

An Immunohistochemical Investigation of The Effect of Sambucus Nigra on Chymase-, Tryptase- and Ghrelin- Positive Cells in Rat Lung

Tuğrul Ertuğrul^{1*}, Gökçen Sevilgen²

¹Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Histology and Embryology, Samsun, Turkey

²Recep Tayyip Erdogan University, Faculty of Medicine, Department of Thoracic Surgery, Rize, Turkey

ABSTRACT

Sambucus nigra (S.nigra) is used in the treatment of many diseases and disorders thanks to its antioxidant, anticarcinogenic, immunostimulating, antiallergic, antiviral, and antibacterial properties. Ghrelin has anti-inflammatory effect on oxidative damage in various organs and cell types. The aim of this study was to immunohistochemically examine the chymase, tryptase, and ghrelin in rat lung after S. nigra administration. A total of 16 male rats were used in the study. The rats were assigned to two groups, control and S. nigra. 1st control group (n=8): No application was made. 2nd S. nigra group (n=8): S. nigra extract was administered at 15 mg/kg by oral gavage for 14 days. Tryptase, chymase, and ghrelin-positive cells were found in the lung tissue in a spindle-shaped, round, or oval shape. When the groups were evaluated within themselves, a significant increase in the number of tryptase, chymase, and ghrelin positive cells was observed in the S. nigra treated group. This study showed that S. nigra, which has an immunomodulatory and antioxidant effect, increases the expression of chymase-, tryptase- and ghrelin-positive cells in the lungs. Additionally, based on our findings, it can be said that mast cells can produce, store and release ghrelin.

Keywords: Chymase, ghrelin, lung, Sambucus nigra, tryptase

Sıçan Akciğerinde Sambucus Nigra'nın Kimaz, Triptaz ve Ghrelin Pozitif Hücrelerin Üzerindeki Etkisinin İmmünohistokimyasal Olarak İncelenmesi

ÖZ

Sambucus nigra (S. nigra) antioksidan, antikanserojenik, immün sistemi uyarıcı, antialerjik, antiviral ve antibakteriyel özellikleri sayesinde birçok hastalık ve rahatsızlığın tedavisinde kullanılmaktadır. Ghrelin, çeşitli organlarda ve hücre tiplerinde oksidatif hasar üzerinde anti-inflamatuar etkilere sahiptir. Bu çalışmanın amacı, S. nigra uygulamasından sonra sıçan akciğerinde kimaz, triptaz ve ghrelin immünopozitif hücrelerinin immünohistokimyasal olarak incelenmesidir. Çalışmada toplam 16 erkek sıçan kullanıldı. Sıçanlar kontrol ve S. nigra olmak üzere iki gruba ayrıldı. 1. kontrol grubu (n=8): herhangi bir uygulama yapılmadı. 2. S. nigra grubu (n=8): S. nigra ekstresi 14 gün süreyle oral gavaj yoluyla 15 mg/kg dozunda uygulandı. Akciğer dokusunda yuvarlak, oval veya mekik şeklinde triptaz, kimaz ve ghrelin pozitif hücreler bulundu. İki grup kendi içinde değerlendirildiğinde S. nigra uygulanan grupta triptaz, kimaz ve ghrelin pozitif hücre sayısında önemli artış gözlemlendi. Bu çalışma, immünomodülatör ve antioksidan etkiye sahip olan S. nigra'nın akciğerde kimaz, triptaz ve ghrelin pozitif hücrelerin ekspresyonunu arttırdığını göstermiştir. Ayrıca mast hücrelerinin diğer bağışıklık hücreleri gibi ghrelin üretebildiği, depolayabildiği ve serbest bırakabildiği açıklanmaya çalışılmıştır.

Anahtar kelimeler: Akciğer, ghrelin, kimaz, Sambucus nigra, triptaz

To cite this article: Ertuğrul T, Sevilgen G. The Immunohistochemical Investigation of Chymase-, Tryptase- and Ghrelin-Positive Cells on The Lung of Rat After an Application of Sambucus Nigra

Kocatepe Vet J. (2022) 15(2)209-216

Submission: 09.02.2022 Accepted: 18.04.2022 Published Online: 30.05.2022

ORCID ID; TE: 0000-0002-9310-1200, GS: 0000-0003-4638-8007.

*Corresponding author e-mail: tugrulerugrul06@hotmail.com

INTRODUCTION

Sambucus nigra (*S. nigra*) is a widespread species of the Caprifoliaceae family that grows in most of Europe, western Asia, northern Africa, and the United States (Fazio et al. 2013). Because of its antioxidant, anticarcinogenic, immune system boosting, antiallergic, antiviral, and antibacterial characteristics, *S. nigra* has been used in folk medicine for generations to treat a variety of ailments and problems (Oniszczuk et al. 2016). It contains high levels of polyphenols, especially flavonols, phenolic acids and anthocyanins. At the same time, these compounds are known as radical scavengers, which protect the body against oxidative stress and lipid peroxidation (Duymuş et al. 2014). Herbal supplements, including *S. nigra*, have been known to be used to boost immunity against respiratory diseases (Wieland et al. 2021). Moreover, it has been suggested that *S. nigra* may help in the treatment of upper respiratory tract symptoms and shorten the duration of the common cold or flu (Hawkins et al. 2018).

