

The role of carbondioxide insufflation in preventing postoperative peritoneal adhesions in rats

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ABSTRACT

Aims: Adhesion is the pathological connections that occur during the healing with scar formation of peritoneal surface defects. CO₂ is used the most frequently in laparoscopic operations for insufflation. It is believed that it causes to changes in the inflammatory reply of the pneumo-peritoneum, defects in acid-base balance and decrease in peritoneal macrophage functions. CO₂ is the only gas whose immunologic effects have been shown. It has been proven in experimental studies that the CO₂ insufflation causes to local peritoneal acidosis without affecting the systemic status. Moreover, it has also been shown that it decreases the pneumo-peritoneum TNF-α and IL-6 production; however, increases the IL-10 production which is an anti-inflammatory cytokine. In the literature, the relation between the laparoscopy and the postoperative adhesions has always been explained by taking the suggestion of its causing to less tissue trauma as a basis when compared with the open surgery. The inflammatory reply of the CO₂ has been less dealt with. In this study, we wanted to find the answer to the question whether the capno-peritoneum has a role in preventing the postoperative adhesion formation only by using CO₂ without a laparoscopic operation.

Methods: 30 female Wistar Albino type rats whose weights varied between 250±20 were used in the study. The rats were divided into 5 groups. Each group had 6 rats. Rats were placed in standard polycarbon cages in groups of 6. The room temperature was kept in 21°C. The rats were fed with standard pellet food during the study and tap water was provided to them. The operational anesthesia was performed by injecting intramuscular Ketamine Hydrochloride (Ketalar, Parke Davis and Eczacıbaşı, İstanbul) 50 mg/kg and Xylazine hydrochloride (Rompun, Bayer HealthCare) 5 mg/kg.

Results: A meaningful difference ($p<0.05$) was determined between the inflammation results of the groups. The inflammation findings become lighter as moved from Group 1 to Group 5. A meaningful difference ($p<0.05$) was determined between the fibrosis results. The fibrosis findings become lower as moved from Group 1 to Group 5. A meaningful difference ($p<0.05$) was determined between the adhesion results of the groups. The adhesion findings become lower as moved from Group 1 to Group 5. A statistically meaningful difference was not determined ($p>0.05$) between the PAI values of the groups. A statistically meaningful difference was not determined ($p<0.05$) between the MDA values of the groups. The difference stems from Group 1 and Group 5. The MDA values of Group 1 is relatively higher than those of other groups; while the MDA values of Group 5 is found to be lower when compared with the other groups

Conclusion: Our results suggest that CO₂ pneumo-peritoneum has positive effects in postoperative intraperitoneal adhesion development. Since we formed a scraping model in our study, we cannot suggest that the adhesion formation is decreased with mechanical effect. The patho-physiological and molecular bases of the postoperative adhesion formation have been documented and described well. However, we consider that the capno-peritoneum and postoperative adhesion formation is prevented with anti-inflammatory effect. We need to conduct more studies to examine this mechanism.

Keywords: Postoperative adhesion, laparoscopy, capnoperitoneum

INTRODUCTION

Adhesion (being stick together) is the pathological connections that occur during the healing with scar formation of peritoneal surface defects. Adhesions may be congenital or acquired later. Congenital adhesions occur after the abnormal development of peritoneal cavity (vitello-intestinal bands etc.). Acquired adhesions are divided into two classes as inflammatory and postoperative. Inflammatory adhesions occur due to appendicitis, cholecystitis, diverticulitis, pelvic inflammatory disease, or after the use of intrauterine contraception devices.^{1,2}

- When the physiopathology of adhesion formation is considered, the following situations are observed;
- Serosal damage,
- Increase in local inflammatory reply, increase in vascular permeability and inflammatory exudate,
- Formation of free radicals such as superoxide, peroxidase and hydroxyl radicals, and these formations causing to defects in cell membrane,
- Activation of the coagulation system (fibrin formation) and the fibrinolytic system (fibrinogen degradation)

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and the mechanisms that move one within the other during the formation of imbalance intraperitoneal adhesions that occur in the balance between these two elements³⁻⁶

Menzies reported that intraabdominal adhesion developed in 79% and 93% of the patients who received intraabdominal operational procedure is his study.² The major morbidities caused by intraabdominal adhesions are intestinal obstruction infertility, difficult and risky re-explorations, ectopic pregnancy and chronic stomachache. Laparoscopy technique is used widely in surgical practices.

