown to play a role in other autoimmune diseases such as systemic lupus erythematosus. In addition, two GWAS studies were conducted to investigate the role of more genes in the disease by screening the entire genome and the relationship of previously unknown genes with SS was shown.

Result and Discussion: Studies are being conducted with spontaneous and genetically modified animal models in order to better reveal the relationship between SS and genes and to reinforce the data obtained from humans. In this study, the relationship between the genes previously studied in other autoimmune diseases and the genes associated with SS in GWAS studies and the possible pathways that may contribute to the pathogenesis of the disease through related genes were investigated.

Keywords: Autoimmune disease, gene polymorphisms, genetic toxic effects, genotoxicity, Sjögren's syndrome

ÖZ

Amaç: Sjögren sendromu hala patogenezi tam olarak aydınlatılamamış, hastalık gelişimini çevresel ve genetik faktörlerin etkilediği kompleks ve yaygın bir otoimmün hastalıktır. Genetik katkının etkisini ortaya koymak için daha önce sistemik lupus eritromatozus gibi diğer otoimmün hastalıklarda rolü gösterilen genler üzerinde bu genlerin SS ile ilişkisini ortaya koymak için çalışmalar yapılmıştır. Ayrıca iki GWAS çalışmasıyla da tüm genom taranarak daha fazla genin hastalıkta rolü incelenmiş ve daha önce SS ile ilişkisi bilinmeyen genlerin SS ile ilişkisi gösterilmiştir.

* Corresponding Author / Sorumlu Yazar: İlker Ateş
e-mail / e-posta: ilkerates976@gmail.com, Tel. / Phone: +903122033007

Submitted / Gönderilme : 17.07.2023

Accepted / Kabul : 09.08.2023

Published / Yayınlanma : 20.09.2023

Sonuç ve Tartışma: *SS'in genlerle ilişkisini daha iyi ortaya koymak ve insanlardan elde edilen verilerin pekiştirilmesi için spontan ve genetiği modifiye edilmiş hayvan modelleriyle de çalışmalar yürütülmektedir. Bu çalışmada daha önce diğer otoimmün hastalıklarda incelenen genler ile GWAS çalışmalarında ilişkili bulunan genlerin SS ile ilişkisi, ilişkili bulunan genler üzerinden hastalığın patogenezisine katkısı olabilecek olası yollar irdelenmiştir.*

Anahtar Kelimeler: *Gen polimorfizmi, genetik toksik etkiler, genotoksisite, otoimmün hastalık, Sjögren sendromu*

INTRODUCTION

Sjögren's syndrome (SS) is a chronic autoimmune condition characterized by lymphocytic infiltration in exocrine glands such as saliva and lacrimal glands. Typical clinical findings in patients with SS due to progressive damage to exocrine glands are dry mouth (xerostomia) and dry eye (keratoconjunctivitis sicca). In SS patients, in addition to symptoms of dryness, extraglandular signs such as Raynaud's phenomenon, fatigue, or arthritis are common [1-4]. Another feature of the disease is B cell hyperactivity. Major autoantibodies in SS target the intracellular antigens Ro52/TRIM21, Ro60/TROVE2 and La/SSB antigens, which are ribonucleoprotein-RNA complexes [5,6].

SS predominantly affects premenopausal women, and the incidence is 9: 1 in women compared to men [4,7]. It is known that SS occurs at 0.4-4.0% of the general population [4,8,9]. Modified European and American diagnostic criteria are the most commonly used diagnostic criteria in clinical practice [4,10]. According to this criterion, when it is accompanied by other rheumatoid diseases such as SLE or RA, it is classified as secondary SS (sSS) and SS alone is classified as primary SS (pSS). SS progresses with or without other autoimmune diseases [4,10,11]. Disease severity is determined according to ESSDAI (The EULAR SS disease activity index) [12]. Although mortality in SS is not different from the general population, quality of life in SS patients is significantly affected by reduced morbidity. SS patients are dependent on palliative methods in order to relieve major symptoms of xerostomia along with immunosuppressive methods. However, there are still no effective therapies to restore the SS process or to repair secretion dysfunction [4,13]. The risk of developing Non-Hodgkin lenfs B cell lymphoma is 44 times higher in SS patients compared to healthy individuals [4,14].

The etiopathology of the disease is unknown. It has been suggested that genetic factors, as well as exogenous agents such as Epstein-Barr virus (EBV), Hepatitis C (HCV) and human T-cell leukemia virus-1 (HTLV-1), hormones and microremism may cause the onset of this disease [15].

SS' Relations with Genes

Genetic predisposition is one of the main features of autoimmune diseases [16,17]. SS is a genetically complex disease and little is known about the contribution of genetic factors to the disease. There are studies on monozygotic twins in other autoimmune diseases to investigate genetic contribution. In these studies, the comorbidity between disease and monozygotic twins was reported to be 25-40% in monozygotic twins [18]. However, there are no studies on monozygotic twins for SS. Case reports were made for case monozygotic and dizygotic SS twins only for SS. However, reliable correlation between twins is not evaluated in these presentations [19-21]. The incidence of other autoimmune diseases in the families of SS patients was reported to be 30-35%. Thyroid diseases, SLE and RA are the most common autoimmune diseases [22,23]. To date, gene studies on SS are included in the study of specific genes that may be genetic risk factors.

Candidate Genes

The first genetic studies of SS were carried out in genes that were previously known in the immune system and which had important functions or were shown to affect other autoimmune diseases such as romataid arthritis and systemic lupus erythromatosis (SLE).

In 1977, HLA genes were shown to be a risk factor for SS [24]. HLA (Human Leukocyte Antigens) complex is located on the short arm of chromosome 6 [25-27]. HLA antigens are expressed on many cell surfaces and have an important role in the recognition of antigenic stimulants, stimulation

of the immune system, and regulation of cellular and humoral immunity [26]. HLA complexes are classified into three classes as Class I, Class II and Class III [27].

HLA Class II proteins have the largest hereditary susceptibility to autoimmune diseases including SS. Reported risk haplotypes differ slightly from phenotype and race. The HLA-DR3 SS relationship was shown primarily in the white race, and the SS relationship of HLA-DR3-DQ haplotypes was shown in different ethnic groups [16,17,28-36] However, a meta-analysis identifies DRB1*0301, DQA1*0501, DQB1*0201, and DRB1*03 alleles as risk factors for SS, while identifying DQA1*0201, DQA1*0301 and DQB1*0501 alleles as preservatives [37]. Recently, a strong association between HLA-DRA, HLA-DQB1 and HLA-DQA1 and SS in 6p21 locus in a large study in Europe was reported [38]. In a study in China, HLA-DRB1/HLA-DQA1 in 6p21.3 locus and two independent signals associated with HLA-DPB1/COL11A2 [39]. Deterioration of autoreactive T cell tolerance through the presence of abnormal antigen demonstrates the key role of HLAs in autoimmune diseases. The disease relationship of HLA-suspected alleles is common in autoimmune diseases and different specific alleles and haplotypes are formed, different alleles direct targeting of specific autoantigens [40]. HLA Class II is associated with autoantibody production in SS, whereas anti-Ro/SSA and anti-La/SSB are significantly higher in HLA-DQ1/HLA-DQ2 heterozygous patients [41] but not related to other clinical features [32]. HLA-DRB1*1501-DRB1*0301 is associated with anti-ACA (anticyclic citrullinated antibodies) [42]. Amino acid variations in the hypervariable region (HVR) region of the HLA complex affect peptide binding and T cell presentation; The association of specific variations in binding wells 7 and 9 of HLA-DRB1 with changes in depth and polarity was shown in the Chinese population [35]. Although HLA Class I and HLA Class III genes were also studied in the following years, studies focused on HLA Class II genes.

On the other hand, polymorphisms in non-HLA genes, which have been shown to be associated with other autoimmune diseases, were also investigated in SS. One of the non-HLA genes is STAT4. STAT4 (signal transducers and activators of transcription-4) is an important transcription factor for the transmission of IL-12, IL-23 and Type 1 interferon-mediated signals involved in Th1 and Th17 differentiation, activation of monocytes and INF γ production [43-45]. STAT4 haplotypes have been shown to be a risk factor for the development of SLE and RA in the caucasians and its relationship with SS [46]. STAT4 polymorphism was investigated in different ethnic groups in different loci such as rs7574865 [47] and rs7582694 [48]. In these three studies, it was determined that rs7582694 polymorphism posed a risk for SS. This polymorphism was found to be poorly correlated with m-RNA levels of various interferon-induced genes in peripheral blood mononuclear cells of SS patients [48].

