

New Trend Med Sci 2020; 1(2): 84-89.

https://dergipark.org.tr/tr/pub/ntms

Medicine Sciences

New Trends in

The Effect of Apilarnil on the Autophagia Against Lipolysaccarite-Based Sepsis in Liver

Züleyha Doğanyiğit^{1*}, Betül Köklü¹, Arda Üner¹, Aslı Okan¹, Emin Kaymak¹, Sibel Silici²

¹Department of Histology and Embryology, Faculty of Medicine, Yozgat Bozok University, 66100 Yozgat, Turkey

²Department of Agricultural Biotechnology, Faculty of Agriculture, Erciyes University, Nutral Therapy Co. Erciyes Technopark, 38039 Kayseri, Turkey

Article History Received 25 July 2020 Accepted 18 Aug 2020 Published Online 30 Sep 2020

**Corresponding Author* Dr Züleyha Doğanyiğit Department of Histology-Embryology, Faculty of Medicine, Yozgat Bozok University, Yozgat, 66100, Turkey, Phone: +90 3542126201, Fax: +90 354 4375285 E-mail: zuleyha.doganyigit@gmail.com ORCID:http://orcid.org/0000-0002-6980-3384 Abstract: Sepsis, triggered by highly bacterial lipopolysaccharide (LPS) endotoxins, exhibits high morbidity and mortality despite medical advances. Damage to the liver occurs due to the production of highly reactive oxygen species (ROS) and the release of various proinflammatory cytokines. It is suggested that autophagy, which regulates inflammation and selectively destroys damaged mitochondria, suppresses apoptosis and provides a possible protective mechanism in the endotoxic liver. However, apilarnil, a bee product, is known to have high antioxidant activity and positive effects against various diseases thanks to its polyphenols. In this study, it is aimed to reveal the potential protective effect of apilarnil on the autophagy mechanism in the endotoxic liver model LPSinduced. 64 male Sprague Dawley rats weighing 200-250 g; control, apilarnil treated groups (0.2, 0.4 and 0.8 g/kg), LPS (30 mg/kg) group and LPS+apilarnil treated groups (LPS+0.2 g/kg, LPS+0.4 g/kg and LPS+0.8 g/kg) are randomly divided into eight groups. Beclin-1, LC3 and P62 proteins were analyzed immunohistochemically in order to determine the activity level of autophagy pathway in the liver tissues taken after the completion of the experiment protocol. The data obtained showed that Beclin-1 immunoreactivity decreased while LC3 and P62 expression increased in the tissues of the LPS group compared to the control group. When apilarnil was applied with LPS, it was determined that there was an increase in Beclin-1 level (p>0.05) and a decrease in P62 levels (p<0.05) depending on the dose increase. Apilarnil increases the activity of the autophagy pathway and shows potential positive effects by providing a significant decrease on LC3 and P62 protein expression increased by LPS. However, the role of apilarnil in the autophagy pathway, which is a possible protector against LPSinduced sepsis, should be further investigated. ©2020 NTMS. Keywords: Beclin-1, LC3, P62, Apilarnil, LPS, Liver.

1. Introduction

Sepsis; an uncontrolled immune response that is triggered by infection, trauma, or toxins and occurs systemically. In the following process, it can lead to death by causing septic shock and multiple organ failure (1, 2). Lipopolysaccharide (LPS) is the main structural component of the outer cell membrane of gram-negative bacteria (3). Bacterial LPS, an endotoxin, is involved as a powerful microbial agent in

Cite this article as: Doğanyiğit Z, Köklü B, Üner A, Okan A, Kaymak E and Silici S. The Effect of Apilarnil on The Autophagia Against Lipolysaccarite-Based Sepsis in Liver, *New Trend Med Sci* **2020**; 1(2): 84-89.

the pathogenesis of sepsis and septic shock. LPS delivered to the blood initiates a potentially fatal series of inflammation mediators and procoagulant factor release.

Widespread endothelial damage in tissues leads to hypoperfusion and intravascular coagulation (4). During sepsis, pro-inflammatory responses occur with activation of the complement system, coagulation system, vascular endothelium, neutrophils and platelets. However, the immune system is suppressed due to the reprogramming of antigen presenting cells and apoptosis and depletion of lymphocytes (5).

Despite the constant advances in medicine, sepsis continues to be a global problem with high morbidity and mortality rates (6, 7). However, it is known that liver dysfunction accompanying sepsis has a significant effect on mortality rates. However, the pathological mechanism of sepsis-related liver dysfunction is very complex and has not been fully elucidated yet (8).

The liver plays a key role in the regulation of a wide range of metabolic, homeostatic and host defense activities under septic conditions (9). The liver provides both the clearance of circulating pathogenic microorganisms and toxins and the release of components of the liver-induced cytokines, inflammatory mediators, and coagulation cascade (10). The irregular inflammatory response that occurs following excessive LPS stimulation can kill bacteria as well as damage the liver, which has an effective function in defense responses (11).

