

AQUATIC SCIENCES AND ENGINEERING

Aquat Sci Eng 2020; 35(4): 94-9 • DOI: https://doi.org/10.26650/ASE2020648340

Original Article

Contribution of Passive Sampling Devices on the Determination of Hydrophobic Organic Contaminant Bioaccumulation in Marine Organisms

Sevil Deniz Yakan¹ 💿

Cite this article as: Yakan, S. D. (2020). Contribution of passive sampling devices on the determination of hydrophobic organic contaminant bioaccumulation in marine organisms. Aquatic Sciences and Engineering, 35(4), 94-9.

ABSTRACT

Hydrophobic Organic Contaminants (HOC) are a group of chemicals needed to determine the health of marine ecosystems, and passive sampling devices are promising tools that offer a convenient monitoring opportunity. Traditional biomonitoring studies involved different types of marine organisms, and it appeared that simultaneous deployment of passive samplers with biomonitoring organisms provided the necessary information for the calculation of the aquatic organisms' bioaccumulation factors (BAF). There was not any other parameter than BAF, that could be used to determine the biomagnification and fate of contaminants in the upper trophic levels, which eventually affect all marine and terrestrial ecosystem health. In the light of the essence of BAF, this study applied a modified version of BAF estimation dependent on the contaminant concentrations both in the passive and active samplers. Thus, BAF parameters could be calculated properly without any need of a contaminant concentration in the surrounding water environment. For this purpose, the HOC concentration detected from the anthropogenic settlements in the coastal regions of Turkey were collocated, evaluated, and represented for different HOC groups. It was concluded that the present method is appropriate and applicable for BAF calculations of different groups of HOCs, where there are simultaneous deployments of both active and passive samplers in the process of biomonitoring studies.

Keywords: Passive sampling, HOC, bioaccumulation, BAF, concentration ratio model

INTRODUCTION

Polyaromatic hydrocarbons (PAH), polychlorinated biphenyls (PCB) and organochloride pesticides all belong to the group of hydrophobic organic contaminants (HOCs). The determination of concentrations of hydrophobic organic contaminants (HOCs) in water samples is difficult because of chemical properties like low water solubility, high lipophilicity, strong adsorption tendency to suspended materials and tendency to accumulate in organisms (David et al., 2010). In order to overcome this obstacle, alternative methods such as biomonitoring and passive sampling have been used for determination of the sources and the fate of HOCs in different matrices of the marine environment (Blasco & Picó, 2009; Bourgeault & Gourlay-Francé, 2013). Bivalve mollusks and Semi-Permeable Membrane Devices (SPMD) are widely used examples of biomonitoring and passive sampling studies for the determination of HOC concentration in a water environment (André Lourenço et al., 2015; Fontenelle, Taniguchi, da Silva, & Lourenço, 2019; Lance, Matz, Reeves, & Verbrugge, 2012; Lourenço et al., 2016; Peven, Uhler, & Querzoli, 1996). A wide geographic distribution and the sessile nature of bivalves makes them the preferable organisms in biomonitoring studies, in addition to the accumulation tendency of HOCs in bivalves (Bayne, 1976). On the other hand, passive samplers have the advantage of not being affected

ORCID IDs of the author: S.D.Y. 0000-0003-2493-680X

¹Istanbul Technical University, Faculty of Naval Architecture and Ocean Engineering, Istanbul, Turkey

Submitted: 19.11.2019

Revision Requested: 05.04.2020

Last Revision Received: 11.04.2020

Accepted: 22.04.2020

Online published: 22.05.2020

Correspondence: Sevil Deniz Yakan E-mail: yakans@itu.edu.tr

©Copyright 2020 The Author(s) Available online at https://dergipark.org.tr/ase by the poor water quality in contrast to the organisms that need to survive to be used in biomonitoring studies. Thus, a type of passive samplers, SPMDs, have become a popular instrument of detecting HOCs in water bodies (Huckins, J.N; Petty, J.D.; Booij, 2006; Huckins, J.N.; Tubergen, M.W.; Manuweera, 1990).

