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Research Article

QSAR Studies of amino-pyrimidine derivatives as *Mycobacterium tuberculosis* Protein Kinase B inhibitors

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Abstract: Quantitative structure activity relationship (QSAR) analysis was applied to a series of amino-pyrimidine derivatives as PknB inhibitors using a combination of various physicochemical and quantum descriptors. A multiple linear regression (MLR) procedure was used to model the relationships between molecular descriptors and the chemotherapeutic activity of the amino-pyrimidine derivatives. Good agreement between experimental and predicted activity values, obtained in the validation procedure, indicated the good quality of the derived QSAR model. The best QSAR model developed show a good predictive correlation coefficient $R^2 = 0.973$ and external predictive ability of prediction $R^2 = 0.778$ was developed by MLR. The proposed model has good robustness and predictability when verified by internal and external validation.

Keywords: QSAR, amino-pyrimidine, PknB, MLR.

1. Introduction

Tuberculosis (TB) is the primary cause of mortality due to an infectious disease in the world today. The causative agent of tuberculosis is the intracellular pathogen *M. tuberculosis* [1]. Recent studies have focused on finding new pathways vulnerable to inhibition by small molecules and previously unexploited by drug discovery efforts [2]. The inhibition of signaling pathways both in *M. tuberculosis* and the host may yield new classes of drug targets and a large amount of recent studies are focused on developing this further [3].

Target based drug discovery, in which there is in vitro high throughput screening of a large number of small molecules against a validated target, has been used on a number of occasions to search for new anti-tuberculosis agents. We sought to find inhibitors of an essential mycobacterium tuberculosis serine/threonine protein kinase, (PknB) [4].

Novel molecules such as chalcones, thiophene, pyrimidine derivatives was synthesis exhibit antimycobacterial activity against other species of *Mycobacterium* [5-8].

Despite the fact that death from TB is often preventable, the rapid increase of multidrug-resistant tuberculosis and extensively drug-resistant tuberculosis has resulted in an urgent need to develop new drug targets for *Mycobacterium tuberculosis* (Mtb) [9, 10].

The discovery and development of new anti-TB therapeutics is widely recognized as one of the major global health emergencies, yet it is also a major pharmaceutical challenge. A theoretical technique such as quantum chemical descriptors have been extensively used in Quantitative Structure-Activity Relationship (QSAR) [11-14] to predict the physiological and biological properties of compounds under study. Whether these compounds (amino-pyrimidine derivatives)

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provide structural precedence and may lead to the generation of novel anti-TB therapeutics.

The main goal of QSAR studies is the development of predictive models which can be used in computer-aided drug discovery and design for prediction of activities or properties of new compounds (i.e., those included in chemical databases or combinatorial libraries). Prior to using the model for external predictions, its predictive power should be established and validated. Thus, model validation has become a standard (and in some laboratories such as ours, mandatory) part of QSAR modeling. In the past, model validation was performed using leave one out (LOO) cross-validation. LOO cross-validation has been determined to yield an overoptimistic estimate of predictive ability (if model feature selection is not redone for each hold out) [15].

External validation is now a “must have” tool for evaluating the reliability of QSAR models [16]. In this procedure, typically the overall set is randomly divided into a training set and a test set. QSAR models are developed based on the training set and then are used to make predictions for the test set. The advantage of this approach is that the test set compounds are “unknown” to the models since they are excluded from the model development procedure, especially the variable selection.

2. Materials and methods

2.1. Data Set

A dataset of 29 amino-pyrimidine derivatives as Mycobacterium tuberculosis PknB inhibitors was selected from the literature based on the biological assay method [3, 17-18]. The structures and their inhibitory activities in pIC50 are listed in Table 1. $pIC_{50} = -\log(IC_{50})$. IC50 represents the compound concentration required for 50% inhibition.

The data set was divided into the two subsets, training set of 22 compounds (75.76%) and test set of 7 compounds (24.24% >20% for external validation). The test set compounds were selected manually considering the distribution of biological data and structural diversity. Training set was used to build a regression model, and the test set was used to evaluate the predictive ability of the model obtained.

