ANTIBACTERIAL AND ANTIOXIDANT ACTIVITY OF MYRTUS COMMUNIS L. GROWING WILD IN MARMARIS

Fikret Keven-Karademir^{1*}, Sibel Avunduk²

¹The Vocational College of Datca, Mugla University, Datca /Mugla, Turkey ²Medical Laboratory Techniques Programme, Vocational School of Health Care, Mugla University, Marmaris, Mugla, Turkey

> *Received* / Geliş tarihi: 19.05.2015 *Received in revised form* / Düzeltilerek Geliş tarihi: 30.05.2015 *Accepted* / Kabul tarihi: 03.06.2015

Abstract

Three myrtus fruit samples collected from different regions of Marmaris were dried, grinded and extracted with n-hexane, CH₂Cl₂ and MeOH respectively. The extracts of *Myrtus communis* L. were screened in vitro for their antimicrobial activities using disc diffusion method against four test bacteria. The antimicrobial test results showed that the inhibition zones have been measured between 7 to 16 mm. *S. aureus* was the most sensitive one to all concentrations of all *M. communis* L. samples. *P. aeruginosa* was the most resistant one to all concentrations of *M. communis* L. sample from Yeşil Belde. The antioxidant activity of MeOH extracts has also been determined by DPPH assay. This is the first report of comparative antimicrobial and antioxidant study for *M. communis* L. samples collected from different regions from Marmaris/ Mugla/ Turkey.

Keywords: M. communis L., disc diffusion method, pathogen bacteria, DPPH method, Marmaris

MARMARİSTE YABANİ OLARAK YETİŞEN MYRTUS COMMUNIS L.'NİN ANTİBAKTERİYEL VE ANTİOKSİDAN AKTİVİTESİ

Özet

Marmaris'in üç farklı bölgesinden toplanan *Myrtus* (Mersin ağacı) meyveleri kurutulmuş, öğütülmüş ve sırasıyla, n-heksan, CH₂CI₂ ve MeOH ile ekstrakte edilmiştir. Elde edilen kurutulmuş, ekstraktlar dört farklı test bakterisine karşı disk difüzyon yöntemi kullanılarak in vitro olarak taranmıştır. Antimikrobiyel test sonuçları inhibisyon zonlarının 7-16 mm çapında oluklarını göstermiştir. *S. aureus, M. communis* L. örneklerinin tüm konsantrasyonlarına karşı en duyarlı olan bakteri iken, *P. aeruginosa* ise Yeşil Belde'den toplanan *M. communis* L. örneklerinin bütün konsantrasyonlarına karşı en dirençli bakteri türü olarak belirlenmiştir. MeOH ekstraktlarının antioksidan aktivitesi, DPPH metodu kullanılarak ölçülmüştür. Bu çalışmada, ilk kez Marmaris'in farklı bölgelerinden toplanan *M. communis* L. örneklerinin karşılaştırmalı olarak antimikrobiyel ve antioksidan aktiviteleri saptanmıştır.

Anahtar kelimeler: M. communis L., disk diffüzyon metodu, patojen bakteri, DPPH metodu, Marmaris

^{*}Yazışmalardan sorumlu yazar / Corresponding author;

fkeven@hotmail.com,
(+90) 252 211 13 00,

^{(+90) 252 211 17 37}

INTRODUCTION

Myrtaceae (*Myrtus communis* L.) is an evergreen shrub growing spontaneously throughout the Mediterranean area. It is a typical annual shrub of the Mediterranean countries including Turkey, Greece, Italy, Algeria, Tunisia, and Morocco. In Turkey, myrtle plants are found within the natural pine forests and riversides in the Mediterranean region, particularly in the Taurus Mountains, 500 to 600 m above sea level (1).

