



Seroprevalence and risk factors for *Toxoplasma gondii* infection in shelter cats in Erzurum province of Türkiye

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Geliş Tarihi / Received: 04.10.2023, Kabul Tarihi / Accepted: 06.12.2023

Abstract: This study was aimed to evaluate the seroprevalence and risk factors of the infection of *Toxoplasma gondii* in cats kept in Animal Shelter and Rehabilitation Center of Erzurum Municipality in Erzurum that collected from stray. Total 70 blood samples were collected from apparently healthy cats and diseased cats such as diarrhea, and ocular and nasal discharge. Sera samples were stored in screw-capped sterile tubes at -20 °C until analysis. In sera samples, IgG antibodies against *T. gondii* were analyzed by Sabin-Feldman Dye test. Of 70 cats, 25 (35.7%) was negative and 45 (64.3%) positive for *T. gondii* infection. An important association was not found between young and adult, female and male, and apparently healthy and diseased cats with *T. gondii* seropositivity. In this study, *T. gondii* seroprevalence was determined at high rates in cats kept in shelter in Erzurum province of Turkey. The prevention measures against *T. gondii* infection in cats should be implemented for animal and public health concerns.

Keywords: *Toxoplasma gondii*, risk factors, seroprevalence, cat

Türkiye'nin Erzurum ilinde barınak kedilerinde *Toxoplasma gondii* enfeksiyonu için seroprevalans ve risk faktörleri

Özet: Bu çalışmada Erzurum ilinde sokaktan toplanan Erzurum Büyükşehir Belediye'sine ait Hayvan Bakımevi ve Rehabilitasyon Merkezi'nde bakılan kedilerde *Toxoplasma gondii* enfeksiyonunun seroprevalansı ve risk faktörlerinin araştırılması amaçlandı. İshal ve göz ve burun akıntısı gibi hastalıklı kediler ve görünüşte sağlıklı kedilerden toplam 70 kan örneği toplandı. Serum örnekleri vida kapaklı steril tüplerde analize kadar -20 °C'de saklandı. Serum örneklerinde *T. gondii*'ye karşı IgG antikorları Sabin-Feldman Dye testi ile analiz edildi. *T. gondii* enfeksiyonu için 70 kedinin 25'i (%35,7) negatif ve 45'i (%64,3) pozitif idi. *T. gondii* seropozitifliğiyle genç ve erişkin, dişi ve erkek ve görünüşte sağlıklı ve hasta kediler arasında önemli bir ilişki belirlenmedi. Bu çalışmada Türkiye'nin Erzurum ilinde barınakta bakılan kedilerde *T. gondii* seroprevalansı yüksek oranlarda belirlendi. Kedilerde *T. gondii* enfeksiyonuna karşı korunma önlemlerinin hayvan ve halk sağlığı sorunları bakımından uygulanması gerekmektedir.

Anahtar kelimeler: *Toxoplasma gondii*, risk faktörleri, seroprevalans, kedi

Introduction

Toxoplasmosis is a protozoan disease occurring in animals and humans all over the world. *Toxoplasma (T.) gondii* is a coccidian parasite that can infect almost all animals. Oocysts are shed in cat feces. Oocysts are sporulated 1-5 days after they are shed with feces (Datz and Evason 2020). They can survive in the environment for months or even years. The soil contamination with oocysts indicates ongoing contamination (Awobode et al., 2020).

Toxoplasmosis is widespread throughout the world, is more common in temperate and tropical climates and less common in dry and cold climates

(Datz and Evason 2020). Cats infected with *T. gondii* are likely to have lifelong tissue cysts. Therefore, the presence of serum antibodies is likely to indicate the current infection. Since the risk of exposure increases over time, the increase in seroprevalence is correlated with advancing age (Lappin 2014).

Common clinical findings are anorexia, weakness, fever, respiratory distress, diarrhea, gastritis and neurological disorders (Lappin 2014; Datz and Evason 2020). In addition, lethal toxoplasmosis occurs when the liver, lung, central nervous system and pancreatic tissues are affected (Lappin 2014).

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Toxoplasmosis in humans is caused by exposure to the contaminated meat or infected cat feces (Dijurkovic-Djakovic et al., 2019), as well as by drinking contaminated water, eating fruits and vegetables contaminated with spore-forming oocysts (Dubey 2009). Especially, the feces of infected cats are an important source of contamination (Montazeri et al., 2020). After transplacental infection in humans, it poses a significant risk in the fetuses and immunocompromised patients. Most people become infected by eating spore-forming oocysts or tissue cysts or transplacentally (Lappin 2014).