Firstly, all the structures of molecules were pre-optimized using the Molecular Mechanics Force

Field (MM+) method included in HyperChem release 8.08 [19]. The resulted minimized structures were further refined using the semi empirical PM3 Hamiltonian, which is implemented also in HyperChem. We chose a gradient norm limit of 0.01 kcal/Å for the geometry optimization.

In the second part, the series of the amino-pyrimidine derivatives was re-optimised by the density functional theory DFT/B3LYP at the 6-311G(d,p) basis set of level, by using Gaussian 09 software [20]. Further, the regression analysis was performed using the SPSS version 21 for Windows [21].

The QSAR properties module from HyperChem (8.08) was used to calculate: molar polarizability (Pol), the molar refractivity (MR), partition coefficient octanol/water(log P), molar volume (MV), Surface area grid (SAG) and molar weight (MW).

The Quantum Chemical descriptors: dipole moment (DM), HOMO-LUMO energy gap (ΔE), total energy (ET) and atomic net charges (Natural bond orbital charges NBO)(qN1, qC2, qN3, qC5 and qN6) were computed using Gaussian 09 software.

2.2. Multiple linear regression (MLR) analysis

Multiple linear regression is the standard method for multivariate data analysis. It is also called as Ordinary Least Squares regression (OLS). This method of regression estimates the values of the regression coefficients by applying least squares curve fitting method. For getting reliable results, dataset having typically 5 times as many data points (molecules) as independent variables (descriptors) is required. The regression equation takes the form:

$$Y = a_0 + a_1 * X_1 + a_2 * X_2 + a_3 * X_3 + \dots + a_n * X_n \quad (1)$$

Where Y is the response or dependent variable, X1, X2, ..., Xn are descriptors (features or independent variables) present in the model with the corresponding regression coefficients a1, a2, ..., an, respectively, and a0 is the constant term of the model.

2.3. Validation Methods for QSAR Models

For the validation of the predictive power of a QSAR model, two basic principles (internal validation and external validation)

2.3.1. Internal validation

Internal model validation was carried out using leave-one-out (LOO- $R_{CV}^2(Q^2)$) method. For

calculating $R^2_{cv}(Q^2)$, each sample in the training set was eliminated once and the activity of the eliminated sample was predicted by using the model developed by the remaining samples. $LOO-R^2_{cv}(Q^2)$ calculated [22] according to the below formula:

$$R^2_{CV}(Q^2) = 1 - \frac{\sum(Y_{pred} - Y_{obs})^2}{\sum(Y_{pred} - Y_{mean})^2} > 0.5 \quad (2)$$

In Eq. (2), Y_{pred} and Y_{obs} indicate predicted and observed activity values accordingly and Y_{mean} signify mean activity value. A model is considered acceptable when the value of $R^2_{CV}(Q^2)$ exceeds 0.5.

2.3.2. External validation

According to Tropsha et al. [23], for considering the validity of the developed models, in addition to the internal validation, the models should be externally validated using the test set compounds. According to their study, the following criteria could be considered as of acceptable predictability:

$$R^2_{pred} = 1 - \frac{\sum(Y_{pred} - Y_{obs})^2}{\sum(Y_{pred} - Y_{mean(train)})^2} > 0.6 \quad (3)$$

where R^2_{pred} is squared correlation coefficient between the experimental and predicted biological activities of dataset compounds (training and test sets).

Y_{pred} and Y_{obs} indicate predicted and observed activity values for the test set and

$Y_{mean(train)}$ indicates mean activity value of the training set. For the predictive QSAR model.

$$\frac{(R^2 - R_0^2)}{R^2} < 0.1 \quad \text{or} \quad \frac{(R^2 - R_0'^2)}{R^2} < 0.1 \quad (4)$$

$$|R_0^2 - R_0'^2| < 0.3 \quad (5)$$

where R^2 is determination coefficient between the experimental and predicted pIC50 values of test set compounds with intercept. R_0^2 and $R_0'^2$ are determination coefficients of predicted versus experimental and experimental versus predicted biological activity values in which their regression line must pass through the origin, respectively.

$$0.85 \leq K \leq 1.15 \quad \text{or} \quad 0.85 \leq K' \leq 1.15 \quad (6)$$

The slope of regression line through the origin which mentioned in the above was indicated by K and K' , respectively.