Mast cells (MCs) are tissue-resident sentinel cells with densely packed secretory granules. (Metcalf et al. 1997). MCs are capable of secreting a variety of biologically active mediators, cytokines, and chemokines. MC-derived mediators can affect the biological activities of adjacent cells and tissues (Mukai et al. 2018). These cells are multifunctional effector cells involved in innate immunity, host defense, hypersensitivity, and allergic disease (Da Silva et al. 2014). MCs exhibit substantial heterogeneity based on their granule content and protease expression patterns (Dwyer et al. 2016). Based on protease content, two types of MCs have been identified immunohistochemically: tryptase positive mast cells (MC_T) and chymase positive mast cells (MC_{TC}) (Tütüncü et al. 2020). Tryptase is stored in the secretory granules of MCs, from which it is released after degranulation following cell stimulation (Schwartz et al. 1981). Under various physiological and pathological settings, tryptase is involved in the activation, proliferation, and migration of many mesenchymal cells, including endothelial cells and fibroblasts (Sonneck et al. 2006). Chymase is an intracellular, granular-associated, neutral serine protease produced mainly by MCs. It plays a role in regulating extracellular matrix proteolysis, which promotes tissue remodeling (Hamada et al. 1999). In addition, MC_{TC} can promote vascular proliferation, atherosclerosis and tissue fibrosis (Miyazaki et al. 2006). In experimental studies, chymase has been shown to reduce fibrosis in lung tissue (Tomimori et al. 2003, Sakaguchi et al. 2004).

Ghrelin is an endogenous peptide that interacts with the growth hormone secretagogue receptor 1a (GHSR1a) (Kojima and Kangawa, 2005). The

presence of ghrelin and its receptor in a wide range of tissues has been determined by gene expression studies in humans and rats (Akalu et al. 2020). Ghrelin regulates growth hormone secretion, cell proliferation, appetite increase, and inflammation through GHSR1a (Nakazato et al. 2001). It has anti-inflammatory effects on oxidative damage in various organs and cell types (Raghay et al. 2020). The expression of ghrelin can be modulated by factors such as peptide hormones, neurotransmitters, glucose, fatty acids, neurotransmitters and enzymes (Akalu et al. 2020). It has been reported that ghrelin is expressed in immune tissues and modulates immune function (Chowen and Argente. 2017). Furthermore, it is stated that ghrelin may have an effect on hematological parameters, which may increase lymphocyte count by stimulating lymphopoiesis (Narin and Çetin, 2010). Moreover, Stefanov et al. (2017) suggested that MCs in rat stomachs can produce, store, and release ghrelin like other immune cells.

The aim of this study was to investigate the effect of *S. nigra* on the immunohistochemical distribution of chymase-, tryptase- and ghrelin-positive cells in rat lung. Also, the lack of data on the ability of MCs to express, store and release ghrelin motivated this study.

MATERIAL and METHODS

Animals

All procedures were approved by the Ethical Committee of Ondokuz Mayıs University (Decision no: 11.03.2020, number 15).

In this study, 16 male rats, weighing 250-300 g, that were used. The rats were kept in a standard cage with 12 hours of light and 12 hours of darkness in a 22°C ambient temperature environment and were given ad libitum and tap water.

Experiment Groups

1st control group (n=8): there was no application made. 2nd *S. nigra* group (n=8): *S. nigra* extract was administered at a dose of 15 mg/kg by oral gavage for 14 days (Bidian et al. 2021). Then, cervical dislocation was performed under anesthesia and lung tissue samples were collected for immunohistochemistry. The lung tissue samples were fixed for 24 h in a 10% formaldehyde solution, and tissue sections were cut from the prepared paraffin blocks with a thickness of 5 µm.

Immunohistochemistry

The lung sections were stained immunohistochemically with Streptavidin biotin complex. Immunopositive cells were determined using anti-rabbit polyclonal chymase (1/200 dilution, Biorbyt, orb11030), mouse monoclonal tryptase

(1/200 dilution, Abcam, ab2378), and rabbit polyclonal anti-ghrelin antibody (1/400 dilution, Abcam, ab129383) (True, 1990). As a secondary antibody, Histostain Plus (Zymed kit: 85-6743) was used. Following deparaffinization, sections were heated in a 700-watt microwave oven in a citrate buffer (pH=6) solution for proteolysis. The tissues were treated in a 3 % hydrogen peroxide solution to inhibit endogenous peroxidase activity. To prevent nonspecific protein binding in sections, serum in the kit was instilled after washing with phosphate buffer solution (PBS). The primary antibody was applied to the samples, which were then kept at +4 OC overnight. The negative control group's tissues were treated with only PBS solution. Following the washing procedure, sections were treated with a biotinylated secondary antibody and incubated with streptavidin-horseradish peroxidase complex. Finally, 3, 3'-diaminobenzidine (DAB) was utilized as chromogen, and the samples were counterstained with hematoxylin and then coated with entellan.

Microscopical Evaluation and Positive Cell Counts

Following histochemical and immunohistochemical staining, tissue samples were examined under an microscope (Nikon Eclipse 50i) in terms of immunoreactivity. Chymase-, tryptase- and ghrelin-positive cell distribution was evaluated semiquantitatively. The following criteria were employed in semiquantitative evaluation: no positive cell in the scanned area (-), 1-2 cells (\pm), 3-4 cells (+), and 5-6 cells (++) (Ertuğrul et al. 2021).