T. gondii seroprevalence was reported to be 5.6%-6.7% in shelter cats in different years in Japan (Oi et al., 2015), 64.48% in stray cats in Iran (Tehrani-Sharif et al., 2015), 19.2% in shelter cats in Scotland (Bennett et al., 2011), 44.1% in cats with free-roaming and indoor in Kars, Turkey (Erkılıç et al., 2016), 41.0% in pet cats in Norway (Saevik et al., 2015), 52.8% in stray cats in Ankara, Turkey (Özkan et al., 2008), 76.4% in stray cats in Niğde (Karatepe et al., 2008), 32.11% in domestic and feral cats in South Africa (Tagwireyi et al., 2019), 14.1% in stray cats in Korea (Par et al., 2020), 86.7% in stray cats in Nigeria (Awobode et al., 2020), and 35% and %59 in domestic and feral cats globally, respectively (Montazeri et al., 2020).

An important risk factor in cats is hunting behavior and ingestion of prey. Eating raw or undercooked meat is also a common mode of transmission. Seroprevalence is higher in adult cats, in free-roaming cats, and in shelter cats (Datz and Evason 2020).

Exposure to cats and litter boxes is thought to be an important risk factor for humans (Datz and Evason 2020). Since cats are in close contact with humans, cats are potential sources of *T. gondii* (Oi et al., 2015). Toxoplasmosis is a zoonotic parasitic disease that is common in animals and humans around the world (Bilgili and Hanedan 2019). It has been reported that more than 1 billion people in the world are infected with *T. gondii* (Hoffmann et al., 2012). Therefore, in order to protect human and animal health, there is a need to know the current situation of toxoplasmosis in cats in our country and to implement ways to prevent this disease.

In Turkey, less studies on the seroprevalence of *T. gondii* in cats are carried out and one study on risk factors is observed (Yücesan et al., 2019). In this study, it was aimed to investigate the seroprevalence and risk factors of *T. gondii* infection in shelter cats in Erzurum, Turkey.

Materials and methods

Animal material

This study was funded by Atatürk University Scientific Research Projects Coordination Unit on 30.09.2021 with the code TKP-2021-9600. In addition, this study was approved by Atatürk University Animal Experiments Local Ethics Committee (27.05.2021/119). In this study, blood samples were collected from 70 cats living in Animal Shelter and Rehabilitation Center of Erzurum Municipality in Erzurum, Turkey, were centrifuged, and sera samples were stored in screw-capped sterile tubes at -20°C until analysis. In the sera samples, IgG antibodies against *T. gondii* were analyzed by Sabin-Feldman Dye test. Young or adult, gender, breed, feeding regimens, place where they were brought, and apparently healthy or diseased conditions for each animal were noted. They were crossbreed and stray cats.

Sabin-Feldman Dye test

The seropositivity of *T. gondii* in cats was analyzed by the Sabin-Feldman Dye Test (SFDT) (Sabin and Feldman 1948; Katsube et al., 1972). In the analysis, the dilutions of positive and negative controls and the test sera were performed using 0.9% saline in a 4-fold serial dilutions (1:4-1:1,024). After diluted, 25 µl was transferred to a tube and an equal volume of activator serum rich in C2, C3, C4 and properdin was added. After 48 hours, tachyzoites (creating a density of approximately 25 tachyzoites per 400x microscope field) were added to each tube. The incubation of tubes were carried out at 37°C for 50 min. Afterwards, an additional 10 min incubation at 37°C was performed in the presence of 25 µl of alkaline methylene blue (pH 11). After incubation, 20 µl of each sample was examined at a 40x objective. If more than 50% of the observed *T. gondii* tachyzoites remained unstained, dilutions were considered positive.

Statistical analysis

Frequency analyzes related to the seropositivity status of cats were calculated. Chi-square independence test was used to investigate the association between risk factors and seropositivity status.

Results

Of 70 cats, 25 (35.7%) were negative and 45 (64.3%) were positive for *T. gondii* seropositivity. The frequency and percentage distributions of cats according to *T. gondii* seropositivity titers are given in Table 1.

