Plasma visfatin concentrations in hypothyroid patients and its relationship with thyroid autoimmunity and atherosclerosis

Hipotiroid hastalarda plazma visfatin konsantrasyonlarının tiroid otoimmünitesi ve ateroskleroz ile ilişkisi

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Abstract

Aim: Visfatin/pre-B cell enhancement factor stimulates the production of inflammatory cytokines released from adipose tissue such as TNF-α and IL-6, which are shown to be related to the pathogenesis of insulin resistance, diabetes, dyslipidemia, inflammation, autoimmune diseases, and atherosclerosis. Our aim was to evaluate plasma visfatin concentrations in patients with autoimmune hypothyroidism and its relationship with thyroid autoimmunity and atherosclerosis.

Material and Method: Thirty-five patients with newly diagnosed Hashimoto’s thyroiditis (32 women/3 men, mean age 43.8±9.6 years) and 18 healthy controls (17 women/1 men, mean age 43.3±5.2 years) were enrolled in the study. Anthropometric measurements, carotid intima-media thickness (CIMT), serum anti-Tg, anti-TPO, hsCRP, homocysteine, lipo(a), ApoA, ApoB1, β-2 microglobulin, insulin, glucose, visfatin, IL6, TNF-α, oxidized-LDL concentrations, lymphocyte subgroups and lipid profile were investigated both before and after thyroid hormone replacement therapy. Serum visfatin concentrations were determined by enzyme-linked immunosorbent assay (ELISA).

Results: Mean serum visfatin concentrations were similar between autoimmune hypothyroidism group and control group (6.29 ± 1.63 ng/ml to 6.36 ± 1.51 ng/ml, p>0.05) and it was not correlated with thyroid autoimmunity and atherosclerosis markers. Plasma visfatin, oxidized LDL, IL-6, and TNF-α concentrations did not differ statistically between the control group and hypothyroid patients both before and after the therapy. The cardiovascular risk factors for systolic and diastolic blood pressure, HOMA-IR index, triglyceride, Apo B and ApoB/ApoA, homocysteine, β-2 microglobulin, and CIMT were found to be elevated in patients. Ox-LDL, cholesterol, homocysteine, and proteinuria concentrations were positively correlated with anti-Tg concentrations.

Conclusion: Serum visfatin concentrations were not associated with thyroid autoimmunity and atherosclerosis.

Keywords: Visfatin, autoimmune hypothyroidism, atherosclerosis
Öz

Amaç: Visfatin/pre-B cell enhancement factor; insulin direnci, diyabet, dislipidemi, inflamasyon, otoimmun hasta-
lıklar ve ateroskleroz patogeneziyle ilişkili adipodan salinan TNF-alfa ve IL-6 gibi inflamatuar sitokinlerin
yapımını uyarır. Bu çalışmada amacımız otoimmün hipotiroidizmli hastalarda plazma visfatin düzeyi ve bunun
tiroid otoimmünitesi ve ateroskleroz ile ilişkisini değerlendiriklerdir.

Gereç ve Yöntem: Çalışmamız 35 yeni tanı almış Hashimoto hipotiroidili olgu (32 kadın/3 erkek, ort. yaş 43.8±9.6)
və 18 sağlıklı kontrol (17 kadın/1 erkek, ort. yaş 43.3±5.2) dahil edildi. Tiroid hormon replasmanı öncesi ve sonrası
antropometrik ölçümler, karotid intima-media kalınlığı (KIMK), serum anti-Tg, anti-TPO, hsCRP, homosistein, lipoida, ApoA, ApoB1, β-2 mikroglobulin, insulin, glukoz, visfatin, IL6, TNF-α, okside-LDL, lenfosit subgrupları
ve lipid profile ölçümleri yapıldı. Serum visfatin konsantrasyonları ELISA yöntemiyle çalışıldı.