In folk medicine, a decoction of leaves and fruits or infusion of myrtle are used for stomachic, hypoglycemic, cough and oral diseases, antimicrobic, for constipation, appetizing, antihaemorrhagic and externally for wound healing (2, 3). In Turkish folk medicine, the leaves and fruits have been used as an antiseptic and for healing wounds as well as in the treatment of urinary diseases (4). Different parts of the plant find various uses in the food industry, such as for flavoring meat and sauces, and in the cosmetic industry (5).

Over the past few years, liqueurs prepared from the berries of myrtle have become popular, (6) while its leaves have been used as a hop substitute in beer and as a cosmetic ingredient in products against hair dandruff (7).

Until now, the majority of studies on myrtle have focused on its volatile fraction (5, 8-18) and of phenolic compounds in leaves and berries (19-27). The leaves contain tannins, flavonoids such as quercetin, catechin and myricetin derivatives and volatile oils (4, 28). The fruits of this plant are mostly composed of volatile oils, tannins, sugars, flavonoids and organic acids such as citric and malic acids (4, 21).

The volatile oil in leaves of *M. communis* L. growing in Turkey contains 1,8-cineole, linalool, myrtenyl acetate and myrtenol as major components (3). In addition, Mansouri et al. (28) reported that a crude methanol extract of *M. communis* L. leaves had potent antibacterial activity against 10 microorganisms, including 6 Gram positive and 4 Gram negative bacteria. A few researches have undertaken the antioxidant activity of myrtle leaf essential oil (28) and extract (30-33).

The main objectives of this study were to investigate the antimicrobial activity of the extracts obtained from *Myrtus communis* L. berries by disc diffusion method against some pathogen bacteria.

As far as our literature survey could ascertain, our study is different from the previous reports on this plant in terms of plant material collected from different region to evaluate regional variety and the extracts obtained by using different solvents.

MATERIAL AND METHODS

Collection of plant material

M. communis L. berries were collected from Bozburun, Çetibeli and Yeşil Belde; Marmaris, Turkey.

Extraction of M. communis L. berries

The dried and grinded berries (25 g of samples) were extracted with n-Hexane (500ml), CH_2Cl_2 (500ml) and MeOH (500ml) using soxlet apparatus respectively. The extracts of berries were evaporated to dryness in vacuum at 50 oC. The yields (%, w/w dry plant material) of dry extracts are presented in Table 1.

Locality	Extract yield (%, w/w)								
	n-Hexane	CH ₂ Cl ₂	MeOH						
Bozburun	1.51	0.71	77.93						
Çetibeli	2.24	1.47	73.67						
Yeşil Belde	2.01	0.68	91.88						

Antimicrobial assay

Staphylococcus aureus, Klebsiella pneumoniae, Escherichia coli and Pseudomonas aeruginosa were used as test bacteria. Bacteria inoculates were prepared by growing cells in Nutrient broth (Merck) for 24 h at 37 °C (34). These cells suspensions were diluted with peptone water to provide initial cell counts of about 105- 106 cfu/ml. The extracts (all extracts were filter-sterilized using a 0.45 µm membrane filter) were prepared at 1%; 2.5%; 5% and 10% concentrations in correspond solvent. 17 ml sterile Mueller-Hinton agar at 45 °C and poured into Petri dishes (9 cm in diameter). Then the agars were allowed to solidify at 4 °C for 1 h. Test bacteria were spread on Muller Hinton Agar. Sterile paper disk of 6 mm diameter were impregnated with this solutions (30 µl) (35). These impregnated disks were applied on solid agar medium in Petri dishes. The treated Petri dishes were left 10-15 minutes at room temperature and then incubated 37 ± 0.1 °C for 24-48 hours. After the incubation period inhibition zones were measured in millimeters. These experiments were carried out in duplicate (36).

Antioxidant activity

DPPH assays/TLC autographic assay

After developing and drying the TLC plates (samples ranging from 0.1 to 100µg) were sprayed with 0.2% (2mg/ml) of DPPH solution in methanol. The plates were examined half an hour after spraying. Active compounds appeared as yellow spots against a purple background. (37-41).