3. Results and Discussion

In an attempt to determine the role of structural features of compounds, which appears to have an effect the antitubercular activity, QSAR models were generated. The structures of studied compounds are shown in Table 1., and descriptors are used in Table 2. To establish the statistical correlation, the physicochemical descriptors were taken as independent variables and antitubercular activity as dependent variable

Table 1. Chemical structure, experimental and predicted activities of the molecules under study.

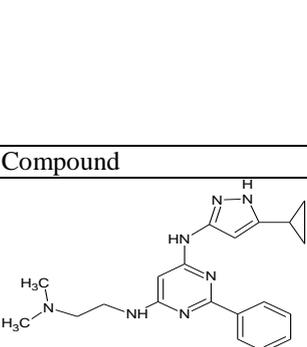
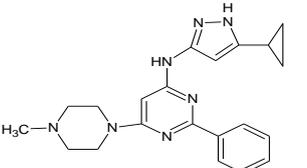
No	Compound	pIC50 Exp.	pIC50 Pred.	Res.
1		7.06	7.118	-0.058
2		6.92	6.873	0.046

Table 1. Continued

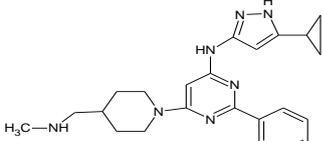
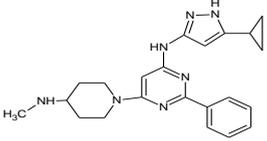
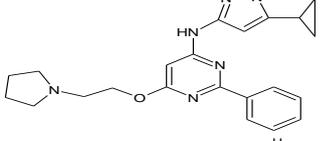
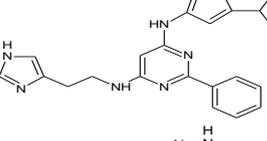
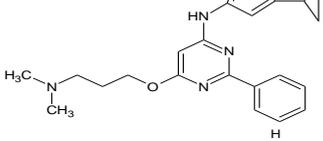
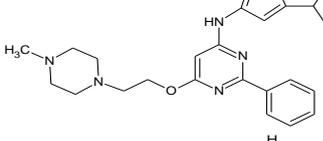
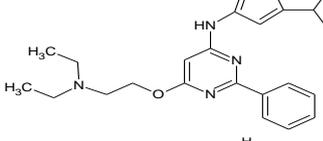
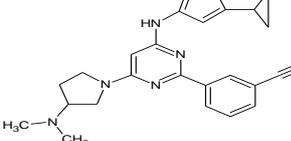
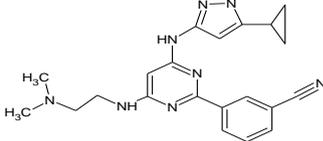
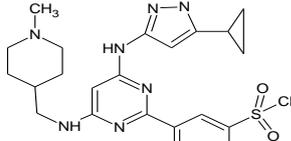
No	Compound	pIC50 Exp.	pIC50 Pred.	Res.
3 ^T		7.14	6.865	0.274
4 ^T		7.28	7.240	0.040
5		7.25	7.165	0.084
6		6.46	6.491	-0.031
7		7.26	7.204	0.055
8		7.02	7.216	-0.196
9		7.04	6.951	0.088
10		7.25	7.182	0.067
11		7.41	7.372	0.037
12		7.40	7.402	-0.002

Table 1. Continued

No	Compound	pIC50 Exp.	pIC50 Pred.	Res.
13		6.43	6.618	-0.188
14		6.42	6.354	0.064
15		7.07	6.951	0.118
16 ^T		7.08	6.327	0.752
17		6.45	6.389	0.061
18 ^T		6.15	6.304	-0.154
19		6.66	6.712	-0.053
20		6.41	6.432	-0.022
21		6.62	6.644	-0.025
22		7.06	7.047	0.013