Laparoscopic operations mean less trauma in the peritonea, less intraoperative bleeding, polite manipulations to intraabdominal organs and tissues, less tissue damage in areas other than the operation area, less exposure to glove powder, gauze, compress and operational tools, and postoperative earlier recovery. These characteristics make the laparoscopy technique superior to conventional surgical methods in terms of adhesion formation status.

Carbon dioxide (CO₂) is used the most frequently in laparoscopic operations for insufflation; and with its biological characteristics, it is still the most suitable gas in this field. It is believed that it causes changes in the inflammatory reply of the pneumo-peritoneum, defects in acid-base balance and decrease in peritoneal macrophage functions. CO₂ is the only gas whose immunologic effects have been shown. It has been proven in experimental studies that the CO₂ insufflation causes to local peritoneal acidosis without affecting the systemic status. Moreover, it has also been shown that it decreases the pneumo-peritoneum TNF-α and IL-6 production; however, increases the IL-10 production which is an anti-inflammatory cytokine. In the literature, the relation between the laparoscopy and the postoperative adhesions has always been explained by taking the suggestion of its causing to less tissue trauma as a basis when compared with the open surgery. The inflammatory reply of the CO₂ has been less dealt with. In this study, we wanted to find the answer to the question whether the capno-peritoneum has a role in preventing the postoperative adhesion formation only by using CO₂ without a laparoscopic operation.

METHODS

This experimental study was carried out with the permission of Gazi University Animal Experiments Local Ethics Committee (Date: 04.09.2011, Decision No: B.30.2.GÜN.O.EU.00.00/38-5869.30). The study adhered to the animal research guidelines of the National Institute of Health.

30 female Wistar Albino type rats whose weights varied between 250±20 were used in the study. The rats were divided into 5 groups. Each group had 6 rats. Rats were placed in standard polycarbon cages in groups of 6. The room temperature was kept in 21°C. The rats were fed with standard pellet food during the study and tap water was provided to them. The operational anesthesia was performed by injecting intramuscular Ketamine Hydrochloride (Ketalar, Parke Davis and Eczacıbaşı, İstanbul) 50 mg/kg and Xylazine hydrochloride (Rompun, Bayer HealthCare) 5 mg/kg.

Operational Procedure

The necessary sterile conditions were provided after the anesthesia, and the rats were fixed to the operation board from their four extremities, and abdominal front walls of the rats were shaved and cleaned with povidone iodine. The cannula was connected to an electronic insufflator (Storz and Co., Tutthufen, Germany) and the CO₂ was insufflated so as it remained in 6 mmHg (Picture 1). After the operation, the rats were put in the cages in groups. The rats were then observed for 10 days in standard feeding and living conditions. Tissue samples were collected and frozen with liquid nitrogen, and kept in -80 °C until the analysis time.



Picture 1. CO₂ was insufflated so as it remained in 6 mmHg

Groups

Group I: The Sham Group (n: 6): Laparotomy was performed from the middle line, and closed without any interventions with 3/0 vicril.

Group II: The Control Group (n: 6): Laparotomy was performed from the middle line, and the petechial bleedings were performed with sterile dry sponge over the cecum, and the ileo-cecal artery was clamped for 1-2 minutes to form the scraping model. After this process, the incision was closed with 3/0 vicril.

Group III (n: 6): Laparotomy was performed from the middle line after 15 minutes carbon dioxide insufflation. Then petechial bleedings were performed with sterile dry sponge over the cecum, and the ileo-cecal artery was

clamped for 1-2 minutes to form the scraping model. After this process, the incision was closed with 3/0 vicril.