LPS increases hepatocyte damage and leukocyte infiltration by interacting with TLR-4, a sub-member of the Toll-like receptor (TLR) family, which is highly expressed in Kupffer cells in the liver (12). Following increased TLR-4 stimulation, chemokine from Kupffer cells, various cytokines (tumor necrosis factor-alpha (TNF- α), interleukin (IL) -1 β , IL-6, IL-12 and IL-18), reactive oxygen species (ROS) and nitric oxide (NO) is secreted (13, 14). However, activation of the TLR-4 signal can interestingly contribute to the destruction of pathogens by inducing autophagy in immune cells (15, 16).

Autophagy is one of the innate and well-protected defense mechanisms against microbial attack. It controls the destruction of damaged organelles and various macro molecules, such as protein, through the formation of double membrane autophagosomes to provide cellular homeostasis (17, 18). It also plays an active role in the elimination of bacteria and pathogens in the cytoplasm (19).

Current studies in the literature reveal that in the case of sepsis, autophagy suppresses immune reactions and inhibits apoptosis by regulating inflammation and metabolism (20, 21). It also reduces cellular stress by preventing high ROS-induced LPS production and accumulation of damaged mitochondria (22). Autophagy can eliminate damaged mitochondria by selectively (23, 24). It also promotes regeneration of damaged proteins and organelles through lysosomal dependent degradation to deal with oxidative stress damage caused by ROS excess (25). Thus, autophagy has been shown to have a protective role against multiple organ failure, including liver damage (26). Various regulatory proteins such as Beclin-1, LC3 and P62, located in the autophagy pathway, allow examination of the autophagy activity in tissues.

Apilarnil is a poorly studied and little known biologically active bee product (27). It is obtained by freezing and breaking the 3-7-day old bee larvae with the nutrients they contain (28). In the analysis of the chemical composition of drones, it was observed that it contains a high rate of water, carbohydrates and lipids (29). In another analysis study conducted in 2019, similarly high protein content was determined, and 16 kinds of amino acids were detected. It has also been shown to be rich in phenolic compounds. It is suggested that it can perform its biologically beneficial activity through its antioxidant and radical inhibitory activity (30). In addition, current studies reveal that apilarnil has positive effects against gastrointestinal diseases, toothache, muscle fatigue, respiratory problems and male infertility (31).

In this study, the potential protective effect of apilarnil against LPS-induced liver sepsis will be examined. It was aimed to reveal the effect of apilarnil on the autophagy mechanism by performing immunohistochemical staining of Beclin-1, LC3 and P62 proteins in the liver tissue of the rats with sepsis model.

2. Material and Methods

2.1. Chemicals

Lipopolysaccharide was provided from Sigma Aldrich (*Escherichia coli* LPS, serotype 0127: B8) and Lyophilized Apilarnil was provided from Nutral Therapy Co Ltd (Erciyes University Technopark, Kayseri).

2.2. Experiment groups

Ethics committee approval was received from Erciyes University Animal Experiments Local Ethics Committee (HADYEK) for this study (Protokol No: 18/063). Animals were housed under controlled conditioning (on a 12-h light/12-h dark cycle at a temperature of 22-25 °C and air humidity of 55 %). Throughout the study, the animals were provided with ad libitum rat feed and drinking water. Lyophilized apilarnil distilled water was also dissolved and administered to animals by oral gavage. Sixty-four adult male *Sprague Dawley* rats (200-250 g weighing) were randomly divided into 8 groups:

- LPS group: 30 mg/kg/body weight (bw) LPS single dose i.p. (32).

- 0.2 g/kg API group: 0.2 g/kg/bw oral gavage for 10 days (33),

- 0.4 g/kg API group: 0.4 g/kg/bw oral gavage for 10 days,

- 0.8 g/kg API group: 0.8 g/kg/bw gavage for 10 days,

⁻ Control group: 1 ml saline (SF) (0.9% NaCl) i.p.

- 0.2 g/kg API+LPS: 0.2 g/kg API for 10 days, after 60 min LPS (single-dose i.p., 30 mg/kg)

- 0.4 g/kg API+LPS: 0.4 g/kg API for 10 days, after 60 min LPS (single-dose i.p., 30 mg/kg)

- 0.8 g/kg API+LPS: 0.8 g/kg API for 10 days, after 60 min LPS (single-dose i.p., 30 mg/kg)

Experimental animals were anesthetized with Ketamine (70 mg/kg/bw) and Xylazine (10 mg/kg/bw) six hours after administration and liver tissues were removed.