The distinctive feature of this disease is B cell hyperactivity. B cell hyperactivity was demonstrated by the presence of autoantibodies and hypergammaglobulinemia. The most risky group in development of lymphoma -especially non-Hodgkin's lymphoma- in all autoimmune diseases is pSS [49].

In addition, the association of BAFF (B-cell activating factor) polymorphisms in the development of other autoimmune diseases has been shown previously [49-55]. BAFF also known as B lymphocyte stimulator, is a member of the TNF superfamily that regulates the immune [56-58]. A cytokine facilitates B cell survival and maturation [56,58]. It is expressed as membrane bound (mBAFF) and soluble protein (sBAFF) [3,58-60]. Many cells are produced by antigen-presenting cells (B cells, monocytes/macrophages, dendritic cells (DC), plasmacytoid DC, follicular DC), epithelial cells, active T lymphocytes) [58,59]. In the presence of type 1 interferon (INF γ , LI-10, TLR3, TLR4, TLR9, etc.), BAFF expression increases [57,58,60]. Binding of BAFF to BAFFR triggers NF- κ B (non-canonical nuclear factor κ B) signaling [58,61]. The relationship of BAFF with overexpression with mature B cell hyperplasia and development of SLE and SS-like symptoms in lymphoid tissues has been previously demonstrated in experimental models [1,58,62].

It is thought that genetic variation of BAFF increases the risk of developing lymphoma [53,63]. It has been reported that various SNP (single nucleotide polymorphism) in various BAFF genes contribute to anti-Ro and/or anti-La positivity or high sBAFF level [64]. In addition, the association of the BAFF receptor with His159Tyr mutation, which causes deregulation of apoptosis by activation of the NF- κ B pathway, has also been demonstrated [65]. It is also known that BAFF affects Type I and Type II interferon regulation and thus its contribution to SS development is bi-directional with its

contribution to B cell hyperactivity as well as its contribution to cytokine production. Studies on the effect of BAFF on both B cell and cytokine production showed a relationship between gene polymorphism and serum BAFF level, blood and salivary gland BAFF transcription level [66].

TNFAIP3 interacting protein 1 (TNIP1) encoded by the TNIP1 gene is an important signaling protein in the NF- κ B pathway. Together with TNFAIP3 (Tumor necrosis factor alpha inducible protein 3), it acts together with the TNFAIP3 protein to suppress NF- κ B activation. The association of TNIP1 gene polymorphism with many autoimmune diseases such as systemic sclerosis, rheumatoid arthritis (RA), psoriasis, SLE [67-76] was determined. In addition, its relationship with SS and anti-Ro/SSA and anti-La/SSB autoantibody seropositivity in SS were also shown [38]. On the other hand, the TNFAIP3 gene has been reported to be associated with diseases such as SLE and RA [66,77-79], and is also associated with pSS in GWAS studies. Allelic variations of the TNFAIP3 gene have been reported to be associated with pSS [80,81].

Thrombospondin-1 (TSP-1) is an adhesion matrix protein encoded by the THBS1 gene, which activates latent TGF β and some anti-inflammatory cytokines and regulates extracellular and intracellular signaling complexes; expressed in the corneal epithelium, stroma and endothelium. Dry eye is one of the most important symptoms in SS. In this respect, the relationship between THBS1 polymorphism and SS has attracted attention and the relationship between THBS1 gene variations and anisotropy and orientation symmetry coefficients of corneal nerve fibers has been shown [82]

None of the candidate genes reported for polymorphism in SS studies have changed the coding sequences of these genes, and only single SNPs have been studied in studies with the candidate gene approach.

GWAS Studies

Genome-wide association studies (GWAS) are a powerful molecular method that scans the entire DNA to determine the relationship between specific disease phenotypes and any loci. It yields SNPs associated with different polymorphic alleles covering the entire genome. GWAS studies are conducted in large patient and control populations, which is important, and allows a good comparison between races and to determine whether the observed relationship is race-bound. In addition, since these studies are clinically studied in a broad spectrum, the selection of participants is better [39].

Unlike other autoimmune diseases, there are only two GWAS studies for SS. One of them is found in the European population by over 10,000 participants. All patients were diagnosed with SS according to European-American Consensus Criteria. In this study, seven genetic regions were identified that could exceed the statistical threshold $p < 5 \times 10^{-8}$, MHC-II loci, IRF5, STAT4, IL12A, BLK, CXCR5 and TNIP1. The strongest association was found in the HLA-II locus, followed by STAT4 and IRF5. The HLA-II locus, STAT4 and IRF5 were previously identified by the candidate gene approach, a stronger statistical value was obtained with this study, more samples were studied. IL12A, BLK and CXCR5 are important genes in immune signaling and their association with SS was demonstrated for the first time. TNIP1 is involved in the NF κ B pathway and is a new gene associated with SS in the GWAS study. TNFAIP3, DGKQ, and FCGRN2 were found to be statistically poorly correlated. In this study, genes associated with SS are important genes in immune system functions [83].

The other GWAS study was performed in 1090 healthy and 597 SS cases in the Chinese population and 642,832 SNPs were detected. In this study, the strongest relationship was found in the GTF2I gene, which is a general transcription factor, and the other related genes are MHC-II, STAT4, and TNFAIP3 genes (Table 1) [84].

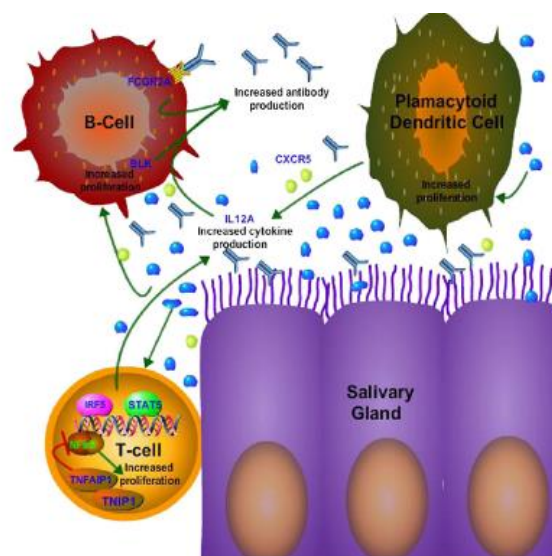
The results of these extensive studies conducted in two different populations show that there are differences and similarities between the European and Chinese populations. GTF2I polymorphism was observed only in Chinese population, while IRF5 polymorphism and other polymorphisms were not seen in Chinese population. The two important genes involved in the NF κ B pathway were found to be related differently in both populations for TNIP1 and TNFAIP3. TNFAIP3 was found to be statistically significant in the Chinese population and weak in the European population. TNIP1 was associated only in the European population. These results indicate different genetic risks for both populations and should be confirmed in further studies.

Table 1. SS Associated Non-HLA genes in GWAS Studies [39]

Alleles with Gene	Function	Determination	References
STAT4	Transcription Factor	2	83
IRF5	Transcription Factor	1	83
IL12A	The cytokine	5	83
BLK	BLK B Cell Kinase	2	83
CXCR5	Chemokine	3	83
TNIP1	NFκB signaling	3	83
GTF2I	Transcription Factor	1	84
TNFAIP3	NFκB signaling	1	84

In these studies, none of the genes associated with SS were lacrimal and salivary glands, proteins associated with nerve conduction in these glands, secretion devices and X-chromosomes. Associated genes are those related to immune system functions. Based on these results, it is seen that the immune system and the differences of activity in the immune system are the most important factors in the pathogenesis of SS.

Although the pathogenesis of SS is still unclear, the mechanisms that may contribute to the pathogenesis of the disease have been proposed based on candidate gene approach and GWAS related genes. The most common pathway is increased interferon signaling and cytokine production. The IRF5, STAT4, and IL12A genes contributing to this pathway were associated with SS in candidate gene approach and GWAS studies. The second possible pathway is B cell production, antibody formation and changes in antibody clearance. BLK, CXCR5 and FCGR2 are also related genes involved in these pathways and involved in these pathways. The third pathway is the NFκB pathway and the genes involved in these studies are TNIP1 and TNFAIP3 (Figure 1) [39].

**Figure 1.** Functional changes in potential paths [39]

Functional Studies

All gene polymorphisms associated with SS were detected in non-coding sequences of the gene of interest. This led to gene studies to evaluate the effect of these polymorphisms on gene expression. In the GWAS study, mRNA expression level of some genes was compared in groups with and without polymorphism. Accordingly, if this polymorphism is found in the transcriptional regulatory region of

the gene, IRF5 and HLA-II gene expression levels are higher in SS patients, and related polymorphisms such as GTF2I have no effect on gene regulation [39].

