*Kocatepe Vet J (2013) 6(2): 29-34* DOI: 10.5578/kvj.6496 Submittion: 20.10.2013 Accepted : 01.11.2013

# **RESEARCH ARTICLE**

# ARAŞTIRMA MAKALESİ

Key Words Campylobacter jejuni Cattle Meat Samples of Bacteriological Meat Inspection Turkish Pastırma Turkish Sausage (Sucuk)

Anahtar Kelimeler Türk Sucuğu Türk Pastırması *Campylobacter jejuni* Bakteriyolojik Et Muayenesi Örneği Sığır eti

> <sup>1</sup>Mustafa Kemal University, Veterinary Medicine Faculty, Food Hygiene and Technology Department, TR-31040, Antakya, Hatay, TURKEY

<sup>2</sup>Adnan Menderes University, Aydın School of Health Sciences, Department of Nutrition and Dietetics, TR-09100, Aydın, TURKEY

<sup>3</sup>Mustafa Kemal University, Veterinary Medicine Faculty, Food Hygiene and Technology Department, TR-31040 Antakya, Hatay, TURKEY

\*Corresponding author E-mail: elmali25erz@gmail.com Telefon: +90 (532) 573 87 34 Determination and Vital Activity of Thermophilic Campylobacter (Campylobacter Jejuni) In The Turkish Pastırma and Turkish Style Fermented Sausage (Sucuk) Commercially Consumed In Market and Experimentally Produced With Inoculation Of Different Levels of Campylobacter Jejuni

Mehmet ELMALI<sup>1</sup>\* Hilmi YAMAN<sup>2</sup>,Süleyman ÖNER<sup>3</sup>

# SUMMARY

In this study, Turkish fermented sucuk (n=30) and Turkish pastirma (n=30) samples sold in retail stores were analyzed in terms of C. jejuni. Afterwards, C. jejuni was experimentally inoculated at different levels (Group I:10<sup>2</sup>, Group II:104, and Group III:106 cfu/ml) into the mixtures of traditional Turkish fermented sucuk and cemen (spice mixture of Turkish pastirma) during the process. C. jejuni was monitored bacteriologically during fifteen days of fermentation. C. jejuni was not detected in the samples of Turkish fermented sucuk (n: 30), Turkish pastırma (n: 30), and raw beef meat samples purchased from the retail markets for use in manufacturing. The numbers of C. jejuni were below the detection level of 10<sup>2</sup> cfu/g in all three experimental Turkish fermented sucuk groups at the end of second day at pH 5.32, and also C. jejuni was not detected by Most Probable Number (MPN) technique (<0.30MPN/g) in the same samples. Likewise, C. jejuni numbers were below the detection level of 10<sup>2</sup> cfu/g at the end of fourth day at pH 5.19 in the samples of all three experimental Turkish pastırma groups, and also C. jejuni was not detected by Most Probable Number (MPN) technique (<0.30MPN/g) in the same samples. In conclusion, the risk of C. jejuni infection through consuming traditional Turkish fermented sucuk and pastirma seems to be very low unless cross contamination occurs.

Piyasada Tüketime Sunulan ve Deneysel Olarak Farklı Düzeylerde *Campylobacter jejuni* İnokule Edilerek Üretilen Türk Fermente Sucuğu ve Pastırmalardaki Termofilik *Campylobacter* (*Campylobacter jejuni*) Varlığı ve Yaşamsal Aktivasyonu

