

Karyotype analysis of Common Cocklebur (*Xanthium strumarium* L.)

Gulden Dogan^{1*} Yasar Kiran¹

Abstract

Objective: The purpose of this study is to determine the mitotic chromosome number, morphometric parameters, and karyotypes of *Xanthium strumarium* and make contributions to other multidisciplinary studies on the genus.

Material and Methods: The seeds were germinated on moist filter paper in Petri dishes at 25°C. Actively growing root tips were pre-treated with aqueous paradichlorobenzene for 4h at room temperature. Then, the root tips were fixed with acetic alcohol (1:3 glacial acetic acid–absolute ethanol) for at least 24 h at 4°C, hydrolysed in 1 N HCl at 60°C for 3 min, then rinsed in tap water for 3–5 min. Finally, they were stained in Feulgen for 1 h and mounted in 45% acetic acid. Digital microphotographs from at least five well-spread metaphase plates were taken using an Olympus BX51 microscope and were recorded with an Olympus Camedia C-4000 digital camera.

Results: The chromosome number is determined as $2n = 36$ for this taxon. The karyotype consists of 16 median region (m) and 2 submedian region (sm) chromosomes. The metaphase chromosome length ranges from 2.30 to 4.03 μm , longest to shortest chromosome ratio is 1.7:1.1, total karyotype length (TKL) 54.76 μm and the karyotype symmetry is type 1A.

Conclusion: The results of this study showed that the chromosome number of *Xanthium strumarium* is $2n=36$. Satellites were not observed in the karyotype of this species. Identifying the chromosome number of this species in this study provides a base for biosystematic studies.

Keywords: Chromosome number, *Xanthium*, Karyotype Analysis.

Introduction

The genus *Xanthium* L. belongs to the family Asteraceae. The members of this genus are distributed globally, however are most frequently in to be found tropical and sub-tropical regions (1). They are widespread in America, Canada, Mexico, Malaysia, Indonesia and India. The taxonomic study of the members of this genus is difficult and confusing. For example, Caius reported that the genus *Xanthium* includes 25 species (2). However, according to Weaver and Lechowich, there are 20 species belonging to this genus. Thus confusion about the species number of this genus still exists (1). Love and Dansureau revised the genus *Xanthium* and reduced the number of species to only 2 (*X. strumarium* L. and *X. spinosum* L.). They mentioned that due to phenotypic plasticity in a number of features, the members showed different phenotypes and creating confusion regarding their taxonomic rank (3). Prain described that the 2 species *i.e.* *X. strumarium* and *X. spinosum* were found in undivided Bengal (4).

However, Oudhia and Dixit (1994) reported *X. indicum* and *X. strumarium* from India (5). In Turkey, only two species, namely *X. strumarium* and *X. spinosum* have so far been reported. *X. strumarium* species found in Turkey has two subspecies (subsp. *strumarium* and *cavanillesii*) (6).

The common names are cocklebur, burr, sheep burr, etc. (7-10). The plant is an annual 30-120 cm in height and is a short-day plant that flowers in July-August. Each cocklebur bur contains two seeds. The seeds are covered by a hard green husk with hooked spines (11). The whole plant (*X. strumarium*), especially its leaf, root and fruit, has been used in traditional medicine for the treatment of rhinitis, malaria, rheumatism, tuberculosis, cancer, and ulcers (12-15). Previous studies indicated that plants of the Asteraceae family are characteristically rich in sesquiterpene lactones, an important class of terpenoids, and the *Xanthium*'s species are rich in such medicinal ingredients.

Received: 04-08-2017 Accepted 22-08-2017 Available Online: 30-09-2017

1 Firat University, Science Faculty, Biology Department, Elazig, Turkey

* Corresponding Author: Gulden Dogan, E-mail: gdogan@firat.edu.tr Phone: +904242370000 - 3771



The pharmacological properties of *X. strumarium* are largely attributed to the presence of xanthanolides (a class of sesquiterpene lactones), which have been reported to possess antifungal, antibacterial, and cytotoxic activities, and exhibit a growth inhibitory activity against insects (16-22).

The plant is used in classical homeopathy (23) and is officially recognized in China and several other countries. The preparation Adenostop is manufactured from cocklebur in Romania and is used to treat prostate adenoma. High anticancer activity of cocklebur (for breast, lung, stomach, and colon cancer) was recently reported (24). This plant is used as a medicine for curing nasal sinusitis, headache, urticaria and arthritis. It has also been reported to possess curative effects against chronic bronchitis, chronic rhinitis, allergic rhinitis, lumbago and other ailments (25) and is used by various native American tribes to relieve constipation, diarrhea and vomiting (26).