Diffusion and partitioning are the main pathways of PAH accumulation in SPMDs, thus freely dissolved contaminants in water can be detected precisely over a specific time interval (Gourlay, Tusseau-Vuillemin, Garric, & Mouchel, 2003; Greenwood, Mills, & Roig, 2007). The deployment of SPMD during a specific time interval has many advantages compared with a spot analysis of water contamination. For example, the decrement of detection limits due to an accumulation tendency of HOCs into the material led eventually to a higher concentration of contaminants accumulated in SPMDs. In addition, the average measurement of a time period instead of a measurement of a moment provides more reliable results of the target zone (Uher, Mirande-Bret, & Gourlay-Francé, 2016). SPMDs are not designed to mimic all metabolic activities of aquatic organisms, but instead to mimic the pathways of diffusion and partitioning in the process of contaminant bioaccumulation through adsorption to the surfaces of the target organism like algae, mussel, fish etc. (Gourlay et al., 2005; Mayer, Philipp; Tolls, Johannes; Hermens, Joop L.M.; Mackay, 2003; Uher et al., 2016). Thus, simultaneous deployment of passive and active sampling devices (for example, SPMDs and bivalves) is promising for more sensitive and reliable measurements as applied in many studies (Amdany et al., 2014; Bourgeault & Gourlay-Francé, 2013; David et al., 2010; Verweij, Booij, Satumalay, Van Der Molen, & Van Der Oost, 2004; Vrana et al., 2014).

The main advantage of simultaneous deployment of active and passive sampling methods is to be able to determine the bioaccumulation tendency of HOCs in selected marine organisms without any need of ambient water concentration data (Booij, Smedes, Van Weerlee, & Honkoop, 2006). The similar uptake processes of HOCs in both bivalves and passive samplers enable researchers to obtain the bioaccumulation data by means of a comparison of accumulated HOC concentrations in both organisms and samplers (Harman, Brooks, Sundt, Meier, & Grung, 2011). The assessment of HOCs in different matrices of the marine environment was performed using this approach in several studies (e.g. Bourgeault & Gourlay-Francé, 2013; David et al., 2010). This study aimed to determine the bioaccumulation factors in marine organisms using the concentrations of HOCs detected in the western and north-western regions of Turkey by means of this simultaneous monitoring type of approach. Within the scope of this study, bioaccumulation of HOCs in selected marine organisms were determined without the necessity of additional HOCS data, like its presence and concentration in the surrounding water environment. Consequently, the results of the simultaneous deployment of the Mediterranean mussel species, Mytilus galloprovincialis, and SPMDs were evaluated using a published concentration ratio model (Booij et al., 2006) that was taken as the basis of a BAF determination, and adapted for evaluation of HOCs in determining bioaccumulation factors of selected marine organisms for PAH, PCB and OCP.

MATERIALS AND METHODS

The concentration data of HOCs (PAHs, PCBs and OCPs) in mussels and SPMDs were collocated from three previously published papers (Karacik, Okay, Henkelmann, Pfister, & Schramm, 2013; O. S. Okay et al., 2014; Oya S. Okay et al., 2017). The data in the referenced papers was reorganized into Excel sheets and used as inputs for the present study. The field studies in the papers were accomplished in total at 13 different stations: four stations along the Istanbul Strait with a deployment time of seven and 21 days; and nine stations in anthropogenic settlements like shipyards and marinas with a deployment time of 30 and 60 days, which are located in the west and north-west coast of Turkey (three marinas in the west, three shipyards and three marinas in the north-west coast of the country). Because the detailed locations of sampling stations were given in the referenced papers (Karacik et al., 2013; O. S. Okay et al., 2014; O.S. Okay et al., 2017) and the choice of sampling stations were out of the scope of the present study, the sampling stations are only shown in general as seen in Figure 1.