2.3. Immunohistochemical analysis

Immunoreactivity of Beclin-1. LC3A/B and P62/SQSTM1 in liver tissues of the LPS-induced rat sepsis model was determined using the Avidin-Biotin peroxidase method (34). Briefly, after deparaffinization of sections taken at 5µm thickness, citrate buffer was used to open the epitopes (pH: 6.0). The slides were then taken into a 3% hydrogen peroxide solution in methanol to prevent endogenous peroxidase activity. Ultra V block solution was applied to prevent nonspecific staining. Sections were then incubated overnight at 4°C with primary antibodies. Biotinylated secondary streptavidin-HRP and DAB chromogens were applied, respectively. And then sections were counterstained with Gill Hematoxylin. It was dehydrated by passing through increasing alcohol series and closed with a concealer called entellan. Sections were examined with Olympus BX53 light microscope. The evaluation of the immunoreactivity levels was done with Image J program. 10 different areas were evaluated for each slide.

2.4. Statistical analysis

Experimental data were statistically analyzed in GraphPad Prism (version 6.0, GraphPad Software Inc., San Diego, California) and presented as Mean \pm SEM. Data were analyzed using one-way ANOVA with Tukey's post hoc tests for multiple comparisons. P<0.05 was considered significant.

3. Results

As shown in Figure 1, it was observed that Beclin-1 decreased while LC3A/B and P62/SQSTM1 were increased in liver samples belonging to LPS group. In groups where LPS and apilarnil were applied together, while Beclin-1 increased with the increase of apilarnil dose (P>0.05), LC3A/B and P62/SQSTM1 decreased with the increase of apilarnil dose (P<0.05).

4. Discussion

During sepsis, cells are exposed to increased oxidative stress and metabolic demands. All of these processes

can lead to cell damage and cell death (35). However, cells and tissues try to protect themselves from such stresses by activating autophagy and autophagy-like mechanisms (36).

Autophagy is critical for maintaining normal human physiology, such as cellular homeostasis, energy balance, development, and cellular defense (37). Autophagy can also play a role in the pathogenesis of cancer, neurodegenerative diseases, aging, muscle diseases, infectious diseases and immune system diseases (38, 39). In addition, the presence of autophagy in liver tissue in the case of sepsis has been reported in studies (35, 40). In this study, we aimed to determine the effects of apilarnil, a natural bee product, on the autophagy mechanism that may occur as a result of endotoxic shock caused by LPS.

In lysosomal activity during autophagy, the P62 protein is considered a biomarker of autophagy and an indicator of autophagic flow. It is also characterized as an indicator of autophagic flow prevention of P62 reduction (41). Immunohistochemical analysis of LC3A or LC3B has also been used to investigate the level of autophagy occurring in the tissue (42). In addition, in clinical and experimental studies, Beclin-1, which is one of the main proteins of autophagy, is examined as a target for the determination of the presence of autophagy (43). Accordingly, in the sepsis model created with LPS, we performed the immunohistochemical analysis of Beclin-1, LC3A/B and P62 parameters as autophagy markers in liver tissue. According to the results obtained, LC3A/B expression was significantly increased only in the groups injected with LPS compared to the control. These results confirmed the presence of autophagy in liver tissue as a result of sepsis caused by LPS endotoxicity (Figure 1C). However, it was determined that Beclin-1 expression decreased significantly and P62 expression increased as a result of LPS injection (Figure 1B and 1D).

In the study performed by Chen et al. (44), LC3A/B, Beclin-1 and P62 proteins were examined to evaluate autophagy in hepatocytes in the sepsis model created by LPS injection in C57BL/6 mice. As a result of the study, significant changes were observed in LC3 and P62, while no increase was observed in Beclin-1 (44).

According to these results, it can be said that autophagy associated with LPS in liver tissue is not related to Beclin-1 and P62 molecules. Since LPS provides its effect with the TLR-4 receptor (45), more detailed studies are needed to clarify the mechanism underlying the relationship between the TLR-4 receptor and the LPS-induced autophagic pathways.



Figure 1. A. Beclin-1, LC3A/B and P62/SQSTM1 images of immunostaining in rat livers belonging to experiment groups. Histogram graphics of Beclin-1 (B), LC3A/B (C) and P62/SQSTM1 (D) represent the intensity values in percent of immunostaining obtained using Image J software. T bar graph data are expressed as mean \pm SEM, and compared by one-way ANOVA and TUKEY's multiple comparisons test (a P<0.05 vs. control group; b P<0.05 vs. LPS group; c P<0.05 vs. 0,2 g/kg API group; d P<0.05 vs. 0,4 g/kg API group; e P<0.05 vs 0,8 g/kg API group; f P<0.05 vs. LPS+0,2 g/kg API group; g P<0.05 vs LPS+0,4 g/kg API group; * P<0.05 statistically different from all other groups).

