

The Antioxidant Effects of *Ziziphus Jujuba* on Neurodegeneration

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Abstract: Oxidative stress has been known to play an important role in the pathogenesis of various neurodegenerative diseases. Dietary polyphenols and other natural antioxidants are the most popular compounds in clinical testing for the elimination of neurodegeneration. It has been demonstrated in recent studies that fruit of *Zizyphus jujuba* possesses several vital biological activities. This study intends to evaluate antioxidant activity of *Zizyphus jujuba* on human glioblastoma cells. Cell survival was quantified by colorimetric viability assay with dose response. Cells were pretreated with 100 μM *Zizyphus jujuba* essential oil for 1h, then 100 μM H_2O_2 was added to the cells for 12 hours. After that, the cell medium were taken after treatment period and replaced with fresh medium. Total oxidant capacity (TOS) and total antioxidant capacity (TAS) levels were estimated using specific colorimetric methods. Oxidative stress index (OSI) was calculated from the ratio of TOS and TAS values. Many researches have been reported that the essential oil from seeds helps to prevent the oxidative stress induced neuronal diseases. The antioxidant potential of *Zizyphus jujuba* may be attributed to the presence of flavonoids and the other constituents present therein. Our data suggested that *Zizyphus jujuba* is effective in preventing H_2O_2 -induced oxidative stress.

Key words: Oxidative stress, Neurodegeneration, *Zizyphus jujuba*

N rodejenerasyonda *Zizyphus Jujuba*'in Antioksidan Etkileri

 zet: Oksidatif stresin eřitli n rodejeneratif hastalıkların patogenezinde  nemli bir rol oynadıđı bilinmektedir. Diyet polifenoller ve diđer dođal antioksidanlar n rodejenerasyonun giderilmesi amacıyla yapılan klinik testlerde en pop ler bileřiklerdir. Yapılan g ncel alıřmalarda, bir eřit yemiř olan *Zizyphus jujuba*'nın birok  nemli biyolojik aktivitelere sahip olduđu rapor edilmiřtir. Bu alıřma, insan glioblastoma h creleri  zerinde *Zizyphus jujuba*'nın antioksidan etkinliđinin arařtırılması amalamaktadır. H cre canlılıđı, doza bađlı kolorimetrik viyabilite testleri ile belirlenmiřtir. H creler, 1 saat 100 μM *Zizyphus jujuba*'nın uucu yađları ile  n-muamele edilip, daha sonra 100 μM H_2O_2 12 saat boyunca h crelere ilave edildi. Uygulamalar sonunda h cre homojenatları uzaklařtırılarak taze besiyeri ile deđiřtirildi. Total oksidan kapasite (TOS) ve total antioksidan kapasite (TAS) d zeyleri spesifik kolorimetrik y ntemler kullanılarak tayin edilmiřtir. Oksidatif stress indeksi (OSI) TOS ve TAS deđerlerinin oranlamasıyla elde edilmiřtir. Birok arařtırmada dođal bitkilerden elde edilen uucu yađların oksidatif stress uyarımlı n ronal hastalıkları  nlemeye yardımcı oldukları rapor edilmiřtir. *Zizyphus jujuba*'nın antioksidan etkinliđinin ierisinde bulunan flavonoidler ve diđer bileřenlere bađlı olduđu d ř n lmektedir. Elde edilen veriler g re, *Zizyphus jujuba*'nın H_2O_2 kaynaklı oksidatif stresi  nlemede etkili olduđu  ng r lmektedir.

Anahtar kelimeler: Oksidatif stres, N rodejenerasyon, *Zizyphus jujube*

Introduction

Cellular and molecular signaling failure is the main reason for many human disorders and signal transduction defects and the proteins involved in these processes are the mayor elements for neurodegenerative disorders such as Alzheimer's disease (AD) and Parkinson's disease (PD) [3,23]. It is stated that many factors like environmental, genetic predisposition and abnormal metal metabolism plays a critical role in neurodegeneration [22]. Free radicals catalyzed by redox metals and oxidative stress are

the most important reasons for the development of neurodegeneration [7].