Depression is a common condition in SS. In a study, the association of platelet serotonin levels with the serotonin transporter gene (5-HTT) polymorphisms in SS patients was investigated and it was reported that platelet serotonin levels were lower in the presence of intronic 5-HTTVNTRin2 (I/s) polymorphism compared to controls [85].

In another study, the relationship between the level of surfactant protein-D (SP-D) and SP-D genotype, which is thought to have an effect on the pathophysiology of the disease, was examined but no relationship was found between them [86].

Protein tyrosine phosphatase non-receptor type 22 (PTPN22) T cells, B cells, natural killer cells, DCs, monocytes and macrophages are expressed in many immune-related cells [87], regulate T cell receptor signaling [88]. PTPN22 in myeloid cells potential TLR-induced Type I interferon (INF) production [89]. PTPN22 allelic variations are risk factors for many autoimmune diseases such as Type 1 diabetes, RA, SLE and hashimoto thyroiditis [87-90]. Many PTPN22W-related diseases have been reported to be associated with impaired adaptive immunity and autoantibody production [88]. The frequency of phenotype PTPN22W* variation in pSS patients with low Type I INF blood levels was reported to be higher than controls and pSS errors with high Type I INF blood levels [91]. Gene variations of the F11R protein, which has many functions such as intracellular signaling, regulation of cellular permeability, stimulation of cell translocation during inflammatory processes, and cytokine production were investigated in SS. In this study, it was reported that some variations were associated with SS and F11R mRNA expression was lower in SS patients compared to healthy controls [92].

Animal Studies

In order to understand SS pathogenesis, human genotyping and genotype and phenotype compatibility studies are conducted concurrently with animal studies. Thus, the results obtained from humans and animals are compared and tried to prove the accuracy of the predicted mechanisms.

Animal models have been developed in order to understand the pathogenesis of SS and studies are being conducted on these models. Animal models are very valuable for elucidating pathogenesis and applying therapeutic approaches. Animal models are important especially in the absence of very clear clinical indicators at the onset of the disease and inability to detect changes in disease onset and help researchers to monitor changes that contribute to pathogenesis at the onset of disease [93]. The animals developed and used for this purpose are genetically similar to humans, such as mice and rats suitable for gene cloning and transgenic modification [94]. However, the data obtained from animal studies are limited in the elucidation of the disease due to factors such as developmental process between humans and mice, differences in adaptive and innate immune response and environmental conditions [95].

Spontaneous animal models allow for an understanding of the tendency or resistance loci of the disease, the time-related profile of disease formation and progression. Many spontaneous animal models used for pSS are derived from non-obese diabetic (NOD) mice. NODs develop not only Type 1 diabetes but SS-like autoimmune exocrinopathy. For this reason, SS is one of the most powerful tools for revealing the pathological mechanism [96]. Infiltration of the salivary and lacrimal glands occurs at 12-16 weeks of age. In addition, autoantibodies such as ANA, anti-SSA/Ro, anti-SSB/La and anti-M3R are also seen in SS patients [97,98].

In addition, genetically modified HTLV-1 tax transgenic (Tg) mouse, IL-6 Tg mouse, IL-10 Tg mouse, IL-12 Tg mouse, IL-14 α Tg mouse, B-cell, to investigate the pathophysiology of SS-like diseases activating factor (BAFF) Tg mouse, retinoblastoma associated protein 48 (RbAp48) Tg transgenic mouse species and transforming growth factor beta 1 (TGF- β 1) KO (knock out)mouse, inhibitor of differentiation 3 (Id3) -/- KO mouse, aromatase-deficient (such as Ar KO) mouse, phosphoinositide 3-kinase (PI3K) KO mouse and thrombospondin-1 (TSP-1) -deficient conjugate mouse are also used [99].

Some of the results from animal experiments are consistent with previous human studies. For example, IL-12 transgenic mouse model expressing both subgroups of IL-12 showed features similar to human SS such as increased SSB autoantibody production, decreased saliva flow, and lymphocytic

infiltration in glands [95]. In the GWAS study, the IL-12 gene is one of the genes associated with SS [39].

RESULT AND DISCUSSION

Despite the studies, the etiopathology of the disease is still unknown. Genetic factors, as well as exogenous and endogenous factors, have been shown to cause the onset of the disease.

Polymorphisms of the HLA, STAT4, BAFF and TNFAIP3 genes and the expression of these genes in tissues such as blood and salivary glands have been shown in the studies on the association of genes associated with SS in other autoimmune diseases such as romataid arthritis and SLE.

Again, two GWAS studies with larger populations showed the association of HLA, STAT4, IRF5, IL12A, BLK, CXCR5, TNIP1, GTF2I and TNFAIP3 genes and the expression of these genes in SS.

In these studies, mechanisms that could contribute to the pathogenesis of the disease were predicted from related genes. The most common of these is increased interferon signaling and cytokine production pathways. The second possible pathway is B cell production, changes in antibody formation and antibody clearance, and the third pathway is the NF κ B pathway. Revealing the mechanisms that may contribute to the pathogenesis is very important in terms of contributing to the development of new drugs for the treatment of the disease.

In addition to human studies, gene polymorphisms which may be related to animal studies and appropriate animal models and the expression of this gene expression in tissues such as blood and salivary gland are compared with human studies and possible mechanisms are explained.

Although studies on suspected genes have shown that many genes may contribute to the pathogenesis of the disease, studies on genes that are not yet studied but that may be related to the disease should also be conducted. So far, gene studies are only SNP studies and there is a need for studies to determine multigenetic factors and to produce more powerful data in larger populations.

AUTHORSHIP CONTRIBUTIONS

Concept: U.T., I.A.; Design: I.A.; Control: I.A.; Sources: U.T., I.A.; Materials: I.A.; Data Collection and/or Processing: U.T., I.A.; Analysis and/or Interpretation: U.T., I.A.; Literature Review: U.T.; Manuscript Writing: U.T., I.A.; Critical Review: I.A.; Other: -

CONFLICT OF INTEREST

The authors declare that there is no real, potential, or perceived conflict of interest for this article.

REFERENCES

1. Mackay, F., Woodcock, S.A., Lawton, P., Ambrose, C., Baetscher, M., Schneider, P., Tschopp, J., Browning, J.L. (1999). Mice transgenic for BAFF develop lymphocytic disorders along with autoimmune manifestations. *Journal of Experimental Medicine*, 190(11), 1697-1710. [\[CrossRef\]](#)
2. Pers, J.O., Daridon, C., Devauchelle, V., Jousse, S., Saraux, A., Jamin, C., Youinou, P. (2005). BAFF overexpression is associated with autoantibody production in autoimmune diseases. *Annals of the New York Academy of Sciences*, 1050, 34-39. [\[CrossRef\]](#)
3. Ittah, M., Miceli-Richard, C., Eric, G.J., Lavie, F., Lazure, T., Ba, N., Sellam, J., Lepajolec, C., Mariette, X. (2006). B cell-activating factor of the tumor necrosis factor family (BAFF) is expressed under stimulation by interferon in salivary gland epithelial cells in primary Sjögren's syndrome. *Arthritis Research & Therapy*, 8(2), R51. [\[CrossRef\]](#)
4. Sarigul, M., Yazisiz, V., Bassorgun, C.I., Ulker, M., Avci, A.B., Erbasan, F., Gelen, T., Gorczyński, R.M., Terzioglu, E. (2010). The numbers of Foxp3 + Treg cells are positively correlated with higher grade of infiltration at the salivary glands in primary Sjogren's syndrome. *Lupus*, 19(2), 138-145. [\[CrossRef\]](#)
5. Higgs, R., Gabhann, J.N., Larbi, N.B., Breen, E.P., Fitzgerald, K.A., Jefferies, C.A. (2008). The E3 ubiquitin ligase Ro52 negatively regulates IFN-beta production post-pathogen recognition by polyubiquitin-mediated degradation of IRF3. *Journal of Immunology*, 181(3), 1780-1786. [\[CrossRef\]](#)