Figure 1. The locations of the sampling stations are marked with the red frame: four stations along the Istanbul Strait; nine stations at the west and north-west coast of Turkey. Detailed maps were given in the papers of (Karacik et al., 2013; O. S. Okay et al., 2014; O.S. Okay et al., 2017).

The variety of stations enhanced the results of this study. For example, sampling stations at the north-west coast of the country are in the most populated city of the country (Istanbul) with about 17 million inhabitants. Marinas and shipyards in this region are on different sides of the city. The east part of the south entrance of the Istanbul Strait is filled with marinas while the east border of the city is filled with several shipyard companies consisting of a higher number of workers. On the other hand, sampling stations at the west coast of the country are located in a less populated area which is popular with summer tourism. As mentioned in the referenced papers, the Mediterranean mussel, Mytilus galloprovincialis, was used as the selected mussel species, and simultaneously deployed with a commercial type of passive sampler, SPMD. The data from these two different types of samplers, named as active and passive samplers, was used as inputs in the present study in order to determine the bioaccumulation potential of HOCs.

The analyzed results of HOCs in mussels and SPMDs were used in the slightly modified version of the mussel/SPMD concentration ratio model (Booij et al., 2006). The modification included the parameter of initial contaminant concentration in transplanted mussels, which is not a zero value naturally, at the time of deployment. The fitting of field data was implemented using the NonlinearModelFit method in Mathematica (Wolfram Research, version 10.0). The kinetic rate constants were matched with the results of mussel and SPMD analysis. Not all the results were used in the NonlinearModelFit method because of the detection limitations of some HOCs. Among all HOCs, the data of 14 PAHs and 11 PCBs were used for the implementation of the NMF method. Regression equations presented in the referenced publication were used for determination of the SPMD-water partitioning coefficient (K $_{\scriptscriptstyle SW'}$ L kg^-1) and elimination rate constant of the SPMD (k_a, d⁻¹) (Booij et al., 2006). The bioaccumulation factors (BAF, L kg⁻¹) were obtained by replacing the determined coefficients and constants in Equation (1).

$$C_{M} = \frac{\rho_{S}C_{S}(BAF(1-e^{-k_{2}t})) + C_{M,0}e^{-k_{2}t}}{K_{SW}(1-e^{-k_{e}t})}$$
(1)

In Equation (1), C_M (ng g⁻¹), $C_{M,0}$ (ng g⁻¹), C_S (ng g⁻¹ triolein), ρ_S (g mL⁻¹) represents the mussel concentration at any day, the mussel concentration at the initial day (day 0), the SPMD concentration and its density, respectively. Because the initial concentrations of mussels are not always zero, Equation (1) consists of an additional term compared with the original equation in reference (Booij et al., 2006). After fitting Equation (1), the parameters of BAF were evaluated with respect to their octanol-water partition coefficient (K_{ow}) dependency. Thus, BAFs (L kg⁻¹) of mussels for a series of HOCs were obtained without any need of ambient water concentration.

RESULTS AND DISCUSSION

Log K_{ow} values for a series of HOCs range within 3.37 and 7.41; specifically, between 3.37 - 6.9 for PAHs, 5.66 - 7.41 for PCBs and 3.7 - 7.13 for OCPs. This variation brought about inherently different patterns of accumulation. It was seen that Log K_{ow} values positively correlate with the accumulation of PAH and OCP both in the passive and active samplers. On the contrary, the accumulation of PCBs followed a different trend. This difference was probably the result of having comparatively higher Log K_{ow} values. The different nature of HOCs reflected their accumulation patterns.

