

A New Eco-friendly Approach for Fabrication of Electrically Conductive and Antibacterial Polyamide Yarns

Seyma Kanara¹  0000-0002-0596-3311

Suat Cetiner¹  0000-0002-6604-145X

¹Kahramanmaraş Sutcu Imam University, Department of Textile Engineering, Kahramanmaraş, Türkiye

ABSTRACT

In this paper, an environmentally friendly method to produce electrically conductive and antibacterial silver nanoparticles (AgNPs) coated polyamide (PA) yarns was reported. A new AgNPs coated composite PA yarn has been synthesized in the presence of carboxy methyl starch (CMS) reducing agent. Silver nitrate (AgNO₃) was reduced on the surface of the PA fibers which allowed formation of silver nanoparticles. Scanning electron microscopy (SEM) images indicated distribution of silver nanolayer formation on the composite fiber surface. Electrical conductivity of composite yarns was measured by four point probe technique and was changed from 1.015x10⁻⁵ to 1.282x10⁰ S/cm. The ecofriendly coating method used in this study enabled to produce multifunctional composite PA yarns with antistatic and antibacterial properties.

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1. INTRODUCTION

Increasing the awareness towards green chemistry and other biological processes evoked the interest to develop an eco-friendly approach for the synthesis of nanoparticles which has several advantages such as simplicity, cost effectiveness, compatibility for textile application as well as for large scale commercial production [1-5]. Utilization of nontoxic chemicals, environmentally benign solvents and renewable materials are some of the key issues that merit important consideration in a green synthesis strategy. Nanocomposites are used in many applications such as medical textiles, cosmetics, agriculture, optics, food packaging, semiconductor devices, aerospace, construction and catalysis. New properties are imparted to textiles using nanotechnology such as anti-bacteria, UV-protection, antistatic, improvement of dye ability, flame retardancy, water repellency, soil resistance and wrinkle resistance [3]. There are various materials used in the production of multifunctional nanocomposite textiles. The AgNP is a well-known effective antibacterial agent, which is commonly used in commercial products including in the textile industry to avoid microbial contamination.

Nowadays, silver nanoparticles (AgNPs) are increasingly used in various fields such as medical device coatings, food industry, health care, protective clothing, medical textiles, optical sensors, cosmetic product, seat covers. Silver nanoparticles easily interact with other particles due to their physical, chemical, thermal, optical, high electrical conductivity and biological properties [6-11]. Biologically-mediated synthesis of nanoparticles have been shown to be simple, cost effective, dependable, and environmentally friendly approaches. Several studies reported the synthesis of AgNPs using green, cost effective, and biocompatible methods without the use of toxic chemicals in biological methods. Compared to chemical methods, biological methods allow for more ease in the control of shape, size, and distribution of the produced nanoparticles by optimization of the synthesis methods, including the amount of precursors, temperature, pH, and the amount of reducing and stabilizing parameters [11, 12]. The present study involves the incorporation of silver nanoparticles on polyamide fibres by “green synthesis strategy”. The advantages of this process are: (a) no need to have extra reducing agent(s), (b) the process can be conducted at room temperature, under normal pressure in aqueous solution,

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and (c) finally the developed silver nanoparticles by this process have excellent properties, including long term dispersion stability [1-5].

Polyamide (PA) is the third largest textile fiber consumed in the world, after polyester and cotton. PA-knitted goods are used in underwear, sports and swimwear, stockings and tights, bed linen, car upholstery, nets, and others. Besides, the polyamide textiles with multifunctional properties can be applied in various commercial usages due to good electrical conductivity, light weight, and corrosion resistance along with enhanced mechanical properties [10,13]. In order to obtain a multi-functional yarn with an extremely high improvement in adhesion of silver nanolayer, the polyamide yarn has been chosen as the substrate. Polyamide is an electron-rich and polar synthetic polymer (polyamide) usually made of adipoyl chloride and hexamethylene diamine monomers to form a linear molecular chain[13].