6. Higgs, R., Lazzari, E., Wynne, C., Gabhann, J.N., Espinosa, A., Wahren-Herlenius, M., Jefferies, C.A. (2010). Self protection from anti-viral responses-Ro52 promotes degradation of the transcription factor IRF7 downstream of the viral Toll-Like receptors. *PloS One*, 5(7), e11776. [\[CrossRef\]](#)
7. Oke, V., Wahren-Herlenius, M. (2012). The immunobiology of Ro52 (TRIM21) in autoimmunity: A critical review. *Journal of Autoimmunity*, 39(1-2), 77-82. [\[CrossRef\]](#)
8. Yoshimi, R., Chang, T.H., Wang, H., Atsumi, T., Morse, H.C., Ozato, K. (2009). Gene disruption study reveals a nonredundant role for TRIM21/Ro52 in NF-kappaB-dependent cytokine expression in fibroblasts. *Journal of Immunology*, 182(12), 7527-7538. [\[CrossRef\]](#)
9. Mallery, D.L., McEwan, W.A., Bidgood, S.R., Towers, G.J., Johnson, C.M., James, L.C. (2010). Antibodies mediate intracellular immunity through tripartite motif-containing 21 (TRIM21). *Proceedings of the National Academy of Sciences of the United States of America*, 107(46), 19985-19990. [\[CrossRef\]](#)
10. Jauharoh, S.N.A., Saegusa, J., Sugimoto, T., Ardianto, B., Kasagi, S., Sugiyama, D., Kurimoto, C., Tokuno, O., Nakamachi, Y., Kumagai, S., Kawano, S. (2012). SS-A/Ro52 promotes apoptosis by regulating Bcl-2 production. *Biochemical and Biophysical Research Communications*, 417(1), 582-587. [\[CrossRef\]](#)
11. Espinosa, A., Zhou, W., Ek, M., Hedlund, M., Brauner, S., Popovic, K., Horvath, L., Wallerskog, T., Oukka, M., Nyberg, F., Kuchroo, V.K., Wahren-Herlenius, M. (2006). The Sjogren's syndrome-associated autoantigen Ro52 is an E3 ligase that regulates proliferation and cell death. *Journal of Immunology*, 176(10), 6277-6285. [\[CrossRef\]](#)
12. Seror, R., Ravnaud, P., Bowman, S.J., Baron, G., Tzioufas, A., Theander, E., Gottenberg, J.E., Bootsma, H., Mariette, X., Vitali, C., EULAR Sjögren's Task Force. (2010). EULAR Sjogren's syndrome disease activity index: Development of a consensus systemic disease activity index for primary Sjogren's syndrome. *Annals of the Rheumatic Diseases*, 69(6), 1103-1109. [\[CrossRef\]](#)
13. Espinosa, A., Dardalhon, V., Brauner, S., Ambrosi, A., Higgs, R., Quintana, F.J., Sjöstrand, M., Eloranta, M.L., Gabhann, J.N., Winqvist, O., Sundelin, B., Jefferies, C.A., Rozell, B., Kuchroo, V.K., Wahren-Herlenius, M. (2009). Loss of the lupus autoantigen Ro52/Trim21 induces tissue inflammation and systemic autoimmunity by disregulating the IL-23-Th17 pathway. *Journal of Experimental Medicine*, 206(8), 1661-1671. [\[CrossRef\]](#)
14. Verhagen, A.P.M., Pruijn, G.J.M. (2011). Are the Ro RNP-associated Y RNAs concealing microRNAs? Y RNA-derived miRNAs may be involved in autoimmunity. *BioEssays*, 33(9), 674-682. [\[CrossRef\]](#)
15. Sandhya, P., Kurien, B.T., Danda, D., Scofield, R.H. (2017). Update on pathogenesis of Sjögren's syndrome. *Current Rheumatology Reviews*, 13(1), 5-22. [\[CrossRef\]](#)
16. Bolstad, A.I., Wassmuth, R., Haga, H.J., Jonsson, R. (2001). HLA markers and clinical characteristics in Caucasians with primary Sjogren's syndrome. *The Journal of Rheumatology*, 28(7), 1554-1562.
17. Perez, P., Anaya, J.M., Aguilera, S., Urzúa, U., Munroe, D., Molina, C., Hermoso, M.A., Cherry, J.M., Allende, C., Olea, N., Ruiz-Narvaez, E., Gonzalez, M.J. (2009). Gene expression and chromosomal location for susceptibility to Sjogren's syndrome. *Journal of Autoimmunity*, 33(2), 99-108. [\[CrossRef\]](#)
18. Bogdanos, D.P., Smyk, D.S., Rigopoulou, E.I., Mytilinaiou, M.G., Heneghan, M.A., Selmi, C., Gershwin, M.E. (2012). Twin studies in autoimmune disease: Genetics, gender and environment. *Journal of Autoimmunity*, 38(2-3), J156-J169. [\[CrossRef\]](#)
19. Besana, C., Salmaggi, C., Pellegrino, C., Pierro, L., Vergani, S., Faravelli, A., Rugarli, C. (1991). Chronic bilateral dacryo-adenitis in identical twins: A possible incomplete form of Sjögren syndrome. *European Journal of Pediatrics*, 150(9), 652-655. [\[CrossRef\]](#)
20. Bolstad, A.I., Haga, H.J., Wassmuth, R., Jonsson, R. (2000). Monozygotic twins with primary Sjogren's syndrome. *The Journal of Rheumatology*, 27(9), 2264-2266.
21. Houghton, K.M., Cabral, D.A., Petty, R.E., Tucker, L.B. (2005). Primary Sjögren's syndrome in dizygotic adolescent twins: One case with lymphocytic interstitial pneumonia. *The Journal of Rheumatology*, 32(8), 1603-1606.
22. Reveille, J.D., Wilson, R.W., Provost, T.T., Bias, W.B., Arnett, F.C. (1984). Primary Sjögren's syndrome and other autoimmune diseases in families. Prevalence and immunogenetic studies in six kindreds. *Annals of Internal Medicine*, 101(6), 748-756. [\[CrossRef\]](#)
23. Anaya, J.M., Tobon, G.J., Vega, P., Castiblanco, J. (2006). Autoimmune disease aggregation in families with primary Sjögren's syndrome. *The Journal of Rheumatology*, 33(11), 2227-2234.
24. Chused, T.M., Kassan, S.S., Opelz, G., Moutsopoulos, H.M., Terasaki, P.I. (1977). Sjögren's syndrome association with HLA-Dw3. *The New England Journal of Medicine*, 296(16), 895-897. [\[CrossRef\]](#)
25. Klein, J., Sato, A. (2000). The HLA system. First of two parts. *The New England Journal of Medicine*, 343(10), 702-709.