Oxidative stress occurs as a result of the release of reactive oxygen species (ROS) [26]. Free radicals can be produced from endogenous sources, such as from mitochondria, peroxisomes, and inflammatory cell activation and exogenous sources, including environmental agents, pharmaceuticals, and industrial chemicals [19]. Brain is particularly sensitive to free radicals because of having antioxidant enzymes in low concentration and the consumption about 20% of the body's total oxygen [9]. Recent studies have

indicated that ROS causes oxidative stress and programmed cell death in neuronal cells [27].

Herbs contain a wide variety of molecules including phenolic compounds (flavonoids, quinones, tannins etc.), nitrogen compounds (alkaloids, amines), vitamins and terpenoids. These compounds exert strong free radical scavenging and antioxidant properties [2,30]. The nutritional *Ziziphus jujuba* Mill., (ZJ) is a herbal plant used in traditional medicine, belongs to the *Rhamnaceae* family and it is one of the most important *Ziziphus* species [6,14]. Recent phytochemical studies of jujuba fruits have shed some light on their biological effects, such as the anticancer, anti-inflammatory, antiobesity, immunostimulating, antioxidant, hepatoprotective, and gastrointestinal protective activities and inhibition of foam cell formation in macrophages [10].

Herbal medicines are generally low in cost, plentiful, and show very little toxicity or side effects in clinical practice. Therefore, our main objective in this study is to investigate antioxidant effects of essential oil of *Ziziphus Jujuba* fruit on human glioblastoma cells.

Materials and Methods

Human glioblastoma (U87MG) was obtained from American Type Culture Collection (Manassas, VA) and maintained in DMEM supplemented with 10% fetal bovine serum, 2 mM glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin, at 37°C with 5% CO₂. Briefly, cells were plated in 24-well plates (0.4 × 10⁵ cells) and pretreated with 100 µM *Ziziphus jujuba* essential oil for 1h then 100 µM H₂O₂ was added to each well for 12h. After the incubation, the supernatant was replaced by fresh medium.

The cell survival was quantified by the colorimetric MTT (4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay [13]. Following incubation period, cell culture at 37°C, 1 ml/well of MTT (5 mg/ml-Sigma) was added to the wells, followed by incubation for an additional 2 h for each experiment time. The viable cells produced a dark blue formazan product, whereas no such staining was formed in the dead cells. The resulting formazan product was solubilized in 1 ml/well of acidic

isopropanol, and absorbance read at 570nm with ELISA reader (µQuant-USA).

The novel total antioxidant status (TAS) and total oxidant status (TOS) assays have been shown to be stable, reliable and sensitive to determine antioxidant and oxidant capacity of the biologic samples, respectively [8]. Total antioxidant level of the sample was calculated according to ABTS (dark blue colored radical) reducing capacity of antioxidants at 660 nm. Results were given as Trolox equivalent (mmol/l) which is a vitamin E analog. Additionally, oxidants in the sample oxidize ferrous-ion chelator complex to ferric ion. Briefly, total oxidant level was measured by colorimetric methods according to absorbance change of formed colored complex at 530 nm. Results were given as H₂O₂ equivalent (µmol/l). OSI was calculated as the ratio of TOS and TAS values.

Statistical analysis

The one-way analysis of variance (ANOVA) and post hoc Duncan tests were performed on the data to examine the differences among groups using the SPSS statistical software package. The results are presented as average ±SE. A value of p<0.05 was considered significant.

Results

We used H₂O₂ treatment in order to model oxidative stress in our cellular system. Effect of H₂O₂ on cell viability were performed by MTT analysis. Viability of U87 cells were decreased straightly in a concentration-dependent manner over the range of 5 to 250 µM following 12 h H₂O₂ treatment. The data showed that 100 µM H₂O₂ (0.190± 0.008) killed about 42% of cells at the end of the incubation according to the control group (0.329± 0.011) (p<0.05) (Fig. 1A). However, 10 µM ZJ (0.367±0.026) increased the number viable cells by 17% as compared to control group (0.3142±0.010). For this reason 10 µM ZJ was used as a cell protective concentration for further experiments. Treatments utilizing 50 µM and higher concentrations of ZJ decreased cell viability (Fig 1B). Moreover, 10 µM ZJ (0.240±0.017) pre-treatment for 1h prevented 25 % of cell death caused by H₂O₂ (0.193±0.044) (Fig 1C).