Group IV (n: 6): Laparotomy was performed from the middle line after 15 minutes carbon dioxide insufflation. Then petechial bleedings were performed with sterile dry sponge over the cecum, and the ileo-cecal artery was clamped for 1-2 minutes to form the scraping model. After this process, the incision was closed with 3/0 vicril. Then carbon dioxide insufflation was performed for 45 minutes.

Group V (n: 6): Laparotomy was performed from the middle line. Then petechial bleedings were performed with sterile dry sponge over the cecum, and the ileo-cecal artery was clamped for 1-2 minutes to form the scraping model. After this process, the incision was closed with 3/0 vicril. Then carbon dioxide insufflation was performed for 45 minutes.

Parameters

Tissue rat malondialdehyde (MDA) level measurement:

The quantitative measurement of the MDA was performed with the ready-made Csabio® tPA (American Diagnostica Inc.) kit according to the introductions of the producer company with the ELISA (Enzyme Linked-Immuno-Sorbent Assay) method. The values recorded were as pg/ml.

Tissue plasminogen activator inhibitor-1 (PAI-1) level measurement: To measure the amount of the Plasminogen activator inhibitor-1 quantitatively, the Cusabio® Plasma PAI-1 (American Diagnostica Inc.) ready-made kit was used according to the introductions of the producer company with the ELISA (Enzyme Linked-Immuno-Sorbent Assay) method. The values were recorded as pmol/ml.

Intraabdominal adhesion scoring: After 10 days, the rats were anesthetized with 75 mg /kg ketamine hydrochloride. An incision with the shape of a reverse-U was performed in the front wall of the abdomen to obtain a better vision, and all the adhesions in the abdomen were examined, recorded and classified (Picture 2). The Mazuji Classification was used in macroscopic classification.⁷ The formed adhesions were taken out without damaging them, and put in 10% formalin solution, and sent to the Gazi University Faculty of Medicine Pathology Department for histopathological assessment. Some tissue samples were also sent to the biochemistry laboratory for MDA and PAI level measurements. After the exemplification, the rats were sacrificed under anesthesia by taking intracardiac blood and hypotensive collapse.

Histopathological Examination

The specimens, dehydrated and embedded in paraffin, were dyed with H&E in 1 mm thickness, and assessed in

light microscope in terms of fibrosis and inflammation (Table 1,2). The fibroblast cell density (Table 1) and inflammation scoring systems (Table 2) were used in histopathological examination.⁸



Picture 2. Limited adhesion in an area thick (group 2 adhesion)

Table 1. Macroscopic adhesion evaluation scoring (Mazuji et al.)

Score	Evaluation
0	No fibrosis
1	Minimally, loose
2	Moderate
3	Fluoride,intense

Table 2. Fibrosis assessment scoring (Hooker et al.)

Score	Evaluation
0	Not inflammation
1	Giant cells, scattered mononuclear inflammatory cells (Chronic inflammation)
2	An increased number of giant cells mixed with lymphocytes, neutrophils, eosinophils plasma cells (acute inflammation)
3	Many mixed inflammatory cells, presence mikroapse (acute suppurative inflammation)

Statistical Analysis

The data were entered into the SPSS 11.0 package program. In determining the difference between the groups, the Kruskal-Wallis variance analysis was used; and in determining the groups with the differences, the Mann-Whitney U non-parametric test was used. The p<0.05 value was accepted as statistically meaningful. The averages were given as average±standard deviation.

RESULTS

A meaningful difference (p<0.05) was determined between the inflammation results of the groups. The inflammation findings become lighter as moved from Group 1 to Group 5 (Table 3). A meaningful difference (p<0.05) was determined between the fibrosis results. The fibrosis findings become lower as moved from Group 1 to Group 5 (Table 4). A meaningful difference (p<0.05) was

determined between the adhesion results of the groups. The adhesion findings become lower as moved from Group 1 to Group 5 (**Table 5**). A statistically meaningful difference was not determined ($p>0.05$) between the PAI values of the groups. A statistically meaningful difference was not determined ($p<0.05$) between the MDA values of the groups. The difference stems from Group 1 and Group 5. The MDA values of Group 1 is relatively higher than those of other groups; while the MDA values of Group 5 is found to be lower when compared with the other groups.