Antioxitant capacity

One ml of 500 μ M (0.2 mg/ml) DPPH in methanol was mixed with equal volumes of test compounds at various concentrations, mixed well and kept in the dark for 30 min. The absorbance at 517 nm was monitored in the presence of different concentrations of the samples. Blank experiment was also carried out, with just solvent and DPPH (i.e., 2 ml of 500 μ M in methanol), to determine the absorbance of DPPH before interacting with the compounds. The amount of sample in μ g/ml at which the absorbance at 517 nm decreases to half its initial value was used as the IC₅₀ value of the MeOH extracts (36, 42, 43).

The samples were done in triplicate and the mean value of three was recorded.

RESULTS AND DISCUSSION

The antibacterial activities of *M. communis* L extracts at different concentrations in vitro test against different pathogenic bacteria were shown in Table 2, 3 and 4.

The inhibition zones were varied related to different concentrations of *M. communis* L extracts. Our results have shown remarkable antimicrobial activity for the n-hexane extract of *M. communis* L (especially BH and ÇH) against all microorganisms tested with inhibition zones.

Table 2: The antimicrobial activity (diameters of growth inhibition zones) of the crude extracts of three *M. communis* L. samples from Bozburun

Microorganisms	Extracts	S										
		n-Hexa	ane (BH)		CH ₂ Cl ₂ (BC)				MeOH (BM)			
	10%	5%	2.5%	1%	10%	5%	2.5%	1%	10%	5%	2.5%	1%
S. aureus	14	14	14	12	14	12	12	12	12	11	7	7
P. aeruginosa	12	10	8	8	-	-	-	-	-	-	-	-
E. coli	10	8	7	7	7	-	-	-	8	8	8	8
K. pneumonia	12	-	-	-	-	-	10	8	-	-	-	-

Table 3: The antimicrobial activity (diameters of growth inhibition zones) of the crude extracts of three *M. communis* L. samples from Çetibeli

Microorganisms	Extracts	;										
		n-Hex	ane (ÇH)		CH ₂ Cl ₂ (ÇC)				MeOH (ÇM)			
	10%	5%	2.5%	1%	10%	5%	2.5%	1%	10%	5%	2.5%	1%
S. aureus	14	14	14	12	12	_	_	_	10	8	7	_
P. aeruginosa	12	10	8	8	-	-	-	-	-	-	-	-
E. coli	10	8	7	7	7	-	-	-	-	-	-	-
K. pneumonia	12	-	-	-	-	-	-	-	-	-	-	-

Table 4: The antimicrobial activity (diameters of growth inhibition zones) of the crude extracts of three *M.communis* L. samples from Yeşil Belde

	Extract	S										
Microorganisms	n-Hexane (YH)				CH ₂ Cl ₂ (YC)				MeOH (YM)			
	10%	5%	2.5%	1%	10%	5%	2.5%	1%	10%	5%	2.5%	1%
S. aureus	10	10	10	8	16	16	14	14	10	8	-	-
E. coli	- 7 15	- 14	- 14	- 10	7	_	_	_	-	-	-	_
n. prieurioriia	15	14	14	12	_	_	_	_	-	-	_	_

The inhibition zones of M. communis L. collected from Bozburun and Çetibeli have shown a similarity. The diameters of growth inhibition zones ranged from 7 to 16 mm, with the highest inhibition zone values observed against the medically important pathogens S. aureus (16 mm), K. pneumoniae (15 mm) and P. aeruginosa (12 mm) at 10% concentrations. Though, dichloromethane extracts (YC) were found to have strong activity against S. aureus (16 mm) at 10% concentrations, dichloromethane extracts (CC) were exhibited moderate activity against S. aureus (12 mm) and E. coli (7 mm) at 10% concentrations. Except for S. aureus, the MeOH extract of M. communis L. collected from Cetibeli, showed no antimicrobial activity against the other microorganisms.