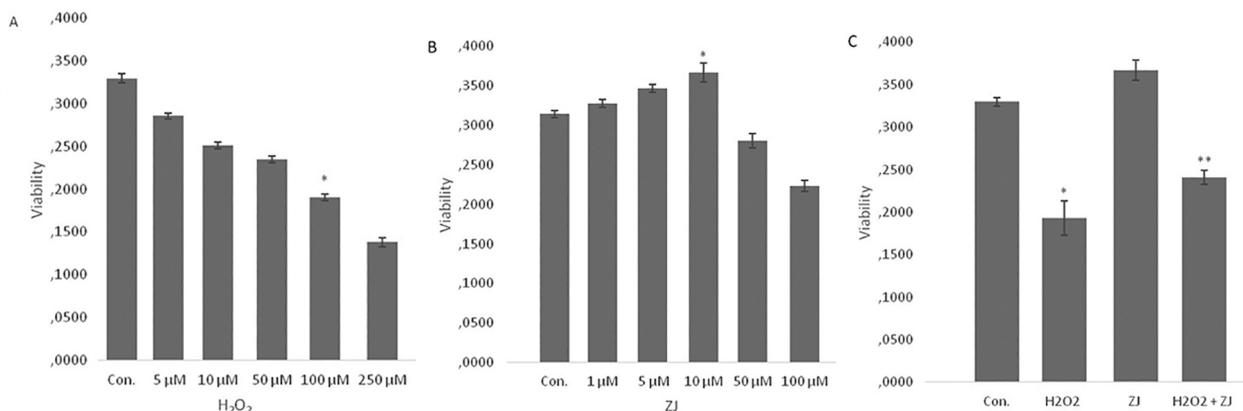


Figure 1. The effect of *Z. Jujuba* on cell viability in hydrogen peroxide induced oxidative stress. A. Dose dependent effect of H_2O_2 on U87 cell viability. B. Dose dependent effect of ZJ in viability. C. Dose dependent protective effect of ZJ on H_2O_2 -induced cytotoxicity in cells. The data is represented as mean \pm SD of five independent experiments. * $p < 0.05$ versus control group. ** $p < 0.05$ versus H_2O_2 group.

Hydrogen peroxide significantly enhanced total oxidant status (TOS) 3.88 times in human glioblastoma cell line U87 ($p < 0.05$). The addition of *Z. jujuba* essential oil pretreatment decreased TOS by 18.67% in only jujuba group and in 13.22% ZJ plus H_2O_2 group against control. The ZJ oil also has important effects (by 6.69%) on ZJ plus H_2O_2 group when compared only group (Table 1). Total antioxidant status levels significantly decreased by

H_2O_2 pretreatment but ZJ oil prevented the situation by 22.93% against control and by 68% against only H_2O_2 group. The antioxidant status of ZJ oil decreased ZJ plus H_2O_2 group. Hydrogen peroxide (3.0 ± 0.300) increased OSI as compared to control group (0.141 ± 0.014) (Table 1). However, ZJ pretreatment (0.142 ± 0.015) significantly recovered this increase in OSI according to only H_2O_2 added group.

Table 1. Effects of CAPE treatment on total oxidant (TOS) and antioxidant (TAS) status in cells exposed to H_2O_2 .

