

RESEARCH ARTICLE

Antioxidant Activity of Chamomile Essential Oil and Main Components

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Abstract

The objective of this study was to characterize the antioxidant capacity of the chamomile (*Matricaria recutita* L.) essential oil (EO) as well as its major constituents. For this purpose, Pharmacopoeia grade Matricariae aetheroleum was used after the analytical evaluation, which was confirmed by GC/MS and GC/FID, respectively. The main components were identified as α -bisabolol oxide A and B, (*E*)- β -farnesene, α -bisabolone oxide A, chamazulene, and α -bisabolol according the European Pharmacopoeia, respectively. The major constituents α -bisabolol oxide A and chamazulene were purified using prep-TLC from the EO, and the identification and quantification of the constituents were confirmed by GC/MS and GC/FID analyses. Chamomile EO, α -bisabolol oxide A, (*E*)- β -farnesene, chamazulene, and α -bisabolol were evaluated comparatively for their *in vitro* antioxidant activities using the spectrophotometric 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical-scavenging microdilution assay. The highest free radical inhibitory activity was observed for chamazulene followed by α -bisabolol oxide A, the chamomile EO and (*E*)- β -farnesene, respectively.

Keywords: Chamomile essential oil, Matricaria recutita L., DPPH antioxidant activity, chamazulene, sesquiterpene

Introduction

In general, antioxidant substances present in foods or in the biological system at low concentrations compared with oxidative substrates may delay or prevent the oxidation process. Assessments of antioxidant properties of natural compounds including essential oils (EOs) are important due to their uses in food, medicine, and cosmetics (Halliwell et al., 1995; Mishra et al., 2012).

It is well known that EOs have various properties, and the use of EOs as natural antioxidants is common practice in several cases. Essential oils are mainly composed of terpenes such as monoterpenes and sesquiterpenes (Amorati et al., 2013; Baser and Demirci, 2007). The sesquiterpenes are subject for a vast number of biological evaluations and activities including antioxidant potential (Bartikova et al., 2014).

Chamomile, *Matricaria recutita* L. of the Asteraceae family, is an annual herbaceous plant, growing in Germany, Hungary, France, Russia, Yugoslavia, India including Turkey, which is cultivated in several countries such as Egypt, Algeria, Hungary, Germany, among others (Das, 2015). Chamomile EO is highly demanded, which is obtained by steam distillation, and is also described in several pharmacopoeias for its specifications. Pharmaceutically it is commonly used for the treatment of various diseases associated to inflammation, infection, and spasms among others. The EO is characterised by the presence of sesquiterpenes such as chamazulene, farnesene and α -bisabolol and oxygenated sesquiterpenes were the most characteristic chamomile EO such as bisabolone oxide A and bisabolol oxide A and B. The sesquiterpenes are the major biologically active substances in the EO (Rhind, 2012; Schilcher, 2005; Singh et al., 2011; Srivastava et al.,

2010). Among other uses chamomile EO has been reported as a natural antioxidant (Abdoul-Latif et al., 2011; Ayoughi et al., 2011; Owlia et al., 2007; Stanojevic et al., 2016) and also is dark blue coloured oil that contains chamazulene, which is an important factor for the antioxidant power of chamomile EO (Buckle, 2015; Capuzzo et al., 2014; Ornano et al. 2013; Rekka et al., 1996).

The aim of this present study was to evaluate the biological activity of Pharmacopoeia grade chamomile EO. For this purpose, the EO was subjected to *in vitro* antioxidant activity by microdilution spectroscopy. In addition to the EO the major components α -bisabolol oxide A, (*E*)- β -farnesene, chamazulene, α -bisabolol; where the purified bisabolol oxide A and chamazulene were used and evaluated using the DPPH radical scavenging activity.

Materials and Methods

EO and chemicals

Pharmacopeia (PhEur) grade chamomile EO (*Matricaria recutita* L.) was acquired from (Phatrade, Cairo in Eygpt). α -Bisabolol and (*E*)- β -farnesene are purchased from Sigma-Aldrich. All chemicals used were of high purity and analytical grade which were supplied by Sigma Aldrich, Fluka and Merck and used without further purification unless otherwise stated. The chamomile EO composition was previously reported in detail (see Goger et al., 2018).