In the case of the MeOH extract of *M. communis* L. from Bozburun, the diameters of growth inhibition zones ranged from 7 to 12 mm, with the highest inhibition zone values observed against the medically important pathogens *S. aureus* (12 mm at 10% concentrations) and *E. coli* (8 mm for all concentrations). As can be seen in Table 4 all extracts from Yeşil Belde did not exhibit antimicrobial activity against *P. aeruginosa*. According to the results of this study, *S. aureus* (ranged from 14 to 16 mm) was the most sensitive one to all concentrations for YC and BC. All extracts of M. communis L. samples from Yeşil Belde showed no activity against *P. aeruginosa* at all concentrations.

As a result of the present study, all samples from different regions, showed significant activity against *S. aureus*, weak activity against *P. aeruginosa*. The least active region against *E. coli* was Yeşil Belde.

The free radical scavenging activity of the methanolic extracts of Myrtus communis L. tested were determined through the DPPH method and results are presented in Table 5. DPPH is a useful reagent for investigating the free radical scavenging activities of compounds. In the

DPPH test, the extracts were able to reduce the stable radical DPPH to the yellow coloured diphenylpicrylhydrazine. The method is based on the reduction of an alcoholic DPPH solution in the presence of a hydrogen donating antioxidant due to the formation of the non-radical form DPPH– H by the reaction (44).

The methanolic extracts of Myrtus berries collected from Bozburun and Çetibeli (IC_{50} =1.22 mg/ml) showed slightly lower scavenging ability on DPPH radicals than the methanolic extract of Yeşil Belde Myrtus berries (IC_{50} = 1.24). However, when compared to quercetin (IC_{50} = 0.007), all the tested extracts showed significantly lower antioxidant activity.

Gortzi et all.(8, 45), has also studied the methanolic extract of *M. communis* leaves for its antimicrobial activity, they have found 14 mm diameter against *S. aureus* (14 mm) and 12 mm diameter against *P. aeruginosa, K. pneumoniae, E. coli.* We found also same result for n-hexane extracts of M. communis L. berries from Bozburun and Çetibeli.

Cherrat et all. (9), have reported that *S. aureus* (24.2 mm) and *Escherichia coli* (7.4-10.8 mm) for essential oil of *M. communis* L. leaves whereas our n-hexane extracts have less active against *S. aureus* (8-14 mm) and they have similar activity for *Escherichia coli* (7-10 mm).

Messaoud et all (46) have studied antioxidant activity of mature dark blue and white berries from two Tunisian Myrtus communis. They obtained their essential oil and they made GC and GC/MS analyses. The total phenol, flavonoid, and flavonol contents and the concentration of the eight anthocyanins, identified by HPLC analysis, were significantly higher in the dark blue fruits. All extracts showed a substantial antioxidant activity, assessed by the free radical scavenging activity and the ferric reducing power, with the dark blue fruit extracts being more effective which we reported our methanol extract of *M. communis* L. berries exhibited strong DPPH scavenging activity, with IC50 values of 1.22 mg/mL.

Table 5: The antioxidant activity (mg/ml) of the MeOH extracts of three M.communis L. samples from different regions

Extracts	MeOH			Control
Plant Samples	Bozburun (BM)	Çetibeli (ÇM)	Yeşil Belde (YM)	Quercetin
IC ₅₀ (mg/ml)	1.22	1.22	1.24	0.007

As for as our literature survey, we could reach a report that showed that the IC_{50} values of the methanol extract of myrtle fruit, sampled from Tunisia, was 2.1 mg/ml, (46) which supported our results.

Previous studies on the antibacterial and antioxidant activity of *M. communis* L. involved its' leaves, berries, seeds, essential oil, flower. However, it is difficult to compare the results of different studies on *M. communis* L. Because our samples have been collected from Marmaris-Mugla.

We hope that our results will provide a starting point for the investigations to exploit new natural food additive and ingredient substances present in the extracts of the plant studied.