Table 3. Comparison of the rate of inflammation between groups

Group	Inflammation			P value
	0 n (%)	1 n (%)	2 n (%)	
1	-	-	-	6 (100)
2	-	-	6 (100)	-
3	-	4 (66.7)	2 (33.3)	-
4	-	6 (100)	-	-
5	4 (66.7)	2 (33.3)	-	-

Table 4. The comparison between groups fibrosis

Group	Fibrosis			P value
	0 n (%)	1 n (%)	2 n (%)	
1	-	1 (16.7)	5 (83.3)	
2	-	5 (83.3)	1 (16.7)	
3	-	3 (50.0)	3 (50.0)	0.0001
4	-	6 (100)	-	
5	6 (100)	-	-	

(%)*: Row percent, Kruskall Wallis Test was used.

Table 5. Comparing the adhesion between groups

Group	Adhesion				P value
	1 n (%)	2 n (%)	3 n (%)	4 n (%)	
1	-	2 (33.3)	2 (33.3)	2 (33.3)	
2	-	1 (16.7)	2 (33.3)	3 (50.0)	
3	1 (16.7)	3 (50.0)	2 (33.3)	-	0.022
4	1 (16.7)	4 (66.7)	1 (16.7)	-	
5	2 (33.3)	3 (50.0)	1 (16.7)	-	

(%)*: Row percent, Kruskall Wallis Test was used.

A statistically meaningful difference was not determined ($p>0.05$) between the PAI values of the groups. A statistically meaningful difference was not determined ($p<0.05$) between the MDA values of the groups. The difference stems from Group 1 and Group 5. While the MDA values of Group 1 are relatively higher than those of other groups, the MDA values of Group 5 are found to be lower than those of other groups. A positive linear relation in medium level was determined between the PAI and MDA values (correlation coefficient: 0,381, $p<0.05$). As the PAI values increase, so do the MDA values. As the inflammation findings become severe, so do the severeness of fibrosis findings. A positive linear relation in medium level was determined between the inflammation findings and MDA values (correlation coefficient: 0,382, $p<0.05$). As the inflammation findings become severe, the MDA values increase. A strong positive linear relation was determined between the fibrosis findings and PAI values (correlation coefficient: 0,513, $p<0.05$). As the fibrosis findings become severe, the PAI values increase. A positive linear relation in medium level was determined between the fibrosis findings and MDA values (correlation coefficient: 0,399, $p<0.05$). As the PAI values increase, so do the MDA values. As the fibrosis findings become severe, the MDA values increase. A positive linear relation in medium level was determined between the adhesion findings and PAI values (correlation coefficient: 0,324, $p<0.05$). As the PAI values increase, so do the MDA values. As the adhesion findings become severe, the PAI values increase (**Table 6**).

DISCUSSION

Peritoneal adhesions occur after the single-layered mesothelial cells that form the peritonea are harmed due to some reasons (mechanical, ischemic, chemical, infective, inflammatory, etc.).

Table 6. Inflammation, fibrosis, adhesion, PAI 1 and MDA value relationship between

	Inflammation	Fibrosis	Adhesion	PAI	MDA
Inflammation					
Correlation composition (tau)	1	0.454*	0.193	0.111	0.382*
n	30	30	30	30	30
P	.	0.005	0.238	0.433	0.007
Fibrosis					
Correlation composition (tau)	0.454*	1	0.193	0.513*	0.399*
n	30	30	30	30	30
P value	0.005	.	0.238	0.0001	0.007
Adhesion					
Correlation composition (tau)	0.294	0.193	1	0.324*	0.051
n	30	30	30	30	30
P	0.114	0.238	.	0.024	0.719

The general incidence of re-application to the hospital due to adhesion when compared with operation procedures was reported as 4.6%.^{9,10}

Various models such as damaged uterus horn model, peritonea damage model, colon anastomose model, ileal transection model, bacterial peritonitis model and clamping model were formed for experimental adhesion examination purposes. The clamping model that is applied in our study is proper for its defects being two-staged. The first stage is forming mechanical serosal intestine wall damage with direct sponge application; the second stage is forming ischemic damage after the vein is clamped. The reasons for our preference of this model is due to its similarity to surgical methods in the abdomen.