#### ÖZET

Bu araştırmada, piyasada tüketime sunulan 30 adet Türk fermente sucuğu ve 30 adet Türk pastırması C. jejuni bakımından analiz edildi. Takiben, üretim aşamasında geleneksel Türk fermente sucuğunun hamuruna ve Türk pastırmasının çemenine C. jejuni deneysel olarak farklı gruplarda inoküle edildi. (Grup I:10<sup>2</sup>, Grup II:10<sup>4</sup>, ve Grup III:10<sup>6</sup> kob/ml). On beş gün süren fermentasyon süresince C. jejuni düzeyi incelendi. Piyasadan toplanan 30 adet Türk fermente sucuğu ve 30 adet Türk pastırması ve marketlerden üretimde kullanılmak amacıyla temin edilen çiğ sığır etlerinde C. jejuni saptanmadı. Deneysel olarak üretilen Türk fermente sucuklarında pH 5.32'de 2. günün sonunda her 3 grupta C. jejuni düzeyi 102 cfu/g'ın altında saptandı. Aynı örneklerde, C. jejuni düzeyi En Kuvvetle Muhtemel Sayı Tekniği ile (EMS) (<0.30MPN/g) olarak saptandı. Benzer biçimde, deneysel olarak üretilen Türk pastırmalarında C. jejuni düzeyi her 3 grupta 4. günün sonunda pH 5.19'da 10<sup>2</sup> cfu/g'in altında saptandı. Aynı örneklerde C. jejuni düzeyi En Kuvvetle Muhtemel Sayı Tekniği ile (EMS) (<0.30MPN/g) olarak saptandı. Sonuç olarak, C. jejuni infeksiyonunun çapraz kontaminasyonlar olmadığı sürece Türk fermente sucuğu ve Türk pastırmasında çok düşük düzeyde risk yaratabileceği sonucuna varıldı.

#### INTRODUCTION

Turkish pastirma and Turkish fermented sausage (Sucuk) are consumed all over the Turkey, produced using traditional methods in several cities particularly Kayseri and Afyonkarahisar, Although ingredients like garlic, different spices, salt, nitrite or nitrate combinations, decreasing humidity and water activity in the Turkish pastirma and Turkish fermented sucuk production (especially fermentation process) are the essential factors for lowering the presence of risk factors in both meat products in terms of food borne microorganisms, Since no heat treatment is applied during their production processes. Futher more, these products are also consumed raw which could constitute a health risk. Campylobacter jejuni has a minimal infection dose (MID) of  $5.0 \times 10^2$  cfu/g (Black et al 1988). Although it possesses very weak competitive characteristics in nature (Griffiths and Park 1990), it has been isolated from red meat (Elmalı 2001, Grau 1998, Little et al 2008, Stern 1981, Stern et al 1984, Stern et al 1985), broiler meat (Atabay and Corry 1997, Elmalı and Yaman 2004, James et al 2007, Kwiatek et al 1990), milk (Kalman et al 2000, Yaman and Elmalı 2004), and fish and water (Yaman et al 2005) in epidemiological studies.

In this study, Turkish fermented sucuk (n=30) and Turkish pasturma (n=30) samples sold in retail stores were analyzed in terms of *C. jejuni*. Afterwards, *C. jejuni* was experimentally inoculated at different levels (10<sup>2</sup>, 10<sup>4</sup>, and 10<sup>6</sup> cfu/ml) into the mixtures of traditional Turkish fermented sucuk and çemen (spice mixture of Turkish pasturma) during the process. Doing that, this study aimed to monitor the presence of *C. jejuni* in the retail samples and detect the survival of different inoculation numbers of *C. jejuni* during the ripening period of raw traditional Turkish fermented sucuk and processing of Turkish pasturma in terms of risk potential of *C. jejuni* for public health since these products are especially consumed raw as well as cooked.

#### MATERIAL AND METHODS

#### Samples:

Thirty traditional Turkish fermented sucuk and thirty Turkish pastirma samples were randomly collected from retail stores, and markeds stored at 4°C until analyzed.

#### Experimental culture:

*C. jejuni* strain was grown under microaerophilic conditions using Campylobacter Enrichment Broth (CEB) and inoculated at different

levels  $(10^2, 10^4, \text{ and } 10^6 \text{ cfu/ml})$  into the experimentally traditional Turkish fermented sucuk and pastirma cemen mixtures. *C. jejuni* strain used experimentally in this research was isolated and identified from the previous research of Elmali (2001).