REFERENCES

1. Davis PH. 1982. *Flora of Turkey and the East Aegean Islands*. Vol. 4. Edinburgh: University Press.

2. Baytop T. 1984. *Plant remedies in Turkey*. Istanbul University (in Turkish): Press No:3255, Faculty of Medicine; p. 444.

3. Ozek T, Demirci B, Baser KH. 2000. Chemical composition of Turkish myrtle oil. *J Essential Oil Res.* 12, 541-544.

4. Baytop, T. 1999. *Therapy with medicinal Plants in Turkey* (Past and Present). Nobel Tıp Kitapevleri Press, Istanbul.

5. Chalchat JC, Garry RF, Michet A. 1998. Essential oils of Myrtle (*Myrtus communis* L.) of the Miterranean littoral. *J Essential Oil Res.* 10, 613-617.

6. Nuvoli F. 2004. *Il Mirto della Sardegna*; Zonza Editore: Cagliari, Italy; pp 7-22. Also St. John's wort (*Hypericum perforatum* L.), another plant containing antibacterial phloroglucinols, has been used as a hop substitute for the production of beer (see: Buhner, S. H. 1998 *Sacred and Herbal Healing Beers*; Brewer Publications: Boulder, CO,).

7. Puybaret C, David B, Charveron M, Mamatas S. 1990. FR 98-11619 19980917, *Chem Abstr.* 112, 195-230.

8. Aleksic V, Knezevic P. 2014. Antimicrobial and antioxidative activity of extracts and essential oils of *Myrtus communis L. Microbiol* Res. 169(4): 240-54.

9. Cherrat L, Espina L, Bakkali M, Garc a-Gonzalo D, Pagán R, Laglaoui A. 2014. Chemical composition and antioxidant properties of *Laurus nobilis* L. and *Myrtus communis* L. essential oils from Morocco and evaluation of their antimicrobial activity acting alone or in combined processes for food preservation. *J Sci Food Agric*. 94(6): 1197-1204

10. Weyerstahl P, Marschall H, Rustaiyan A. 1994. Constituents of the essential oil of *Myrtus communis* L. from Iran. *Flavour Fragr J.* 9, 333-337.

11. Pirisino G, Mulè A, Moretti MDL, Satta M. 1996. Yield and chemical composition

of essential oil from self-sown *Myrtus communis* L. from Cuglieri (Sardinia). *Riv. Ital. EPPOS* 7, 159-169.

12. Bradesi P, Tomi F, Casanova J, Costa J, Bernardini AF. 1997. Chemical composition of myrtle leaf essential oil from Corsica (France). *J Essential Oil Res.* 9, 283–288.

13. Koukos PK, Papadopoulou KI, Papagiannopoulos AD, Patiaka D. 2001. Chemicals from Greek forestry biomass: constituents of leaf oil of *Myrtus communis* L. grown in Greece. *J Essential Oil Res.* 13, 245-246.

14. Jerkovic I, Radonic A, Borcic I. 2002. Comparative study of leaf, fruit and flower essential oils from Croatian *Myrtus communis* L. during a one-year vegetative cycle. *J Essential Oil Res.* 14, 266-270.

15. Tuberoso CIG, Barra A, Angioni A, Sarritzu E, Pirisi FM. 2006. Chemical composition of volatiles in Sardinian myrtle (*Myrtus communis* L.) alcoholic extracts and essential oils. *J Agric Food Chem.* 54, 1420-1426.

16. Lawrence BM. 2007. Progress in essential oils. *Perfume and Flavor.* 32, 54-62.

17. Aidi Wannes W, Mhamdi B, Marzouk, B. 2007. Essential oil composition of two *Myrtus communis* L. varieties grown in North Tunisia. *Ital J Biochem.* 56, 180–186.