Bulgular: Serum visfatin konsantrasyonları otoimmün hipotiroidizm grubu ile kontrol grubu arasında benzerdi
(6.29 ± 1.63 ng/ml to 6.36 ± 1.51 ng/ml, p>0.05), tiroid otoimmünitesi ve ateroskleroz belirteçleriyle korelasyonu
saptanmadı. Tedavi öncesi ve sonrası hipotiroidi hastalarının plazma visfatin, okside LDL, IL-6 ve TNF-α kons-
santrasyonları ile sağlıklı kontrol gruba istatistiksel farklılık görülmedi. Sistolik ve diyastolik kan basıncı,
HOMA-IR, trigliserid, Apo B ve ApoA/ApoA, homosistein, β-2 mikroglobulin, KIMK gibi kardiyovasküler risk
faktörleri hasta grubunda yüksek olarak bulundu. Okside-LDL, kolesterol, homosistein ve proteinuria konsanstras-
yonları anti-Tg ile pozitif korele idi.

Sonuç: Serum visfatin konsantrasyonları ile tiroid otoimmünitesi ve ateroskleroz arasında ilişki bulunmadı.

Anahtar Kelimeler: Visfatin, otoimmün hipotiroidi, ateroskleroz

Introduction

“Hashimoto’s thyroiditis” is the most common cause of
hypothyroidism. Loss of immune tolerance and the effect of
antigen-producing cells cause autoreactive cells to invade
the thyroid gland and induce apoptosis of thyrocytes [1].

Hypothyroidism increases the risk of cardiovascular disease
[2]. Visfatin/pre-B cell enhancement factor, found in visceral
adipose tissue, stimulates the growth of B cell precursors in
the presence of IL-7 and stem cell factor. It is also found in
activated lymphocytes, monocytes and neutrophils, bone
marrow, skeletal muscles and the liver [3]. Visfatin is a
cytokine produces the production of proinflammatory cytokines
including IL-6, TNF-alfa and IL-1[4]and it was found to be
elevated in sepsis, autoimmune disorders and in low-grade
inflammation, i.e. atherosclerosis [5]. Visfatin concentrations
were demonstrated to be increased in several autoimmune
diseases such as rheumatoid arthritis, systemic lupus
erythematosus, and psoriasis [6-8]. Both hypothyroidism and
hyperthyroidism could affect serum visfatin concentrations
however, the results are controversial [9].

In this study, we hypothesized that visfatin/pre-B cell
enhancement factor as an adipocytokine released from
visceral adipose tissue might provide a bridging function
between thyroid autoimmunity and atherosclerosis in
autoimmune hypothyroidism. Therefore, we aimed to
evaluate plasma visfatin concentrations in patients with
autoimmune hypothyroidism and its relationship with
thyroid autoimmunity and atherosclerosis.

Material and Method

We enrolled 35 patients with newly diagnosed Hashimoto’s
thyroiditis (32 women/3 men, mean age 43.8±9.6 years)
and 18 healthy controls (17 women/1 men, mean age
43.3±5.2 years) in the study. Ethics committee permission
was obtained from Eskişehir Clinical Research Ethics
Committee and informed consent was provided for all
participants.

Hashimoto’s thyroiditis was determined as having the high
anti-Tg (> 115 IU/ml) and/or anti-TPO (> 34 IU/ml) values
and heterogeneous thyroid parenchyma and reduction in
echogenicity on thyroid ultrasonography. Subjects
with endocrinologic disease (Diabetes mellitus, Cushing syndrome, Addison’s disease, etc.), clinically obvious atherosclerotic vascular disease, hypertension, hepatic or renal dysfunction, infectious diseases, previous diagnosis of Hashimoto’s disease and/or another thyroid disease who is on treatment, normal concentrations of thyroid auto-antibodies diagnosed with autoimmune disease apart from Hashimoto’s thyroiditis, drug use such as medications that disrupt efficacy of L-thyroxine treatment (cholestyramine, furosemide, coumadin, proton pump inhibitor, etc.), steroid or immunosuppressive treatment were excluded.