# Making traditional Turkish fermented sausage (Sucuk):

Turkish fermented sucuk production was experimentally made according to the Turkish Standards Institute (TSE 1070) (Anon 2002) and the Meat and Fish Office (EBK) (Anon 1988). No C. jejuni was detected in the microbiological analysis of raw beef meat used for making Turkish fermented sucuk and Turkish pastırma. Initially, 3.250 kg red cattle meat with less fat was minced in a meat grinder (Tefal). The temperature was below 4°C during this process. Subsequently, 500 g grinded frozen renal fat, spice content (20 g red pepper+32 g cumin+20 g black pepper), garlic (50 g) and saccharose (20 g) were added to the minced meat. Finally, nitrite-salt conbination (500 mg sodium nitrite-95 g salt) was also mixed into the sucuk mixture. This mixture was divided into three parts to form experimental groups of I, II and III which were experimentally inoculated at different levels of  $10^2$ ,  $10^4$ , and  $10^6$  cfu/ml with C. jejuni strain, respectively. Then, sucuk coils, placed in 1% water solution of lactic acid-salt combination, was filled up with these inoculated mixtures using a sucuk making machine (Tefal). Finally, these filled sucuk were rinsed with cold water and stored at 16-20°C for 15 days for fermentation and drying in the room conditions. Rinsing was carried out for 5 min for three days after filling. Unlike industrial fermented sausage technology, starter culture is not used in the processing of traditional Turkish fermented sucuk in the most of Anatolian cities. Similarly, No starter culture was used for making experimental sucuk in this study.

#### Making traditional Turkish pastırma:

Turkish pastirma production was experimentally made according to Elmali et al (2005). Raw beef meat pieces (5 kg) were purchased from a supermarket. Spices, fenu-greek (*Trigonella foenum* graceum), garlic and powdered red pepper used in the çemen mixture (spice paste) were bought from a market in Kayseri, Turkey. To prepare meat pieces, excessive fat and nerves were taken away and removed initially. Then, 1 cm of deeply parallel cuts were conducted on both sides of the meat pieces. An amount of 125 ppm sodium nitrite was added to the salt, After that, at an amount of 5% salt of the

weight of meat, was distributed evenly onto the surfaces of meat pieces. Then after keeping at +4°C for 24 h., the excesive water of meat pieces was removed. Then, The salted meat pieces were turned upside down and were salted second time using the same amount of salt. Following 24 h resting at +4 °C, the salted meat pieces were rinsed again under cold tap water for 1 h to remove excessive salt. Subsequently, the meat pieces were placed in an designed particularly for pastirma apparatus production. The meat pieces were squeezed between two marble layers +4 °C for 48 h using the apparatus to remove excessive water and to obtain a firm product. Following, they were dried in a well ventilated room by hanging on a metal bar for ten days. After that, they were squeezed second time in the apparatus at +4 °C for 24 h and dried again in the same condition. Then, the dried meat pieces were marinated in the cemen mixture containing 50% water, 15% fenu-greek (Trigonella foenum graceum), 20% garlic, and 15% red pepper powder, for 2 h as indicated by Dogruer et al (1998). During the process, the pastirma cemen mixture was divided into three parts to form experimental cemen groups of I, II and III which were experimentally inoculated with C. jejuni at the levels of 10<sup>2</sup>, 10<sup>4</sup>, and 10<sup>6</sup> cfu/ml, respectively. Finally, the marinated meat pieces were divided into three groups of I, II and III. Then, each group was embeded into the corresponding experimentally inoculated cemen mixture. These final experimental pastirma samples were dried for 15 days at room temperature.