There are various methods to obtain conductive textiles such as the use of conductive agent and particles, metallic fibers, coating with conductive polymers. The use of conductive agents and particles is a cost effective process that has been attracted much attention because of its low cost and fiber structural features do not change. Hasan et al. studied to develop chitosan-mediated green silver nanoparticles for polyester fabric coloration. They developed that a novel route for the coloration of polyester fabric with green synthesized silver nanoparticles using chitosan as a natural eco-friendly reductant and explained that the proposed coating process using green nanoparticles can contribute to low-cost production of sustainable textiles [14]. Gawish et al. investigated that antibacterial and antifungal properties against *Staphylococcus aureus*, *Klebsiella pneumonia* and *Candida albicans* by using standard methods (AATCC TM100). Surface electrical conductivity and color changes are measured for the treated fabrics and they determined nano silver coated polyamide fabric displayed electrical conductivity better than the untreated one [3]. Sang Su et al. fabricated highly conductive graphene/Ag hybrid fibers for flexible fiber type transistors. This fibers have flexible, and electrically conductive textiles are highly suitable for use in wearable electronic applications. They emphasised that exhibited excellent flexibility, high electrical performance and outstanding device performance stability [15]. Montazer et al. studied a new method to produce conductive polyester fabric by using a novel electroless plating of silver nanoparticles. They investigated silver nanowires and cupro fabrics together using a dipping-drying method to prepare highly stretchable electrically conductive textiles [10]. Dong et al. reported the biosynthesis of silver nanoparticles via a single-step reduction of silver ions and emphasized that green synthesis of silver nanoparticles, using hydroxy propyl methyl cellulose and glucose as capping agents and reducing agents respectively, is an environmental friendly, simple and efficient route for synthesis of metallic nanoparticles. By they investigated using

HPMC, the effects of the reaction time, temperature, concentration of silver ion, reducing agents on the particle size were investigated and measured AgNP size range from 3 nm to 17 nm according to SEM images [16]. Ahmed H.B., et al studied the silver nanoparticles-carboxy methyl cellulose (AgNPs-CMC) composite prepare by interaction of silver nitrate with CMC using pad-dry-cure method. They found electrical resistance as 172 M Ω after application of 10 CMC layers with excellent antibacterial property [17]. Hebeish A. et al. (2011) investigated that green synthesis of nano sized colloidal silver solution using hydroxypropyl starch (HPS). Also, It has determined the optimum conditions formation of AgNP as temperature, time and concentration on cotton fabrics. A similar approach based on the use of silver nanoparticle is presented by Hebeish A. et al. (2011) In this case, Ag NPs have been prepared using hydroxypropyl starch (HPS) as a reducing agent through green synthesis which reduce silver ions (Ag⁺) to nanoparticles (Ag⁰). In addition, it was determined that optimum conditions as 0.9 g HPS (MS: 0.84), AgNO₃; 0.078 g, pH; 12, temperature; 70°C, duration; 15 min. They determined in the range of 6–8 nm of AgNPs colloids with excellent size and distribution according to SEM images [18].

In this study, an environmentally friendly coating method was introduced to fabricate a multifunctional silver nanoparticle coated composite polyamide yarn with desirable electrically conductivity and antibacterial properties in the presence of carboxy methyl starch reducing agent. The advantage of this method is preparing of AgNPs without any organic solvents or other chemical reducing agents and contributing to sustainable textile production. The composite PA yarns can be used in a wide range of applications such as wound dressings, hygienic clothing and medical applications where the presence of bacteria is hazardous.

2. MATERIAL AND METHOD

2.1 Material

Polyamide multifilament yarns (technical properties; 70 denier, two ply, 10 filament (70/2/10)) were supplied from Karacasu Textile, Turkey. AgNO₃ (>99 %) were purchased from Tekkim Chemicals. Carboxy methyl starch (CMS) was synthesized from potato starch waste in ÜSKİM laboratory by Dolaz et al [19]. Ethanol and acetone were supplied from Sigma Aldrich and were used as received.

2.2 Method

2.2.1. Preparation of silver coated polyamide yarns

The silver-coated PA yarn was fabricated by in-situ reduction of silver nitrate. In the first set of optimization studies, the effect of AgNO₃ concentration and process time were investigated in the development of silver coated PA yarns (Table 1).

Table 1. Coating receipt of AgNPs polyamide yarns

Sample	AgNO ₃ (gr)	CMS (gr)	Time (h)
F025	0,25	0,05	1,2,3,4,5
F05	0,625	0,125	1,2,3,4,5
F1	1	0,2	1,2,3,4,5
F2	2	0,4	1,2,3,4,5
F5	5	1	1,2,3,4,5

Selected concentrations of AgNO₃ (i.e.; 0,25 g/100 mL, 0,5 g/100 mL, 1 g/100mL, 2 g/100mL and 5 g/100mL) were dissolved in distilled water at room temperature and PA multifilament yarn was wetted with AgNO₃ solution. Then, PA multifilament yarn was treated with prepared CMS solution at room temperature for selected process time and concentration to obtain a thin silver nanolayer formation on the fiber surface.