Gas chromatography/flame ionization detection (GC/FID) and gas chromatography/mass spectrometry (GC-MS) analyses

The GC analysis was carried out using an Agilent 6890N GC system. FID detector temperature was 300 °C. In order to obtain the same elution order with GC-MS, simultaneous auto-injection was done by using same column and an appropriate operational conditions. The GC/MS analysis was carried out with an Agilent 5975 GC/MSD system as previously described in detail (Demirci et al, 2015; Goger et al., 2018).

Analytical and preparative thin layer chromatography (TLC, prep-TLC)

Chamomile EO was analysed by TLC. TLC analysis was performed on silica gel 60 G_F 254 (Merck) using chloroform-toluene (3:1, v/v) as a mobile phase by spraying with anisaldehyde/sulphuric acid and heating for 5 minutes at 110 °C. α -Bisabolol oxide A and chamazulene were isolated using successive prep-TLC techniques from the chamomile EO and the identification and quantification of the constituents were by GC/MS and GC/FID analysis.

The DPPH free radical scavenging activity

The DPPH free radical scavenging activity is based on the ability of 1,1-Diphenyl-2-picryl-hydrazyl (DPPH), a stable free radical, to be decolorized in the presence of antioxidants (Kumarasamy et al., 2007). Briefly, stock solutions and DMSO (control) were prepared and added 80 μ g/ml DPPH solution in MeOH. The mixtures were shaken vigorously and left to stand in the dark for 30 min at room temperature, then absorbance was read at 517 nm. Radical scavenging capacity was expressed as percentage inhibition (I %) and calculated using the following equation:

(I %) =[(Abs control – Abs sample) / Abs control] × 100 (formulae)

Ascorbic acid was used as a positive control. The DPPH IC_{50} values (IC_{50} value is the concentration of the sample required to inhibit 50 % of radical) of the samples were calculated and are listed in Table 2.

Results and Discussion

Chamomile EO composition

The chemical composition of the study material chamomile EO was recently reported where α - bisabolol oxide A (1) (47.7 %), (*E*)- β -farnesene (2) (21.5 %), α -bisabolol oxide B (3) (6.2 %), α -bisabolone oxide A (4) (5.7 %), chamazulene (5) (4.1 %) and α -bisabolol (6) (2.1 %) (Goger et al., 2018), were identified as the main components. The MS fragmentation of the main components are shown in Figure 1. which was detailed and listed in Table 1.

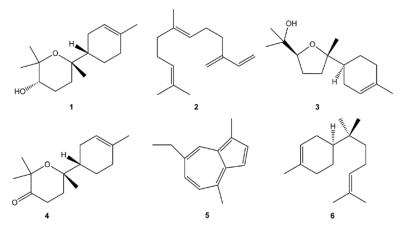
Stanojevic et al. (2016), identified 52 components consisting of β -farnesene (29.8 %), α -farnesene (9.3 %), α bisabolol and its oxide (15.7 %), chamazulene (6.4 %), germacrene D (6.2 %) and spiroether (5.6 %) in chamomile EO. In another study, major compounds were identified as chamazulene (31.2 %), 1,8-cineole (15.2 %), β -pinene (10.1 %), α -pinene (8.14 %), α -bisabolol (7.5 %) and terpinen-4-ol (4.1 %) (Farhoudi, 2013). Also the chamomile EO consisted of α -bisabolol oxide A (48.2 %), α -bisabolol oxide B (23.31 %), α -bisabolol (12.1 %) and β -farnesene (5.2 %), chamazulene (2.4 %) sesquiterpenes as major constituents (Roby et al., 2013). Ayoughi et al. (2010), reported that (*E*)- β -farnesene (24.2 %), guaiazulene (10.6 %), α - bisabolol oxide A (10.2 %), α -farnesene (8.7 %) and α -bisabolol (7.3 %) were present in *M. recutita* EO.