The immunopositive cells were scored from 0 to 3 semi-quantitatively (Samrao et al. 2012) as follows. A histoscore was derived from the immunopositive cell

distribution, 0: no positive cell in the scanned area (-), +1: 1-2 cells (\pm), +2: 3-4 cells (+), +3: 5-6 cells (++)

Statistical Analysis

The IBM SPSS Statistics Version 22.0 statistical software program was used for all statistical analyses. The Shapiro-Wilk W test was used to determine whether the distribution was normal. Depending on the normality of the data, comparisons between control and *S. nigra* groups were performed by independent Student's t-test for parametric data. The findings were presented as mean \pm SEM (standard error of the mean), and statistical significance was accepted at $p < 0.05$.

RESULTS

Tryptase- and Chymase- Immunopositive Cells

In the light microscopic examination, MC_T and MC_{TC} with positive immune reactions were clearly distinguished in brown color. Tryptase and chymase positive cells were found in the lung tissue in a spindle-shaped, round, or oval shape. (Figures 1A, 2A). MC_T and MC_{TC} were observed around the sacculus alveolaris, in the visceral pleura of the lung, the bronchial wall, and connective tissue in the terminal and respiratory bronchioles' walls (Figures 1B, 2B). Also, MC_{TC} and MT_C were observed to be mostly located near the blood vessels and bronchus-associated lymphatic tissue (BALT). When the two groups are evaluated among themselves, a significant increase in the number of MC_T and MC_{TC} was observed in the *S. nigra* treated group. MC_{TC} and MT_C were seen individually or in groups in the lung tissue (Table 1).

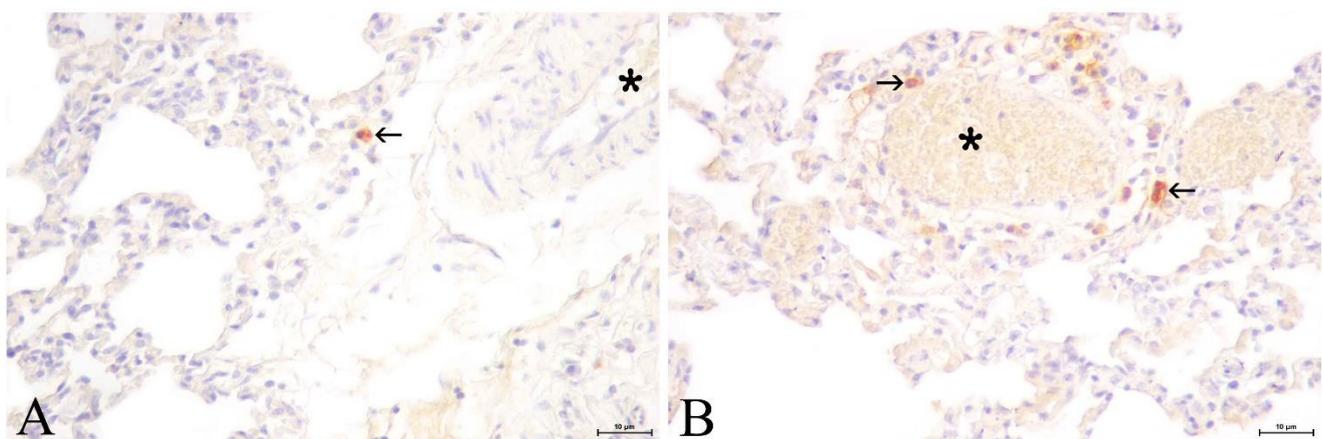


Figure 1: Lung tissue immunostained with antibodies against tryptase; (A) Control group, (\rightarrow): tryptase immunopositive cell, (asterix): blood vessel, (B) *S. nigra* group, (\rightarrow): tryptase immunopositive cell, (asterix): blood vessel, original magnification X40; range bar, 10 μ m.