The purpose of this study is to determine the mitotic chromosome number, morphometric parameters, and karyotypes of *Xanthium strumarium* and make contributions to other multidisciplinary studies on the genus.

Material and Methods

Plant material was collected from natural habitats during the fruiting season in Elazig in 2015. Voucher specimen was deposited at the Firat University Herbarium (FUH). Karyological studies were conducted on meristematic cells obtained from the root tips.

The seeds were germinated on moist filter paper in Petri dishes at 25°C. Actively growing root tips were pre-treated with aqueous paradichlorobenzene for 4h at room temperature. Then, the root tips were fixed with acetic alcohol (1:3 glacial acetic acid-absolute ethanol) for at least 24 h at 4°C, hydrolysed in 1 N HCl at 60°C for 3 min, then rinsed in tap water for 3–5 min.

Finally, they were stained in Feulgen for 1 h and mounted in 45% acetic acid. Digital microphotographs from at least five well-spread metaphase plates were taken using an Olympus BX51 microscope (Olympus Optical Co. Ltd., Tokyo, Japan), and were recorded with an Olympus Camedia C-4000 digital camera (Olympus Optical Co. Ltd., Tokyo, Japan). The short arm (s), long arm (l) and total lengths (tl) of each chromosome were measured and the relative lengths, arm ratios, and centromeric indices were determined from images of selected cells. Chromosomes were classified according to the nomenclature of Levan et al. (27). The intra-chromosomal asymmetry index (A1) and the inter-chromosomal asymmetry index (A2) followed those of Romero-Zarco (28). The karyotype symmetry nomenclature followed Stebbins (29). Also, relevant literature the online chromosome number databases, Index to Plant Chromosome Numbers (IPCN) (30) and Index to Chromosome Numbers in Asteraceae (31) were checked.

Results and Discussion

The results of this study showed that the chromosome number of *Xanthium strumarium* is $2n=36$. Karyotype analysis of this species to reveal the many values were calculated. The number of somatic chromosome, ploidy level, karyotype formula, chromosome length range, total karyotype length (TKL), Stebbins C and asymmetry indexes (A1, A2) are presented in Table 1; relative length, arm ration, centromeric index, type, in Table 2.

Haploid ideograms of *X. strumarium* has been shown in Fig. 1 and metaphase chromosomes in Fig. 2. The chromosome number is determined as $2n = 36$ for this taxon. The karyotype consists of 16 median region (m) and 2 submedian region (sm) chromosomes. The metaphase chromosome length ranges from 2.30 to 4.03 μm , longest to shortest chromosome ratio is 1.7:1.1, total karyotype length (TKL) 54.76 μm and the karyotype symmetry is type 1A. Satellites were not observed in the karyotype of this species.

Table 1. Somatic chromosome number, ploidy level, karyotype formula, chromosome length range, total karyotype length (TKL), asymmetry indexes (A1, A2) of Romero Zarco (1986) and symmetry classes (SC) of Stebbins (1971) of *Xanthium strumarium*.

| Taxon | 2n | Ploidy level | Karyotype formula | Chromosome length range (μm) | TKL (μm) | A1 | A2 | SC |
|-----------------------------------|----|--------------|-------------------|---|-----------------------|------|------|----|
| <i>Xanthium strumarium</i> | 36 | 4x | 16m+2sm | 2.30-4.03 | 54.76 | 0.21 | 0.15 | 1A |