Table 1. Continued

No	Compound	pIC50 Exp.	pIC50 Pred.	Res.
23 ^T		6.69	6.649	0.041
24 ^T		6.94	7.225	-0.285
25 ^T		5.21	5.147	0.062
26		6.28	6.313	-0.033
27		5.95	5.941	0.009
28		5.70	5.683	0.016
29		7.40	7.451	-0.051

Exp: Experimental activity, Pred: Predicted activity, T: Test Set, Res: Residual

Table 2. Values of molecular descriptors used in the regression analysis.

Comp number	MW (amu)	SAG (Å ²)	MV (Å ³)	Pol (Å ³)	MR (Å ³)	HE (Kcal/mol)	LogP	MD Debye
1	363.47	676.89	1137.04	41.62	113.02	-8.87	1.81	4.74
2	375.48	666.97	1130.58	42.69	116.35	-7.28	1.99	5.71
3	403.53	727.93	1234.55	46.36	125.27	-7.95	2.23	2.58
4	389.50	685.27	1175.67	44.52	120.46	-7.93	1.81	1.74
5	390.49	692.41	1182.78	43.81	118.62	-7.51	2.45	2.50
6	386.46	648.19	1128.08	43.43	118.95	-17.6	0.42	2.00
7	378.48	700.00	1178.93	42.75	115.94	-7.95	2.18	3.72
8	419.53	720.61	1250.68	46.99	127.13	-7.72	1.93	2.07
9	392.50	721.55	1221.92	44.58	120.57	-7.45	2.81	4.16
10	414.51	715.62	1240.43	46.37	125.86	-9.83	1.84	6.11
11	388.47	690.78	1175.96	43.48	118.00	-9.81	1.53	5.90
12	481.62	791.24	1369.02	49.33	138.94	-12.26	0.64	9.65
13	309.35	561.65	917.18	33.33	91.97	-7.37	2.06	3.94
14	334.38	580.68	982.60	36.69	99.66	-11.52	1.18	1.45
15	369.44	604.52	1035.66	36.39	106.69	-9.22	-0.12	4.28
16	312.42	604.63	996.87	35.34	92.11	-6.44	3.41	2.68
17	300.41	593.65	985.85	34.28	89.52	-6.57	3.36	2.83
18	320.40	603.62	999.47	36.60	99.85	-9.38	2.84	2.84
19	330.39	617.61	1115.49	37.10	83.04	-7.44	2.72	2.94
20	384.46	645.49	1080.42	37.74	109.99	-12.29	-0.16	2.30
21	398.48	675.55	1141.48	39.58	112.97	-11.81	1.54	9.70
22	316.37	588.60	962.93	35.27	96.82	-13.21	2.39	8.99
23	292.34	552.48	895.67	32.71	88.34	-9.69	1.32	8.39
24	293.39	571.88	943.99	33.51	88.70	-6.85	1.83	2.95
25	360.46	637.86	1092.89	41.92	114.92	-9.00	2.30	3.60
26	428.46	682.82	1173.21	43.48	120.33	-4.58	3.25	7.75
27	390.49	688.88	1173.45	44.39	121.30	-7.30	1.31	5.34
28	385.47	669.46	1153.85	43.77	116.52	-10.49	3.15	5.61
29	453.47	694.49	1209.04	45.33	125.12	-9.85	2.59	8.48

The MLR QSAR model was developed by applying the selected molecular descriptors to the “active compounds” of the training data (22 compounds).

The best model was selected on the basis of statistical parameters via observed squared correlation coefficient ($R^2 > 0.6$) which is a relative measure of quality of fit. Standard error of estimate (SEE < 0.3) representing absolute measure of quality of fit, Fischer’s value (F), is the Fisher ratio, reflects the ratio of the variance explained by the model and the variance due to the error in the regression. High values of the *F*-test indicate that the model is statistically significant [24].