	Control	H_2O_2	ZJ	$H_2O_2 + ZJ$
TOS ($\mu\text{mol } H_2O_2 \text{ equiv./lt}$)	2.570 ± 0.171	$9.990 \pm 0.267^*$	2.090 ± 0.114	$2.230 \pm 0.153^{**}$
TAS (mmol Trolox equiv./lt)	1.818 ± 0.083	$0.330 \pm 0.056^*$	2.235 ± 0.141	$1.563 \pm 0.182^{**}$
OSI ($\mu\text{mol } H_2O_2 \text{ equiv./lt}$) / (mmol Trolox equiv./lt x 10)	0.141 ± 0.014	3.0 ± 0.300	0.093 ± 0.010	0.142 ± 0.015

The data are represented as mean \pm SE. * $p < 0.05$ compared to control group; ** $p < 0.05$ compared to H_2O_2 -treated group. The cells were preincubated for 1 h with 10 μM of ZJ then incubated for a further 12 h at 37 $^\circ\text{C}$ in the presence of 100 μM H_2O_2 ($n = 5$). OSI: Oxidative stress index.

Discussion

Hydrogen peroxide can swiftly penetrate the cell membrane, reacting with intracellular metal ions such as iron or copper to form highly toxic hydroxyl

radicals, which cause DNA alteration. Thus, even at lower concentration, H_2O_2 can cause heavy damage to the cultured cells. Some natural antioxidant products may be useful to protect neurons from oxidative injury. Clinically, malignant gliomas are among the least responsive of human tumors and for tumors of higher grades, complete remission and/or long term survival is rare [24]. Kitamura et al., [18] demonstrated that in human A172 cells, hydrogen peroxide (H_2O_2) caused cell death in a time- and concentration-dependent manner, accompanied by nucleosomal DNA fragmentation and chromatin

condensation. Similar to our data obtained from this study, Tavakkoli et al., [29] reported that 75µM H₂O₂ treatment applied to the PC12 neuronal like cells for 1h reduced cell viability significantly and triggered oxidative stress and apoptosis.

Herbal medicine in recent years gained a momentum in the treatment of many diseases, especially cancer and neurodegenerative diseases [4,15]. Phenolic compounds derived from jujuba in have been reported to show beneficial properties in neuronal tissue. Considering that treatment H₂O₂ with results in excess ROS, the present study suggests that oxidative stress may play a critical role in oxidative stress induced neuronal injury [17]. Our results indicated essential oil of jujuba prevented neuronal cell loss induced by H₂O₂. It has been reported in recent studies that although ZJ aqueous extract demonstrated proliferative effect on *in vitro* diabetic neuropathy model of on PC12 cells [15].

Medicinal plants have curative properties due to presence of various complex chemical substances of different composition which contain secondary metabolites such as alkaloids, flavonoids, terpenoids, saponin and phenolic compounds distributed in different parts of the plants. Studies demonstrated that, an indigenous plant possesses terrific medicinal properties, attributed by a diverse group of secondary metabolites. Also *Z. Jujuba* has photo-chemical, pharmacological, medicinal properties and biological activities [5,21]. Gao et al., [10] which studies five variations of *Z. jujuba*, demonstrated the antioxidative and free radical scavenging effect of the this plant. Taatil et al., [28] showed that *Z. jujuba* fruit extract improved spatial memory impairment induced by ethanol, due in part, by its antioxidant activities such as GSH level content. The possible antioxidant activities of extracts were due to the presence of tannins [1], carotenes [11] and flavonoids [25]. Additionally, Chen et al., [4] verified the bidirectional immune-modulatory roles of jujuba by regulating the expressions of pro-inflammatory cytokines in macrophages. Park et al., [24] suggested that *Zizyphus jujuba* Mill var. *Spinosa* prevents N-methyl-d-aspartate (NMDA)-induced neuronal cell damage *in vitro*. Studies also showed that it is used traditionally as tonic and aphrodisiac and sometimes as hypnotic-sedative and anxiolytic,

anticancer, antifungal, antibacterial, antiulcer, anti-inflammatory and wound healing properties [12,20].

In a recent study performed by Chen et al., [4] ZJ extract was found to increase anti-oxidant enzyme levels in cultured astrocytes. In another study of the carbon tetrachloride-induced hepatitis by Kandimalla et al., [16] ZJ was reported to exert antioxidant activity in particular by increasing activities of catalase and superoxide dismutase enzymes and also reducing TBARS level, an important biomarker of lipid peroxidation, significantly.