The activities that originated from human actions were the main source of PAHs. The field monitoring results in the areas of the marina and shipyard were evidence of this fact (O. S. Okay et al., 2014; Oya S. Okay et al., 2017; Yilmaz et al., 2014). In addition, the products of industrial waste were the sources of PCBs (Cardellicchio et al., 2007; Cetin et al., 2017; Helou, Harmouche-Karaki, Karake, & Narbonne, 2019; Hong, Yim, Shim, Li, & Oh, 2006), and this was seen clearly in the comparison between the shipyard and marina zones. The machinery and equipment used in the construction, maintenance, and repair of ships in shipyards seemed to directly affect this pattern difference. The main source of OCPs are the products related to agriculture (Ahmad, Salem, & Estaitieh, 2010; Fenik, Tankiewicz, & Biziuk, 2011; Helou et al., 2019), thus they are generally detected at low rates. The results of HOCs were grouped into two groups, depending on their deployment times. The reason for this grouping was to help in the evaluation of different features at the sampling stations. The first group deployment time was 7 and 21 days, and the second group was 30 and 60 days. Therefore, NonlinearModelFit function was run separately for the two groups, and the output was represented as separate figures. Figures 2 and 3 are results of the first group while Figure 5 represents the output of the second group.





The relationship between the ratios of SPMD and mussel concentrations (C_s/C_M) and Log K_{OW} values are shown in Figure 2 for the first group. An inverse trend was observed for the group of PAHs. The accumulation of PAHs decreased with the increase of Log K_{OW} values. In contrast, the group of PCBs, which have higher K_{OW} values, show the opposite trend. The trend of the data is represented with Equation (1) for PAH (in green), PCB (in red) and OCP (in blue) separately, as shown in Figure 2. The combined evaluation of PAH, PCB and OCP data (black spline in Figure 2) shows a plateau, which could be explained by the steric hindrance of higher molecular weight HOCs of Log K_{ow} > 5.0, especially during the accumulation of PAHs through the pores of SP-MDs (Luellen & Shea, 2002). This fact of uptake restriction due to the morphology was also mentioned in several observations (Huckins et al., 1999; Luellen & Shea, 2002).

Bioaccumulation factors (BAFs) were evaluated versus Log K_{OW} values of contaminants. The data are matched separately and together for the groups of PAHs, PCBs and OCPs, as shown in Figure 3. Separate PAH and PCB data were matched well with linear regressions. Although the data of OCPs were scattered, they were also matched and represented with linear regression. Moreover, the combined data of PAH, PCB and OCP were fitted by a hyperbolic function. The characteristics of a fitted hyperbol-



Figure 3. Log BAF versus Log K_{OW} values and model fits for PAH, PCB and OCP separately (in green, red, and blue) and for the combined data set (in black).

ic function was not affected by the scattered data of OCPs. However, the data of PCBs was the main factor affecting the characteristics of the fitted function due to its decreasing trend with the increasing Log K_{ow} values. This fact can also be considered as a probable inhibition of high Log K_{ow} valued PCB accumulation (Baskaran, Armitage, & Wania, 2019; Qiu, Qiu, Zhang, & Li, 2019).



Figure 4. The ratio of contaminant concentrations in SPMD (C_s) and transplanted mussels (C_M) versus octanolwater coefficients (Log K_{ow}) of related contaminants is shown. Each contaminant group is represented with a different color: PAH in green, PCB in red and OCP in blue for the deployment durations of 30 and 60 days, formed with open and closed symbols, respectively. The black vertical line at the y-scale (Log 10=1) indicates the equality of SPMD and mussel concentrations.

In Figure 4, y-scale of Log C_s/C_M enabled researchers to observe the different accumulation trends of the active and passive samplers, and the black line at Log 10 indicates the equality of their concentrations. The upper part of the black line signifies that the metabolization of HOCs in the selected marine organism is higher, whereas the lower part of the black line indicates that the contaminants in the particular phase were higher than their dissolved forms, as a reminder that the dissolved phase of contaminants accumulated in the passive sampler (Bourgeault & Gourlay-Francé, 2013; Gourlay-Francé, Lor-

geoux, & Tusseau-Vuillemin, 2008; Kim, Kim, Alvarez, Lee, & Oh, 2014; Lance et al., 2012; Luellen & Shea, 2002; Taylor, Fones, Vrana, & Mills, 2019; Zhao et al., 2018).