Table 2. Karyomorphological parameters of *Xanthium strumarium*

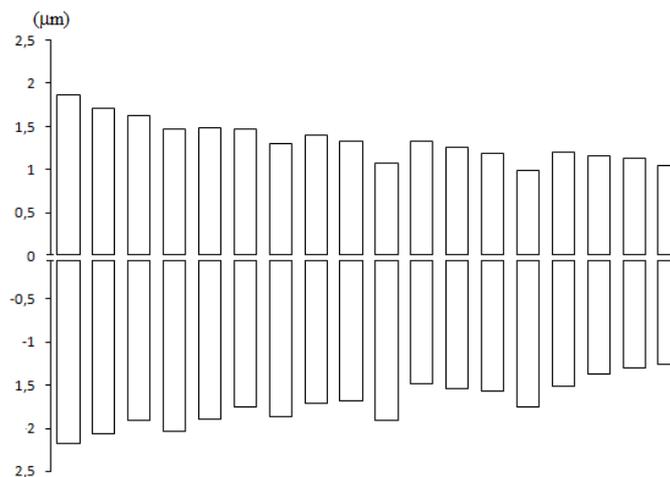
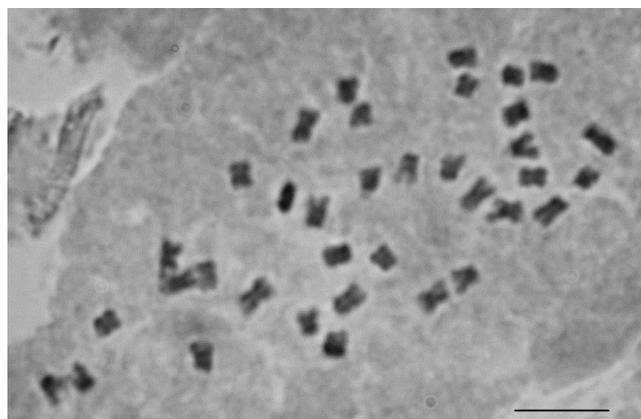
| <i>Xanthium strumarium</i> | | | | |
|----------------------------|-----------------|------------|-------------------|------|
| Pair No | Relative Length | Arm Ration | Centromeric Index | Type |
| 1 | 7.36 | 1.16 | 0.46 | m |
| 2 | 6.89 | 1.20 | 0.45 | m |
| 3 | 6.43 | 1.17 | 0.45 | m |
| 4 | 6.40 | 1.38 | 0.41 | m |
| 5 | 6.16 | 1.26 | 0.44 | m |
| 6 | 5.88 | 1.19 | 0.45 | m |
| 7 | 5.77 | 1.44 | 0.40 | m |
| 8 | 5.67 | 1.23 | 0.44 | m |
| 9 | 5.49 | 1.25 | 0.44 | m |
| 10 | 5.43 | 1.78 | 0.35 | sm |
| 11 | 5.15 | 1.11 | 0.47 | m |
| 12 | 5.11 | 1.22 | 0.44 | m |
| 13 | 5.01 | 1.32 | 0.43 | m |
| 14 | 4.99 | 1.78 | 0.35 | sm |
| 15 | 4.94 | 1.25 | 0.44 | m |
| 16 | 4.62 | 1.18 | 0.45 | m |
| 17 | 4.42 | 1.15 | 0.46 | m |
| 18 | 4.21 | 1.20 | 0.45 | m |

Meiotic chromosome number is $n=18$ this species was reported in the literature (32-36). Besides, in de book of "Flora der Schweiz und angrenzender Gebiete" reported that chromosome number of *X. strumarium* $n=18$.

$2n=36$ chromosomes of this species was reported in the literature (37-54). Therefore, the present count confirmed the earlier reports on $2n$ chromosomes number.

Conclusion

According to our knowledge, chromosome number and morphology report for *Xanthium strumarium* does not exist. Identifying the chromosome number of this species in this study provides a base for biosystematic studies.

**Figure 1.** Haploid idiograms of *Xanthium strumarium*.**Figure 2.** Metaphase chromosomes of *Xanthium strumarium*. Scale bar=10 µm

Conflict of Interest: The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Author's Contributions: GD, YK: Collecting of data, writing and revision of article,

Ethical issues: All Authors declare that Originality of research/article etc... and ethical approval of research, and responsibilities of research against local ethics commission are under the Authors responsibilities. The study was conducted due to defined rules by the Local Ethics Commission guidelines and audits.