## Isolation and Identification of C. jejuni:

Isolation and identification of the C. jejuni were performed according to the Food and Drug Administration (FDA) (Anon 1998). In order to isolate C. jejuni, each 25 g Turkish fermented sucuk and Turkish pastirma samples were added into 225 ml Campylobacter Enrichment Broth (CEB). They were initially incubated at 37°C for 2-4 hours and then at 42°C for 48 h in a microaerobic atmosphere using Campygen (CN025A, Oxoid, Basingstoke, UK). To enumerate C. jejuni by culture, 0.1 ml of each enrichment broth sample was streaked onto Campylobacter Selective Agars (modified CCDA, CM 739, Oxoid) and incubated microaerobically at 42°C for 48 h. Grown colonies demonstrating the characteristics of Campylobacter spp. were picked, purified and spesified further by Gram stain, motility test, growth in media containing 3.5 % NaCI, and 25 °C, 35-37°C and 42°C, oxidase reaction, hippurate hydrolysis, sensitivity to nalidixic acid (CT0031C, Oxoid) and cephalothin (CT0010B, Oxoid). The

evaluation of MPN table was done according to De man (1983).

## RESULTS

*C. jejuni* was not detected in the samples of Turkish fermented sucuk (n: 30), Turkish pastirma (n: 30), and raw beef meat samples purchased from the retail markets for use in manufacturing.

The initial numbers of *C. jejuni* were  $10^2$ ,  $10^4$ , and  $10^6$  cfu/ml in the experimental Turkish fermented sucuk groups of I, II and III, respectively at pH 5.72 on the production day. The numbers of *C. jejuni* were found to be  $<10^2$ ,  $10^2$ , and  $10^4$  cfu/g in the experimental Turkish fermented sucuk groups of I, II, and III, respectively, indicating a two log reduction at pH 5.47 on day one. At the end of second day, pH was 5.32, and the numbers of *C. jejuni* were below the detection level of  $10^2$  cfu/g in all three experimental Turkish fermented sucuk groups and *C. jejuni* was not also detected by Most Probable Number (MPN) technique (<0.30MPN/g) in the same samples.

In the experimental Turkish pastirma samples, the number of *C. jejuni* was below the detection level of 10<sup>2</sup> cfu/g in group I while one log of reduction was observed in the groups of II and III at pH 5.58 on day one. These one log reductions continued in the groups of II and III untill the end of day three at pH 5.32. *C. jejuni* numbers were below the detection level of 10<sup>2</sup> cfu/g at the end of fourth day at pH 5.19 in the samples of all three experimental Turkish pastirma groups, and also *C. jejuni* was not detected by Most Probable Number (MPN) technique (<0.30MPN/g) in the same samples. The result of *C. jejuni* numbers in the experimental traditional Turkish fermented sucuk and Turkish pastirma samples are summarized in Table 1.

## DISCUSSION

Red meat consumption is considered as an identified but minor risk factor for exposure to *Campylobacter* spp. As a medium for microbial growth, meat is suitable because of the high water and protein content. Not much information has been informed about the behaviour of *Campylobacter* spp. on beef or lamb meat during processing. Some information is available on retail fresh or frozen raw red and minced meat but not much about other red meat products. On meat cuts or minced meat, important factors for the behaviour of *C. jejuni* are temperature, medium, microaerophilic atmosphere, and pH. *Campylobacter* spp. usually survives poorly at atmospheric oxygen concentrations. Although *C.* 

| Elmalı M, Yaman H, Öner S | Determination and Vital Activity of Thermophilic Campyloba |
|---------------------------|--|
|                           | In The Turkish Pastırma and Turkish Style Fermente         |

*jejuni* may not multiply readily in or on meat (Turnbull and Rose 1982), *Campylobacter* spp. can survive and even grow when initially packed under normal atmospheric conditions, as the metabolic activity of the food, for instance, raw meat may create a different gaseous environment (ICMSF 1996). Several researchers have reported that retail minced meat and sausages were contaminated with campylobacters (Bolton et al 1985, Turnbull and Rose 1982).

d Sausage (Sucuk)