18. Aidi Wannes W, Mhamdi B, Sriti J, Ben Jemia M, Ouchikh O, Hamdaoui G, Kchouk ME, Marzouk B. 2010. Antioxidant activities of the essential oils and methanol extracts from myrtle (*Myrtus communis* var. italica L.) leaf, stem and flower. *Food Chem Toxicol.* 48(5):1362-70.

19. Martin T, Villaescusa L, De Sotto M, Lucia A, Diaz AM. 1990. Determination of anthocyanic pigments in *Myrtus communis* berries. *Fitoterapia*. 61, 85.

20.Martin T, Villaescusa L, Diaz AM, Ollivier E, Delmas F. 1997. Screening for antiparasitic activity of *Myrtus communis. Fitoterapia.* 68, 276-277.

21. Martin T, Rubio B, Villaescua L, Fernandez L, Diaz AM. 1999. Polyphenolic compounds from pericarps of *Myrtus communis. Pharml Biol.* 37, 28-31.

22. Rosa A, Deiana M, Casu V, Corona G, Appendino G, Bianchi F, Ballero M, Dessi M A. 2003. Antioxidant activity of oligomeric acylphloroglucinols from *Myrtus communis* L. *Free Radic Res.* 37, 1013-1019.

23. Mulas M, Spano D, Biscaro S, Parpinello L. 2000. Parametri di qualità dei frutti di mirto (*Myrtus communis* L.) destinati all'industria dei liquori. *Ind Delle Bevande*. 29, 494-498.

24. Montoro P, Tuberoso CIG, Perrone A, Piacente S, Cabras P, Pizza C. 2006. Characterisation by liquid chromatography electrospray tandem mass spectrometry of anthocyanins in extracts of *Myrtus communis* L. berries used for the preparation of myrtle liqueur. *J Chromatogr A*. 1112, 232-240.

25. Montoro P, Tuberoso CIG, Piacente S, Perrone A, De Feo V, Cabras P, Pizza C. 2006. Stability and antioxidant activity of polyphenols in extracts of *Myrtus communis* L. berries used for the preparation of myrtle liqueur. *J Pharm Biomed Anal.* 41, 1614–1619.

26. Tuberoso CIG, Melis M, Angioni A, Pala M, Cabras P. 2007. Myrtle hydroalcoholic extracts obtained from different selections of *Myrtus communis* L. *Food Chem.* 101, 806-811.

27. Alipour G, Dashti S, Hosseinzadeh H. 2014. Review of pharmacological effects of *Myrtus communis* L. and its active constituents. *Phytother Res.* 28(8):1125-36.

28. Mansouri S, Foroumadi A, Ghaneie T, Najar AG. 2001. Antibacterial activity of the crude extracts and fractionated constituents of *Myrtus communis. Pharm Biol.* 39, 399-401.

29. Yadegarinia D, Gachkar L, Rezaei MB, Taghizadeh M, Alipoor-Astaneh S, Rasooli I. 2006. Biochemical activities of Iranian *Mentha piperita* L. and *Myrtus communis* L. essential oils. Phytochemistry. 67, 1249-1255.

30. Hayder N, Abdelwahed A, Kilani S, Ben Ammar R, Mahmoud A, Ghedira K, Chekir-Ghedira L. 2004. Anti-genotoxic and free radical scavenging activities of extracts from (Tunisian) *Myrtus communis. Mut Res.* 564, 89–95.

31. Sacchetti G, Muzzoli M, Statti GA, Conforti F, Bianchi A, Agrimonti C, Ballero M, Poli F. 2007. Intra-specific biodiversity of Italian myrtle (*Myrtus communis*) through chemical markers profile and biological activities of leaf methanolic extracts. *Nat Prod Res.* 21 (2), 167-179.

32. Gardeli C, Papageorgiou V, Mallouchos A, Theodosis K, Komaitis M. 2008. Essential oil composition of Pistacia lentiscus L. and *Myrtus communis* L. evaluation of antioxidant capacity of methanolic extracts. *Food Chem.* 107, 1120-1130.