Table 1. GC/MS analysis results of chamomile EO

Rt (min)	<i>m/z</i> (Relative intensity, %)	%
33,66	M ⁺ 204 (4), 189 (3), 175 (1), 161 (18), 148 (4), 133 (33), 120 (23), 107 (11), 93 (67), 79 (26), 69 (100), 55 (16), 41 (52)	21.5
46,38	M⁺ 238 (2), 220 (5), 205 (3), 179 (27), 161 (55), 143 (100), 134 (39), 125 (34), 107 (24), 105 (74), 95 (27), 85 (58), 71 (35), 59 (47), 43 (55)	
47,45	M⁺ 236 (2), 218 (3), 200 (5), 178 (11), 169 (6), 150 (28), 141 (96), 132 (16), 121 (54), 107 (38), 93 (100), 79 (34), 67 (46), 55 (20), 43 (47)	5.7
48,06	06 M ⁺ 222, 204 (33), 189 (6), 161 (15), 147 (6), 134 (11),119 (97), 109 (100), 93 (46), 79 (20), 69 (85), 55 (23), 43 (63)	
mazulene 52,73 M ⁺ 184 (97), 169 (100), 153 (34), 141 (13), 128 (19), 115 (13), 89 (6), 77 (6), 63 (3), 45 (6)		4.1
52,96	M ⁺ 238 (1), 220 (2), 180 (3), 159 (3), 143 (100), 134 (18),125 (36), 107 (26), 93 (37), 81 (12), 71 (24), 59 (17), 43 (40)	47.7
	33,66 46,38 47,45 48,06 52,73	33,66 M ⁺ 204 (4), 189 (3), 175 (1), 161 (18), 148 (4), 133 (33), 120 (23), 107 (11), 93 (67), 79 (26), 69 (100), 55 (16), 41 (52) 46,38 M ⁺ 238 (2), 220 (5), 205 (3), 179 (27), 161 (55), 143 (100), 134 (39), 125 (34), 107 (24), 105 (74), 95 (27), 85 (58), 71 (35), 59 (47), 43 (55) 47,45 M ⁺ 236 (2), 218 (3), 200 (5), 178 (11), 169 (6), 150 (28), 141 (96), 132 (16), 121 (54), 107 (38), 93 (100), 79 (34), 67 (46), 55 (20), 43 (47) 48,06 M ⁺ 222, 204 (33), 189 (6), 161 (15), 147 (6), 134 (11), 119 (97), 109 (100), 93 (46), 79 (20), 69 (85), 55 (23), 43 (63) 52,73 M ⁺ 184 (97), 169 (100), 153 (34), 141 (13), 128 (19), 115 (13), 89 (6), 77 (6), 63 (3), 45 (6) 52,96 M ⁺ 238 (1), 220 (2), 180 (3), 159 (3), 143 (100), 134 (18), 125 (36), 107 (26),

Rt: Retention time; *m/z:* Mass-to-charge ratio; % calculated from FID data.

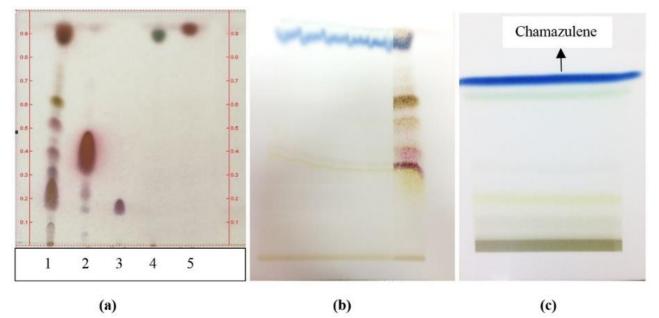
Figure 1. Chamomile EO major constituents



Isolation of the EO components

 α -Bisabolol oxide A and chamazulene were purified by prep-TLC eluting with chloroform-toluene (3:1, v/v) from the chamomile EO (Figure 2), and the identification-quantification of the constituents were performed by GC/MS and GC/FID analyses.

Figure 2. (a) TLC of analytical standard, (1) chamomile EO, (2) α -bisabolol, (3) α -bisabolol oxide A, (4) chamazulene, (5) (*E*)- β -farnesene; (b) preparative TLC of chamomile EO; (c) isolation of high-purity chamazulene.



As previously reported (Ashnagar et al., 2009), three major components were separated, purified and identified namely as chamazulene ($R_f = 0.93$), bisabolonoxide ($R_f = 0.75$) and bisabololoxide A ($R_f = 0.3$) from *Matricaria recutita* EO. The chemical structures of the components were determined by their IR, ¹H NMR and MS spectra. These data also comply with our results.

Results of the DPPH free radical scavenging assay

In the DPPH assay the radical scavenging ability of the chamomile EO, and its four main components as listed in (Table 2), and also the positive control (ascorbic acid) was compared spectrophotometrically.