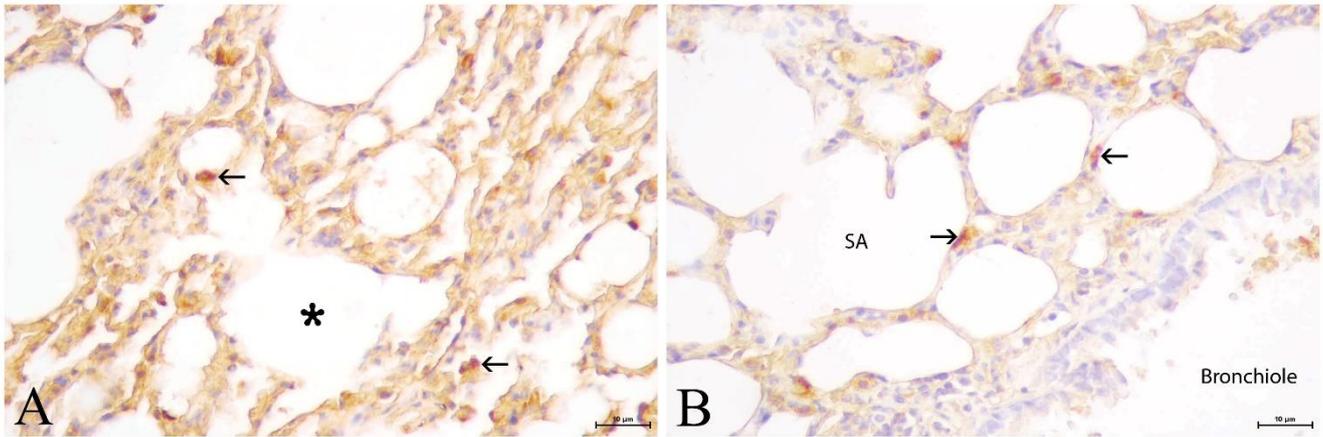


Figure 2: Lung tissue immunostained with antibodies against chymase; (A) Control group, (→): chymase immunopositive cell, (asterix): alveolar space, (B) *S. nigra* group, (→): chymase immunopositive cell, (SA): sacculus alveolaris, original magnification X40; range bar, 10 µm.

Table 1. Tryptase-, chymase- and ghrelin- immunopositive cell numerical density

	Control group (mean ± SEM)	<i>S. nigra</i> group (mean ± SEM)
Tryptase immunopositive cell	0.73 ± 0.06	1.28 ± 0.09**
Chymase immunopositive cell	0.72 ± 0.05	1.31 ± 0.11**
Ghrelin immunopositive cell	1.82 ± 0.07	2.15 ± 0.11*

* p<0.05 and **p<0.001

Ghrelin- Immunopositive Cells

Immunohistochemistry revealed that brown ghrelin-positive cells were scattered throughout the lung tissue, including perivascular areas. In lung tissue, ghrelin-positive cells were spindle-shaped, round, or oval. Ghrelin-positive cells were observed around the sacculus alveolaris, in the interalveolar septal connective tissue, on the periphery of the bronchi, and around blood vessels (Figure 3A). Ghrelin-positive cells were also observed inside and at the margin of the bronchus-associated lymphatic tissue in the lungs. In the lung tissue, ghrelin-positive cells were mostly seen separately or in groups (Figure 3B).

A significant increase in ghrelin positive cells was observed after *S. nigra* administration compared to the control group (Table 1).

When we compared ghrelin-positive cells with MC_T and MC_{TC}, we also found that ghrelin-positive cells had similar morphology (similar size and shape) and localization to MC_T and MC_{TC}, which reacted positively to immune staining. In addition, ghrelin-positive cell density increase after *S. nigra* application allowed us to suggest that MCs can produce, store and release ghrelin in rat lung.

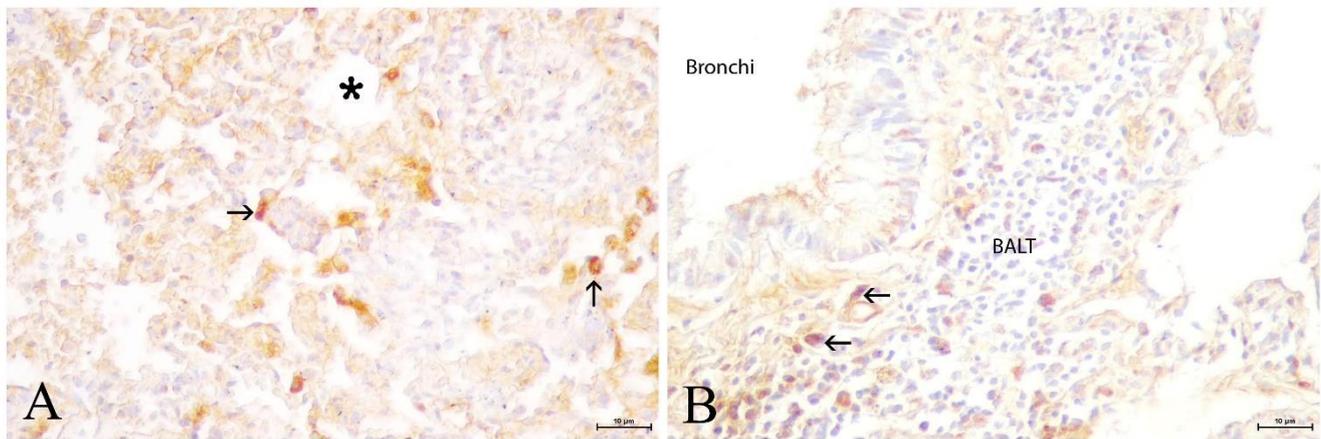


Figure 3: Lung tissue immunostained with antibodies against ghrelin; (A) Control group, (→): ghrelin immunopositive cell, (asterix): alveolar space, (B) *S. nigra* group, (→): ghrelin immunopositive cell, (BALT): bronchus-associated lymphatic tissue, original magnification X40; range bar, 10 μ m.

DISCUSSION

Numerous food components, due to their immunomodulatory properties, have been shown to activate MCs as well as modulate the synthesis of MC mediators (Uranga et al. 2020). Cells of the immune system are highly sensitive to changes in metabolism status. They can affect by changes in circulating hormones, which can affect immune responses and cytokine expression (Baatar et al. 2011).