Height, weight, waist and hip circumference, systolic and diastolic blood pressures of all participants were measured. All patients right and left carotid intima-media thickness were determined using Toshiba SSA - 240 (Toshiba, Tokyo, Japan) ultrasound with 7.5 MHz linear array transducer by the same radiologist in the radiology department. Venous blood samples were taken from an antecubital vein before and after treatment (at 8th week) after an 8-hour overnight fasting period. After blood samples were centrifuged at 4000 x g for 5 minutes, the following tests were performed on the same day at Eskişehir Osmangazi University Faculty of Medicine Biochemistry laboratories, using Roche original kits on a Roche Modular autoanalyzer: serum TSH (µIU/ml), free T3 (pg/ml), free T4 (ng/ml), anti-Tg antibody, anti–TPO antibody, hsCRP (mg/L), homocysteine (µmol/L), lipid profile, lipo(a) (mg/dl), ApoA (mg/dl), ApoB1 (mg/dl), beta 2 microglobulin (ng/ml), insulin (µU/ml), and glucose (mg/dl). To evaluate visfatin (ng/ml), IL-6 (pg/ml), TNF-alpha (pg/ml) and oxidized-LDL (ng/ml) concentrations, serum samples were stored at -80 °C until analysis. Fibrinogen was studied at ESOGU hematology laboratory using a Fibrinogen – C XL (HemosIL, USA) kit on an ACL TOP device, with lymphocyte subgroups (CD4, CD8, CD19, CD56) studied using Diaclone antibody on a Becton Dickinson FACS Calibur device in the same day blood samples were taken.

L-thyroxine treatment half an hour before breakfast was started to all cases newly diagnosed with hypothyroidism. During follow-up, patients were asked whether they regularly use the medication or use any medications that could affect treatment. After achievement of euthyroidism, blood samples were taken to check the same parameters evaluated before the beginning of therapy. HOMA-IR index developed by Matthews et al. was used (fasting blood sugar (mg/dl) X insulin (µU/ml)/405) for diagnosis of insulin resistance. HOMA index values above 2.7 are accepted as the presence of insulin resistance [10].

**Measurement of Serum Visfatin**

The concentration of Visfatin was measured using enzyme-linked immunosorbent assay (ELISA) method. Commercially available human Visfatin ELISA kit (Phoenix Pharmaceuticals, USA) was used. The procedure for the ELISA method was carried out according to the instructions provided by the manufacturer. The concentrations of Visfatin are presented as ng/ml.

**Statistical Analysis**

All statistical analysis was performed by using SPSS Version 13.0 (SPSS, Inc, Chicago, IL, USA). Data were expressed as a mean ± standard deviation. Data with normal distribution before and after treatment were assessed by using the sample paired t-test, while data with non-normal distribution were assessed by the Wilcoxon t-test. Correlations were analyzed with Pearson test. P values of less than 0.05 were accepted as statistically significant.

**Results**

Demographics and laboratory values of patients and control group were shown in Table 1. Mean age and BMI were similar between groups (p>0.05). ApoB, ApoB/ApoA1, homocysteine and beta 2-microglobulin concentrations and mean CIMT was significantly higher in the patient group before treatment when compared with healthy controls (p<0.05) (Table 1). Mean serum visfatin concentrations were similar between autoimmune hypothyroidism group and control group (6.29 ± 1.63 ng/ml to 6.36 ± 1.51 ng/ml, p>0.05) and it was not correlated with thyroid autoimmunity and atherosclerosis markers. Serum IL-6, TNF-α and oxidized-LDL concentrations measured before treatment in hypothyroid patients were compared with control group and statistically significant difference was not found (p>0.05) (Table 2).
Table 1: Demographics and laboratory values of patients and control group