The stations in group 2 were differentiated according to their type (marinas and shipyards) and locations (west and north-west). Bioaccumulation factors were calculated by using Equation (1) and represented separately for the marinas at the west, shipyards at the north-west and marinas at the north-west region of the country as shown in Figure 5. In addition, combination of all data belonging to all stations with a deployment time of 30 and 60 days were also evaluated and shown in Figure 5, using different colors and shapes for different type of contaminants (green circles for PAH, red stars for PCB and blue diamonds for OCP).

It is clearly seen from Figure 5 that PAHs were abundant in all stations, whether the sampling stations were in an industrialized zone, a highly populated city or a low-populated district famous for its summer tourism. On the contrary, the difference in the distribution of PCBs is clearly seen in Figure 5, with the abundance of PCBs especially in the sampling stations located in the highly industrialized areas like the zones of shipyards compared with sampling stations located in the zones of marinas. Apart from this fact, the observed presence of PCBs in marinas located in the north-west region of the country can be explained because of its proximity to the industrialized and higher populated areas. In addition, the presence of OCPs at all stations can be explained due to its persistent nature, previous usage, and potential runoffs from the observed regions, although its usage has been banned for a long time (Ozcan & Aydin, 2009).

Furthermore, another evaluation was performed using a different deployment period of time for the samplers. It is clearly seen from Figure 6 that deployment time affects the range of results although the data trend remains the same. Different times of deployment were selected for this purpose. As a reference for the deployment duration, the necessity and importance of the exposure time is a minimum of five days for SPMDs as stated in previous studies (Luellen & Shea, 2002). The outputs represented in Figure 6, range from seven days to an unlimited period of time. The results of different deployment times (7, 21, 30, 60, 360 and unlimited days), as shown in Figure 6, could be used for the evaluation and estimation of an efficient deployment time for prospective field monitoring studies. The importance of deployment time should not be undervalued for the design of field studies in an effort to reduce expenses such as disposables and chemicals used both in the field and laboratory and the addition of travel and transportation expenses in distance sampling.



CONCLUSION

The deployment of passive sampling devices in coordination with the field studies of marine organisms is a promising tool for the determination of bioaccumulation factors. The simultaneous usage of both active and passive sampling devices is increasing worldwide for the monitoring of hydrophobic organic contaminants in coastal zones. This study points out an additional feature of this simultaneous deployment: Determination of bioaccumulation factors in marine organisms without any need of HOC concentration data from the surrounding water environment. For this purpose, the concentration ratio model was modified successfully in the implementation of field data for Turkish coastal zones. Additionally, this same model equation was used for the comparison of different periods of deployment time, that could be a guide for an efficient planning of field studies with fewer expenses in consumables and transportation.

Conflict of interests: The author declares no financial, commercial and legal conflict of interest.

Ethics committee approval: In this study, the mathematical evaluation of a number of biomonitoring studies has been performed. Thus, the author declares that it does not need any ethics committee approval.

Funding: This study has been supported by Wageningen Research University (WUR) WIMEK Fellowship (The Wageningen Institute for Environment and Climate Research) and Istanbul Technical University Scientific Research Projects Unit (ITU – BAP Project Code: MAB-2018-41619).

Acknowledgments: The author thanks to Dr. Andreas Focks from WUR for his valuable contributions on the development of the code used in the study. The author also appreciates the anonymous reviewers for their comments and suggestions on the improvement of the manuscript.