Samples	DPPH	
Samples	IC₅₀ (mg/mL)	
Chamomile EO	2.20	
α-Bisabolol	43.88	
α-Bisabolol oxide A	1.50	
(<i>E</i>)-β-Farnesene	7.45	
Chamazulene	0.27	
Ascorbic acid (reference)	0.015	

Table 2. Antioxidant activity of chamomile EO and its four main components on DPPH[•] assay

The results showed that the scavenging activity of the main substances were in the following order; chamazulene > α -bisabolol oxide A > chamomile EO > (*E*)- β -farnesene > α -bisabolol, respectively, accordingly to their IC₅₀ values compared to the positive control ascorbic acid, which exhibited high antioxidant activity with IC₅₀ value of 0.015 mg/mL in the same experimental system.

Previous antioxidant activity of chamomile EO showed the best antioxidant properties after 90 minutes of incubation with the EC₅₀value of 2.07 mg/mL by DPPH free radical method (Stanojevic et al., 2017). In another antioxidant activity study by DPPH free radical scavenging and β -carotene/linoleic acid methods, the EO EC₅₀ value was determined as 5.63 ± 0.20 mg/mL as reported by Ayoughi et al. (2010). In the β -carotene bleaching test, the EO gave the best inhibition result of 82.5 % after 120 minutes, supporting the antioxidant activity (Owlia et al., 2007).

Interestingly, Capuzzo et al. (2014), reported that chamazulene was unable to react with DPPH[•]. According to the literature, it is necessary to avoid DPPH assay to test chamazulene radical scavenging activity owing to its interference with the nitrogen-centred DPPH, colour factor and thus suggests the use of ABTS assay instead. Actually when chamazulene was evaluated using ABTS[•], a strong and significantly higher free radical scavenging activity was observed (IC₅₀ 3.7 \pm 0.7 µg/mL).

Conclusion

The results showed that the *in vitro* scavenging activity of chamazulene was the highest after α -bisabolol oxide A, and the chamomile EO followed by the other chamomile major volatile constituents. To the best of our knowledge this is the first report on the antioxidant activity of α -bisabolol oxide A and (*E*)- β -farnesene constituents, which may responsible of the total activity of the EO.

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REFERENCES

Abdoul-Latif, F.M., Mohamed, N., Edou, P., Ali, A.A., Djama, S.O., Obame, L.C., Bassole, I.H.N. & Dicko, M.H. (2011). Antimicrobial and antioxidant activities of essential oil and methanol extract of *Matricaria chamomilla* L. from Djibouti. *Journal of Medicinal Plants Research*, 5(9), 1512-1517.

Amorati, R., Foti, M.C. & Valgimigli, L. (2013). Antioxidant activity of essential oils. *Journal of Agricultural and Food Chemistry*, 61(46), 10835-10847.

Ashnagar, A., Naseri, N.G. & Alavi, S.Y. (2009). Isolation and identification of the major chemical components in the capitula of *Matricaria chamomilla* grown in Khuzestan Province of Iran. *Asian Journal of Chemistry*, 21(7), 4981-4986.

Ayoughi, F., Barzegar, M., Sahari, M.A. & Naghdibadi, H. (2011). Chemical compositions of essential oils of *Artemisia dracunculus* L. and endemic *Matricaria chamomilla* L. and an evaluation of their antioxidative effects. *Journal of Agricultural Science and Technology*, 13, 79-88.

Bartikova, H., Hanusova, V., Skalova, L., Ambroz, M. & Bousova, I. (2014). Antioxidant, pro-oxidant and other biological activities of sesquiterpenes. *Current Topics in Medicinal Chemistry.*, 14(22), 2478-2494.

Baser, K.H.C. & Demirci, F. (2007). Chemistry of essential oils. In R.G. Berger, (Ed.), *Chemistry of essential oil in flavours and fragrances, chemistry, bioprocessing and sustainability* (pp. 43-86). Springer Berlin Heidelberg.

Buckle, J. (2015). Basic plant taxonomy, basic essential oil chemistry, extraction, biosynthesis, and analysis. In J. Barlow (Ed.), *Clinical aromatherapy* (pp. 37-72). St. Louis: Churchill Livingstone.