Conclusion

Ziziphus jujuba is a widely traditionally used and potent medicinal plant amongst all the thousands of medicinal plants. It is an important source of compounds with their chemical structures as well as pharmacological properties. This study may be useful for predicting other medicinal uses and potential drug or food interactions and may be beneficial for people living where the *jujuba* fruits are prevalent and health care resources are scarce.

References

- Adzu B, Amos S, Wambebe C, and Gamaniel K, (2001). *Antinociceptive activity of Ziziphus spina christi root bark extract*. Fitoterapia. 72, 344-350.
- Cai YZ, Sun M, Corke H, (2003). *Antioxidant activity of betalains from plants of the amaranthaceae*. J Agric Food Chem. 51(8), 2288-2294.
- Castellani RJ, Zhu X, Lee HG, Smith MA and Perry G, (2009). "Molecular pathogenesis of Alzheimer's disease: reductionist versus expansionist approaches," International Journal of Molecular Sciences. 10(3), 1386-1406.
- Chen J, Maiwulanjiang M, Lam KY, Zhang WL, Zhan JY, Lam CT, Xu SL, Zhu KY, Yao P, Lau DT, Dong TT, Tsim KW, (2014). *A standardized extract of the fruit of Ziziphus jujuba (Jujuba) induces neuronal differentiation of cultured PC12 cells: a signaling mediated by protein kinase*. A. J Agric Food Chem. 62, 1890-1897.
- Cheng G, Bai Y, Zzhao Y, Tao J, Liu Y, Tu G, Ma L, Liao N, Xu X, (2000). *Flavonoids from Ziziphus jujuba Mill var. Spinosa*. Tetrahedron, 56(45), 8915-8920.
- Choi SH, Ahn JB, Kozukue N, Levin CE, Friedman M, (2011). *Distribution of Free Amino Acids, Flavonoids, Total Phenolics, and Antioxidative Activities of Jujuba (Ziziphus jujuba) Fruits and Seeds Harvested from Plants Grown in Korea*. J Agric Food Chem. 59(12), 6594-6604.
- Emerit J, Edeas M, Bricaire F, (2004). *Neurodegenerative diseases and oxidative stress*. Biomed Pharmacother. 58, 39-46.