Figure 1. Preparation of AgNPs coated composite PA yarn

The color of reducing solution turned light brown to grey depending on time and amount of silver ions. After the reducing process, the PA yarns were rinsed with distilled water and ethanol. At last, the PA yarns were dried in an oven at 60 °C (Figure 1).

2.2.2. Characterization

In order to investigate the formation of silver nanoparticles on the PA fibers, a Carl Zeiss Evo LS10 scanning electron microscope is used with an accelerating voltage of 3 kV. The samples were gold coated and placed on the aluminum stub and observed under vacuum. Energy dispersive X-ray (EDX) measurements for element analysis were analyzed using Carl Zeiss Evo LS10 instrument. A UV/visible spectrometer (Lambda 750, Perkin- Elmer) was also used to investigate the mono dispersity of the nanosilver particles. XRD patterns were obtained using an X'Pert PRD PAN alytical instrument. Thickness of the AgNPs coated PA

yarns was measured by Mitutoyo Digimatic Outside Micrometer (MDC-25SB). The direct current (DC) electrical conductivity measurements were carried out by a DC conductivity meter using four point probe method (Entek FPP 510). The antimicrobial activity of AgNPs coated PA yarns was determined by evaluating of inhibition zones after 24 h of incubation at 37 °C via disc diffusion method. Martindale Abrasion Tester was used to determine the abrasion resistance of PA compositiye yarns with ISO 12945-1.

3. RESULTS AND DISCUSSION

3.1 UV–Vis Spectroscopy

Figure 2 shows the change in color of silver nanoparticle solutions at different AgNO₃/CMS concentrations.

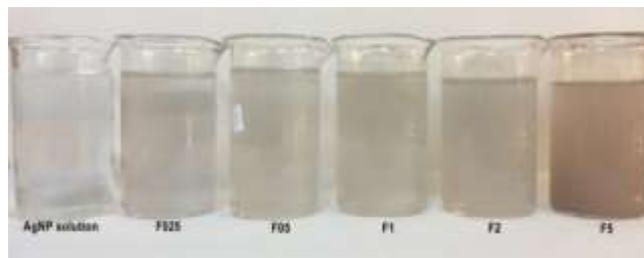


Figure 2. Silver nanoparticle coating solutions at different AgNO₃/CMS concentrations.

It is observed that the color of the colloid solutions were increasingly changed grey to purple with the increasing of AgNO₃/CMS concentration (Figure 2). Figure 3 presents UV–visible spectra of the AgNPs colloid solutions which were obtained in the wavelength range 200–600 nm.

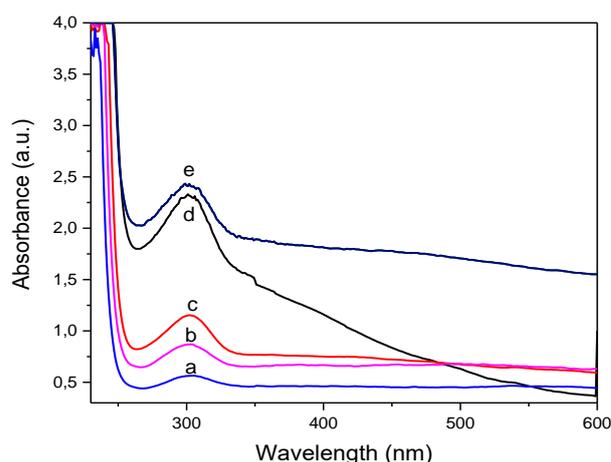


Figure 3. UV–Visible spectra of AgNP coating solutions at different AgNO₃/CMS concentrations (a.F025, b.F05, c. F1, d. F2, e.F5)

As a result of the UV-Vis analysis of the coating solutions containing silver nanoparticles, a distinct peak at 300 nm wavelength was observed. It was determined that the absorbance value of this peak increased due to the increase

in the amount of silver in the solution which reflects the formation of silver nanoparticles. All the solutions exhibited characteristic silver surface plasmon resonance (SPR) typically located in between 280-340 nm [1,20, 21, 22, 23].

Comparing with Fig. 3 a,b,c,d and e for the Ag nanoparticles, a clear increase in the intensity of maximum absorption peak as well as a slight shift in the peak wavelength was noticed with increase of concentration. This study shows that the absorption intensity increases as the concentration of AgNP increases, which reflects in the formation of more Ag nanoparticles. The low wavelength and narrow peak width reflect a small particle size distribution [24] The narrower widths of the SPR bands confirm the smaller size and more uniform size distribution of the silver nano-particles by increasing concentration [4]. It seemed that the AgNPs absorbed radiation in the visible regions of 280–340 nm which agrees with the previous studies [21,24].