26. Sumitran-Holgersson, S. (2008). Beyond ABO and human histocompatibility antigen: Other histocompatibility antigens with a role in transplantation. *Current Opinion in Organ Transplantation*, 13(4), 425-429. [\[CrossRef\]](#)
27. Howell, W.M., Carter, V., Clark, B. (2010). The HLA system: Immunobiology, HLA typing, antibody screening and crossmatching techniques. *Journal of Clinical Pathology*, 63(5), 387-390. [\[CrossRef\]](#)
28. Guggenbuhl, P., Jean, S., Jego, P., Grosbois, B., Chalès, G., Semana, G., Lancien, G., Veillard, E., Pawlotsky, P.Y., Perdriger, A. (1998). Primary Sjögren's syndrome: Role of the HLA-DRB1*0301-*1501 heterozygotes. *The Journal of Rheumatology*, 25(5), 900-905.
29. Jean, S., Quelvennec, E., Alizadeh, M., Guggenbuhl, P., Birebent, B., Perdriger, A., Grosbois, B., Pawlotsky, P.Y., Semana, G. (1998). DRB1*15 and DRB1*03 extended haplotype interaction in primary Sjögren's syndrome genetic susceptibility. *Clinical and Experimental Rheumatology*, 16, 725-728.
30. Loiseau, P., Lepage, V., Djelal, F., Busson, M., Tamouza, R., Raffoux, C., Menkes, C.J., Meyer, O., Charron, D., Goldberg, D. (2001). HLA class I and class II are both associated with the genetic predisposition to primary Sjögren syndrome. *Human Immunology*, 62(7), 725-731. [\[CrossRef\]](#)
31. Anaya, J.M., Correa, P.A., Mantilla, R.D., Arcos-Burgos, M. (2002). TAP, HLA-DQB1, and HLA-DRB1 polymorphism in Colombian patients with primary Sjögren's syndrome. *Seminars in Arthritis and Rheumatism*, 31(6), 396-405. [\[CrossRef\]](#)
32. Gottenberg, J.E., Busson, M., Loiseau, P., Cohen-Solal, J., Lepage, V., Charron, D., Sibilia, J., Mariette, X. (2003). In primary Sjögren's syndrome, HLA class II is associated exclusively with autoantibody production and spreading of the autoimmune response. *Arthritis and Rheumatism*, 48(8), 2240-2245. [\[CrossRef\]](#)
33. Nakken, B., Jonsson, R., Brokstad, K.A., Omholt, K., Nerland, A.H., Haga, H.J., Halse, A.K. (2008). Associations of MHC class II alleles in Norwegian primary Sjögren's syndrome patients: Implications for development of autoantibodies to the Ro52 autoantigen. *Scandinavian Journal of Immunology*, 54, 428-433. [\[CrossRef\]](#)
34. Hernández Molina, G., Vargas Alarcón, G., Rodríguez Pérez, J., Martínez-Rodríguez, N., Lima, G., Sánchez-Guerrero, J. (2015). High resolution HLA analysis of primary and secondary Sjögren's syndrome: A common immunogenetic background in Mexican patients. *Rheumatology International*, 35(4), 643-649. [\[CrossRef\]](#)
35. Huang, R., Yin, J., Chen, Y., Deng, F., Chen, J., Gao, X., Liu, Z., Yu, X., Zheng, J. (2015). The amino acid variation within the binding pocket 7 and 9 of HLA-DRB1 molecules are associated with primary Sjögren's syndrome. *Journal of Autoimmunity*, 57, 53-59. [\[CrossRef\]](#)
36. Lagha, A., Messadi, A., Boussaidi, S., Kochbati, S., Tazeghdenti, A., Ghazouani, E., Almawi, W.Y., Yacoubi-Loueslati, B. (2016). HLA DRB1/DQB1 alleles and DRB1-DQB1 haplotypes and the risk of rheumatoid arthritis in Tunisians: A population-based case-control study. *HLA*, 88, 100-109. [\[CrossRef\]](#)
37. Cruz-Tapias, P., Rojas-Villarraga, A., Maier-Moore, S., Anaya, J.M. (2012). HLA and Sjögren's syndrome susceptibility. A meta-analysis of worldwide studies. *Autoimmunity Reviews*, 11(4), 281-287. [\[CrossRef\]](#)
38. Lessard, C.J., Li, H., Adrianto, I., Ice, J.A., Rasmussen, A., Grundahl, K.M., Kelly, J.A., Dozmorov, M.G., Miceli-Richard, C., Bowman, S., Lester, S., Eriksson, P., Eloranta, M.L., Brun, J.G., Goransson, L.G., Harboe, E., Guthridge, J.M., Kaufman, K.M., Kvarnström, M., Jazebi, H., Graham, D.S.C., Grandits, M.E., Nazmul-Hossain, A.N.M., Patel, K., Adler, A.J., Maier-Moore, J.S., Farris, A.D., Brennan, M.T., Lessard, J.A., Chodosh, J., Gopalakrishnan, R., Hefner, K.S., Houston, G.D., Huang, A.J.W., Hughes, P.J., Lewis, D.M., Radfar, L., Rohrer, M.D., Stone, D.U., Wren, J.D., Vyse, T.J., Gaffney, P.M., James, J.A., Omdal, R., Wahren-Herlenius, M., Illei, G.G., Witte T., Jonsson, R., Rischmueller, M., Rönnblom, L., Nordmark, G., Ng, W.F., UK Primary Sjögren's Syndrome Registry, Mariette, X., Anaya, J.M., Rhodus, N.L., Segal, B.M., Scofield, R.H., Montgomery, C.G., Harley, J.B., Sivils, K.L. (2013). Variants at multiple loci implicated in both innate and adaptive immune responses are associated with Sjögren's syndrome. *Nature Genetics*, 45(11), 1284-1292. [\[CrossRef\]](#)
39. Li, Y., Zhang, K., Chen, H., Sun, F., Xu, J., Wu, Z., Li, P., Zhang, L., Du, Y., Luan, H., Li, X., Wu, L., Li, H., Wu, H., Li, X., Li, X., Zhang, X., Gong, L., Dai, L., Sun, L., Zuo, X., Xu, J., Gong, H., Li, Z., Tong, S., Wu, M., Li, X., Xiao, W., Wang, G., Zhu, P., Shen, M., Liu, S., Zhao, D., Liu, W., Wang, Y., Huang, C., Jiang, Q., Liu, G., Liu, B., Hu, S., Zhang, W., Zhang, Z., You, X., Li, M., Hao, W., Zhao, C., Leng, X., Bi, L., Wang, Y., Zhang, F., Shi, Q., Qi, W., Zhang, X., Jia, Y., Su, J., Li, Q., Hou, Y., Wu, Q., Xu, D., Zheng, W., Zhang, M., Wang, Q., Fei, Y., Zhang, X., Li, J., Jiang, Y., Tian, X., Zhao, L., Wang, L., Zhou, B., Li, Y., Zhao, Y., Zeng, X., Ott, J., Wang, J., Zhang, F. (2013). A genome-wide association study in Han Chinese identifies a susceptibility locus for primary Sjögren's syndrome at 7q11.23. *Nature Genetics*, 45(11), 1361-1365. [\[CrossRef\]](#)

40. Fernando, M.M.A., Stevens, C.R., Walsh, E.C., De Jager, P.L., Goyette, P., Plenge, R.M., Vyse, T.J., Rioux, J.D. (2008). Defining the role of the MHC in autoimmunity: A review and pooled analysis. *PLoS Genetics*, 4(4), e1000024. [\[CrossRef\]](#)
41. Harley, J.B., Reichlin, M., Arnett, F.C., Alexander, E.L., Bias, W.B., Provost, T.T. (1986). Gene interaction at HLA-DQ enhances auto-antibody production in primary Sjögren's syndrome. *Science*, 232(4754), 1145-1147. [\[CrossRef\]](#)
42. Mohammed, K., Pope, J., Le Riche, N., Brintnell, W., Cairns, E., Coles, R., Bell, D.A. (2009). Association of severe inflammatory polyarthritis in primary Sjögren's syndrome: Clinical, serologic, and HLA analysis. *The Journal of Rheumatology*, 36(9), 1937-1942. [\[CrossRef\]](#)
43. Mathur, A.N., Chang, H.C., Zisoulis, D.G., Stritesky, G.L., Yu, Q., O'Malley, J.T., Kapur, R., Levy, D.E., Kansas, G.S., Kaplan, M.H. (2007). Stat3 and Stat4 direct development of IL-17-secreting Th cells. *Journal of Immunology*, 178(8), 4901-4907. [\[CrossRef\]](#)
44. Korman, B.D., Kastner, D.L., Gregersen, P.K., Remmers, E.F. (2008). STAT4: Genetics, mechanisms, and implications for autoimmunity. *Current Allergy and Asthma Reports*, 8(5), 398-403. [\[CrossRef\]](#)
45. Liu, Q.F., Li, Y., Zhao, Q.H., Wang, Z.Y., Hu, S., Yang, C.Q., Ye, K., Li, L. (2015). Association of STAT4 rs7574865 polymorphism with susceptibility to inflammatory bowel disease: A systematic review and meta-analysis. *Clinics and Research in Hepatology and Gastroenterology*, 39(5), 627-636. [\[CrossRef\]](#)
46. Korman, B.D., Alba, M.I., Le, J.M., Alevizos, I., Smith, J.A., Nikolov, N.P., Kastner, D.L., Remmers, E.F., Illei, G.G. (2008b). Variant form of STAT4 is associated with primary Sjögren's syndrome. *Genes and Immunity*, 9(3), 267-270. [\[CrossRef\]](#)
47. Palomino-Morales, R.J., Diaz-Gallo, L.M., Witte, T., Anaya, J.M., Martin, J. (2010). Influence of STAT4 polymorphism in primary Sjogren's syndrome. *The Journal of Rheumatology*, 37(5), 1016-1019. [\[CrossRef\]](#)
48. Gestermaan, N., Mekinian, A., Comets, E., Loiseau, P., Puechal, X., Hachulla, E., Gottenberg, J.E., Mariette, X., Miceli-Richard, C. (2010). STAT4 is a confirmed genetic risk factor for Sjögren's syndrome and could be involved in type 1 interferon pathway signaling. *Genes and Immunity*, 11(5), 432-438. [\[CrossRef\]](#)
49. Novak, A.J., Slager, S.L., Fredericksen, Z.S., Wang, A.H., Manske, M.M., Ziesmer, S., Liebow, M., Macon, W.R., Dillon, S.R., Witzig, T.E., Cerhan, J.R., Ansell, S.M. (2009). Genetic variation in B-cell-activating factor is associated with an increased risk of developing B-cell non-Hodgkin lymphoma. *Cancer Research*, 69 (10), 4217-4224. [\[CrossRef\]](#)
50. Lavie, F., Miceli-Richard, C., Quillard, J., Roux, S., Leclerc, P., Mariette X. (2004). Expression of BAFF (BLYS) in T cells infiltrating labial salivary glands from patients with Sjogren's syndrome. *The Journal of Pathology*, 202(4), 496-502. [\[CrossRef\]](#)
51. Nossent, J.C., Lester, S., Zahra, D., Mackay, C.R., Rischmueller, M. (2008). Polymorphism in the 5' regulatory region of the B-lymphocyte activating factor gene is associated with the Ro/La autoantibody response and serum BAFF levels in primary Sjogren's syndrome. *Rheumatology*, 47(9), 1311-1316. [\[CrossRef\]](#)
52. Hildebrand, J.M., Luo, Z., Manske, M.K., Price-Troska, T., Ziesmer, S.C., Lin, W., Hostager, B.S., Slager, S.L., Witzig, T.E., Ansell, S.M., Cerhan, J.R., Bishop, G.A., Novak, A.J. (2010). A BAFF-R mutation associated with non-Hodgkin lymphoma alters TRAF recruitment and reveals new insights into BAFF-R signaling. *Journal of Experimental Medicine*, 207(12), 2569-2579. [\[CrossRef\]](#)
53. Baek, A., Park, H.J., Na, S.J., Shim, D.S., Moon, J.S., Yang, Y., Choi, Y.C. (2012). The expression of BAFF in the muscles of patients with dermatomyositis. *Journal of Neuroimmunology*, 249(1-2), 96-100. [\[CrossRef\]](#)
54. Ohmatsu, H., Sugaya, M., Miyagaki, T., Suga, H., Fujita, H., Asano, Y., Sato, S. (2012). BAFF levels are increased in lesional skin and sera in patients with cutaneous T-cell lymphoma. *The British Journal of Dermatology*, 167(2), 359-367. [\[CrossRef\]](#)
55. Zhai, K., Tian, X., Wu, C., Lu, N., Chang, J., Huang, L., Zhang, T., Zhou, Y., Qiao, Y., Yu, D., Tan, W., Chen, J., Lin, D. (2012). Cytokine BAFF gene variation is associated with survival of patients with T-cell lymphomas. *Clinical Cancer Research*, 18(8), 2250-2256. [\[CrossRef\]](#)
56. Mackay, F., Browning, J.L. (2002). BAFF: A fundamental survival factor for B cells. *Nature Reviews Immunology*, 2(7), 465-475. [\[CrossRef\]](#)
57. Vincent, F.B., Saulep-Easton, D., Figggett, W.A., Fairfax, K.A., Mackay, F. (2013). The BAFF/APRIL system: emerging functions beyond B cell biology and autoimmunity. *Cytokine & Growth Factor Reviews*, 24(3), 203-215. [\[CrossRef\]](#)
58. Novak, A.J., Grote, D.M., Ziesmer, S.C., Kline, M.P., Manske, M.K., Slager, S., Witzig, T.E., Shanafelt, T., Call, T.G., Kay, N.E., Jelinek, D.F., Cerhan, J.R., Gross, J.A., Harder, B., Dillon, S.R., Ansell, S.M.