Chymase plays an important role in the regulation of coagulation by activating and catalyzing the degradation of thrombin and plasmin (Dell'Italia LJ and Husain, 2002). MC_{TC} has been shown to contribute to tissue remodeling, fibroblast mitogenicity, and angiogenesis in lung tissue (Mitani et al. 1999). Increased numbers of MC_{TC} have been found in chronic asthma (Van der Velden et al. 2012), lungs with interstitial pneumonia (Hirata et al. 2007), and blunt lung trauma (Tütüncü et al. 2020). It is also known to cause an increase in MC_{TC} in many conditions such as viral infections, asthma, chronic obstructive pulmonary syndrome, pulmonary hypertension, and fibrosis in the lungs (Kosanovic et al. 2015). However, although there are studies on some active substances with protective effects on the organism in the literature, we have not found any studies on the effects of *S. nigra* on chymase expression in the lung. Because of this, our study is the first one conducted in this field. There are studies in the literature on the effects of other active ingredients on the number of MC_{TC} s. For example, in a study evaluating the effects of thymoquinone (TQ) application method and dose on the expression of the cytokines in the rat spleen, it was observed that TQ, which has an immunomodulatory effect, did not directly affect the number of MC_{TC} (Ertuğrul et al. 2021). However, Hayiroğlu et al. (2016) emphasized that chymase expression increases in metabolic diseases such as diabetes and hormonal changes. Moreover, it was found in the study conducted by

Tütüncü et al. (2020) that MC counts were closely related to polyphenolic antioxidants with anti-inflammatory effects. It was reported that there is a close relationship between increased chymase expression and resveratrol application. In this study, we found that *S. nigra*, an immune system stimulating product, may have positive effects on MC_{TC} in the lung. It is well known that MCs play a key role in lung pathophysiology. We think that increased MC_{TC} with *S. nigra* application may have a beneficial effect on possible lung disorders.

Tryptase is a neutral protease that plays an important role in allergic diseases, cytokine release, adhesion molecule expression, and smooth muscle bronchi contraction (Pejler et al. 2010). Also, tryptase is a known potent growth factor for epithelial and smooth muscle cells of the airways in lung tissue (Payne and Kam, 2004). It has been observed that the number of MC_{TS} increases in diseases acute lung injury, sepsis, pneumonia (Zhao et al. 2014). Montelukast (Çetinel et al. 2011), which is used in the treatment of asthma, and ketamine (Li et al. 2014), which can be used for pain relief and sedation, have been shown in studies to cause an increase in the number of MC_{TS} . Furthermore, resveratrol, an active element in the structure of many plants that has an antibacterial effect and can be utilized against infections, has been shown to increase MC_{TS} (Tütüncü et al. 2020). Additionally, it was demonstrated that capsaicin increased the density of tryptase-positive cells. (Tutuncu and Ertuğrul, 2019). Immunostaining against tryptase found in the granules of MCs is one of the most effective methods for identifying MCs. Parallel to the above studies, it has been demonstrated that MC_{TS} behavior can vary according to different active substances. In this study, it was observed that *S. nigra*, effectiveness was investigated recently can also affect the number of MC_{TS} . It is known that MCs are multifunctional

effector cells involved in host defense. Based on the study's findings, it can be said that *S. nigra* may be effective in protecting the lung tissue indirectly by increasing the number of MC_{TS}.

Ghrelin regulates many cellular functions and physiological processes, including apoptosis, vascular permeability, and both innate and adaptive immunity. It also contributes to the healing of various lung diseases such as pulmonary edema, emphysema, cystic fibrosis, and pneumonia (Chen et al. 2008, Schwenke et al. 2008). Ghrelin is known to regulate the expression of inflammatory cytokines and plays an important role in immune cell function (Xia et al. 2004). Ghrelin-producing cells are found in various tissues such as the hypothalamus, pituitary, stomach, heart, lung, pancreas, intestine, kidney, testes, and ovaries (Gnanapavan et al. 2002). Volante et al. (2002) observed by immunohistochemistry using a specific antibody that ghrelin-producing cells have a polygonal or elongated shape and are found in small clusters in fetal, infant, and adult human lungs. It has also been shown that ghrelin-positive cells in the lung are found mainly around the bronchi, in the bronchiolar wall, and in the alveolar septa (Ivanova et al. 2021). Stefanov et al. (2017) reported that in double immunofluorescence staining, both ghrelin and tryptase are expressed by the same MCs in rat stomachs. Additionally, it was demonstrated that ghrelin-positive cells have a similar morphology and location to MC_{TS}. Moreover, co-localization with tryptase immunoreactivity in serial sections from the porcine bile duct suggested that most ghrelin-positive cells might also be MC_{TS} (Stefanov, 2021). Stefanov et al. (2021) used immunohistochemistry to investigate the distribution of ghrelin cells and MC_{TS} in the domestic pig and found that the numbers of ghrelin and tryptase immunoreactive positive cells both varied in parallel to the tunics of the common hepatic duct. Furthermore, it was shown that the percentages of ghrelin- and tryptase-positive cells increased in parallel with age in the interalveolar septa of the rat lung (Ivanova and Stefanov, 2021). Our data showed that *S. nigra* increased ghrelin immunopositive cell expression. In addition, an increase in tryptase immunopositive cells was also observed in our study. Also, we found that ghrelin- and tryptase-positive cells were morphologically similar to each other. This approach does not allow for precise identification but considering in our study, ghrelin, tryptase positive cells, which increased in density with *S. nigra* application, and previous studies, it might be concluded that MCs can produce and secrete ghrelin.