The associations between cats' *T. gondii* seropositivity status and risk factors were given in Table 2. Cats were classified according to age as young and adult. Of the youngs, 19 were positive, 14 were negative, and 26 of the adult cats were positive and 11 were negative. 21 of the female cats were positive, 14 were negative. 24 of the male cats were positive and 11 were negative. Of the ill cats, 20 were positive and 11 were negative, and 25 of the apparently healthy cats were positive and 14 were negative. It was determined that there was no significant relationship between *T. gondii* seropositivity and age, gender and health status.

Table 1. Frequency and percentage distributions of cats according to *T. gondii* seropositivity titers

| Titer | Frequency | Percent (%) |
|----------|-----------|-------------|
| 1/16 | 35 | 50 |
| 1/64 | 7 | 10 |
| 1/256 | 3 | 4.3 |
| Negative | 25 | 35.7 |

Table 2. Results of associations between *T. gondii* seropositivity and risk factors

| Variable | Category | Positive | Negative | Total | Prevalence (%) |
|--|--------------------|----------|----------|-------|----------------|
| Age | Young | 19 | 14 | 33 | 57.6 |
| | Adult | 26 | 11 | 37 | 70.3 |
| P = 0.26; df = 1; chi square value = 1.224 | | | | | |
| Gender | Female | 21 | 14 | 35 | 60.0 |
| | Male | 24 | 11 | 35 | 68.6 |
| P = 0.45; df = 1; chi square value = 0.560 | | | | | |
| Status of health | Apparently healthy | 25 | 14 | 39 | 64.1 |
| | ill | 20 | 11 | 31 | 64.5 |
| P = 0.97; df = 1; chi square value = 0.001 | | | | | |

Discussion and Conclusion

This study was first on *T. gondii* seropositivity and risk factors in shelter cats in Erzurum, Turkey. The seroprevalence of *T. gondii* in shelter cats in Erzurum, Turkey was determined to be 64.3%. The seroprevalence rate in shelter cats in this study was higher than 44.1% in cats with outdoor access in Kars city (Erkiliç et al., 2016), 52.8% in stray cats in Ankara city (Özkan et al., 2008), 42-48% in 1021 apparently healthy stray cats and 37.84% in stray cats in İzmir city (Can et al., 2014; Karakavuk et al., 2021), but was lower than 76.4% in stray cats in Nigde city (Karatepe et al., 2008).

In the studies investigating the seropositivity of *T. gondii* in cats around the world, the seroprevalences of infection have been reported to be 64.48% in stray cats in Iran (Tehrani-Sharif et al., 2015), 86.7% in stray cats in Nigeria (Awobode et al., 2020), 82% in feral cats in Qatar (Boughattas et al., 2017), 57.1% in shelter cats in Estonia (Must et al., 2015), 22.3% in stray cats in Greece (Kokkinaki et al., 2023), 20.9% in stray cats in China (Xia et al., 2022), and 14.1% in stray cats in Korea (Park et al., 2020). In addition, globally, the seroprevalence of *T. gondii* has been

determined to be 4.8% to 97.4% in domestic cats (Jittapalapong et al., 2010; Al-Kappany et al., 2010).

Accordingly, *T. gondii* seropositivity is appeared to be high in stray cats in Turkey and in many countries in the world. In this study, when the positivity titers of infected cats were examined, it was determined that there were 35 cats with 1/16 titer, 7 cats with 1/64 titer and 3 cats with 1/256 titer. High-titer antibody responses do not always require cats to have been recently infected (Afonso et al., 2007).

Cat species are highly susceptible to *T. gondii* infection and a single tissue cyst bradyzoite can cause infection (Dubey 2009). Raw or undercooked food and hunting rodents and birds are the important risk factors (Saevik et al., 2015).

In this study, no significant association was found between *T. gondii* seropositivity and gender in cats consistent with the study by Boughattas et al. (2017) in stray cats in Qatar. However, Saevik et al. (2015) have reported a significant association with seropositivity in male cats compared to female cats. Finding different results between infection positivity and gender in different studies may be due to the geographical region studied, study population, and

statistical methods (Jittapalapong et al., 2007; Saevik et al., 2015), as well as male cats hunting more and exposure to infection more (Afonso et al., 2007).