We did not determine a statistically meaningful difference among the inflammation results, the adhesion results and the fibrosis results in groups. We found that the inflammation findings became less severe as moved from Group 1 to Group 5. We found that the fibrosis findings became less severe as moved from Group 1 to Group 5. We also found that the adhesion findings became less severe as moved from Group 1 to Group 5. The findings of the study conducted by Scott-Coombes et al.¹¹ support our findings. In their studies, they determined PAI-1 in high density in peritoneal tissues of the patients with adhesion. We did not determine a statistically meaningful difference between the PAI-1 values among the groups in our study. We did not determine a statistically meaningful difference between the MDA values in groups. The difference stems from Group 1 and Group 5. The MDA values of Group 1 is higher than those of other groups, while the MDA values of Group 5 are lower than those of other groups. As the PAI-1 values increase, so do the MDA values. As the inflammation findings become severe, so do the severeness of fibrosis findings. As the inflammation findings become severe, the MDA values increase. As the fibrosis findings become severe, the PAI-1 values increase. As the fibrosis findings become severe, the MDA values increase. As the adhesion findings become severe, the PAI-1 values increase. As the adhesion findings in Group 1 become severe, the PAI-1 values increase. As the adhesion findings become severe in Group 3, the MDA values decrease. No relation was determined among the other findings and values in other groups.

The history of laparoscopy dates back to one century ago.¹² In early 1900s, endoscopy was used mainly in bladder, rectum, larynx, esophagus examinations; in 1901, a Russian Gynecologist named Dimitri Von Ott reported that he had entered to the peritoneal cavity with a minimal incision and examined the inside of the abdomen, and this process was later called as Ventroscopy by him.^{13,14}

Laparoscopy is related with a small entry and manipulation injuries; however, pneumo-peritoneum is also responsible for the decrease in acute phase reply.¹⁵ Laparoscopic surgery in abdominal explorations ensures that the blind dissection and retractors and compresses are used less. Moreover, when compared with traditional surgery, it decreases the washing and drying of peritonea, and contact of operational objects, tissue damage, and bleeding in operational area.^{16,17} For these reasons, it is possible to suggest that laparoscopy causes to less adhesion. Carbon dioxide is the most frequently used gas in today's world.¹⁸ The advantage of carbon dioxide is that it is not inflammable and explosive; its diffusion to the tissues is fast, and it is relatively cheaper. Gas embolism is the most important complication of the gasses that are used for pneumo-peritoneum. A gas such as carbon dioxide that is absorbed fast causes less persistent airway obstructions in right ventricular outlet. The most important disadvantage of carbon dioxide is that it dissolves in solutions quickly; and its causing to unwanted biological effects such as hypercapnia and acidosis less.¹⁹ However, these effects are in minimal level in people who do not have further stage respiration problems and cardiac pathology. For these reasons, carbon dioxide is the most frequently preferred gas today. CO₂ is known to change the systemic immune reply in a proper way during laparoscopic surgery of the pneumo-peritoneum. It has been proven that CO₂ insufflations cause to local peritoneal acidosis. CO₂ is a strong anti-inflammatory agent.²⁰⁻²² It has been proven that CO₂ decreases the systemic inflammatory reply and capno-peritoneum macrophage function.^{22,23} In case the peritoneal pH decreases to 6,1 after the CO₂ insufflation, the macrophage intracellular pH values also decrease.²⁴ It has been shown that superoxide production as well as phagocyte and cytokines are suppressed with the decrease in intracellular pH.^{25,26} The decreasing pro-inflammatory cytokine production after the laparoscopy in peritoneal tissues may be responsible for the decrease in postoperative pain and adhesion formation.²⁵⁻²⁷