Table 1. The numbers of *C. jejuni* in the experimentally made traditional Turkish fermented sucuk and Turkish pastirma.

| Days of production | Turkish fermented sucuk<br>Level of inoculation (cfu/ml) |            |            | Turkish pastırma<br>Level of inoculation (cfu/ml) |                 |            |            |      |
|--------------------|--|------------|------------|---|-----------------|------------|------------|------|
|                    |  |            |            |   |                 |            |            |      |
|                    | 10 <sup>2</sup>  | 104        | 106        | 5.72  | 10 <sup>2</sup> | 104        | $10^{6}$   | 5.75 |
| 1                  | <102   | 102        | 104        | 5.47  | <102            | 103        | 105        | 5.58 |
| 2                  | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.32  | $< 10^{2}$      | $10^{2}$   | $10^{3}$   | 5.42 |
| 3                  | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.24  | $< 10^{2}$      | $< 10^{2}$ | $10^{2}$   | 5.32 |
| 4                  | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.03  | $< 10^{2}$      | $< 10^{2}$ | $< 10^{2}$ | 5.19 |
| 5                  | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.00  | $< 10^{2}$      | $< 10^{2}$ | $< 10^{2}$ | 5.10 |
| 7                  | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.02  | $< 10^{2}$      | $< 10^{2}$ | $< 10^{2}$ | 5.08 |
| 10                 | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.02  | $< 10^{2}$      | $< 10^{2}$ | $< 10^{2}$ | 5.07 |
| 15                 | $< 10^{2}$   | $< 10^{2}$ | $< 10^{2}$ | 5.03  | $< 10^{2}$      | $< 10^{2}$ | $< 10^{2}$ | 5.08 |

G1: Group I (inoculated with 10<sup>2</sup> cfu/ml C. jejum); G2: Group I (inoculated with 10<sup>4</sup> cfu/ml C. jejum)

G3: Group I (inoculated with 106 cfu/ml C. jejuni); \*Initial pH

In this study, a very particular meat products of traditional Turkish fermented sucuk and Turkish pastirma were inoculated experimentally with C. jejuni at different levels of  $10^2$ ,  $10^4$  and  $10^6$  cfu/g to cover a broad range of numbers. Even though C. jejuni contamination of raw meat is generally considered low (Pezzotti et al 2003, Whyte et al 2004, Wong et al 2007) cross-contaminations occur as a risk in terms of uncooked meat products. During making Turkish fermented sucuk and Turkish pastırma, meat is neither cooked nor refrigerated, these products must be safe to consume yet. C. jejuni has been isolated with particular frequency in food products including ground meat and sausage (Baffone et al 1995, Bolton et al 1985). In minced meat, C. jejuni survived better and viable counts were relatively unchanged in minced beef at refrigerator temperatures and 22°C, but showed a decrease in corresponding samples of sausage mixture (Barrell 1984). However when it was stored at 20°C, C. jejuni also decreased rapidly and could not be isolated after 3 days (Svedhem 1981). In the Chinese-style sausages, no viable C. jejuni was detected in either vacuum-packed or non-vacuum-packed sausage after 7 days of storage at 25°C (Yeh 1994). In the study of Bostan et al (2001), C. jejuni numbers on the 3rd day of fermentation was <10 cfu/g in the experimentally made Turkish fermented sucuk which were experimentally inoculated with C. jejuni at the numbers of  $10^5$  cfu/g. They, also reported that C. jejuni was not detected in their Turkish fermented sucuk samples on the 4th day of fermentation. On

the other hand, in our study, *C. jejuni* was inactivated on the  $2^{nd}$  day of fermentation. While inactivation period was shorter in our study, inactivation pH level (pH 5.32) of this study was in paralell with those of Bostan et al (2001).