In this study, no significant association was found between *T. gondii* infection positivity and age consistent with the study implemented in stray cats by Matsuu et al. (2017) in Japan. However, in other studies, it has been reported that infection positivity increases significantly in adult cats (Tehrani-Sharif et al., 2015; Must et al., 2015; Bawm et al., 2020). In this study no significance between age and seropositivity might be attributed to similar exposure of *T. gondii* via ingesting oocysts and/or tissue cysts in youngs and adults living outdoors or IgG positivity of *T. gondii* in youngs until 12 weeks of age might be derived from colostral IgG transfer (Gauss et al., 2003; Kokkinaki et al., 2023) or transplacental infection (Xia et al., 2022).

It can be expected that aging, free-roaming and hunting behavior in cats would increase infection rates. Similarly, Brennan et al. (2020) have found the risk factors for *T. gondii* infection in cats as older than 1 year, eating raw meat, and hunting behaviors. Eating cystic meat containing *T. gondii* bradyzoites causes an increase in the infection rates (Salman et al., 2018).

In this study, no significant association was found between the health status of cats and *T. gondii* positivity. Consistent with this study, Saevik et al. (2015) have reported that they do not detect a significant relationship between *T. gondii* positivity and health status in cats. The rates of seropositivity prevalence in apparently healthy and diseased cats (diarrhea and/or respiratory problems) were highly increased as 64.1% and 64.5%, respectively.

As the infected cats shed *T. gondii* oocysts with feces, they cause contamination of soil and water and health problems in animals and humans (Bawm et al., 2020). Cats develop antibodies against *T. gondii* 1-2 weeks after infection (Dubey and Lappin 2012). Therefore, it is important to investigate the presence of *T. gondii* in feces, body fluids, tissue samples and antibodies in serum together in the evaluation of *T. gondii* infections in cats. Investigation of *T. gondii* oocysts in the feces of cats with high-sensitivity techniques such as PCR, colorimetric loop-mediated isothermal magnification analysis (Karakavuk et al., 2022) and making the necessary treatments of positive cases are important in protecting other vertebrate animals and humans against *T. gondii* infections.

In this study, *T. gondii* seroprevalence was determined at high levels in cats living in shelter in Erzurum, Turkey. This suggests that the distribution of *T. gondii* oocysts is prevalent in the environment. The prevention measures against *T. gondii* infection in cats should be implemented.

Acknowledgments: The authors thank Atatürk University Scientific Research Projects Coordination Unit that funded this study.

Conflict of Interest: This study authors have no conflicts of interest.

Ethics Committee Approval: This study was approved by Atatürk University Animal Experiments Local Ethics Committee (27.05.2021/119).

References

- Afonso E, Thulliez P, Pontier D, Gilot-Fromont E. (2007) Toxoplasmosis in prey species and consequences for prevalence in feral cats: not all prey species are equal. *Parasitology* 2007, 134(Pt.14), 1963-1971.
- Al-Kappany YM, Rajendran C, Ferreira LR, Kwok OCH, Abu-Elwafa SA, Hilali M, Dubey JP. (2010) High prevalence of toxoplasmosis in cats from Egypt: isolation of viable *Toxoplasma gondii*, tissue distribution, and isolate designation. *J Parasitol.* 96(6), 1115-1118.
- Awobode HO, Ohiolele JA, Adekeye TA, Adeyi AO, Anumudu CI. (2020) Shedding proportion of *Toxoplasma gondii*-like oocysts in feral cats and soil contamination in Oyo State, Nigeria. *Parasite Epidemiol Control* 11, e00181.
- Bennett AD, Gunn-Moore DA, Brewer M, Lappin MR. (2011) Prevalence of *Bartonella* species, haemoplasmas and *Toxoplasma gondii* in cats in Scotland. *J Feline Med Surg.* 13(8), 553-557.
- Bilgili A, Hanedan B. (2019) Importance of toxoplasmosis for human and animal health, present condition, problems and solution proposals in Turkey and the world. *WJARR.* 4, 61-74.
- Bawm S, Phyu AZ, Chel HM, Htun LL, Nakao R, Katakura K. (2020) Seroprevalence of *Toxoplasma gondii* in household cats in Myanmar and molecular identification of parasites using feline faecal oocysts. *Food Waterborne Parasitol.* 20, e00094.
- Boughattas S, Behnke J, Sharma A, Abu-Madi M. (2017) Seroprevalence of *Toxoplasma gondii* infection in feral cats in Qatar. *BMC Vet Res.* 13, 26.
- Brennan A, Hawley J, Dhand N, Boland L, Beatty JA, Lappin MR, Barrs VR. (2020) Seroprevalence and risk factors for *Toxoplasma gondii* infection in owned domestic cats in Australia. *Vector Borne Zoonotic Dis.* 20 (4), 275-280.
- Can H, Döşkaya M, Ajzenberg D, Özdemir HG, Caner A, İz SG, Döşkaya AD, Atalay E, Çetinkaya Ç, Ürgen S, Karaçalı S, Ün C, Darde M-L, Gürüz Y. (2014) Genetic characterization of *Toxoplasma gondii* isolates and toxoplasmosis seroprevalence in stray cats of Izmir, Turkey. *PLoS One* 9(8), e104930.
- Datz C, Evason M. (2020) *Toxoplasma gondii* (Toxoplasmosis). In: Weese JS, Evason M. (eds). *Infectious Diseases of the Dog and Cat.* 1st ed. CRC Press, Taylor and Francis Group, p. 282-286.
- Dijurkovic-Djakovic O, Dupouy-Camet J, Van der Giessen J, Dubey JP. (2019) Toxoplasmosis: overview from a One Health perspective. *Food Waterborne Parasitol.* 15, e00054.
- Dubey JP. (2009) History of the discovery of the life cycle of *Toxoplasma gondii*. *Int J Parasitol.* 39, 877-882.