Adhesion formation is a complicated process. Macrophages, fibroblasts and mesothelial cells have major roles in the stages of adhesion formation. The studies conducted so far have shown that there is a dynamic balance between the pro- and anti-inflammatory cytokines in these stages of adhesion formation.^{28,29} Surgery, inflammation and other similar factors cause to the collapse of this balance. It is known that when to defective sides come together, intraperitoneal adhesion formation is triggered. Miyano et al.³⁰ and Jansen et al.³¹ reported that CO₂ insufflation and the mechanical effect of the pneumo-peritoneum decrease the contact between the defective sides. They determined that PAI-1 concentrations increased in the laparoscopic group as

a secondary process to the CO₂ insufflation.^{32,33} In our study, as the severeness of the inflammation and fibrosis increased, so did the PAI-1 concentration.

Miyano et al.³⁰ reported that CO₂ insufflation decreased the contact between the damaged surfaces. It was suggested that this was due to the increase in the pressure after the pneumo-peritoneum duration and insufflation pressure causing to an increase in adhesion in hypoxia.^{33,34}

Corona et al.³⁵ conducted a study and reported that adhesion and inflammation increased in CO₂ models. Unlike this study, it was determined in our study that capno-peritoneum decreased the adhesion and inflammation especially in the long-term group.

We did not observe a clinical side effect or histopathological abnormality due to CO₂ in our study and this result supports the findings of Miyano et al.³⁰ Bustorff-Silva et al.³⁶

CONCLUSION

Our results suggest that CO₂ pneumo-peritoneum has positive effects in postoperative intraperitoneal adhesion development. Since we formed a scraping model in our study, we cannot suggest that the adhesion formation is decreased with mechanical effect. The patho-physiological and molecular bases of the postoperative adhesion formation have been documented and described well. However, we consider that the capno-peritoneum and postoperative adhesion formation is prevented with anti-inflammatory effect. We need to conduct more studies to examine this mechanism.

ETHICAL DECLARATIONS

Ethics Committee Approval: The study was carried out with the permission of Gazi University Animal Experiments Local Ethics Committee (Date: 04.09.2011, Decision No: B.30.2.GÜN.O.EU.00.00/38-5869.30).

Informed Consent: This experimental animal study does not require informed consent.