CONCLUSION

This study showed that *S. nigra*, which has an immunomodulatory and antioxidant effect, increases

the expression of chymase-, tryptase- and ghrelin-positive cells in lung. Also, it has been tried to explain that MCs can produce, store and release ghrelin like other immune cells. In conclusion, we think that the findings we obtained in this study will contribute to the literature on the potential role of *S. nigra*, the MC, and ghrelin in the respiratory and immune systems.

Conflict of Interest: The authors declare that there is no conflict of interest for this article and no financial support has been received.

Ethics Committee Information: The tissue samples used in our study were obtained from the project named “*Sambucus Nigranın* diyabetli rat dalağında mast hücre ve Vasküler Endotelial Büyüme faktörü (VEGF) üzerine etkilerinin histokimyasal ve immunohistokimyasal olarak incelenmesi” which is approved by the Animal Ethics Committee of Ondokuz Mayıs University (Decision no: 11.03.2020, number 15).

Financial Support: --

Acknowledgment: --

Description: --

Authors Contribution Rate: TE:%65, GS%35

REFERENCES

- Akalu Y, Molla MD, Dessie G, Ayelign B. Physiological effect of ghrelin on body systems. *Int J Endocrinol.* 2020; 25:1385138. doi: 10.1155/2020/1385138.
- Baatar D, Patel K, Taub DD. The effects of ghrelin on inflammation and the immune system. *Mol Cell Endocrinol.* 2011; 340(1): 44-58. doi: 10.1016/j.mce.2011.04.019.
- Bidian C, Mitrea DR, Tatomir C, Perdeschrepler M, Lazăr C, Chetan I, Bolfa P, David L, Clichici S, Filip GA, Mureşan M, Micle O. *Vitis Vinifera* L. And *Sambucus Nigra* L. extracts attenuate oxidative stress and inflammation in femoral ischemia. *Farmacía.* 2021; 69:1. doi.org/10.31925/farmacía.2021.1.8.
- Chen J, Liu XJ, Shuo QL, Li SQ, Luo FM. Ghrelin attenuates lipopolysaccharide-induced acute lung injury through no pathway. *Med Sci Monitor.* 2008; 14(7): 141-146.
- Chowen JA, Argente J. Ghrelin: A Link Between Energy Homeostasis and the Immune System. *Endocrinology.* 2017; 158(7): 2077-2081. doi: 10.1210/en.2017-00350.
- Çetinel S, Çamilhoğlu YE, Çikler E, Sener G, Ercan F. Leukotriene D4 receptor antagonist montelukast alleviates protamine sulphate-induced changes in rat urinary bladder. *BJU Int.* 2011; 107(8): 1320-1325. doi: 10.1111/j.1464-410X.2010.09532.x.
- Da Silva EZM, Jamur MC, Oliver C. Mast cell function: a new vision of an old cell. *J Histochem Cytochem.* 2014; 62(10): 698-738. doi: 10.1369/0022155414545334.
- Dell'Italia LJ, Husain A. Dissecting the role of chymase in angiotensin II formation and heart and blood vessel diseases. *Curr Opin Cardiol.* 2002; 17(4): 374-379. doi: 10.1097/00001573-200207000-00009.