3.2. Electrical Conductivity and Surface Morphology

DC electrical conductivity measurements were performed by four point probe technique and calculated by Van der Pauw equation [25]:

$$\sigma \text{ (S/cm)} = (\ln(2)i) / (\pi dV) \quad (1)$$

where d is coating thickness (cm), V is potential (Volt) and I is associated with current passing through the material (Ampere). The electrical conductivity measurements were repeated five times with different parts of the composite yarns and the arithmetic average of the results was used. The DC conductivity results of AgNPs coated PA composite yarns depending on AgNO₃/CMS concentration was given in Table 2.

Table 2. DC conductivity of AgNPs coated composite PA yarns depending on AgNO₃/CMS wt%

Yarn Code	Thickness (µm)	Coating (%)	Conductivity (S/cm)
Uncoated	590	-	-
F025	597	0,7	1.015x10 ⁻⁵
F05	601	1,1	3.782x10 ⁻⁴
F1	602	1,2	1.347x10 ⁻²
F2	605	1,5	6.541x10 ⁻²
F5	609	1,9	1.282x10 ⁰

In the coating process of composite polyamide yarns; it was determined that the electrical conductivity increased from 1.015x10⁻⁵ S/cm to 1.282x10⁰ S/cm depending on the increase in AgNO₃/CMS concentration in the solution. A positive correlation was found between the coating thickness and electrical conductivity (Table 2). Associated with it, the classification of samples was carried out according to their resistivity (ohm/sq) as given in table 3

[26]. It has been determined that AgNPs coated PA composite yarns can be evaluated in the conductive class according to the DC electrical measurements.

Table 3. Surface Resistivity Classification

Classification	Surface resistivity (ohm/sq)
Conductive	<10 ⁵
Dissipative	10 ⁵ to 10 ¹²
Insulating	>10 ¹²

The AgNO₃ concentration has a major role in fabrication of electrically conductive fibers. Carboxyl sites in polyamide chains cause the formation of a continuous silver nanoparticle network producing a nanolayer on the fiber surface with increasing of the concentration. In order to generate a conductive surface, homogeneous distribution of conductive silver particles without agglomeration is the key factor [13]. Therefore, scanning electron microscopy analysis was preferred to analyze the distribution of silver nanoparticles. Figure 4 shows the SEM images of uncoated PA yarn and silver coated PA yarns that were processed at different concentrations.

When the uncoated PA yarn has a smooth surface with the average diameter of 18.4 µm, the agglomerated formations are observed with the increasing of AgNO₃/CMS concentration. When the concentration increased, although the coating thickness increased, coating heterogeneity were observed especially for F2 ve F5-coded PA yarns. Similar results were obtained in the literature [10, 13, 27, 28,29]. However, homogeneously dispersed silver-coated thin layers were observed in the presence of F1-coded yarn (Figure 4). According to the electrical and morphological analysis, the concentration of AgNO₃/CMS was determined as [1:0,2] and F1-coded yarn was selected for the process time optimization studies in terms of both homogeneous distribution and sufficient conductivity level (Table 2, Figure 4).

The DC conductivity of F1-coded PA composite yarns depending on process time was given in Table 4.

In the coating process of composite PA yarns; it was determined that the electrical conductivity increased from 2.107x10⁻⁵ S/cm to 4.442x10⁻² S/cm depending on the increase in process time. According to the Table 4, higher conductivity values were measured especially at the processing times of 3 hours and after. A positive correlation was found between the coating thickness and electrical conductivity with time (Table 4).

Figure 5 shows the SEM images of F1-coded yarns which were processed at different time.