There are several factors that might effect the survival of C. jejuni in traditional Turkish fermented sucuk. The spices, garlic and nitrite-salt combination in the sucuk and cemen mixture may have inhibited C. jejuni. Thermophilic Campylobacter spp. are very The maximum NaCl sensitive to NaCl. concentration in which Campylobacter spp. which can multiply is 1.5% (Anon 1998) while optimum NaCl concentration for them is 0.5% (Doyle and Roman 1982, Hanninen 1981). Reduction due to organic and inorganic acids such as lactic acid occuring during the fermantation process and decreasing water activity are among other essential factors for lowering C. jejuni numbers, but it has been demonstrated that a strong bactericidal effect of organic acids on campylobacters occured much more between the pH values of 4.0 and 4.5 while bactericidal activity was low at values of pH 5.0 and above (Chaveerach et al 2002). In week acid conditions (pH between 5.0 and 6.0) Campylobacter spp. survived well and only minor decreases in culturability were found (Alter et al 2006, Waterman and Small 1988). Competitive flora such as lactobacilli may not have showed an antogonistic effect on the survival of C. jejuni as Koidis and Doyle (1983), and Moore and Madden (2001) reported that Campylobacter spp. behave differently than other food

pathogens in the presence of competitive flora. On the contrary, Campylobacter spp. are very sensitive to particularly at ambient temperatures drving, (Fernandez 1985, Lake et al 2007). In the study of Alter et al (2006), in paralel to reduction in water activity after 24-48 h ripening, detection of C. jejuni was only possible after enrichment in the German style fermented turkey sausages. Therefore, C. jejuni is likely to be inhibited at the temperatures of 20°C and during drving at room temperature in our Turkish fermented sucuk and Turkish pastırma. There are other reports with either similiar or different results on inactivation periods and bacteria levels (Yeh and Chou 1994, Warburton, 1987). The varriations in inactivation periods may be due to different fermentation processes, ingredients and variations in the parameters of these process.

conclusion, Bacterial In carcass meat inspection is an important indicator for meat inspection. It must be done for public health because Turkish pastirma and Turkish style fermented sausage (sucuk) are raw meat products. Considering the result of this study and previous a few relevant studies, the risk of C. jejuni infection through consuming of traditional Turkish fermented sucuk and Turkish pastirma seems to be very low unless croos contamination occurs. However, due to very low infectious dose of Campylobacter spp. consuming unripened Tukish fermented sucuk and pastirma at the early days of fermentation (1-7 days) should be avoided without cooking.

#### REFERENCES

- Alter T. Bori A. Hamedi A. Ellerbroek L. Fehlhaber K. 2006. Influence of inoculation levels and processing parameterson the survival of *Campylobacter jejuni* in German style fermented turkey sausage. Food Microbiol., 23: 701-707.
- Anonymous. 1988. Et ve balık Kurumu (Meat and Fish Office) et mamüller idairesi işletme ve imalat yönetmeliği, Et ve balık kurumu, yönetmelik, sıra no 33, second edit., Ankara, Turkey, pp. 16.
- Anonymous. 1998. Bacteriological Analytical Manual., F.D.A.: 8<sup>th</sup> ed. published AOAC. Chapt: 7.*Campylobacter*, USA.
- Anonymous. 2002. Turkish Standard Institute (TS 1070) Turk Sucugu - Sucuk (Turkish style fermented sausage, Necatibey Cad. 112 06100, Ankara, Turkey.
- Atabay HI. Corry JEL. 1997. The prevalence of campylobacters and arcobacters in broiler chickens. J. Appl. Microbiol., 83: 619-626.