Disclosure: -

REFERENCES

- Ahmad, R., Salem, N. M., & Estaitieh, H. (2010). Occurrence of organochlorine pesticide residues in eggs, chicken and meat in Jordan. Chemosphere, 78(6), 667–671. [CrossRef]
- Amdany, R., Chimuka, L., Cukrowska, E., Kukučka, P., Kohoutek, J., Tölgyessy, P., & Vrana, B. (2014). Assessment of bioavailable fraction of POPS in surface water bodies in Johannesburg City, South Africa, using passive samplers: An initial assessment. *Environmental Monitoring and Assessment*, 186(9), 5639–5653. [CrossRef]
- André Lourenço, R., Francisco de Oliveira, F., Haddad Nudi, A., Rebello Wagener, Â. de L., Guadalupe Meniconi, M. de F., & Francioni, E. (2015). PAH assessment in the main Brazilian offshore oil and gas production area using semi-permeable membrane devices (SPMD) and transplanted bivalves. *Continental Shelf Research*, 101, 109–116. [CrossRef]
- Baskaran, S., Armitage, J. M., & Wania, F. (2019). Model-based exploration of the variability in lake trout (Salvelinus namaycush) bioaccumulation factors: The influence of physiology and trophic relationships. *Environmental Toxicology and Chemistry*, 38(4), 831– 840. [CrossRef]
- Bayne, B. L. (1976). Watch on mussels. *Marine Pollution Bulletin, 7*(12), 217–218. [CrossRef]
- Blasco, C., & Picó, Y. (2009). Prospects for combining chemical and biological methods for integrated environmental assessment. TrAC -Trends in Analytical Chemistry, 28(6), 745–757. [CrossRef]
- Booij, K., Smedes, F., Van Weerlee, E. M., & Honkoop, P. J. C. (2006). Environmental monitoring of hydrophobic organic contaminants: The case of mussels versus semipermeable membrane devices. *Environmental Science and Technology*, 40(12), 3893–3900. [CrossRef]
- Bourgeault, A., & Gourlay-Francé, C. (2013). Monitoring PAH contamination in water: Comparison of biological and physico-chemical tools. *Science of the Total Environment*, 454–455, 328–336. [CrossRef]
- Cardellicchio, N., Buccolieri, A., Giandomenico, S., Lopez, L., Pizzulli, F., & Spada, L. (2007). Organic pollutants (PAHs, PCBs) in sediments from the Mar Piccolo in Taranto (Ionian Sea, Southern Italy). *Marine Pollution Bulletin*, 55(10–12), 451–458. [CrossRef]
- Cetin, B., Yurdakul, S., Keles, M., Celik, I., Ozturk, F., & Dogan, C. (2017). Atmospheric concentrations, distributions and air-soil exchange tendencies of PAHs and PCBs in a heavily industrialized area in Kocaeli, Turkey. *Chemosphere*, *183*(x), 69–79. [CrossRef]
- David, A., Gomez, E., Aït-Aïssa, S., Bachelot, M., Rosain, D., Casellas, C., & Fenet, H. (2010). Monitoring organic contaminants in small French coastal lagoons: Comparison of levels in mussel, passive sampler and sediment. *Journal of Environmental Monitoring*, 12(7), 1471– 1481. [CrossRef]
- Fenik, J., Tankiewicz, M., & Biziuk, M. (2011). Properties and determination of pesticides in fruits and vegetables. TrAC - Trends in Analytical Chemistry, 30(6), 814–826. [CrossRef]
- Fontenelle, F. R., Taniguchi, S., da Silva, J., & Lourenço, R. A. (2019). Environmental quality survey of an industrialized estuary and an Atlantic Forest Biosphere Reserve through a comparative appraisal of organic pollutants. *Environmental Pollution*, 248, 339–348. [CrossRef]