References

- Weaver SE, Lechowich MJ. The biology of Canadian weeds. 56. *Xanthium strumarium* L. *J Plant Sci.* 1982; 63:211-225.
- Caius JF. Medicinal and poisonous plants of India. *Pbl Sci Publ Jodhpur India.* 1986; 375-376.
- Love D, Dansureau P. Biosystematic studies on *Xanthium*: taxonomic appraisal and ecological status. *Can J Bot.* 1959; 37:173-208.
- Prain D. Bengal plants-II, Botanical Survey of India, Calcutta. Bishen Singh Mahendra Pal Singh, Dehradun, 1903; 607-608.
- Oudhia P, Dixit A. Weeds in Ambimkapur region (Madhya Pradesh) and their traditional use. *Weed News.* 1994; 1:19-21.
- Davis PH. Flora of Turkey and the East Aegean Island, Vol. 5. Edinburgh University Press: Edinburgh; 1975.
- Flora of the Ussr [In Russian]. Vol. 25. Izd. Akad. Nauk SSSR, Moscow, Leningrad. 1959; 521.
- Plant resources of Russia, Wild Flowering Plants, Their Quantitative Composition and Biological Activity [in Russian]. Vol. 5. Part 2. St. Petersburg, Moscow. 2013; 139-141.
- Makhlayuk VP. Medicinal Plants in Folk Medicine [in Russian], Povolzhskoe Kn. Izd. Saratov. 1991; 131-132.
- Ladynina EA. Wisdom of Herbs, Herbal Cures and Homeopathy [in Russian], AIF Print, Moscow. 2003; 216-217.
- Shrivastava KS, Krisnamurthy RS, Haksar CN. *J Sci Ind Res Sect B.* 1957; 16: 427.
- Chandel S, Bagai U, Vashishat N. Antiplasmodial activity of *Xanthium strumarium* against *Plasmodium berghei*-infected BALB/c mice. *Parasitol Res.* 2012; 110(3):1179-83.
- Gautam R, Saklani A, Jachak SM. Indian medicinal plants as a source of antimycobacterial agents. *J Ethnopharmacol.* 2007; 110(2):200-34.
- Ma YT, Huang MC, Hsu FL, Chang HF. Thiazinedione from *Xanthium strumarium*. *Phytochemistry.* 1998; 48(6):1083-1085.
- Yin MH, Kang DG, Choi DH, Kwon TO, Lee HS. Screening of vasorelaxant activity of some medicinal plants used in oriental medicines. *J of Ethnopharmacol.* 2005; 99(1):113-117.
- Rodriguez E, Towers G, Mitchell J. Biological activities of sesquiterpene lactones. *Phytochemistry.* 1976; 15(11):1573-1580.
- Harada A, Sakata K, Ina H, Ina K. Isolation and identification of xanthatin as an anti-attaching repellent against blue mussel. *Agric Biol Chem.* 1985; 49(6):1887-1888.
- Kawazu K, Nakajima S, Ariwa M. Xanthumin and 8-epi-xanthatin as insect development inhibitors from *Xanthium canadense* Mill. *Experientia.* 1979; 35(10):1294-1295.
- Kim H, Lee I, Yeo S, Seong L, Yu T. Isolation and characterization of antitumor agents from *Xanthium strumarium* L. *Korean J Biotechnol Bioeng.* 2003; 18:324-328.
- Lavault M, Landreau A, Larcher G, Bouchara JP, Pagniez F, Le P, et al. Antileishmanial and antifungal activities of xanthanolides isolated from *Xanthium macrocarpum*. *Fitoterapia.* 2005; 76(3):363-366.
- Roussakis C, Chinou I, Vayas C, Harvala C, Verbist J. Cytotoxic activity of xanthatin and the crude extracts of *Xanthium strumarium*. *Planta Med.* 1994; 60(5):473.
- Sato Y, Oketani H, Yamada T, Singyouchi KI, Ohtsubo T, Kihara M, et al. A Xanthanolide with potent antibacterial activity against Methicillin-resistant *Staphylococcus aureus*. *J Pharm Pharmacol.* 1997; 49(10):1042-1044.
- Ladynina EA. Wisdom of Herbs, Herbal Cures and Homeopathy [in Russian], AIF Print, Moscow. 2003; 216-217.
- Klimakhin GI, Fonin VS, Maslyakov VY, Fadeev NB, Semikin VV, Pelgunova LA. Biochemical features of common cocklebur (*Xanthium strumarium* L.) *Pharmaceutical Chemistry J.* 2015; 49(8):32-35.
- Zhu YP. Chinese Materia Medica: Chemistry, Pharmacology and Applications. Harwood Academic Publishers: Amsterdam; 1998.
- Kamboj A, Saluja AK. Phytopharmacological review of *Xanthium strumarium* L. (Cocklebur). *Int J of Green Pharmacy.* 2010; 4:129-139.
- Levan A, Fredga K, Sanberg AA. Nomenclature for centromeric position on chromosomes. *Hereditas.* 1964; 52(2):201-220.
- Romero-Zarco C. A new method for estimating karyotype asymmetry. *Taxon.* 1986; 35: 526-530.
- Stebbins GL. Chromosomal evolution in higher plants. London, UK: Edward Arnold (Publishers) Ltd; 1971.
- <http://www.tropicos.org/Project/IPCN>
- http://www.lib.kobe-u.ac.jp/infolib/meta_pub/G0000003
- Sarker AK, Datta N, Chatterjee U, Hazra D. In Love, A. (ed.): IOPB Chromosome number reports. LXXVI. *Taxon.* 1982; 31(3):574-598.