- Baffone W. Bruscolinl F. Pianetti A. Biffi MR. Brandi G. Salvaggio L. Albano V. 1995. Diffusion of thermophilic *Campylobacter* in the Pesaro-Urbino area (Italy) from 1985 to 1992. Europan J. Epidemiol., 11 (1): 83-86.
- Barrell RAE. 1984. The survival of *Campylobacter jejuni* in red meats stored at different temperatures. Int. J. Food Microbiol., 1(4): 187-196.
- Black RE. Levine MM. Clements ML. Hughes TP. Blaser MJ. 1988. Experimental *Campylobacter jejuni* infection in humans., J. Infect. Dis., 157: 472-479.
- Bolton FJ. Dawkins HC. Hutchinson D. 1985. Biotypes and serotypes of thermophilic campylobacters isolated from cattle, sheep, and pig offal and other red meats. J.Hyg. Camb., 95: 1-6.
- **Bostan K. 2001**. Survival of *Campylobacter jejuni* in Turkish fermented sausage. Arch. Lebensmittelhyg., 52: 119-121.
- Chaveerach P. Keuzenkamp DA. Urlings HAP. Lipman LJA. Van Knapen F. 2002. In vitro study on the effect of organic acids on *Campylobacter jejuni/coli* populations in mixtures of water and feed. Poult. Sci., 81(5): 621-628.
- De man JC. 1983. MPN-tables, corrected. J. Appl.Microbiol., Biotechnol.17:301-305.
- **Dogruer Y. Nizamlıoglu M. Gurbuz U. Kayaardı S. 1998**. Cesitli çemen karisimlarinin pastırma kalitesine etkisi II: Mikrobiyolojik nitelikleri, Turk J. Vet. Anim. Sci., 22: 221- 229.
- Doyle MP. Roman DJ. 1982. Response of *Campylobacter jejuni* to sodium chloride. Appl. Environ. Microbiol., 43: 561.
- Elmalı M. 2001. Isolation and identification thermophilic *Campylobacter* species in the bacteriological meat inspection samples of cattle. PhD. Theises, Institute of Health Science, University of Ankara, Ankara.
- Elmalı M. Yaman H. 2004. Thermophilic *Campylobacter* spp. on frozen poultry carcasses. Kafkas Univ. Vet. Fak. Derg., 10(1): 27-30.
- Elmalı M. Yaman H. Ulukanlı Z. Elmalı DA. 2005. Determination of some chemical and microbial parameters of experimental Turkish pastrami during production. Arch. Lebensmittelhyg., 56(6):139-143.
- Fernandez H. Vergara M. Tapia FF. 1985. Dessication resistance in thermotolerant *Campylobacter* species. Infect., 13(4): 197-197.
- Grau F. 1998. Campylobacter jejuni and Campylobacter hyointestinalis in the intestinal tract and on the carcasses of calves and cattle. J. Food Prot., 51(11): 857-861.

Griffiths PL. Park RWA. 1990. Campylobacters

associated with human diarrhoeal disease. J. Appl. Bacteriol., 69:281-301.

- Hanninen ML. 1981. The effect of NaCl on Campylobacter jejuni/coli. Acta Vet.Scand., 22: 578. http://www.ncbi.nih.nlm.gov. Date of arriving: 08.02.2000.
- ICMSF. 1996. Micro-organisms in foods 5. Microbiological specifications of food pathogens. International Commission on Microbiological Specifications for Foods (ICMSF), Blackie Academic and Professional, London,UK.
- James C. James SJ. Hanay N. Purnell G. Barbedo-Pinto C. Yaman H. Araujo M. Gonzalez ML. Calvo J. Howell M. Corry JEL. 2007. Decontamination of poultry carcasses using steam or hot water in combination with rapid cooling, chilling or freezing of carcass surface. Int. J. Food Microbiol., 114: 195-203.
- Kalman M. Szollosi E. Czermann B. Zimayni M. Szekeres S. Kalman M. 2000. Milkborne *Campylobacter* infection in Hungary. J Food Prot.,63(10): 1426-1429.
- Koidis P. Doyle MP. 1983. Survival of *Campylobacter jejuni* in fresh and heated red meat. J.Food Prot., 46(9): 771-774.
- Kwiatek K. Wojton B. Stern N. 1990. Prevalence and distribution of *Campylobacter* spp. on poultry and selected red meat carcasses in Poland. J. Food Prot., 53 (2):127-130.
- Lake R. Hudson A. Cressey P. Gilbert SS. 2007. Risk Profile: *Campylobacterjejuni/coli* in red meat. Client Report FW0485., Institute of Environmental Science & Research Limited Christchurch Science Centre, Christchurch, New Zealand.
- Little CL. Richardson JF. Owen RJ. Pinna E. Threlfall EJ. 2008. *Campylobacter* and *Salmonella* in raw meats in the United Kingdom: Prevalence, characterization and antimicrobial resistance pattern, 2003-2005. Food Microbiol., 25: 538-543.
- Moore JE. Madden RH. 2001. Survival of *Campylobacter coli* in porcine liver. Food Microbiol., 18 (1): 1-10.
- Pezzotti G. Serafin A. Luzzi I. Mioni RM, Perin MR. 2003. Occurrence and resistance to antibiotics of *Campylobacter jejuni* and *Campylobacter coli* in animals and meat in northeastern Italy. Int. J. Food Microbiol., 82:281-287
- Stern NJ. 1981. Recovery rate of *Campylobacter fetus* spp. *jejuni* in eviscerated pork, lamb and beef carcasses. J. Food Sci., 46:1291-1293.