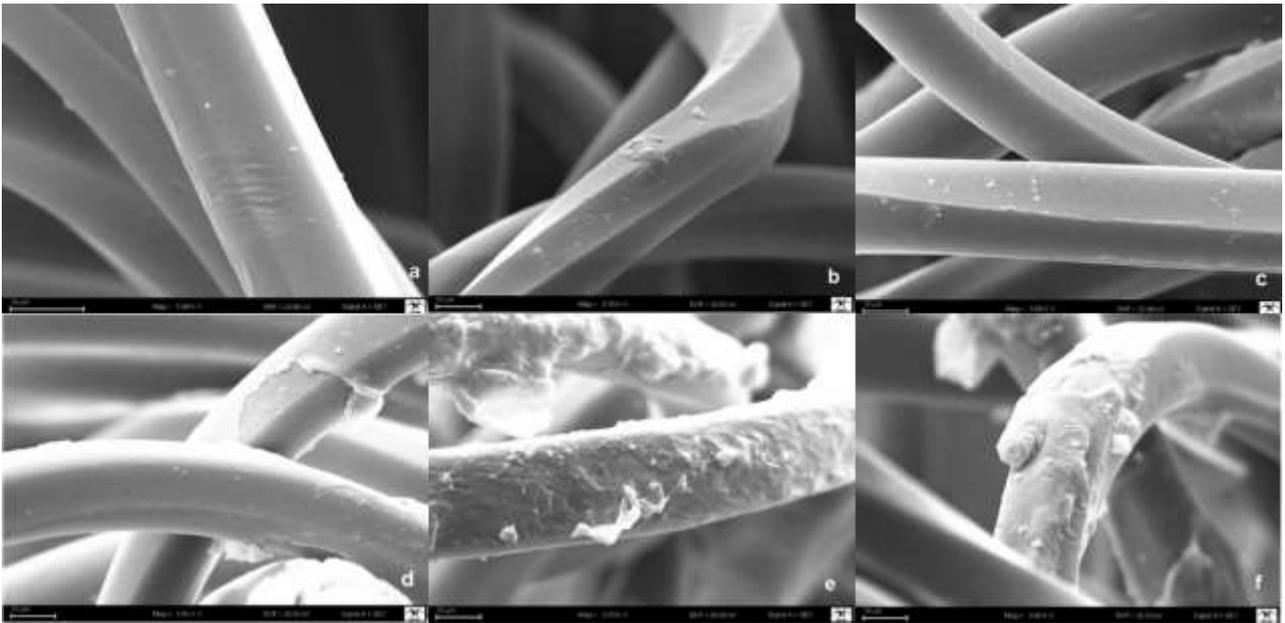


Figure 4. SEM images of AgNPs coated composite PA yarns (a. Uncoated yarn b. F025, c. F05, d. F1, e. F2, f. F5, Scale: 10 μm)

Table 4. DC conductivity of F1-coded composite PA yarns depending on process time

Time (h)	Thickness (μm)	Coating (%)	Conductivity (S/cm)
Uncoated	590	-	-
1	602	1,2	$2,107 \times 10^{-5}$
2	604	1,4	$3,754 \times 10^{-4}$
3	605	1,5	$1,389 \times 10^{-2}$
4	606	1,6	$4,250 \times 10^{-2}$
5	608	1,8	$4,442 \times 10^{-2}$

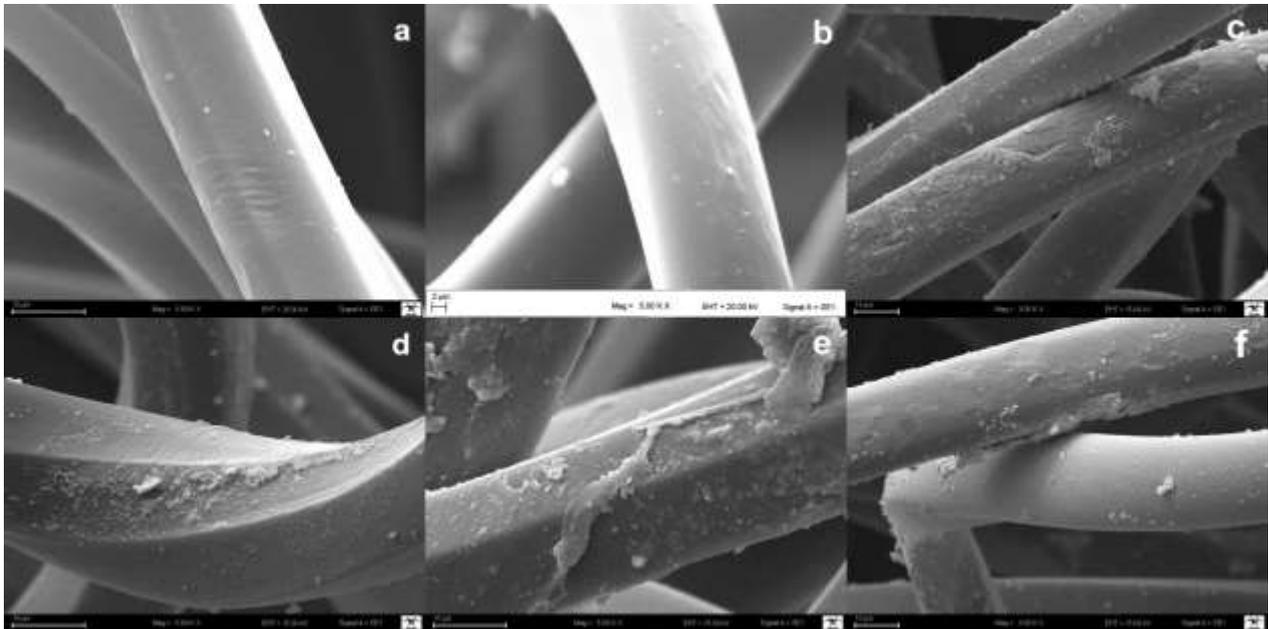


Figure 5. SEM images of AgNPs coated PA composite yarns (a.Uncoated yarn b. 1h, c. 2h, d. 3h, e. 4h f. 5h, Scale: 10 μm)