- Dubey JP, Lappin MR. (2012) Chapter 79. Toxoplasmosis and neosporosis. In: Greene CE (ed.). *Infectious Diseases of the Dog and Cat*. 4th ed. Elsevier Saunders, USA, p. 806-821.
- Erkiliç EE, Mor N, Babür C, Kirmızıgül AH, Beyhan YE. (2016) The seroprevalence of *Toxoplasma gondii* in cats from the Kars Region, Turkey. *IJVM*. 71, 31-35.
- Gauss CBL, Almería S, Ortuño A, Garcia F, Dubey JP. (2003) Seroprevalence of *Toxoplasma gondii* antibodies in domestic cats from Barcelona, Spain. *J Parasitol*. 89(5), 1067-1068.
- Gilot-Fromont E, Lélou M, Dardé ML, Richomme C, Aubert D, Afonso E., Mercier A, Gotteland C, Villena I. (2012). The life cycle of *Toxoplasma gondii* in the natural environment. *Toxoplasmosis-Recent Advances* 10, 10-34.
- Hoffmann S, Batz MB, Morris JG Jr. (2012) Annual cost of illness and quality-adjusted life year losses in the United States due to 14 foodborne pathogens. *J Food Prot*. 75, 1292-1302.
- Jittapalapong S, Nimsupan B, Pinyopanuwat N, Chimnoi W, Kabeya H, Maruyama S. (2007) Seroprevalence of *Toxoplasma gondii* antibodies in stray cats and dogs in the Bangkok metropolitan area, Thailand. *Vet Parasitol*. 145 (1-2), 138-141.
- Jittapalapong S, Inpankaew T, Pinyopanuwat N, Chimnoi W, Kengradomkij C, Wongnarkpet S, Maruyama S, Lekkla A, Sukthana Y. (2010) Epidemiology of *Toxoplasma gondii* infection of stray cats in Bangkok, Thailand. *Southeast Asian J Trop Med Public Health*. 41(1), 13-18.
- Karakavuk M, Can H, Selim N, Yeşilsiraz B, Atlı E, Şahar EA, Demir F, Gül A, Özdemir HG, Alan N, Yalçın M, Özkurt O, Aras M, Çelik T, Can Ş, Döşkaya AD, Gürüz AY, Döşkaya, M. (2021) Investigation of the role of stray cats for transmission of toxoplasmosis to humans and animals living in Izmir, Turkey. *J Infect Dev Ctries*. 15 (1), 155-162.
- Karakavuk M, Can H, Karakavuk T, Gül A, Alak SE, Gül C, Ün C, Gürüz AY, Döşkaya M, Döşkaya AD. (2022) Rapid detection of *Toxoplasma gondii* DNA in cat feces using colorimetric loop-mediated isothermal amplification (LAMP) assays targeting RE and B1 genes. *Comp Immunol Microbiol Infect Dis*. 81, 101745.
- Karatepe B, Babür C, Karatepe M, Kiliç S, Dündar B. (2008) Prevalence of *Toxoplasma gondii* antibodies and intestinal parasites in stray cats from Niğde, Turkey. *Ital J Anim Sci*. 7, 113-118.
- Katsube Y, Hagivara T, Imaizumi K. (1972) Reliability of the dye and modified hemagglutination test for the latent infection of *Toxoplasma*. *Nihon Juigaku Zasshi*. 34(3), 123-133.
- Kokkinaki KCG, Saridomichelakis MN, Mylonakis ME, Leontides L, Xenoulis PG. (2023) Seroprevalence of and risk factors for *Toxoplasma gondii* Infection in cats from Greece. *Animals*, 13(7), 1173.
- Lappin MR. (2014) Chapter 72: Toxoplasmosis. In: Sykes JE (ed.). *Canine and Feline Infectious Diseases*. 1st ed. Elsevier, Saunders, p. 693-703.