- (2006). Elevated serum B-lymphocyte stimulator levels in patients with familial lymphoproliferative disorders. *Journal of Clinical Oncology*, 24(6), 983-987. [\[CrossRef\]](#)
59. Mackay, F., Schneider, P. (2009). Cracking the BAFF code. *Nature Reviews Immunology*, 9(7), 491-502. [\[CrossRef\]](#)
60. Manetta, J., Bina, H., Ryan, P., Fox, N., Witcher, D.R., Kikly, K. (2014). Generation and characterization of tabalumab, a human monoclonal antibody that neutralizes both soluble and membrane-bound B-cell activating factor. *Journal of Inflammation Research*, 2014(7), 121-131. [\[CrossRef\]](#)
61. Schneider, P. (2005). The role of APRIL and BAFF in lymphocyte activation. *Current Opinion in Immunology*, 17(3), 282-289. [\[CrossRef\]](#)
62. Gross, J.A., Johnston, J., Mudri, S., Enselman, R., Dillon, S.R., Madden, K., Xu, W., Parrish-Novak, J., Foster, D., Lofton-Day, C., Moore, M., Littau, A., Grossman, A., Haugen, H., Foley, K., Blumberg, H., Harrison, K., Kindsvogel, W., Clegg, C.H. (2000). TACI and BCMA are receptors for a TNF homologue implicated in B-cell autoimmune disease. *Nature*, 404(6781), 995-999. [\[CrossRef\]](#)
63. Nezos, A., Gravani, F., Tassidou, A., Kapsogeorgou, E.K., Voulgarelis, M., Koutsilieris, M., Crow, M.K., Mavragani, C.P. (2015). Type I and II interferon signatures in Sjögren's syndrome pathogenesis: Contributions in distinct clinical phenotypes and Sjögren's related lymphomagenesis. *Journal of Autoimmunity*, 63, 47-58. [\[CrossRef\]](#)
64. Thompson, N., Isenberg, D., Jury, E.C., Ciurtin, C. (2016). Exploring BAFF: Its expression, receptors and contribution to the immunopathogenesis of Sjögren's syndrome. *Rheumatology*, 55, 15. [\[CrossRef\]](#)
65. Novak, A.J., Bram, R.J., Kay, N.E., Jelinek, D.F. (2002). Aberrant expression of B-lymphocyte stimulator by B chronic lymphocytic leukemia cells: a mechanism for survival. *Blood*, 100(8), 2973-2979. [\[CrossRef\]](#)
66. Graham, R.R., Cotsapas, C., Davies, L., Hackett, R., Lessard, C.J., Leon, J.M., Burtt, N.P., Guiducci, C., Parkin, M., Gates, C., Plenge, R.M., Behrens, T.W., Wither, J.E., Rioux, J.D., Fortin, P.R., Graham, D.C., Wong, A.K., Vyse, T.J., Daly, M.J., Altshuler, D., Moser, K.L., Gaffney, P.M. (2008). Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus. *Nature Genetics*, 40(9), 1059-1061. [\[CrossRef\]](#)
67. Thomson, W., Barton, A., Ke, X., Eyre, S., Hinks, A., Bowes, J., Donn, R., Symmons, D., Hider, S., Bruce, I.N., Wellcome Trust Case Control Consortium, Wilson, A.G., Marinou, I., Morgan, A., Emery, P., YEAR Consortium, Carter, A., Steer, S., Hocking, L., Reid, D.M., Wordsworth, P., Harrison, P., Strachan, D., Worthington, J. (2007). Rheumatoid arthritis association at 6q23. *Nature Genetics*, 39(12), 1431-1433. [\[CrossRef\]](#)
68. Wellcome Trust Case Control Consortium (2007). Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*, 447, 661-678. [\[CrossRef\]](#)
69. Musone, S.L., Taylor, K.E., Lu, T.T., Nititham, J., Ferreira, R.C., Ortmann, W., Shifrin, N., Petri, M.A., Kamboh, M.I., Manzi, S., Seldin, M.F., Gregersen, P.K., Behrens, T.W., Ma, A., Kwok, P.Y., Criswell, L.A. (2008). Multiple polymorphisms in the TNFAIP3 region are independently associated with systemic lupus erythematosus. *Nature Genetics*, 40(9), 1062-1064. [\[CrossRef\]](#)
70. Fung, E.Y.M.G., Smyth, D.J., Howson, J.M.M., Cooper, J.D., Walker, N.M., Stevens, H., Wicker, L.S., Todd, J.A. (2009). Analysis of 17 autoimmune disease-associated variants in type 1 diabetes identifies 6q23/TNFAIP3 as a susceptibility locus. *Genes and Immunity*, 10(2), 188-191. [\[CrossRef\]](#)
71. Gateva, V., Sandling, J.K., Hom, G., Taylor, K.E., Chung, S.A., Sun, X., Ortmann, W., Kosoy, R., Ferreira, R.C., Nordmark, G., Gunnarsson, I., Svenungsson, E., Padyukov, L., Sturfelt, G., Jönsen, A., Bengtsson, A.A., Rantapää-Dahlqvist, S., Baechler, E.C., Brown, E.E., Alarcon, G.S., Edberg, J.C., Ramsey-Goldman, R., McGwin Jr, G., Reveille, J.D., Vila, L.M., Kimberly, R.P., Manzi, S., Petri, M.A., Lee, A., Gregersen, P.K., Seldin, M.F., Rönnblom, L., Criswell, L.A., Syvanen, A.-C., Behrens, T.W., Graham, R.R. (2009). A large-scale replication study identifies TNIP1, PRDM1, JAZF1, UHRF1BP1 and IL10 as risk loci for systemic lupus erythematosus. *Nature Genetics*, 41(11), 1228-1233. [\[CrossRef\]](#)
72. Nair, R.P., Duffin, K.C., Helms, C., Ding, J., Stuart, P.E., Goldgar, D., Gudjonsson, J.E., Li, Y., Tejasvi, T., Feng, B.J., Ruether, A., Schreiber, S., Weichenthal, M., Gladman, D., Rahman, P., Schrodi, S.J., Prahalad, S., Guthery, S.L., Fischer, J., Liao, W., Kwok, P.-Y., Menter, A., Lathrop, G.M., Wise, C.A., Begovich, A.B., Voorhees, J.J., Elder, J.T., Krueger, G.G., Bowcock, A.M., Abecasis, G.R., Collaborative Association Study of Psoriasis. (2009). Genome-wide scan reveals association of psoriasis with IL-23 and NF-kappa B pathways. *Nature Genetics*, 41(2), 199-204. [\[CrossRef\]](#)
73. Dieude, P., Guedj, M., Wipff, J., Ruiz, B., Riemekasten, G., Matucci-Cerinic, M., Melchers, I., Hachulla, E., Airo, P., Diot, E., Hunzelmann, N., Cabane, J., Mouthon, L., Cracowski, J.L., Ricciardi, V., Distler, J., Meyer, O., Kahan, A., Boileau, C., Allanore, Y. (2010). Association of the TNFAIP3 rs5029939 variant with systemic sclerosis in the European Caucasian population. *Annals of the Rheumatic Diseases*, 69(11), 1958-1964. [\[CrossRef\]](#)