<table>
<thead>
<tr>
<th></th>
<th>Patients group (n:35)</th>
<th>Control group (n:18)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Age (year)</strong></td>
<td>43.85 ± 9.68</td>
<td>43.38 ± 5.27</td>
<td>0.850</td>
</tr>
<tr>
<td><strong>BMI (kg/m²)</strong></td>
<td>28 ± 5</td>
<td>25 ± 4</td>
<td>0.057</td>
</tr>
<tr>
<td><strong>SBP (mmHg)</strong></td>
<td>120.14 ± 17.42</td>
<td>107.22 ± 8.26</td>
<td>0.005</td>
</tr>
<tr>
<td><strong>DBP (mmHg)</strong></td>
<td>77.85 ± 13.24</td>
<td>67.77 ± 10.60</td>
<td>0.007</td>
</tr>
<tr>
<td><strong>FT3 (pg/ml)</strong></td>
<td>3.03 ± 0.90</td>
<td>3.38 ± 0.50</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>FT4 (ng/dl)</strong></td>
<td>0.77 ± 0.24</td>
<td>1.01 ± 0.12</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>TSH (µIU/ml)</strong></td>
<td>30.41 ± 44.3</td>
<td>1.72 ± 0.83</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>FPG (mg/dl)</strong></td>
<td>91.85 ± 11.07</td>
<td>82.05 ± 9.63</td>
<td>0.002</td>
</tr>
<tr>
<td><strong>Insulin (µU/ml)</strong></td>
<td>7.06 ± 7.87</td>
<td>3.20 ± 1.31</td>
<td>0.045</td>
</tr>
<tr>
<td><strong>HOMA-IR</strong></td>
<td>1.52 ± 1.73</td>
<td>0.62 ± 0.32</td>
<td>0.002</td>
</tr>
<tr>
<td><strong>TG (mg/dl)</strong></td>
<td>131.91 ± 57.58</td>
<td>96.38 ± 39.47</td>
<td>0.023</td>
</tr>
<tr>
<td><strong>HDL-C (mg/dl)</strong></td>
<td>49.80 ± 10.63</td>
<td>53.72 ± 11.53</td>
<td>0.222</td>
</tr>
<tr>
<td><strong>LDL-C (mg/dl)</strong></td>
<td>125.60 ± 36.99</td>
<td>121.94 ± 26.75</td>
<td>0.712</td>
</tr>
<tr>
<td><strong>Total cholesterol</strong></td>
<td>199.09 ± 49.39</td>
<td>195.79 ± 31.83</td>
<td>0.775</td>
</tr>
<tr>
<td><strong>Visfatin (ng/ml)</strong></td>
<td>9.20 ± 23.36</td>
<td>84.70 ± 16.46</td>
<td>0.023</td>
</tr>
<tr>
<td><strong>Ox LDL (ng/ml)</strong></td>
<td>94.62 ± 10.63</td>
<td>53.72 ± 11.53</td>
<td>0.222</td>
</tr>
<tr>
<td><strong>ApoB/ApoA1</strong></td>
<td>0.61 ± 0.18</td>
<td>0.50 ± 0.11</td>
<td>0.027</td>
</tr>
<tr>
<td><strong>ApoB/ApoA1</strong></td>
<td>0.72 ± 0.22</td>
<td>0.58 ± 0.02</td>
<td>0.015</td>
</tr>
<tr>
<td><strong>Mean CIMT</strong></td>
<td>9.99 ± 3.63</td>
<td>8.27 ± 3.44</td>
<td>0.009</td>
</tr>
<tr>
<td><strong>Homocysteine</strong></td>
<td>1503.71 ± 510.75</td>
<td>1486.83 ± 282.23</td>
<td>0.029</td>
</tr>
</tbody>
</table>

Comparisons of plasma glucose, insulin, lipid parameters, homocysteine and lipoprotein concentrations before and after treatment were shown in Table 3. ApoB, homocysteine and ApoB/ApoA1 concentrations before treatment were significantly higher than values measured after treatment (p<0.05) (Table 3). Serum visfatin, TNF-α, IL-6, and oxidized-LDL were similar between before and after treatment (p>0.05) (Table-4).

Table 2: The comparison of adipocytokines and oxidised-LDL cholesterol concentrations between groups

<table>
<thead>
<tr>
<th></th>
<th>Patients Group (n=35)</th>
<th>Control Group (n=18)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Visfatin (ng/ml)</strong></td>
<td>6.29 ± 1.63</td>
<td>6.36 ± 1.51</td>
<td>P = 0.600</td>
</tr>
<tr>
<td><strong>Ox LDL (ng/ml)</strong></td>
<td>111.25 ± 156.16</td>
<td>83.83 ± 68.38</td>
<td>P = 0.077</td>
</tr>
<tr>
<td><strong>TNF-α (pg/ml)</strong></td>
<td>5.24 ± 3.08</td>
<td>4.43 ± 1.78</td>
<td>P = 0.225</td>
</tr>
<tr>
<td><strong>IL-6 (pg/ml)</strong></td>
<td>4.32 ± 2.65</td>
<td>3.92 ± 1.12</td>
<td>P = 0.199</td>
</tr>
</tbody>
</table>

Table 3: Comparisons of plasma glucose, insulin, lipid parameters, homocysteine and lipoprotein concentrations before and after treatment