33. Razaq ZA, Vahidy AA, Ali DH. Chromosome numbers in Compositae from Pakistan. *Ann Missouri Bot Gard.* 1994; 81(4):800-808.
34. Bala S, Gupta G. In K. Marhold (ed.): IAPT/IOPB Chromosome data 12. *Taxon.* 2011; 60(6):1784-1796.
35. Bala S, Gupta G. Male meiosis and chromosome number in Asteraceae family from district Kangra of H.P. (Western Himalayas). *Int J Bot Res.* 2013; 3(1):43-58.
36. Singhal VK, Garg P, Kumar P. Cytological studies of some dicots from the hills of Mandi district (Himachal Pradesh) in Northwest Indian Himalayas. *Cytologia.* 2013; 78(1):1-14.
37. Ishikawa M. A list of the number of chromosomes. *Bot Mag Tokyo (J Plant Res.)* 1916; 30(360): 404.
38. Skalinska M, Malecka J, Izmailow R. et al. Further studies in chromosome numbers of polish angiosperms X. *Acta Biol Cracov Ser Bot.* 1974; 17:133-164.
39. Rostoutseva TS. Chromosome numbers of some species of the family Asteraceae Dumort. *Bot Zhurn. (Moscow and Leningrad)* 1979; 64(4):582-589.
40. Sidhu MK. Distributional and cytological studies of the weed flora of cultivable field of Patiala district (Panjab). Ph. D. Thesis, Punjabi University: Patiala -India; 1979.
41. Bir SS, Sidhu M. Cyto-palynological studies on weed flora of cultivable lands of Patiala district (Punjab). *J Palynology.* 1980; 16:85-105.
42. Love A, Love D. In Love, A.(ed.): IOPB Chromosome number reports. LXXV. *Taxon.* 1982; 31(2): 342-368.
43. Lomonosova MN. In K. Marhold and I. Breitwieser (eds.): IAPT/IOPB Chromosome data 16. *Taxon.* 2013; 62(6):1357.
44. Bakale VL, Srinivasu T. Mitotic and karyological studies in *Xanthium strumarium* L. *Adv Plant Sci.* 1988; 1:201-207.
45. Mathew A, Mathew PH. Cytological studies on the south Indian Compositae. *Glimpses Plant Res.* 1998; 8:1-177.
46. Probatova NS, Sokolovskaia AP. Chromosome numbers in some representatives of the families Asclepiadaceae, Asteraceae, Boraginaceae, Chenopodiaceae, Lamiaceae, Oleaceae, Onagraceae, Scrophularoaceae, Solanaceae, Urticaceae from the Soviet Far East. (in Russian). *Bot. Zhurn. (Moscow and Leningrad).* 1990; 75(11):1619-1622.
47. Baltisbarger M. Chromosome numbers in some species from Greece. *Bot Chron (Patras).* 1994; 11:15-29.
48. Jose JC, Mathew PM. Chromosome numbers in the south Indian Heliantheae (Compositae). *Compositae Newslett.* 1995; 27:7-10.
49. Badr A, Kamel EA, Garcia-Jacas N. Chromosome studies in the Egyptian Flora. IV. Karyotype features of some species in subfamily Asteroideae (Asteraceae). *Compositae Newslett.* 1997; 30:15-28.
50. Bennett MD, Leitch IJ, Hanson L. DNA amounts in two samples of Angiosperm weeds. *Ann Bot.* 1998; 82 (Suppl. A):121-134.
51. Joshi KK, Joshi SD. Genetic heritage of medicinal and aromatic plants of Nepal Himalayas, Buddha Academic Publishers and Distributors Pvt. Ltd, Kathmandu: Nepal; 2001.
52. Alam SS, Sukur, MB, Zaman, MY. Karyotype analysis in 2 morphological forms of *Xanthium strumarium* L. *Cytologia.* 2011; 76(4):483-488.
53. Gupta RC, Kataria V, Mehra A. Cytomorphological studies in some gamopetalous species of Western Himalaya: An attempt to add new or varied cytotypes. *Chromosome Botany.* 2012; 7(2):59-65.
54. Chepinoga VV, Gnutikov AA, Lubogoschinsky PI, Isaikina MM, Kononov AS. In Marhold, K.(ed.):IAPT/IOPB Chromosome data 13. *Taxon.* 2012; 61(4):891.