- Matsuu A, Yokota SI, Ito K, Masatani T. (2017) Seroprevalence of *Toxoplasma gondii* in free-ranging and feral cats on Amami Oshima Island, Japan. *J Vet Med Sci*. 79(11), 1853-1856.
- Montazeri M, Mikaeili Galeh T, Moosazadeh M, Sarvi S, Dodangeh S, Javidnia J, Sharif M, Daryani A. (2020) The global seroprevalence of *Toxoplasma gondii* in felids during the last five decades (1967-2017): a systematic review and meta analysis. *Parasit Vectors*. 13, 82.
- Must K, Lassen B, Jokelainen P. (2015) Seroprevalence of and risk factors for *Toxoplasma gondii* infection in cats in Estonia. *Vector Borne Zoonotic Dis*. 15(10): 597-601.
- Oi M, Yoshikawa S, Maruyama S, Nogami S. (2015) Comparison of *Toxoplasma gondii* seroprevalence in shelter cats and dogs during 1999-2001 and 2009-2011 in Tokyo, Japan. *PLoS One* 10:e0135956.
- Özkan AT, Çelebi B, Babür C, Lucio-Forster A, Bowman DD, Lindsay DS. (2008) Investigation of anti-*Toxoplasma gondii* antibodies in cats of the Ankara region of Turkey using the Sabin-Feldman Dye test and an indirect fluorescent antibody test. *J Parasitol*. 94, 817-820.
- Park Y, Noh J, Seo HJ, Kim KH, Min S, Yoo MS, Yun BR, Kim JH, Choi EJ, Cheon DS, Hong SJ, Yoon SS, Cho YS. (2020) Seroprevalence and B1 gene phylogeny of *Toxoplasma gondii* of dogs and cats in Republic of Korea. *Korean J Parasitol*. 58(3), 257-265.
- Sabin AB, Feldman A. (1948) Dyes as microchemical indicators of a new immunity phenomenon affecting a protozoan parasite (*Toxoplasma*). *Science* 108(2815), 660-663.
- Saevik BK, Krontveit RI, Eggen KP, Malmberg N, Thoresen SI, Prestud KW. (2015) *Toxoplasma gondii* seroprevalence in pet cats in Norway and risk factors for seropositivity. *J Feline Med Surg*. 17, 1049-1056.
- Salman D, Pumidonming W, Oohashi E, Igarashi M. (2018) Prevalence of *Toxoplasma gondii* and other intestinal parasites in cats in Tokachi sub-prefecture, Japan. *J Vet Med Sci*. 80(6), 960-967.
- Tagwireyi WM, Etter E, Neves L. (2019) Seroprevalence and associated risk factors of *Toxoplasma gondii* infection in domestic animals in southeastern South Africa. *Onderstepoort J Vet Res*. 86, e1-e6.
- Tehrani-Sharif M, Jahan S, Alavi SM, Hodami M. (2015) Seroprevalence of *Toxoplasma gondii* antibodies of stray cats in Garmsar, Iran. *J Parasit Dis*. 39, 306-308.
- Xia N, Ji N, Li L, Huang Y, Yang C, Guo X, Guo Q, Shen B, Xiao L, Feng Y. (2022) Seroprevalence and risk factors of *Toxoplasma gondii* in urban cats from China. *BMC Vet Res*. 18(1), 331.
- Yücesan B, Babür C, Koç N, Sezen F, Kılıç S, Gürüz Y. (2019). Ankara yöresindeki kedilerde 2016 yılında Sabin-Feldman Dye testi (SFDT) ile anti-*Toxoplasma gondii* antikorlarının araştırılması. *Türkiye Parazitoloj Derg*. 43(1), 5.