74. Allamore, Y., Saad, M., Dieude, P., Avouac, J., Distler, J.H., Amouyel, P., Matucci-Cerinic, M., Riemekasten, G., Airo, P., Melchers, I., Hachulla, E., Cusi, D., Wichmann, H.E., Wipff, J., Lambert, J.C., Hunzelmann, N., Tiev, K., Caramaschi, P., Diot, E., Kowal-Bielecka, O., Valentini, G., Mouthon, L., Czirjak, L., Damjanov, N., Salvi, E., Conti, C., Müller, M., Müller-Ladner, U., Riccieri, V., Ruiz, B., Cracowski, J.L., Letenneur, L., Dupuy, A.M., Meyer, O., Kahan, A., Munnich, A., Boileau, C., Martinez, M. (2011). Genome-wide scan identifies TNIP1, PSORS1C1, and RHOB as novel risk loci for systemic sclerosis. *PLoS Genetics*, 7(7), e1002091. [\[CrossRef\]](#)
75. Musone, S.L., Taylor, K.E., Nititham, J., Chu, C., Poon, A., Liao, W., Lam, E.T., Ma, A., Kwok, P.Y., Criswell, L.A. (2011). Sequencing of TNFAIP3 and association of variants with multiple autoimmune diseases. *Genes and Immunity*, 12(3), 176-182. [\[CrossRef\]](#)
76. Nordmark, G., Wang, C., Vasaitis, L., Eriksson, P., Theander, E., Kvarnström, M., Forsblad-d'Elia, H., Jazebi, H., Sjöwall, C., Reksten, T.R., Brun, J.G., Jonsson, M.V., Johnsen, S.J., Wahren-Herlenius, M., Omdal, R., Jonsson, R., Bowman, S., Ng, W.F., Eloramta, M.L., Syvanen, A.C., UK Primary Sjögren's Syndrome Registry. (2013). Association of genes in the NF- κ B pathway with antibody-positive Primary Sjögren's Syndrome. *Scandinavian Journal of Immunology*, 78(5), 447-454. [\[CrossRef\]](#)
77. Shimane, K., Kochi, Y., Horita, T., Ikari, K., Amano, H., Hirakata, M., Okamoto, A., Yamada, R., Myouzen, K., Suzuki, A., Kubo, M., Atsumi, T., Koike, T., Takasaki, Y., Momohara, S., Yamanaka, H., Nakamura, Y., Yamamoto, K. (2010). The association of a nonsynonymous single-nucleotide polymorphism in TNFAIP3 with systemic lupus erythematosus and rheumatoid arthritis in the Japanese population. *Arthritis and Rheumatism*, 62(2), 574-579. [\[CrossRef\]](#)
78. Ramos, P.S., Criswell, L.S., Moser, K.L., Comeau, M.E., Williams, A.H., Pajewski, N.M., Chung, S.A., Graham, R.R., Zidovetzki, R., Kelly, J.A., Kaufman, K.M., Jacob, C.O., Vyse, T.J., Tsao, B.P., Kimberly, R.P., Gaffney, P.M., Alarcon-Riquelme, M.E., Harley, J.B., Langefeld, C.B., The International Consortium on the Genetics of Systemic Erythematosus. (2011). A comprehensive analysis of shared loci between systemic lupus erythematosus (SLE) and sixteen autoimmune diseases reveals limited genetic overlap. *PLoS Genetics*, 7(12), e1002406. [\[CrossRef\]](#)
79. Zhou, X.J., Lu, X.I., Nath, S.K., Lv, J.C., Zhu, S.N., Yang, H.Z., Qin, L.X., Zhao, M.H., Su, Y., Shen, N., Li, Z.G., Zhang, H. (2012). International Consortium on the Genetics of Systemic Lupus Erythematosus. (2012). Gene-gene interaction of BLK, TNFSF4, TRAF1, TNFAIP3, and REL in systemic lupus erythematosus. *Arthritis and Rheumatism*, 64(1), 222-231. [\[CrossRef\]](#)
80. Nezos, A., Gkioka, E., Koutsilieris, M., Voulgarelis, M., Tzioufas, A.G., Mavragani, P.M. (2018). TNFAIP3 F127C coding variation in Greek Primary Sjogren's Syndrome patients. *Journal of Immunology Research*, 2018, 6923213. [\[CrossRef\]](#)
81. Ciccacci, C., Latini, A., Perricone, C., Conigliaro, P., Colafrancesco, S., Ceccarelli, F., Priori, R., Conti, F., Perricone, R., Novelli, G., Borgiani, P. (2019). TNFAIP3 gene polymorphisms in three common autoimmune diseases: Systemic lupus erythematosus, rheumatoid arthritis, and primary Sjogren Syndrome-association with disease susceptibility and clinical phenotypes in Italian patients. *Journal of Immunology Research*, 2019, 6728694. [\[CrossRef\]](#)
82. Safonova, T.N., Surnina, Z.V., Zaitseva, G.V., Burdennyi, A.M., Loginov, V.I. (2020). The role of polymorphic markers rs1478604, rs2292305, and rs2228262 in THBS1 gene in the development of autoimmune dry eye syndrome. *Bulletin of Experimental Biology and Medicine*, 169(5), 707-709. [\[CrossRef\]](#)
83. Burbelo, P.D., Ambatipudi, K., Alevizos, I. (2014). Genome-wide association studies in Sjögren's syndrome: What do the genes tell us about disease pathogenesis? *Autoimmunity Reviews*, 13(7), 756-761. [\[CrossRef\]](#)
84. Markeljevic, J., Sarac, H., Bozina, N., Henigsberg, N., Simic, M., Cicin Sain, L. (2015). Serotonin transporter gene polymorphisms: Relation with platelet serotonin level in patients with primary Sjogren's syndrome. *Journal of Neuroimmunology*, 282, 104-109. [\[CrossRef\]](#)
85. Soto-Cárdenas, M.J., Gandía, M., Brito-Zerón, P., Arias, M.T., Armiger, N., Bové, A., Bosch, X., Retamozo, S., Akasbi, M., Perez-De-Lis, M., Gueitasi, H., Kostov, B., Perez-Alvarez, R., Siso-Almirall, A., Lozano, F., Ramos-Casals, M. (2015). Etiopathogenic role of surfactant protein D in the clinical and immunological expression of Primary Sjögren Syndrome. *The Journal of Rheumatology*, 42(1), 111-118. [\[CrossRef\]](#)
86. Burn, G.L., Svensson, L., Sanchez-Blanco, C., Saini, M., Cope, A.P. (2011). Why is PTPN22 a good candidate susceptibility gene for autoimmune disease? Why is PTPN22 a good candidate susceptibility gene for autoimmune disease? *FEBS Letters*, 585(23), 3689-3698. [\[CrossRef\]](#)
87. Ivashkiy, L.B. (2013). PTPN22 in autoimmunity: Different cell and different way. *Immunity*, 39(1), 91-93. [\[CrossRef\]](#)