- Dwyer DF, Barrett NA, Austen KF.** Expression profiling of constitutive mast cells reveals a unique identity within the immune system. *Nat Immunol.* 2016; 17(7):878-887. doi: 10.1038/ni.3445.
- Duymuş HG, Göger F, Başer KHC.** In vitro antioxidant properties and anthocyanin compositions of elderberry extracts. *Food Chem.* 2014; 155: 112-119. doi: 10.1016/j.foodchem.2014.01.028.
- Ertuğrul T, Tutuncu Ş, Ozdemir B, Delice N.** Possible effect of thymoquinone on mast cell number and chymase, IL-4 and IFN- γ expression in rat spleen. *Med Weter* 2021; 77(10): 484-490. doi.org/10.21521/mw.6580.
- Fazio A, Plastina P, Meijerink J, Witkamp RF, Gabriele B.** Comparative analyses of seeds of wild fruits of *Rubus* and *Sambucus* species from southern Italy: fatty acid composition of the oil, total phenolic content, antioxidant and anti-inflammatory properties of the methanolic extracts. *Food Chem.* 2013; 140(4): 817-824. doi: 10.1016/j.foodchem.2012.11.010.
- Gnanapavan S, Kola B, Bustin SA, Morris DG, McGee P, Fairclough P, Bhattacharya S, Carpenter R, Grossman AB, Korbonits M.** The tissue distribution of the mRNA of ghrelin and subtypes of its receptor, GHS-R, in humans. *J Clin Endocrinol Metab.* 2002; 87(6): 2988. doi: 10.1210/jcem.87.6.8739.
- Hamada H, Terai M, Kimura H, Hirano K, Oana S, Niimi H.** Increased expression of mast cell chymase in the lungs of patients with congenital heart disease associated with early pulmonary vascular disease. *Am J Respir Crit Care Med.* 1999; 160(4): 1303-1308. doi: 10.1164/ajrccm.160.4.9810058.
- Hawkins J, Baker C, Cherry L, Dunne E.** Black elderberry (*Sambucus nigra*) supplementation effectively treats upper respiratory symptoms: a metaanalysis of randomized, controlled clinical trials. *Complement Ther Med.* 2019; 42:3 61-65. doi.org/10.1016/j.ctim.2018.12.004.
- Hayiroğlu AE, Karaca T, Demirtaş S.** Streptozotosin ile deneysel diyabet oluşturulan sıçanlarda östrus siklusunun değişik evrelerinde ovarium ve uterus dokularında mast hücrelerinin dağılımının histokimyasal ve immünohistokimyasal olarak incelenmesi. *Kafkas J Med Sci.* 2016; 6(1):29-37. doi: 10.5505/kjms.2016.30074.
- Hirata K, Sugama Y, Ikura Y, Ohsawa M, Inoue Y, Yamamoto S, Kitaichi M, Ueda M.** Enhanced mast cell chymase expression in human idiopathic interstitial pneumonia. *Int J Mol Med.* 2007; 19(4): 565-570.
- Ivanova K, Stefanov I, Ivanova I, Ananiev J, Gulubova M.** Ghrelin expression in mast cells of infant lung with respiratory distress syndrome. *Acta Med. Bulg.* 2021; 48: 40-45. doi.org/10.2478/amb-2021-0006.
- Ivanova IG, Stefanov IS.** Tryptase- and ghrelin positive mast cells in the interalveolar septa of rat's lung. *Bulg J Vet Med.* 2021; 24(4): 469-477.
- Kojima M, Kangawa K.** Ghrelin: structure and function. *Physiol Rev.* 2005; 85(2): 495-522. doi: 10.1152/physrev.00012.2004.
- Kosanovic D, Luitel H, Dahal BK, Cornitescu T, Janssen W, Danser AH, Garrelds IM, De Mey JGR, Fazzi G, Schiffers P, Iglarz M, Fischli W, Ghofrani HA, Weissmann N, Grimminger F, Seeger W, Reiss I, Schermuly RT.** Chymase: a multifunctional player in pulmonary hypertension associated with lung fibrosis. *Eur Respir J.* 2015; 46(4): 1084-1094. doi: 10.1183/09031936.00018215.
- Li M, Yang K, Wang X, Xu X, Zhu L, Wang H.** Mast cells infiltration and decreased E-cadherin expression in ketamine-induced cystitis. *Toxicol Rep.* 2014; 8(2): 205-209. doi: 10.1016/j.toxrep.2014.12.003.
- Metcalfe DD, Baram D, Mekori YA.** Mast cells. *Physiol Rev.* 1997; 77(4):1033-1079. doi: 10.1152/physrev.1997.77.4.1033.
- Miyazaki M, Takai S, Jin D, Muramatsu M.** Pathological roles of angiotensin II produced by mast cell chymase and the effects of chymase inhibition in animal models. *Pharmacol Ther.* 2006; 112(3):668-676. doi: 10.1016/j.pharmthera.2006.05.008.
- Mitani Y, Ueda M, Maruyama K, Shimpo H, Kojima A, Matsumura M, Aoki K, Sakurai M.** Mast cell chymase in pulmonary hypertension *Thorax.* 1999; 54(1): 88-90. doi: 10.1136/thx.54.1.88.
- Mukai K, Tsai M, Saito H, Galli SJ.** Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol Rev.* 2018; 282(1): 121-150. doi: 10.1111/imr.12634.
- Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, Matsukura S.** A role for ghrelin in the central regulation of feeding. *Nature.* 2001; 409(6817):194-8. doi: 10.1038/35051587.
- Narin N, Çetin E.** Effect of ghrelin administration on some hematological parameters in rats. *J Health Sci.* 2010; 19(3): 202-208.
- Oniszcuk A, Olech M, Oniszcuk T, Wojtunik-Kulesza K, Wójtowicz A.** Extraction methods, LC-ESI-MS/MS analysis of phenolic compounds and antiradical properties of functional food enriched with elderberry flowers or fruits. *Arabian Journal of Chemistry.* 2016; 12(8): 4719-4730. doi.org/10.1016/j.arabjc.2016.09.003.
- Payne V, Kam PC.** Mast cell tryptase: a review of its physiology and clinical significance. *Anaesthesia* 2004; 59(7): 695-703. doi: 10.1111/j.1365-2044.2004.03757.x.
- Pejler G, Ronnberg E, Waern I, Wernersson S.** Mast cell proteases: multifaceted regulators of inflammatory disease. *Blood.* 2010; 115(24): 4981-4990. doi: 10.1182/blood-2010-01-257287.
- Raghay K, Akki R, Bensaid D, Errami M.** Ghrelin as an anti-inflammatory and protective agent in ischemia/reperfusion injury. *Peptides.* 2020; 124:170226. doi: 10.1016/j.peptides.2019.170226.
- Sakaguchi M, Takai S, Jin D, Okamoto Y, Muramatsu M, Kim S, Miyazaki M.** A specific chymase inhibitor, NK3201, suppresses bleomycin-induced pulmonary fibrosis in hamsters. *Eur J Pharmacol.* 2004; 493(1-3): 173-176. doi: 10.1016/j.ejphar.2004.04.024.
- Samrao D, Wang D, Ough F, Lin YG, Liu S, Menesses T, Yessaian A, Turner N, Pejovic T, Mhawech-Fauceglia P.** Histologic parameters predictive of disease outcome in women with advanced stage ovarian carcinoma treated with neoadjuvant chemotherapy. *Transl Oncol.* 2012; 5(6): 469-474. doi: 10.1593/tlo.12265.
- Schwartz LB, Lewis RA, Seldin D, Austen KF.** Acid hydrolases and tryptase from secretory granules of dispersed human lung mast cells. *J Immunol.* 1981; 126(4): 1290-1294.
- Schwenke DO, Tokudome T, Shirai M, Hosoda H, Horio T, Kishimoto, Kangawa K.** Exogenous ghrelin attenuates the progression of chronic hypoxia-induced pulmonary hypertension in conscious rats. *Endocrinology.* 2008; 149(1): 237-44. doi: 10.1210/en.2007-0833.
- Sonneck K, Florian S, Böhm A, Krauth MT, Kondo R, Hauswirth AW, Gleixner KV, Aichberger KJ, Derdak S, Pickl WF, Sperr WR, Schwartz LB, Valent P.** Evaluation of biologic activity of tryptase secreted from blast cells in acute myeloid leukemia. *Leuk Lymphoma.* 2006; 47(5): 897-906. doi: 10.1080/10428190500513652.