8. Erel OA, (2005). *New colorimetric method for measuring total oxidant status*. Clin Biochem. 38(12):1103-11.
9. Floyd RA, Hensley K, (2002). *Oxidative stress in brain aging. Implications for therapeutics of neurodegenerative diseases*. Neurobiol Aging. 23(5), 795-807.
10. Gao QH, Wu PT, Liu JR, Parry JW, Wang M, (2011). *Physico-chemical properties and antioxidant capacity of different jujuba (Ziziphus jujuba Mill.) cultivars grown in loess plateau of China*. Scientia Horticulturae. 130(1), 67-72.
11. Guil-Guerrero JL, Diaz Delgado A, Gonzalez MCM, and Torija Isasa ME, (2004). *Fatty acids and carotenenes in some ber (Ziziphus jujuba mill) varieties*. Plant Foods Hum Nutr, 59, 23-27.
12. Gupta RB, Sharma S, Sharma JR, and Goyal R, (2004). *Study on the physicochemical characters of fruits of some wild and cultivated forms/spp. (Ziziphus spp.)*. Haryana Journal of Horticultural Sciences. 33(3/4), 167-169.
13. Hansen MB, Nielsen SE, Berg K, (1989). *Re-examination and further development of a precise and rapid dye method for measuring cell growth/cell kill*. Journal of Immunology Methods. 119, 203-210.
14. Huang YL, Yen GC, Sheu F, Chau CF, (2008). *Effects of water-soluble carbohydrate concentrate from Chinese jujuba on different intestinal and fecal indices*. J Agric Food Chem. 56, 1734-9.
15. Kaeidi A, Taati M, Hajjalizadeh Z, Jahandari F, Rashidipour M, (2015). *Aqueous extract of Zizyphus jujuba fruit attenuates glucose induced neurotoxicity in an in vitro model of diabetic neuropathy*. Iran J Basic Med Sci. 18, 301-306.
16. Kandimalla R, Dash S, Kalita S, Choudhury B, Malampati S, Kalita K, Kalita B, Devi R, Kotoky J, (2016). *Protective Effect of Bioactivity Guided Fractions of Ziziphus jujuba Mill. Root Bark against Hepatic Injury and Chronic Inflammation via Inhibiting Inflammatory Markers and Oxidative Stress*. Front. Pharmacol. 7, 298.
17. Kim MC, Cui FJ, Kim Y, (2013). *Hydrogen Peroxide Promotes Epithelial to Mesenchymal Transition and Stemness in Human Malignant Mesothelioma Cells*. Asian Pacific Journal of Cancer Prevention. 14(6), 3625-3630.
18. Kitamura Y, Ota T, Matsuoka Y, Tooyama I, Kimura H, Shimohama S, Nomura Y, Gebicke-Haerter PJ, Taniguchi T, (1999). *Hydrogen peroxide-induced apoptosis mediated by p53 protein in glial cells*. Glia. 25(2), 154-164.
19. Klaunig JE, Kamendulis LM, (2004). *The role of oxidative stress in carcinogenesis*. Annu Rev Pharmacol Toxicol. 44, 239-67.
20. Kumar S, Ganachari MS, Banappa, V, Nagoor S, (2004). *Anti-inflammatory activity of Ziziphus jujuba Lam leaves extract in rats*. Journal of Natural Remedies, 4(2), 183-185.
21. Mahajan RT, Chopda MZ, (2009). *Phyto-Pharmacology of Ziziphus jujuba Mill-A plant review*. Phcog Rev. 3(6), 320-329.
22. Mark PM, (2004). *Metal-catalyzed disruption of membrane protein and lipid signaling in the pathogenesis of neurodegenerative Disorders*. Ann NY Acad Sci. 1012, 37-50.
23. Mitra A, and Dey B, (2013). *"Therapeutic interventions in Alzheimer disease," in Neurodegenerative Diseases*. InTech, Rijeka, chapter 12, 291-317.
24. Park JH, Lee HJ, Koh SB, Ban JY, (2004). *Protection of NMDA-induced neuronal cell damage by methanol extract of Zizyphi Spinosi Semen in cultured rat cerebellar granule cells*. Journal of Ethnopharmacology, 95 (1), 39-45.
25. Pawlowska AM, Camangi F, Bader A, and Braca A, (2009). *Flavonoids of Zizyphus jujuba L. and Zizyphus spina-christi (L.) wild (Rhamnaceae) fruits*. Food Chem. 112, 858-862.
26. Poon HF, Calabrese V, Scapagnini G, Butterfield DA, (2004). *Free radicals: key to brain aging and heme oxygenase as a cellular response to oxidative stress*. J Gerontol A Biol Sci Med Sci. 59(5), 478-93.
27. Salganik RI, (2001). *The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population*. J Am Coll Nutr. 20, 464-72.
28. Taatil M, Alirezaei M, Meshkatsadat MH, Rasouljan B, Moghadasi M, Sheikhzadeh F, Sokhtezari A, (2011). *Protective effects of Ziziphus jujuba fruit extract against ethanol-induced hippocampal oxidative stress and spatial memory impairment in rats*. Journal of Medicinal Plant Research. 5(6), 915-921.
29. Tavakkoli M, Miri R, Jassbi AR, Erfani N, Asadollahi M, Ghasemi M, Saso L, Firuzi O, (2014). *Salvia and Stachys species protect neuronal cells against oxidative stress-induced apoptosis*. Pharm Biol. 52(12), 1550-7.
30. Zheng W, Wang SY, (2001). *Antioxidant activity and phenolic compounds in selected herbs*. J Agric Food Chem. 49 (11), 5165-70.