When F1-coded yarns were subjected to the coating process at different times, the presence of densely silver nanoparticles on the yarn surfaces were observed with the increase of the processing time. Especially in the coating

times between 3 and 5 hours, the presence of more dense and heterogeneous coating regions were observed. SEM images clearly depicted the formation of a nanolayer around the PA fibers and approved the importance of

process time on the surface morphology. According to the electrical conductivity and surface morphology analysis, optimum coating time was determined as 3 hours for F1-coded yarn (Figure 5).

The SEM-EDX results of F1-coded yarn is presented in Figure 6.

Spectrum: Acquisition

Element	Series	unn. C [wt.%]	norm. C [wt.%]	Atom. C [at.%]	Error [wt.%]
Carbon	K-series	1.52	37.23	55.80	0.3
Oxygen	K-series	1.44	35.19	39.59	0.3
Silver	L-series	1.13	27.57	4.60	0.1
Total:		4.10	100.00	100.00	

Figure 6. SEM-EDX results of F1-coded yarn

Figure 6 presents the EDX spectrum of F1-coded yarn, and both weight percentages (wt %) and atomic percentages (at %) of the detected elements. Carbon and oxygen were detected in EDX spectrum due to the chemical structure of the polyamide yarns [13]. According to the EDX results of the F1-coded yarn, the percentage of silver atoms in the coated polyamide yarn was measured as 4.6% which shows a certain amount of silver nanoparticle coating on the PA yarn surface (Figure 6).

Polyamides are produced from a diamine and a dicarboxylic acid [30]. There is both electrostatic attraction and coordination interaction between Ag⁺ carboxylic and amine groups on the PA yarn surface. In the AgNPs process, the synthesized AgNPs can interact with the amine groups of the PA chains via electrostatic and coordination interaction [10]. Therefore, the AgNPs were finally attached on the PA chains. On the other hand, Ag NPs are formed on the surface by the reduction of CMS and immobilized by the coordination bonds between the hydroxyl groups of CMS and silver atoms on the surface of the Ag NPs [31,41]. The schematic presentation of the formation of silver NPs on the surface of PA yarns via reduction of Ag ions with CMS is shown in Figure 7.

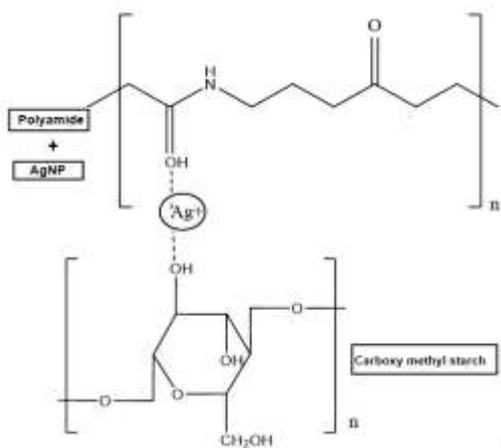


Figure 7. Schematic formation of AgNPs coated PA yarns

3.3. XRD Analysis

Basing on XRD analyses, the average size (DAg) of the Ag NPs in the modified cotton fabrics were calculated using the Debye–Scherrer equation:

$$D = K\lambda (\beta \cos\theta)^{-1} \quad (2)$$

where λ is the wavelength of the X-ray radiation (Cu K α = 0.15418 nm), K is the constant, taken as 0.89, β is the line width at half-maximum height, and θ is the diffracting angle [6,31].

Figure 8 shows XRD pattern of uncoated PA yarn and F1-coded composite PA yarn.