Stern NJ. Green SS. Thaker N. Krout DJ. Chiy

**J. 1984**. Recovery of *Campylobacter jejuni* from fresh and frozen meat and poultry collected at slaughter. J Food Prot., 47(5): 372-374.

- Stern NJ. Hernandez MP. Blankenship L. Deibel KE. Doores S. Doyle MP. Ng H. Pierson MD. Sofos JN. Sveum WH. Westhoff DC 1985. Prevalence and distribution of *Campylobacter jejuni* and *Campylobacter coli* in retails meats. J. Food Prot., 48(7): 595-599.
- Svedhem A. Kaijser B. Sjogren E. 1981. The Occurrence of *Campylobacter jejuni* in fresh food and survival under different conditions. J. Hygiene., 87 (3): 421-425.
- **Turnbull PCB. Rose P. 1982**. *Campylobacter jejuni* and *Salmonella* in raw red meats. J. Hyg. Camb., 88:29-37.
- Warburton DW. Weiss KF. Purvis U. Hill RW. 1987. The microbiological quality of fermented sausage produced under good hygienic practices in Canada. Food Microbiol., 4:187-197.
- Waterman SR. Small PL 1988. Acid-sensitive enteric pathogens are protected from killing under extremely acidic conditions of pH 2.5 when they are inoculated onto certain solid food sources. Appl. Environ. Microbiol., 64: 3882-3886.
- Whyte P. McGill K. Cowley D. Madden RH. Moran L. Scates P. Carroll C. O'Leary A. Fanning S. Collins JD. McNamara E. Moore JE. Cormican M. 2004. Occurence of *Campylobacter* in retail foods in Ireland. Int. J. Food Microbiol., 95: 111-118.
- Wong TL. Hollis L. Cornelius A. Nicol C. Cook R. Hudson JA. 2007. Prevalence, numbers and subtypes of *Campylobacter jejuni* and *Campylobacter coli* in uncooked retail meat samples. J. Food Prot., 70: 566-573.
- Yaman H. Elmali M. 2004. The Occurence of Thermophilic *Campylobacter* (*C. jejuni*) in Raw Milk. Kafkas Univ. Vet. Fak. Derg., 10(1): 37-40.
- Yaman H. Elmalı M. Ulukanlı Z. Tekinsen KK. 2005. Presence of *Campylobacter*. (*C. Jejuni*) in Recreational, Lake, Stream Water and Fresh Fish in Turkey. Arch Lebensmittelhyg., 56(4): 83-86.
- Yeh C. Chou C. 1994. Behaviour of *Campylobacter jejuni* during the manufacture and storage of Chineese-style sausage. Food Microbiol., 11: 461-466.