Pearson’s correlation matrix has been performed on all descriptors by using SPSS statistics 21 Software. The analysis of the matrix revealed ten descriptors for the development of MLR model.

The correlation between the biological activities and descriptors expressed by the following relation:

$$pIC50 = -50.585 - 0.002SAG + 0.065H - 0.260 \text{Log } P + 0.128MD + .001ET - 49.366qN1 - 11.471qC2 + 14.051qN3 + 107.556qC4 + 11.553qC5 \quad (7)$$

$$N=22 \quad R=0.986 \quad SEE=0.110 \quad F=39.341 \quad Q=8.963$$

where N is the number of compounds (training set). The model shows a good correlation coefficient (*R*) of 0.986 between descriptors (Log *P*, SAG, HE, MD, ET, qN₁, qC₂, qN₃, qC₅ and qC₄).

In this model the values of qC₄, qN₃, qC₅, ET, MD and HE suggest that the activity increases with the increase of these descriptors values, on the other hand the activity decreases with increasing the values of qN₁, qC₂, log *P* and SAG. Since they have a negative value in this equation (7).

Table 2. Continued.

Compound	ET a.u	ΔE a.u	qN1	qC2	qN3	qC4	qC5	qC6
1	-1160.695	0.234	-0.581	0.469	-0.558	0.460	-0.406	0.455
2	-1198.813	0.162	-0.593	0.466	-0.585	0.453	-0.399	0.469
3	-1277.461	0.234	-0.588	0.474	-0.592	0.460	-0.396	0.463
4	-1238.144	0.241	-0.593	0.475	-0.592	0.449	-0.394	0.463
5	-1258.010	0.239	-0.604	0.476	-0.577	0.454	-0.375	0.591
6	-1251.789	0.242	-0.597	0.474	-0.592	0.455	-0.388	0.455
7	-1219.888	0.237	-0.597	0.472	-0.542	0.451	-0.378	0.593
8	-1352.677	0.240	-0.608	0.475	-0.578	0.453	-0.374	0.591
9	-1259.211	0.237	-0.595	0.473	-0.542	0.451	-0.378	0.594
10	-1330.400	0.242	-0.592	0.467	-0.593	0.459	-0.391	0.460
11	-1252.970	0.247	-0.583	0.465	-0.592	0.464	-0.399	0.456
12	-1865.431	0.251	-0.601	0.466	-0.579	0.455	-0.398	0.464
13	-1031.279	0.244	-0.557	0.468	-0.570	0.458	-0.336	0.282
14	-1100.770	0.052	-0.557	0.464	-0.564	0.458	-0.332	0.279
15	-1519.949	0.244	-0.556	0.461	-0.566	0.458	-0.324	0.276
16	-991.024	0.251	-0.613	0.648	-0.611	0.464	-0.382	0.288
17	-952.905	0.252	-0.612	0.645	-0.613	0.464	-0.380	0.290
18	-1026.712	0.255	-0.612	0.645	-0.611	0.464	-0.380	0.290
19	-912.333	0.250	-0.617	0.645	-0.612	0.464	-0.383	0.289
20	-1575.339	0.253	-0.547	0.460	-0.536	0.460	-0.338	0.279
21	-1614.662	0.244	-0.550	0.462	-0.532	0.456	-0.337	0.280
22	-1024.280	0.052	-0.556	0.459	-0.557	0.460	-0.349	0.292
23	-948.055	0.245	-0.548	0.453	-0.562	0.454	-0.341	0.291
24	-951.687	0.247	-0.620	0.644	-0.612	0.464	-0.382	0.288
25	-1143.499	0.229	-0.428	0.362	-0.424	0.462	-0.237	0.464
26	-1480.623	0.226	-0.576	0.471	-0.549	0.451	-0.387	0.463
27	-1258.055	0.230	-0.585	0.466	-0.588	0.447	-0.390	0.465
28	-1235.762	0.241	-0.579	0.470	-0.598	0.454	-0.384	0.463
29	-1572.897	0.233	-0.580	0.467	-0.578	0.448	-0.400	0.472

The cross-validated $R_{CV}^2(Q^2)$ was found to be very close to the value of R_{CV}^2 for the training set and hence this model can be termed as statistically significant. The QSAR model expressed by (Eq.7) were cross validated by the high value of $R_{CV}^2=0.973$ obtained by leave one out (LOO).The value of $R_{CV}^2(Q^2)$ is indeed greater than 0.5 which is the essential condition to qualify a QSAR model as valid [22], and RMSE of cross-validation was 0.078, which indicates reliability of the proposed model.