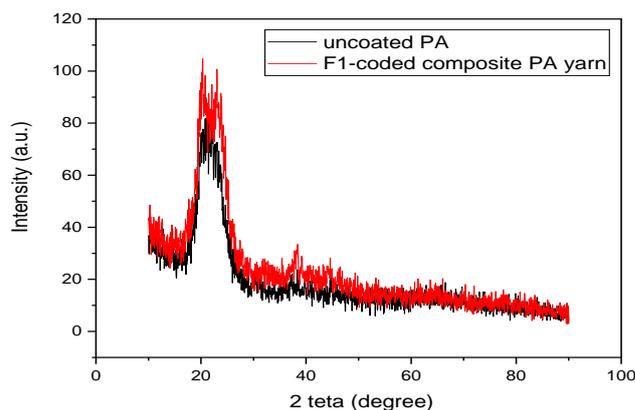


Figure 8. XRD patterns of uncoated and F1-coded composite PA yarn

It is observed that uncoated and coated PA yarns had two distinct diffraction peaks at 20.03° and 23.37° that related to nylon 6 crystals [3]. After AgNPs coating, it was observed that three different diffraction peaks appeared at 2 θ values of about 38, 44 and 44 which could be assigned to the (111) and (200) diffractions of metallic silver, respectively (Figure 8). Additionally, no characteristic peaks were observed for the other impurities such as Ag₂O indicating synthesis of pure silver metal particles [13]. This leads to the high electrical conductivity of the fabric. Also, the sharp peaks suggest the formation of highly crystalline silver particles [6]. These results in compatible with literature which suggests silver nanoparticle crystals were successfully obtained [24, 31, 32, 33].

3.4. Antibacterial Properties

The antibacterial activity of AgNPs coated composite PA yarns are determined against Escherichia Coli (E Coli), Staph Aureus, Salmonella and Bacillus Cereus by disc diffusion or Kirby-Bauer method [14, 35, 36] according to (ISO 20645:2004) (Qualitative methods) [36-38].

According to this standard, a mixture of nutrient broth and nutrient agar in 1 L distilled water at pH 7.2 as well as the empty Petri plates were autoclaved. The agar medium was then cast into the Petri plates and cooled in laminar airflow.

Approximately 10^5 colony-forming units of each bacterium were inoculated on plates [37,39].

The test specimen and pure control samples were cut into a disk shape with 6.5 mm diameter, sterilized by autoclaving for 20 min at 121°C . The assessment of antibacterial activity was based on the observation of the presence of bacterial growth in the contact zone between agar and specimen (inhibition zone), on the appearance and size of the inhibition zone formed around the specimens and on the evaluation of bacterial growth under the sample [4,36].

Figure 9 and Table 5 present antibacterial activity of AgNPs coated composite PA yarns before and after washing in the presence of different AgNO_3/CMS concentrations.

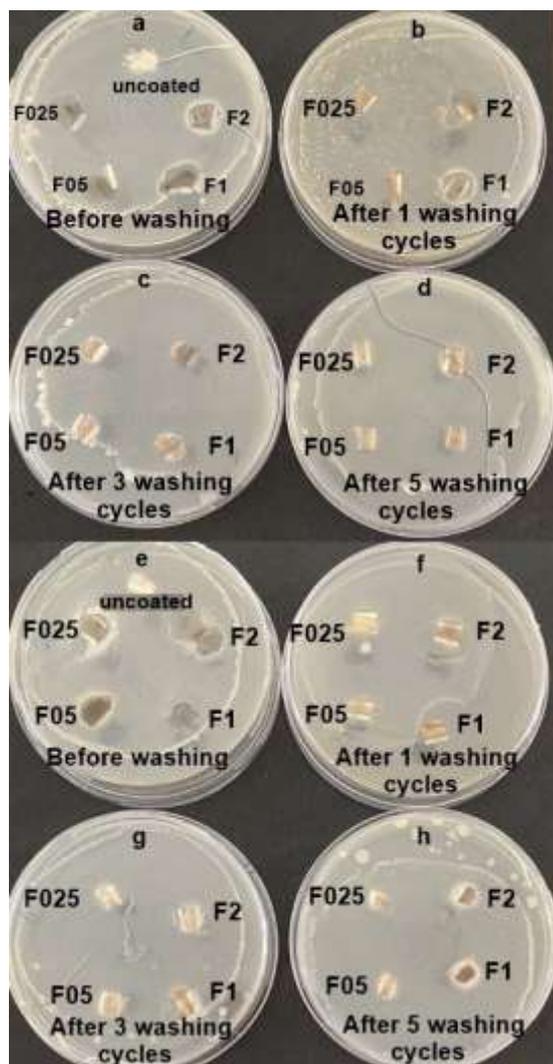


Figure 9. The antibacterial activity of AgNPs coated composite PA yarns and after different washing cycles in the presence of *Escherichia coli* (a, b, c, d) and *Staphylococcus aureus* (e, f, g, h)