- Stefanov IS, Ananiev JR, Ivanovab KV, Tolekovic AN, Vodenicharovd AP, Gulubova MV.** Distribution of ghrelin-positive mast cells in rat stomach. *Biotechnol Biotechnol Equip.* 2017; 31(4): 774–781. doi.org/10.1080/13102818.2017.1326013.
- Stefanov I.** Mast cell distribution in porcine common bile duct with special reference to ghrelin. *Bulg J Vet Med.* 2021; (online first). doi: 10.15547/bjvm.2020-0127.
- Stefanov I, Vodenicharov A, Atanassova P, Hrishev P, Vulkova I, Stoyanov D, Tsandev N, Hristov HA.** Mast cell density in domestic swine common hepatic duct. *Bulg. J. Vet. Med.* 2021; (online first). doi: 10.15547/bjvm.2020-0145.
- Tomimori Y, Muto T, Saito K, Tanaka T, Maruoka H, Sumida M, Fukami H, Fukuda Y.** Involvement of mast cell chymase in bleomycin-induced pulmonary fibrosis in mice. *Eur J Pharmacol.* 2003; 478(2-3): 179-185. doi: 10.1016/j.ejphar.2003.08.050.
- True LD.** Principles of immunohistochemistry, Gower Medical Publishing, New York. 1990; p. 67-75.
- Tutuncu S, Ertuğrul T.** Immunohistochemical expression of tryptase-chymase and mast cell heterogeneity in capsaicin-treated rat ovaries. *IJVAR.* 2019; 2(2): 25-31.
- Tütüncü Ş, Torun AÇ, Ertuğrul T.** The role of resveratrol on mast cell and chymase and tryptase expression in blunt-chesttrauma-induced acute lung injury in rats. *Turk J Vet Anim Sci.* 2020; 44: 1260-1268. doi:10.3906/vet-2005-23.
- Xia Q, Pang W, Pan H, Zheng Y, Kang JS, Zhu SG.** Effects of ghrelin on the proliferation and secretion of splenic T lymphocytes in mice. *Regul Pept.* 2004; 122(3):173-178. doi: 10.1016/j.regpep.2004.06.016.
- Uranga JA, Martínez V, Abalo R.** Mast cell regulation and irritable bowel syndrome: effects of food components with potential nutraceutical use. *Molecules.* 2020; 25(18): 4314. doi: 10.3390/molecules25184314.
- Van der Velden J, Barker D, Barcham G, Koumoundouros E, Snibson K.** Increased mast cell density and airway responses to allergic and non-allergic stimuli in a sheep model of chronic asthma. *PLoS One.* 2012; 7(5): e37161. doi: 10.1371/journal.pone.0037161.
- Volante M, Fulcheri E, Allia E, Cerrato M, Pucci A, Papotti M.** Ghrelin expression in fetal, infant, and adult human lung. *J Histochem Cytochem.* 2002; 50(8): 1013-1021. doi: 10.1177/002215540205000803.
- Wieland LS, Piechotta V, Feinberg T, Ludeman E, Hutton B, Kanji S, Seely D, Garritty C.** Elderberry for prevention and treatment of viral respiratory illnesses: a systematic review. *BMC Complement Med Ther.* 2021; 21(1): 112. doi: 10.1186/s12906-021-03283-5.
- Zhao W, Gan X, Su G, Wanling G, Li S, Hei Z, Yang C, Wang H.** The interaction between oxidative stress and mast cell activation plays a role in acute lung injuries induced by intestinal ischemia-reperfusion. *J Surg Res.* 2014; 187 (2): 542-552. doi.org/10.1016/j.jss.2013.10.033.