<table>
<thead>
<tr>
<th></th>
<th>Before Treatment (n=35)</th>
<th>After Treatment (n=35)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>FT3 (pg/ml)</strong></td>
<td>3.03 ± 0.90</td>
<td>3.56 ± 0.49</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>FT4 (ng/dl)</strong></td>
<td>0.77 ± 0.24</td>
<td>1.32 ± 0.29</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>TSH (µIU/ml)</strong></td>
<td>30.41 ± 44.3</td>
<td>0.78 ± 0.59</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td><strong>FPG (mg/dl)</strong></td>
<td>91.8 ± 11.07</td>
<td>90.4 ± 10.8</td>
<td>0.440</td>
</tr>
<tr>
<td><strong>Insulin (µU/ml)</strong></td>
<td>7.0 ± 7.88</td>
<td>4.7 ± 2.8</td>
<td>0.069</td>
</tr>
<tr>
<td><strong>HOMA-IR</strong></td>
<td>1.5 ± 1.7</td>
<td>1.0 ± 0.7</td>
<td>0.125</td>
</tr>
<tr>
<td><strong>Total cholesterol</strong></td>
<td>199.08 ± 49.39</td>
<td>190.71 ± 35.17</td>
<td>0.249</td>
</tr>
<tr>
<td><strong>LDL-C (mg/dl)</strong></td>
<td>125.60 ± 36.99</td>
<td>118.00 ± 33.36</td>
<td>0.170</td>
</tr>
<tr>
<td><strong>HDL-C (mg/dl)</strong></td>
<td>49.80 ± 10.63</td>
<td>48.00 ± 10.98</td>
<td>0.196</td>
</tr>
<tr>
<td><strong>TG (mg/dl)</strong></td>
<td>131.91 ± 57.58</td>
<td>120.60 ± 44.70</td>
<td>0.087</td>
</tr>
<tr>
<td><strong>ApoB (mg/dl)</strong></td>
<td>94.20 ± 23.36</td>
<td>87.37 ± 21.60</td>
<td>0.023</td>
</tr>
<tr>
<td><strong>ApoB/ApoA1</strong></td>
<td>0.61 ± 0.18</td>
<td>0.56 ± 0.11</td>
<td>0.006</td>
</tr>
<tr>
<td><strong>Homocysteine</strong></td>
<td>9.99 ± 3.63</td>
<td>8.54 ± 2.79</td>
<td>0.009</td>
</tr>
</tbody>
</table>


Table 4: Comparisons of adipocytokines and oxidised-LDL cholesterol concentrations before and after treatment

<table>
<thead>
<tr>
<th></th>
<th>Before Treatment (n=35)</th>
<th>After Treatment (n=35)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Visfatin (ng/ml)</strong></td>
<td>6.29 ± 1.63</td>
<td>6.08 ± 1.85</td>
<td>0.600</td>
</tr>
<tr>
<td><strong>Ox LDL (ng/ml)</strong></td>
<td>111.25 ± 156.16</td>
<td>143.53 ± 251.95</td>
<td>0.077</td>
</tr>
<tr>
<td><strong>TNF-α (pg/ml)</strong></td>
<td>5.24 ± 3.08</td>
<td>7.10 ± 7.98</td>
<td>0.225</td>
</tr>
<tr>
<td><strong>IL-6 (pg/ml)</strong></td>
<td>4.32 ± 2.65</td>
<td>17.86 ± 6.97</td>
<td>0.199</td>
</tr>
</tbody>
</table>

Separate comparisons of lymphocyte subgroups in the patient group before treatment were similar with the control group and they were not affected by levothyroxine treatment (Table 5).

Table 5: The comparison of lymphocyte subgroups before and after treatment with control group