Low value of standard error of estimate (<0.3) indicates the accuracy of the statistical fit. All the

values of the *t*-statistic are significant which confirms the significance of each descriptor.

The calculated *F* value for the generated QSAR model exceeds the tabulated *F* value by large margin as desired for a meaningful regression. Furthermore, the calculated *F* Value also determines a confidence limit superior to 95% for this model. The positive value of quality factor (Q) for QSAR model suggests its high predictive power and lack of over fitting, low standard deviation of the model demonstrates accuracy of the model.

Table 3. Cross-validation parameters.

Model	PRESS	SSY	PRESS/SSY	S_{PRESS}	R_{cv}^2	R_{adj}^2	6PE
	0.135	4.963	0.027	0.077	0.973	0.948	0.023

The present QSAR study can be successfully applied to predict inhibitor activity of aminopyrimidine in these molecule generations. To investigate the presence of a systematic error in developing the QSAR models, in the figure 1 are shown the residuals of predicted values of the

biological activity pIC50 which are plotted against the experimental values. The propagation of the residuals on both sides of zero indicates that no systematic error exists, as suggested by Jalali-Heravi and Kyani [26].

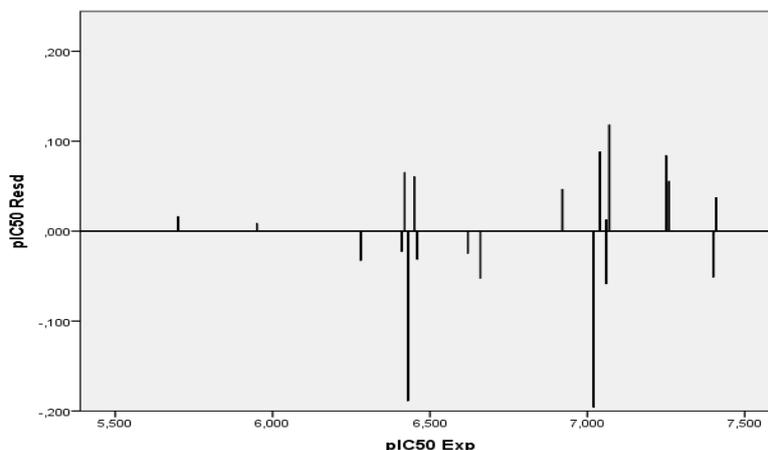


Fig. 1. Plots of the residual values against the experimentally observed.

In order to test the validity of the predictive power of selected MLR model (Eq.7), the leave-one out technique (LOO technique) was used. The developed models were validated by calculation of the following statistical parameters: predicted residual sum of squares (PRESS), total sum of squares deviation (SSY), the predictive error of the coefficient of correlation (PE) and cross-validated correlation coefficients (R^2_{adj} and R^2_{cv}) (Table 3).

PRESS is an important cross-validation parameter as it is a good approximation of the real predictive error of the models. Its value being less than SSY points out that the model has a good predictive power and can be considered statically significant. The smaller PRESS value means the better model predictability. From the results reported in Table 3, this model is statistically significant. Furthermore, for reasonable QSAR model, the PRESS/SSY ratio should be lower than 0.4. [25]

The data presented in Table 3 indicate that for the developed model this ratio is 0.027. The resulted value of R^2_{CV} for these QSAR model is 0.973. The high value of R^2_{CV} and R^2_{adj} are essential criteria for the best qualification of the QSAR model.