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REFERENCES

1. Menzies D. Postoperative adhesions: their treatment and relevance in clinical practice. *Ann R Coll Surg Engl.* 1993;75(3):147-153.
2. ten Broek RP, Issa Y, van Santbrink EJ, et al. Burden of adhesions in abdominal and pelvic surgery: systematic review and meta-analysis. *BMJ.* 2013;347:f5588.
3. Hellebrekers BW, Trimbos-Kemper TC, Trimbos JB, Emeis JJ, Kooistra T. Use of fibrinolytic agents in the prevention of postoperative adhesion formation. *Fertil Steril.* 2000;74(2):203-212. doi:10.1016/s0015-0282(00)00656-7
4. Günay C, Sağlıyan A, Yaman İ. Ratlarda deneysel olarak oluşturulan intraabdominal adezyonların önlenmesinde aprotinin ile metilen mavisinin etkinliğinin karşılaştırılması. *FÜ Sağlık Bil Derg.* 2005;19(1):51-55.
5. Reed KL, Stucchi AF, Becker JM. The peritoneal fibrinolytic response to conventional and prolonged surgery is similar. *J Surg Res.* 2009;152(2):175-177. doi:10.1016/j.jss.2008.04.042
6. Rahimi VB, Shirazinia R, Fereydouni N, et al. Comparison of honey and dextrose solution on post-operative peritoneal adhesion in rat model. *Biomed Pharmacother.* 2017;92:849-855. doi:10.1016/j.bioph.2017.05.114
7. Holmdahl L, Eriksson E, Eriksson BI, Risberg B. Depression of peritoneal fibrinolysis during operation is a local response to trauma. *Surgery.* 1998;123(5):539-544. doi:10.1067/msy.1998.86984
8. Raftery AT. Effect of peritoneal trauma on peritoneal fibrinolytic activity and intraperitoneal adhesion formation. An experimental study in the rat. *Eur Surg Res.* 1981;13(6):397-401. doi:10.1159/000128208
9. Schnüriger B, Barmparas G, Branco BC, Lustenberger T, Inaba K, Demetriades D. Prevention of postoperative peritoneal adhesions: a review of the literature. *Am J Surg.* 2011;201(1):111-121. doi:10.1016/j.amjsurg.2010.02.008
10. Raisi A, Dezfoulian O, Davoodi F, Taheri S, Ghahremani SA. Salvia miltiorrhiza hydroalcoholic extract inhibits postoperative peritoneal adhesions in rats. *BMC Complement Med Ther.* 2021;21(1):126. doi:10.1186/s12906-021-03300-7
11. Scott-Coombes D, Whawell S, Vipond MN, Thompson J. Human intraperitoneal fibrinolytic response to elective surgery. *Br J Surg.* 1995;82(3):414-417. doi:10.1002/bjs.1800820346
12. Poerwosusanta H, Gunadi, Noor Z, et al. The effect of laparoscopy on mast cell degranulation and mesothelium thickness in rats. *BMC Surg.* 2020;20(1):111. doi:10.1186/s12893-020-00775-y
13. Lau WY, Leow CK, Li AK. History of endoscopic and laparoscopic surgery. *World J Surg.* 1997;21(4):444-453. doi:10.1007/pl00012268
14. Valdivieso E, Saenz R, Claudio N. Natural orifice transluminal endoscopic surgery: putting together minimally invasive techniques for a new era. *Gastrointest Endosc.* 2007;66(2):340-342. doi:10.1016/j.gie.2007.03.1039
15. Are C, Talamini MA, Murata K, De Maio A. Carbon dioxide pneumoperitoneum alters acute-phase response induced by lipopolysaccharide. *Surg Endosc.* 2002;16(10):1464-1467. doi:10.1007/s00464-001-8305-5
16. Ellis H, Moran BJ, Thompson JN, et al. Adhesion-related hospital readmissions after abdominal and pelvic surgery: a retrospective cohort study. *Lancet.* 1999;353(9163):1476-1480. doi:10.1016/S0140-6736(98)09337-4
17. Nagelschmidt M, Gerbecks D, Minor T. The impact of gas laparoscopy on abdominal plasminogen activator activity. *Surg Endosc.* 2001;15(6):585-588. doi:10.1007/s004640010282
18. Brokelman WJ, Lensvelt M, Borel Rinkes IH, Klinkenbijl JH, Reijnen MM. Peritoneal changes due to laparoscopic surgery. *Surg Endosc.* 2011;25(1):1-9. doi:10.1007/s00464-010-1139-2
19. Orhurhu VJ, Gao CC, Ku C. Carbon Dioxide Embolism. In: StatPearls. Treasure Island (FL): StatPearls Publishing; November 28, 2022.
20. Fuentes JM, Hanly EJ, Aurora AR, et al. CO₂ abdominal insufflation pretreatment increases survival after a lipopolysaccharide-contaminated laparotomy. *J Gastrointest Surg.* 2006;10(1):32-38. doi:10.1016/j.gassur.2005.07.031