<table>
<thead>
<tr>
<th></th>
<th>Before Treatment (n=35)</th>
<th>After Treatment (n=35)</th>
<th>Control group (n=18)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>CD4+ (%)</strong></td>
<td>43.07 ± 5.90</td>
<td>43.60 ± 5.09</td>
<td>45.89 ± 8.72</td>
<td>&gt; 0.05</td>
</tr>
<tr>
<td><strong>CD8+ (%)</strong></td>
<td>26.67 ± 7.69</td>
<td>26.70 ± 8.03</td>
<td>25.01 ± 5.17</td>
<td>&gt; 0.05</td>
</tr>
<tr>
<td><strong>CD19+ (%)</strong></td>
<td>10.47 ± 3.51</td>
<td>9.97 ± 2.96</td>
<td>10.53 ± 3.09</td>
<td>&gt; 0.05</td>
</tr>
<tr>
<td><strong>CD56+ (%)</strong></td>
<td>13.20 ± 6.28</td>
<td>13.11 ± 6.52</td>
<td>13.72 ± 6.29</td>
<td>&gt; 0.05</td>
</tr>
<tr>
<td><strong>CD4+/CD8+ (%)</strong></td>
<td>1.76 ± 0.72</td>
<td>1.74 ± 0.62</td>
<td>1.90 ± 0.67</td>
<td>&gt; 0.05</td>
</tr>
</tbody>
</table>

*Both for Before treatment versus control group and before treatment versus after treatment
Anti-thyroglobulin antibody (anti-Tg) concentrations were positively correlated with Ox-LDL ($r=0.373$, $p=0.027$), total cholesterol ($r=0.365$, $p=0.031$), HDL-C ($r=0.347$, $p=0.041$), homocysteine ($r=0.390$, $p=0.021$).

**Discussion**

A variety of studies has evaluated the relationship between visfatin and thyroid dysfunction. Caixas et al. [11] investigated plasma visfatin concentrations in hyperthyroidism and hypothyroidism both before and after treatment. The patients with hyperthyroidism had higher visfatin concentrations which increased after treatment. Hypothyroid patients had higher visfatin concentrations, which further increased after treatment regardless of changes in anthropometric and insulin resistance parameters. They could not find a correlation between visfatin and any other parameters. Ozkaya et al. [12] also evaluated the serum visfatin concentrations in hypothyroidism and hyperthyroidism before and after treatment. The patients with hyperthyroidism had lower visfatin concentrations compared with the hypothyroid group and controls. A significant degree of decrease in plasma visfatin concentrations was observed following treatment of patients with hypothyroidism. In patients with hyperthyroidism, an increase in plasma visfatin concentrations was observed with anti-thyroid treatment. A positive correlation between visfatin and TSH concentrations and a negative correlation between visfatin concentrations and fT3 and fT4 values were observed. Han et al. [13] examined the regulation of visfatin by thyroid hormones both in vivo and in-vitro and they found both hyperthyroidism and hyperthyroidism were associated with elevated visfatin concentrations. Sawicka-Gutaj et al. [14] investigated whether coexisting autoimmune inflammation in hypothyroidism could have an effect on serum visfatin concentrations or not. They demonstrated that visfatin serum concentration in hypothyroid patients is associated with both autoimmunity and free thyroid hormones level. In our study, serum visfatin concentrations were similar between autoimmune hypothyroidism group and control group and, it was not correlated with thyroid autoimmunity and atherosclerosis markers. We also could not find a difference in visfatin concentrations by levothyroxine treatment. The idea of a possible involvement of visfatin in the pathogenesis of autoimmune thyroiditis was not suggested by our study. As the measurement of these values was completed 6-8 weeks after the beginning of treatment, we still do not know whether the visfatin concentrations are affected by treatment in the long term or not. Together with this, our results showed that visfatin concentrations were not affected by the hypothyroid situation, and as it is secreted by adipose tissue, it may have similar concentrations to healthy controls with similar BMI.

In many studies, clinical or subclinical hypothyroidism has been demonstrated as a risk factor for atherosclerosis and cardiovascular disease [15]. Increased concentrations of LDL-C, VLDL-C, HDL-C, ApoB and lipoprotein (a) are seen in patients with hypothyroidism and thyroid hormone replacement provides a decrease in plasma cholesterol, ApoB, homocysteine and ApoB/ApoA1 concentrations [16]. Studies have shown plasma visfatin concentrations are associated with BMI, diabetes mellitus, metabolic syndrome and coronary artery disease [17-20]. Dahl et al. [4] stated that visfatin may have a direct role in atherosclerosis and plaque destabilization. In our study, ApoB, ApoB/ApoA1, homocysteine and beta 2-microglobulin concentrations and mean CIMT were significantly higher in the patient group before treatment. ApoB, homocysteine and ApoB/ApoA1 concentrations were decreased by levothyroxine treatment. LDL-C, total cholesterol, TG concentrations were higher in the patient group before treatment. Although there is a trend towards a decrease in LDL-C, total cholesterol, TG concentrations by levothyroxine treatment, these concentrations were statistically not affected by levothyroxine treatment. These findings are consistent with the literature.