33. Amensour M, Sendra E, Abrini J, Bouhdid S, Pérez-Alvarez JA, Fernández-L pez J. 2009. Total phenolic content and antioxidant activity of myrtle (*Myrtus communis*) extracts. *Nat Prod Com.* 4 (6), 819–824.

34. Halkman AK (ed). 2005. Merck Gıda Mikrobiyolojisi Uygulamaları. Başak Matbaacılık. Ankara, Türkiye, 358 p.

35. Kim J, Marshall M R, Wei C. 1995. Antibacterial activity of some essential oil components against five foodborne pathogens. *J Agric Food Chem.* 43, 2839-2845.

36. Bradshaw LJ. 1992. *Laboratory Microbiology*. Fourth Edition. Printed in USA. p. 435.

37. Chacha M, Gojase-Moletta G, Majinda RRT. 2005. Antimicrobial and radical scavenging flavonoids from the stem wood of *Erythrina latissima*. *Phytochem.* 66, 99-104.

38. Cuendet, M, Hostettmann K, Potterat O. 1997. Iridoid Glucosides with Free Radical Scavenging Properties from *Fagraea blumei*. *Helvetica Chimica Acta*. 80, 1144-1152.

39. Cuendet M, Potterat O, Salvi A, Testa B, Hostettmann K. 2000. A stilbene and dihydrochalcones with radical scavenging activities from *Loiseleuria procumbens. Phytochem.* 54, 871-874.

40. Takao T, Kitatani F, Wanatabe N, Yagi A, Sakata K. 1994. A Simple Screening Method for Antioxidants and Isolation of Several Antioxidants Produced by Marine Bacteria from Fish and Shellfish. *Biosci Biotechnol Biochem.* 58, 1780-1783. 41. Torres R, Faini F, Modak B, Urbina F, Labbe C, Guerrero J. 2006. Antioxidant activity of coumarins and flavonols from the resinous exudate of *Haplopappus multifolius*. *Phytochem.* 67, 984-987.

42. Erasto P, Bojase-Moleta G, Majinda RRT. 2004. Antimicrobial and antioxidant flavonoids from the root wood of *Bolusanthus speciosus*. *Phytochem.* 65, 875-880.

43. Naik GH, Priyadarsini KI, Satav JG, Banavalikar MM, Sohoni DP, Biyani MK, Mohan H. 2003. Comparative antioxidant activity of individual herbal components used in Ayurvedic medicine. *Phytochem.* 63, 97-104. 44. Shon MY, Kim TH, Sung NJ. 2003. Antioxidants and free radical scavenging activity of *Phellinus baumii* (Phellinus of Hymenochaetaceae) extracts. *Food Chem.* 82, 593-597.

45. Gortzi O, Lalas S, Chinou I, Tsaknis J. 2008. Reevaluation of bioactivity and antioxidant activity of *Myrtus communis* extract before and after encapsulation in liposomes. Eur. *Food Res Technol.* 226:583-590.

46. Messaoud C, Boussaid M. 2011. *Myrtus communis* Berry Color Morphs: A Comparative Analysis of Essential Oils, Fatty Acids, Phenolic Compounds, and Antioxidant Activities. *Chem. & Biodivers.* 8,300-310





Yazım Kuralları GIDA (2009) 34 (1): 55-58 www.gidadernegi.org/ Gıda Dergisi / Yayın kuralları

Makale Gönderimi ve Telif Hakkı Devir Formu GIDA (2009) 34 (1): 65 www.gidadernegi.org/ Gıda Dergisi / Makale Gönderimi ve Telif Hakkı Devir Formu

Son Kontrol Listesi GIDA (2009) 34 (1): 66 www.gidadernegi.org/ Gıda Dergisi / Son Kontrol Listesi

adreslerinden erişilebilir. Yazarlar, makale göndermeden önce yazım kurallarını tam olarak okumalı ve makalelerini burada verilen kurallara göre hazırlamalıdırlar.