88. Wang, Y., Shaked, I., Stanford, S.M., Zhou, W, Curtsinger, J.M., Mikulski, Z., Shaheen, Z.R., Cheng, G., Sawatzke, K., Campbell, A.M., Auger, J.L., Bilgic, H., Shoyama, F.M., Schmeling, D.O., Balfour Jr, H.H., Hasegawa, K., Chan, A.C., Corbett, J.A., Binstadt, B.A., Mescher, M.F., Ley, K., Bottini, N., Peterson, E.J. (2013). The autoimmunity-associated gene PTPN22 potentiates toll-like receptor-driven, type 1 interferon-dependent immunity. *Immunity*, 39(1), 111-122. [\[CrossRef\]](#)
89. Zheng, J., Ibrahim, S., Petersen, F., Yu, X. (2012). Meta-analysis reveals an association of PTPN22 C1858T with autoimmune diseases, which depends on the localization of the affected tissue. *Genes and Immunity*, 13(8), 641-652. [\[CrossRef\]](#)
90. Koutsilieris, M., Moutsopoulos, H.M., Mavragani, C.P. (2016). Increased frequency of the PTPN22W* variant in primary Sjögren's Syndrome: Association with low type I IFN scores. *Clinical Immunology*, 173, 157-160. [\[CrossRef\]](#)
91. Lavoie, T.N., Lee, B.H., Nguyen, C.Q. (2011). Current concepts: mouse models of Sjögren's syndrome. *Journal of Biomedicine and Biotechnology*, 2011, 549107. [\[CrossRef\]](#)
92. Fang, T.J., Li, R.N., Lin, Y.Z., Lin, C.H., Tseng, C.C., Sung, W.Y., Qu, T.T., Wu, C.C., Yen, J.H. (2021). Association of F11R polymorphisms and gene expression with primary Sjögren's syndrome patients. *International Journal of Rheumatic Diseases*, 24(5), 681-686. [\[CrossRef\]](#)
93. Gibbs, R.A., Weinstock, G.M., Metzker, M.L., Muzny, D.M., Sodergren, E.J., Scherer, S., Scott, G., Steffen, D., Worley, K.C., Burch, P.E., Okwuonu, G., Hines, S., Lewis, L., DeRamo, C., Delgado, O., Dugan-Rocha, S., Miner, G., Morgan, M., Hawes, A., Gill, R., Celera, Holt, R.A., Adams, M.D., Amanatides, P.G., Baden-Tillson, H., Barnstead, M., Chin, S., Evans, C.A., Ferriera, S., Fosler, C., Glodek, A., Gu, Z., Jennings, D., Kraft, C.L., Nguyen, T., Pfannkoch, C.M., Sitter, C., Sutton, G.G., Venter, J.C., Woodage, T., Smith, D., Lee, H.-M., Gustafson, E., Cahill, P., Kana, A., Doucette-Stamm, L., Weinstock, K., Fechtel, K., Weiss, R.B., Dunn, D.M., Green, E.D., Blakesley, R.W., Bouffard, G.G., De Jong, P.J., Osoegawa, K., Zhu, B., Marra, M., Schein, J., Bosdet, I., Fjell, C., Jones, S., Krzywinski, M., Mathewson, C., Siddiqui, A., Wye, N., McPherson, J., Zhao, S., Fraser, C.M., Shetty, J., Shatsman, S., Geer, K., Chen, Y., Abramzon, S., Nierman, W.C., Havlak, P.H., Chen, R., Durbin, K.J., Egan, A., Ren, Y., Song, X.-Z., Li, B., Liu, Y., Qin, X., Cawley, S., Worley, K.C., Cooney, A.J., D'Souza, L.M., Martin, K., Wu, J.Q., Gonzalez-Garay, M.L., Jackson, A.R., Kalafus, K.J., McLeod, M.P., Milosavljevic, A., Virk, D., Volkov, A., Wheeler, D.A., Zhang, Z., Bailey, J.A., Eichler, E.E., Tuzun, E., Birney, E., Mongin, E., Ureta-Vidal, A., Woodwark, C., Zdobnov, E., Bork, P., Suyama, M., Torrents, D., Alexandersson, M., Trask, B.J., Young, J.M., Huang, H., Wang, H., Xing, H., Daniels, S., Gietzen, D., Schmidt, J., Stevens, K., Vitt, U., Wingrove, J., Camara, F., Alba, M.M., Abril, J.F., Guigo, R., Smit, A., Dubchak, I., Rubin, E.M., Couronne, O., Poliakov, A., Hübner, N., Ganten, D., Goesele, C., Hummel, O., Kreitler, T., Lee, Y.-A., Monti, J., Schulz, H., Zimdahl, H., Himmelbauer, H., Lehrach, H., Jacob, H.J., Bromberg, S., Gullings-Handley, J., Jensen-Seaman, M.I., Kwitek, A.E., Lazar, J., Pasko, D., Tonellato, P.J., Twigger, S.M., Ponting, C.P., Duarte, J.M., Rice, S., Goodstadt, L., Beatson, S.A., Emes, R.D., Winter, E.E., Webber, C., Brandt, P., Nyakatura, G., Adetobi, M., Chiaromonte, F., Elnitski, L., Esvara, P., Hardison, R.C., Hou, M., Kolbe, D., Makova, K., Miller, W., Nekrutenko, A., Riemer, C., Schwartz, S., Taylor, J., Yang, S., Zhang, Y., Lindpaitner, K., Andrews, T.D., Caccamo, M., Clamp, M., Clarke, L., Curwen, V., Durbin, R., Eyas, E., Searle, S.M., Cooper, G.M., Batzoglou, S., Brudno, M., Sidow, A., Stone, E.A., Venter, J.C., Payseur, B.A., Bourque, G., Lopez-Otin, C., Puente, X.S., Chakrabarti, K., Chatterji, S., Dewey, C., Pachter, L., Bray, N., Yap, V.B., Caspi, A., Tesler, G., Pevzner, P.A., Haussler, D., Roskin, K.M., Baertsch, R., Clawson, H., Furey, T.S., Hinrichs, A.S., Karolchik, D., Kent, W.J., Rosenbloom, K.R., Trumbower, H., Weirauch, M., Cooper, D.N., Stenson, P.D., Ma, B., Brent, M., Arumugam, M., Shteynberg, D., Copley, R.R., Taylor, M.S., Riethman, H., Mudunuri, U., Peterson, J., Guyer, M., Felsenfeld, A., Old, S., Mockrin, S., Collins, F., Rat Genome Sequencing Project Consortium. (2004). Genome sequence of the Brown Norway rat yields insights into mammalian evolution. *Nature*, 428(6982), 493-521. [\[CrossRef\]](#)
94. Delaleu, N., Nguyen, C.Q., Peck, A.B., Jonsson, R. (2011). Sjögren's syndrome: Studying the disease in mice. *Arthritis Research & Therapy*, 13(3), 217. [\[CrossRef\]](#)
95. Kontinen, Y.T., Tensing, E.K., Laine, M., Porola, P., Tornwall, J., Hukkanen, M. (2005). Abnormal distribution of aquaporin-5 in salivary glands in the NOD mouse model for Sjögren's syndrome. *The Journal of Rheumatology*, 32(6), 1071-1075.
96. Winer, S., Astsaturov, I., Cheung, R., Tsui, H., Song, A., Gaedigk, R., Winer, D., Sampson, A., McKerlie, C., Bookman, A., Dosch, H.M. (2002). Primary Sjögren's syndrome and deficiency of ICA69. *Lancet*, 360(9339), 1063-1069. [\[CrossRef\]](#)
97. Cha, S., Nagashima, H., Brown, V.B., Peck, A.B., Humphreys-Beher, M.G. (2002). Two NOD Idd-associated intervals contribute synergistically to the development of autoimmune exocrinopathy (Sjögren's syndrome) on a healthy murine background. *Arthritis and Rheumatism*, 46(5), 1390-1398. [\[CrossRef\]](#)

98. Park, Y.S., Gauna, A.E., Cha, S. (2015). Mouse models of primary Sjögren's syndrome. *Current Pharmaceutical Design*, 21(18), 2350-2364. [\[CrossRef\]](#)
99. Vosters, J.L., Landek-Salgado, M.A., Yin, H., Swaim, W.D., Kimura, H., Tak, P.P., Caturegli, P., Chiorini, J.A. (2009). Interleukin-12 induces salivary gland dysfunction in transgenic mice, providing a new model of Sjögren's syndrome. *Arthritis and Rheumatism*, 60(12), 3633-3641. [\[CrossRef\]](#)