- Gourlay-Francé, C., Lorgeoux, C., & Tusseau-Vuillemin, M. H. (2008). Polycyclic aromatic hydrocarbon sampling in wastewaters using semipermeable membrane devices: Accuracy of time-weighted average concentration estimations of truly dissolved compounds. *Chemosphere*, 73(8), 1194–1200. [CrossRef]
- Gourlay, C., Miège, C., Noir, A., Ravelet, C., Garric, J., & Mouchel, J. M. (2005). How accurately do semi-permeable membrane devices measure the bioavailability of polycyclic aromatic hydrocarbons to Daphnia magna? *Chemosphere*, *61*(11), 1734–1739. [CrossRef]
- Gourlay, C., Tusseau-Vuillemin, M. H., Garric, J., & Mouchel, J. M. (2003). Effect of dissolved organic matter of various origins and biodegradabilities on the bioaccumulation of polycyclic aromatic hydrocarbons in Daphnia magna. *Environmental Toxicology and Chemistry*, 22(6), 1288–1294. [CrossRef]
- Greenwood, R., Mills, G. A., & Roig, B. (2007). Introduction to emerging tools and their use in water monitoring. *TrAC - Trends in Analytical Chemistry*, 26(4), 263–267. [CrossRef]
- Harman, C., Brooks, S., Sundt, R. C., Meier, S., & Grung, M. (2011). Field comparison of passive sampling and biological approaches for measuring exposure to PAH and alkylphenols from offshore produced water discharges. *Marine Pollution Bulletin*, 63(5–12), 141–148. [CrossRef]
- Helou, K., Harmouche-Karaki, M., Karake, S., & Narbonne, J. F. (2019). A review of organochlorine pesticides and polychlorinated biphenyls in Lebanon: Environmental and human contaminants. *Chemosphere*, 231, 357–368. [CrossRef]
- Hong, S. H., Yim, U. H., Shim, W. J., Li, D. H., & Oh, J. R. (2006). Nationwide monitoring of polychlorinated biphenyls and organochlorine pesticides in sediments from coastal environment of Korea. *Chemosphere*, 64(9), 1479–1488. [CrossRef]
- Huckins, J. N., Petty, J. D., & Booij, K. (2006). Monitors of Organic Chemicals in the Environment: Semipermeable Membrane Devices.
- Huckins, J. N., Tubergen, M. W., & Manuweera, G. K. (1990). Semipermeable-membrane devices containing model lipid - a new approach to monitoring the bioavailability of lipophilic contaminants and estimating their bioconcentration potential. *Chemosphere*, 20(5), 533–552. [CrossRef]
- Huckins, J. N., Petty, J. D., Orazio, C. E., Lebo, J. A., Clark, R. C., Gibson, V. L., Echols, K. R. (1999). Determination of uptake kinetics (sampling rates) by lipid-containing semipermeable membrane devices (SPMDs) for polycyclic aromatic hydrocarbons (PAHs) in water. *Environmental Science and Technology*, 33(21), 3918–3923. [CrossRef]
- Karacik, B., Okay, O. S., Henkelmann, B., Pfister, G., & Schramm, K. W. (2013). Water concentrations of PAH, PCB and OCP by using semipermeable membrane devices and sediments. *Marine Pollution Bulletin*, 70(1–2), 258–265. [CrossRef]
- Kim, U. J., Kim, H. Y., Alvarez, D., Lee, I. S., & Oh, J. E. (2014). Using SPMDs for monitoring hydrophobic organic compounds in urban river water in Korea compared with using conventional water grab samples. *Science of the Total Environment*, 470–471, 1537–1544. [CrossRef]
- Lance, E. W., Matz, A. C., Reeves, M. K., & Verbrugge, L. A. (2012). Petroleum hydrocarbon contamination in Nelson Lagoon, Alaska, sampling three different matrices. *Marine Pollution Bulletin*, 64(10), 2129–2134. [CrossRef]
- Lourenço, R. A., de Oliveira, F. F., de Souza, J. M., Nudi, A. H., de Luca Rebello Wagener, Â., de Fátima Guadalupe Meniconi, M., & Francioni, E. (2016). Monitoring of polycyclic aromatic hydrocarbons in a produced water disposal area in the Potiguar Basin, Brazilian equatorial margin. *Environmental Science and Pollution Research*, 23(17), 17113–17122. [CrossRef]