Figure 9 shows the zones of inhibition around AgNPs coated composite PA yarns after 24 h of incubation in dark at 37°C . The test was repeated three times and the average value of inhibition zones are given in Table 5. Although

antibacterial activity against all selected bacteria was good for all AgNPs coated composite PA yarns, inhibition zone measurements showed variation. The uncoated PA yarn showed no antibacterial activity. However, the zone of inhibitions was evidenced the antibacterial activity of selected bacteria such as *Escherichia coli* and *Staphylococcus aureus* after coating process of PA yarn by AgNO_3 . According to inhibition zone measurements, antibacterial activity increased with increasing AgNO_3/CMS concentration against all bacterial species. The highest inhibition zone values in two different bacterial species were obtained in F1 and F2 coded samples. The antibacterial property of coated yarns can be attributed to the combination of chemical and physical interactions of bacteria with silver particles. The results indicated that all tested yarns after 1, 3, and 5 washing cycles exhibited an antimicrobial activity (Table 5). The activity reduces gradually after washing with the increase in the washing cycle. These results are compatible with the literature [40].

3.5. Abrasion Resistance

Martindale abrasion tester determines the abrasion resistance of all kinds of textile structures with ISO 12945-2. Samples are rubbed against known abrasives at low pressures and in continuously changing directions. The amount of abrasion is compared against standard parameters [41].

F1 coded yarn with the highest electrical conductivity was selected for the abrasion resistance test. Table 6 shows the abrasion tester classification of F1 coded yarns.

The abrasion test results are given for AgNPs coated composite PA yarns in Table 6. After the abrasion test, bead formation was observed a little at 3000 cycles especially after coating. It has been determined that the abrasion resistance of the F1 coded yarn is not adversely affected after AgNPs coating.

4. CONCLUSION

In this study, an environmentally friendly green synthesis method without any organic chemicals was introduced to develop multifunctional AgNPs coated composite PA yarns with desirable electrical conductivity and antibacterial properties. The electrical conductivity increased depending on the increase of AgNO_3/CMS concentration. A positive correlation was found between the coating thickness and electrical conductivity. According to the electrical and morphological analysis, the concentration of AgNO_3/CMS was determined as [1:0,2] and F1-coded yarn was selected for the time optimization studies in terms of both homogeneous distribution and sufficient conductivity level. Antibacterial activity increased with increasing AgNO_3/CMS concentration against all bacterial species. The highest inhibition zone values in two different bacterial species were obtained in F1 and F2 coded samples. Our

study indicates that the in situ approach of silver nanoparticles to yarns shows a strong antimicrobial property and durability against washing even with low concentrations of AgNO₃. In summary, an environmentally friendly coating method has been developed that allows the formation of the silver nanoparticles on the PA yarn surface. The composite yarns can be used in a wide range of applications such as wound dressings, hygienic clothing and smart and technical textiles where the presence of bacteria is hazardous.

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Table 5. Inhibition zone of AgNPs coated composite PA yarns for different AgNO₃/CMS concentrations in the presence of E Coli and Staphaureus

Yarn Code	Escherichia Coli	Staphaureus	Escherichia Coli	Staphaureus	Escherichia Coli	Staphaureus	Escherichia Coli	Staphaureus
	Inhibition Zone (mm)							
	Before washing		After 1 washing cycles		After 3 washing cycles		After 5 washing cycles	
Uncoated	-	-	-	-	-	-	-	-
F025	10,3	10,1	10,1	9,1	7	8	-	5,6
F05	12,1	10,2	12	10	7,2	7,2	3,68	6,1
F1	12,9	12	11,1	11	8	9,1	5	7
F2	12,8	19,3	11,2	17	8,1	9,3	7,8	8

Table 6. The abrasion tester classification of F1 coded yarns

Test	Before the application		After the application		Test Method
	Cycle	Classification	Cycle	Classification	
Determination of abrasion resistance of textile surface	250	5	250	5	TS EN ISO 12945-2
	500	5	500	5	
	1000	5	1000	5	
	1500	5	1500	5	
	2000	5	2000	4/5	
	2500	5	2500	4	
	3000	4	3000	3/4	

- 1 During friction the fibres constantly collide with each other and fibre fragments break due to fatigue damage, resulting in the breakage of the yarn.
- 2 The fibers are pulled out of the fabric, causing a loosening of the yarn/fabric structure. After repeated action the fibres may be pulled out, leading to thinning of the yarn and fabric, and even disintegration.
- 3 Fibers are cut and broken, leading to breakage of the yarn.
- 4 The fibre surface is worn and fragments are lost from the fibre surface.
- 5 Friction generates high temperatures, causing melt or plastic deformation of the fibers. Affects the structural and mechanical properties of the fibers.

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