21. Hanly EJ, Mendoza-Sagaon M, Murata K, Hardacre JM, De Maio A, Talamini MA. CO₂ Pneumoperitoneum modifies the inflammatory response to sepsis. *Ann Surg.* 2003;237(3):343-350. doi:10.1097/01.SLA.0000055271.58945.E2
22. Hanly EJ, Aurora AR, Fuentes JM, et al. Abdominal insufflation with CO₂ causes peritoneal acidosis independent of systemic pH. *J Gastrointest Surg.* 2005;9(9):1245-1252. doi:10.1016/j.jgassur.2005.09.007
23. O'Boyle CJ, deBeaux AC, Watson DI, et al. Helium vs carbon dioxide gas insufflation with or without saline lavage during laparoscopy. *Surg Endosc.* 2002;16(4):620-625. doi:10.1007/s00464-001-8218-3
24. West MA, Hackam DJ, Baker J, Rodriguez JL, Bellingham J, Rotstein OD. Mechanism of decreased in vitro murine macrophage cytokine release after exposure to carbon dioxide: relevance to laparoscopic surgery. *Ann Surg.* 1997;226(2):179-190. doi:10.1097/00000658-199708000-00010
25. Kopernik G, Avinoach E, Grossman Y, et al. The effect of a high partial pressure of carbon dioxide environment on metabolism and immune functions of human peritoneal cells-relevance to carbon dioxide pneumoperitoneum. *Am J Obstet Gynecol.* 1998;179(6 Pt 1):1503-1510. doi:10.1016/s0002-9378(98)70016-x
26. Douvdevani A, Rapoport J, Konforty A, Yulzari R, Moran A, Chaimovitz C. Intracellular acidification mediates the inhibitory effect of peritoneal dialysate on peritoneal macrophages. *J Am Soc Nephrol.* 1995;6(2):207-213. doi:10.1681/ASN.V62207
27. Guzmán-Valdivia Gómez G, Tena-Betancourt E, Angulo Trejo M. Different doses of enoxaparin in the prevention of postoperative abdominal adhesions. experimental study. *Ann Med Surg (Lond).* 2021;73:103132. doi:10.1016/j.amsu.2021.103132
28. Menger MD, Vollmar B. Surgical trauma: hyperinflammation versus immunosuppression? *Langenbecks Arch Surg.* 2004;389(6): 475-484. doi:10.1007/s00423-004-0472-0
29. Romeo C, Crucetti A, Turiaco A, et al. Monocyte and neutrophil activity after minor surgical stress. *J Pediatr Surg.* 2002;37(5):741-744. doi:10.1053/jpsu.2002.32268
30. Miyano G, Yamataka A, Doi T, et al. Carbon dioxide pneumoperitoneum prevents intraperitoneal adhesions after laparotomy in rats. *J Pediatr Surg.* 2006;41(5):1025-1028. doi:10.1016/j.jpedsurg.2005.12.048
31. Soltany S. Postoperative peritoneal adhesion: an update on physiopathology and novel traditional herbal and modern medical therapeutics. *Naunyn Schmiedebergs Arch Pharmacol.* 2021;394(2):317-336. doi:10.1007/s00210-020-01961-8
32. Cheong YC, Laird SM, Li TC, Shelton JB, Ledger WL, Cooke ID. Peritoneal healing and adhesion formation/reformation. *Hum Reprod Update.* 2001;7(6):556-566. doi:10.1093/humupd/7.6.556
33. Bryant LR. an evaluation of the effect of fibrinolysis on intraperitoneal adhesion formation. *Am J Surg.* 1963;106:892-897. doi:10.1016/0002-9610(63)90152-1
34. Moris D, Chakedis J, Rahnemai-Azar AA, et al. Postoperative abdominal adhesions: clinical significance and advances in prevention and management. *J Gastrointest Surg.* 2017;21(10):1713-1722. doi:10.1007/s11605-017-3488-9
35. Corona R, Verguts J, Schonman R, Bindu MM, Mailova K, Koninkx PR. Postoperative inflammation in the abdominal cavity increases adhesion formation in a laparoscopic mouse model. *Fertil Steril.* 2011;95(4):1224-1228. doi:10.1016/j.fertnstert.2011.01.004
36. Dixon CT, Rixford EL. Cytologic response to peritoneal irritation in man: a protective mechanism. *Am J Surg.* 1934;25(3):504-505.