The predictive error of the coefficient of correlation (PE) is yet another parameter used to evaluate the predictive power of the proposed models. We have calculated the PE value of the proposed model and they are reported in Table 3. For the developed

model the condition $R = 0.986 > 6PE$ is confirmed and hence, they could be considered as good predictive power. Also, for the validation of the QSAR model we went through the test set via training set. The test set contained seven compounds.

The model (Eq. 7) also passed successfully Tropsha's[23] recommended tests for predictive ability:

$$R^2_{pred} = 0.778 > 0.6$$

$$\frac{(R^2 - R_0^2)}{R^2} = -0.225 < 0.1$$

$$\frac{(R^2 - R_0'^2)}{R^2} = -0.218 < 0.1$$

$$k = 1.014 \quad k' = 0.983 \quad (0.85 \leq k \leq 1.15)$$

$$|R_0^2 - R_0'^2| = 0.006 < 0.3$$

These results might be considered as an indicator of good external predictability.

The built model produced good results for the training set and the test set. It is noteworthy, that the MLR equation has acceptable quality and can predict the activity of training and test set with $R^2_{CV} = 0.973$ and $R^2_{pred} = 0.778$, respectively.

The plots of the predicted pIC50 versus the experimental pIC50, obtained by the MLR modeling, are demonstrated in Figure 2. Therefore; we conclude that the antitubercular activity is related to the physicochemical molecular descriptors and Quantum descriptors.

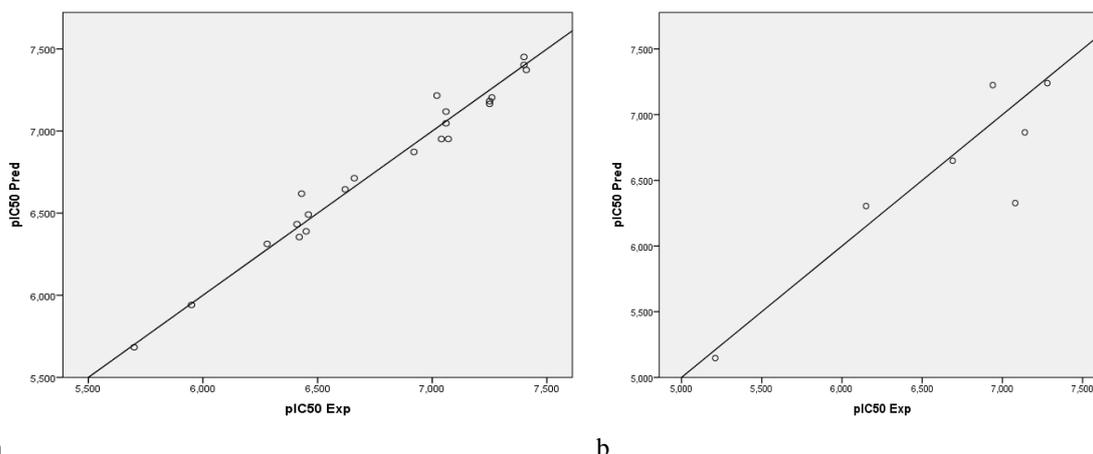


Fig. 2. Data fitness plot of Best Multiple linear regression (a) for the training set, (b) for the test set

4. Conclusion

The QSAR analysis was conducted with a series of 29 amino-pyrimidine derivatives mycobacterium tuberculosis (PknB) inhibitors, the model depending on the (eq.7). MLR is the best produced model with very good statistical fit as evident, $R = 0.986$ and $F=39.341$. The physicochemical molecular and Quantum descriptors were found to have a key role in governing the change in activity. The model was validated using LOO cross-validation, and external test set. The robustness of the constructed model used in this study has great

predictive power, as assessed by the internal and external validations. It indicates the model can be successfully applied to predict the antitubercular activity of these classes of molecules. The applicability domain served as a valuable tool to filter out “dissimilar” compounds. Due to its predictive ability, the proposed model could be a useful aid to the costly and time consuming experiments for determining antitubercular activity. The proposed QSAR workflow aims to help researchers to design a novel chemistry driven molecules with desired biological activity.

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