Capuzzo, A., Occhipinti, A. & Maffei, M.E. (2014). Antioxidant and radical scavenging activities of chamazulene. *Natural Product Research. Form. Nat. Product Lett.*, 28(24), 2321-2323.

Das, M. (2015).Chamomile: medicinal, biochemical, and agricultural aspects. Boca Raton: CRC Press Taylor and Francis Group.

Demirci, F., Bayramic, P., Göger, G., Demirci, B. & Baser, K.H.C. (2015). Characterization and antimicrobial evaluation of the essential oil of *Pinus pinea* L. from Turkey. *Natural Volatiles & Essential Oils*, 2(2), 39-44.

Farhoudi, R. (2013). Chemical constituents and antioxidant properties of *Matricaria recutita* and *Chamaemelum nobile* essential oil growing wild in the South West of Iran. *Journal of Essential Oil Bearing Plants*, 16(4), 531-537.

Goger, G., Demirci, B., Ilgın, S. & Demirci, F. (2018). Antimicrobial and toxicity profiles evaluation of the chamomile (*Matricaria recutita* L.) essential oil combination with standard antimicrobial agents. *Industrial Crops and Products*, 120, 279-285.

Halliwell, B., Aeschbach, R., Löliger, J. & Aruoma, O.I. (1995). The characterization of antioxidants. *Food and Chemical Toxicology*, 33(7), 601-617.

Kumarasamy, Y., Byres, M., Cox, P.J., Jaspars, M., Nahar, L. & Sarker, S.D. (2007). Screening seeds of some Scottish plants for free-radical scavenging activity. *Phytotherapy Research*, 21(7), 615-621.

Mishra, K., Ojha, H. & Chaudhury, N.K. (2012). Estimation of antiradical properties of antioxidants using DPPH⁻ assay: A critical review and results. *Food Chemistry*, 130, 1036-1043.

Ornano, L., Venditti, A., Ballero, M., Sanna, C., Quassinti, L., Bramucci, M., Lupidi, G., Papa, F., Vittori, S., Maggi, F. & Bianco, A. (2013). Chemopreventive and antioxidant activity of the chamazulene-rich essential oil obtained from *Artemisia arborescens* L. growing on the Isle of La Maddalena, Sardinia, Italy. *Chemistry & Biodiversity*, 10(8), 1464-1474.

Owlia, P., Rasooli, I. & Saderi, H. (2007). Antistreptococcal and antioxidant activity of essential oil from *Matricaria* chamomilla. L. Research Journal of Biological Sciences, 2(2), 155-160.

Rekka, E.A., Kourounakis, P. & Kourounakis, P.N. (1996). Investigation of the effect of chamazulene on lipid peroxidation and free radical processes. *Research Communications in Molecular Pathology and Pharmacology*, 92, 361-364.

Rhind, J. P. (2012). The essential oils, absolutes and resinoids of aromatherapy. In S. Dragon (Ed.), *Essential oils: A handbook for aromatherapy practice* (pp. 111-255). Jessica Kingsley Publishers.

Roby, M.H.H, Sarhan, M.A., Selim, K.A.H. & Khalel, K.I. (2013). Antioxidant and antimicrobial activities of essential oil and extracts of fennel (*Foeniculum vulgare* L.) and chamomile (*Matricaria chamomilla* L.). *Industrial Crops and Products*, 44, 437-445.

Schilcher, H. (2005). Legal situation of German chamomile: monographs. In R. Franke & H. Schilcher (Eds.), *Chamomile industrial profiles* (pp. 7-38). Boca Raton: CRC Press Taylor and Francis Group.

Singh, O., Khanam, Z., Misra, N. & Srivastava, M.K. (2011). Chamomile (*Matricaria chamomilla* L.): An overview. *Pharmacognosy Reviews*, 5, 82-95.

Srivastava, K.J., Shankar, E. & Gupta, S. (2010). Chamomile: A herbal medicine of the past with bright future. *Molecular Medicine Reports*, 3(6), 895-901.

Stanojevic, L.P., Marjanovic-Balaban, Z.R., Kalaba, V.D., Stanojevic, J.S. & Cvetkovic, D.J. (2016). Chemical composition, antioxidant and antimicrobial activity of chamomile flowers essential oil (*Matricaria chamomilla* L.). *Journal of Essential Oil Bearing Plants*, 19(8), 2017-2028