- Luellen, D. R., & Shea, D. (2002). Calibration and field verification of semipermeable membrane devices for measuring polycyclic aromatic hydrocarbons in water. *Environmental Science and Technology*, 36(8), 1791–1797. [CrossRef]
- Mayer, P., Tolls, J., Hermens, J. L. M., & Mackay, D. (2003). Equilibrium Sampling Devices. Environmental Science & Technology, 37(9), 184A-191A. [CrossRef]
- Okay, O. S., Karacik, B., Güngördü, A., Ozmen, M., Yilmaz, A., Koyunbaba, N. C., & Schramm, K. W. (2014). Micro-organic pollutants and biological response of mussels in marinas and ship building/breaking yards in Turkey. Science of the Total Environment, 496, 165–178. [CrossRef]
- Okay, O. S., Karacık, B., Güngördü, A., Yılmaz, A., Koyunbaba, N. C., Yakan, S. D., & Ozmen, M. (2017). Monitoring of organic pollutants in marine environment by semipermeable membrane devices and mussels: accumulation and biochemical responses. *Environmental Science and Pollution Research*, 24(23). [CrossRef]
- Okay, O. S., Karacık, B., Güngördü, A., Yılmaz, A., Koyunbaba, N. C., Yakan, S. D., & Ozmen, M. (2017). Monitoring of organic pollutants in marine environment by semipermeable membrane devices and mussels: accumulation and biochemical responses. *Environmental Science and Pollution Research*, 24(23), 19114–19125. [CrossRef]
- Ozcan, S., & Aydin, M. E. (2009). Organochlorine pesticides in urban air: Concentrations, sources, seasonal trends and correlation with meteorological parameters. *Clean - Soil, Air, Water, 37*(4–5), 343–348. [CrossRef]
- Peven, C. S., Uhler, A. D., & Querzoli, F. J. (1996). Caged mussels and semipermeable membrane devices as indicators of organic contaminant uptake in Dorchester and Duxbury Bays, Massachusetts. *Environmental Toxicology and Chemistry*, 15(2), 144–149. [CrossRef]
- Qiu, Y. W., Qiu, H. L., Zhang, G., & Li, J. (2019). Bioaccumulation and cycling of organochlorine pesticides (OCPs) and polychlorinated biphenyls (PCBs) in three mangrove reserves of south China. *Chemosphere*, 217, 195–203. [CrossRef]
- Taylor, A. C., Fones, G. R., Vrana, B., & Mills, G. A. (2019). Applications for Passive Sampling of Hydrophobic Organic Contaminants in Water-A Review. Critical Reviews in Analytical Chemistry, 8347. [CrossRef]
- Uher, E., Mirande-Bret, C., & Gourlay-Francé, C. (2016). Assessing the relation between anthropogenic pressure and PAH concentrations in surface water in the Seine River basin using multivariate analysis. *Science of the Total Environment*, *557–558*, 551–561. [CrossRef]
- Verweij, F., Booij, K., Satumalay, K., Van Der Molen, N., & Van Der Oost, R. (2004). Assessment of bioavailable PAH, PCB and OCP concentrations in water, using semipermeable membrane devices (SPMDs), sediments and caged carp. *Chemosphere*, 54(11), 1675–1689. [CrossRef]
- Vrana, B., Klučárová, V., Benická, E., Abou-Mrad, N., Amdany, R., Horáková, S., & Gans, O. (2014). Passive sampling: An effective method for monitoring seasonal and spatial variability of dissolved hydrophobic organic contaminants and metals in the Danube river. *Environmental Pollution*, 184, 101–112. [CrossRef]
- Yilmaz, A., Karacik, B., Henkelmann, B., Pfister, G., Schramm, K. W., Yakan, S. D., & Okay, O. S. (2014). Use of passive samplers in pollution monitoring: A numerical approach for marinas. *Environment International*, 73. [CrossRef]
- Zhao, D., Zhang, P., Ge, L., Zheng, G. J., Wang, X., Liu, W., & Yao, Z. (2018). The legacy of organochlorinated pesticides (OCPs), polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs) in Chinese coastal seawater monitored by semi-permeable membrane devices (SPMDs). *Marine Pollution Bulletin*, 137(April), 222–230. [CrossRef]