A positive correlation was found between anti-TG antibody and ox-LDL, cholesterol, homocysteine, and proteinuria. Based on these findings mainly non-traditional CVD risk factors are increased in hypothyroidism linked to Hashimoto’s thyroiditis, accompanied by an increase in the atherosclerosis finding of CIMT, leading to the consideration that in anti-TG antibodies may have a bridging function between autoimmunity and atherosclerosis.
In hypothyroidism, another risk factor for CVD has increased the prevalence of obesity. The increased adipose tissue could lead to a release of adipocytokines such as leptin, adiponectin, resistin, visfatin, TNF-α, and IL-6 [21]. Serum visfatin concentrations were shown to be increased in several autoimmune diseases such as rheumatoid arthritis or inflammatory bowel disease [22]. Visfatin stimulates the production of proinflammatory cytokines such as IL-6 and tumor necrosis factor-α which were higher in patients with autoimmune thyroid dysfunction [23,24]. IL-6 can stimulate the expression of visfatin [25]. Moreover, it can be postulated that visfatin release from adipose tissue might be influenced directly or indirectly by proinflammatory cytokines which have been implicated in thyroid dysfunction. Oxidized-LDL is a marker with a significant role in the formation of atherosclerotic plaque and pathogenesis of atherosclerosis. Many studies have found that ox-LDL concentrations are high in individuals with coronary artery disease [26]. However, serum visfatin, IL-6, TNF-α and oxidized-LDL concentrations were similar in patients with hypothyroidism and control group in our study. Additionally, serum TNF-α, IL-6, and oxidized-LDL were not affected by levothyroxine treatment. These findings could be explained by similar BMI in patients and control groups, many of our patients were not obese.

The mechanisms of the relationship between visfatin and insulin resistance are not clear. Studies have not found a correlation between visfatin and HOMA-IR [27,28]. In our study, there was no correlation between serum visfatin concentrations and HOMA-IR index. Carotid artery intima-media thickness (CIMT) is shown to have a clear correlation with all risk factors for atherosclerosis [29]. In our study, CIMT was higher in patients with hypothyroidism however, we did not find a correlation between visfatin concentrations and CIMT.

Atherosclerotic human and animal studies demonstrated autoantibodies against ox-LDL in circulation and immunoglobulins in atherosclerotic lesions lead to the consideration of B-lymphocytes might play a role in the pathogenesis of atherosclerosis [30]. Since subgroups of T-lymphocytes are related to autoimmune events, there are changes in T-lymphocyte subgroups in these events. There are contradictory data related to an association between numbers of T-lymphocyte subgroups found in peripheral blood and autoimmune diseases. A study by Covas et al. [31] of 44 autoimmune thyroid disease patients found a strong effect of functional concentrations of the thyroid hormones on CD3, CD4 and CD8 values which had a negative correlation with fT4 and positive correlation with TSH. The results of the same study observed that CD4/CD8 ratio was decreased in hypothyroidism related to Hashimoto’s thyroiditis and increased in hyperthyroidism related to Graves’ disease. Moschen et al. [32] showed that recombinant visfatin activated human leukocytes and stimulated cytokine production and it increased the surface expression of costimulatory molecules. In our study, we did not find a difference between lymphocyte subgroups evaluated before and after treatment.

A relatively small sample size with a cross-sectional design is the major limitation of our study.

**Conclusion**

Serum visfatin concentrations were not associated with thyroid autoimmunity and atherosclerosis. On the other hand, non-traditional CVD risk factors are higher in hypothyroid patients with Hashimoto’s thyroiditis accompanied by an increase in CIMT measurement and, the anti-Tg antibody might be a bridge between autoimmunity and atherosclerosis. Our findings did not suggest a pathophysiological relationship between thyroid autoimmunity and visfatin which was shown by previous studies. Extensive studies covering larger populations are needed to enlighten the relationship between visfatin and thyroid autoimmunity.

**Declaration of conflicting interests**

The authors declared no conflicts of interest with respect to the authorship and/or publication of this article.

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