In sepsis, organ damage initially progresses to dysfunction without cell death and structural damage. Therefore, it is possible to restore organ function (45). Therefore, in another study we conducted, we investigated the protective role of apilarnil in the LPSinduced sepsis model in liver tissue and observed its positive effects on TLR-4 receptor-associated inflammatory response (34). In the results obtained from the current study, P62 expression increased in the LPS group compared to the control and only apilarniltreated groups. However, in groups in which apilarnil was administered with LPS, it showed decrease compared to the LPS group. Similarly, LC3A/B expression levels increased statistically in the LPS group compared to the control group and apilarnil-only groups (0.2, 0.4, 0.8 g/kg API). The expression amount of LC3A/B decreased in groups where apilarnil was administered with LPS, although it was not significant. No significant change was observed in Beclin-1 activity. With this result, it can be thought that apilarnil plays a role in the adaptation mechanism by acting on autophagy, LC3A/B and P62 pathways, which occur as a metabolic response to sepsis. Further research is needed to clarify the effects of this role on survival and immune response.

In another study conducted by Carchman et al. (46), adaptation metabolic responses of liver tissue to LPS sepsis model were investigated. Mitochondrial homeostasis and mitophagic pathways have been investigated in relation to the TLR-9 receptor. In the results obtained, it has been reported that TLR-9 receptor and mitophagia play an important role in the adaptation of hepatocytes to sepsis (46). Accordingly, it may be important for future studies to investigate the effects of apilarnil on mitophagia, which is a more featured pathway of TLR-9 and autophagy, and also on mitochondrial homeostasis, one of the key players of oxidative stress.

In the literature review, there was no study evaluating the effect of apilarnil on autophagy in the LPS-induced sepsis model, and the studies evaluating the effects on autophagy in other bee products are quite limited. We believe that this study will contribute to the literature on this subject.

5. Conclusions

In the light of all this information, it is important to develop alternative therapeutic agents targeting the autophagy pathway against sepsis, which results in high mortality. In this study, we investigated the effect of apilarnil on sepsis-induced autophagy in liver tissue through Beclin-1, LC3A/B and P62 molecules. As a result, apilarnil exhibited different effects in terms of different markers on the autophagy pathway. Apilarnil increases the activity of the autophagy pathway and shows potential positive effects by providing a significant decrease on P62 protein. However, an increase in its effect was observed due to the increased dose. In the light of the findings, apilarnil, which is a natural bee product, can be targeted in further studies as an alternative treatment method by playing a which is a natural bee product, can be targeted in further studies as an alternative treatment method by playing a protective role against sepsis, especially through the LC3A/B and P62 pathways.

Conflict of interest statement

There is no conflict of interest.

Acknowledgement

This work was supported by Yozgat Bozok University Project Coordination Application and Research Center (Project Number: 6602a-TF/20-375).

References

- 1. Yan J, Li S, Li S. The role of the liver in sepsis. *Int Rev Immunol* **2014**; 33(6): 498-510.
- 2. Hotchkiss RS, Moldawer LL, Opal SM, Reinhart K, Turnbull IR, Vincent JL. Sepsis and septic shock. *Nat Rev Dis Primers* 2016; 2: 16045.
- **3.** Maldonado RF, Sa-Correia I, Valvano MA. Lipopolysaccharide modification in Gram-negative bacteria during chronic infection. *FEMS Microbiol Rev* **2016**; 40(4): 480-493.
- 4. Opal SM. Endotoxins and other sepsis triggers. *Contrib Nephrol* 2010; 167: 14-24.
- van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol* 2017; 17(7): 407-420.
- 6. Cecconi M, Evans L, Levy M, Rhodes A. Sepsis and septic shock. *Lancet* 2018; 392(10141): 75-87.

- 7. Sakr Y, Jaschinski U, Wittebole X, et al. Sepsis in Intensive Care Unit Patients: Worldwide Data From the Intensive Care over Nations Audit. *Open Forum Infect Dis* **2018**; 5(12): 313.
- Woznica EA, Inglot M, Woznica RK, Lysenko L. Liver dysfunction in sepsis. *Adv Clin Exp Med* 2018; 27(4): 547-551.
- **9.** Nesseler N, Launey Y, Aninat C, Morel F, Malledant Y, Seguin P. Clinical review: The liver in sepsis. *Crit Care* **2012**; 16(5): 235.
- 10. Srivastava B, Gimson A. Hepatic changes in systemic infection. *Best Pract Res Clin Gastroenterol* 2013; 27(4): 485-495.
- **11.** Marshall JC. New translational research provides insights into liver dysfunction in sepsis. *PLoS Med* **2012**; 9(11): e1001341.
- De Nardo D. Toll-like receptors: Activation, signalling and transcriptional modulation. *Cytokine* 2015; 74(2): 181-189.
- Kolios G, Valatas V, Manousou P, Xidakis C, Notas G, Kouroumalis E. Nitric oxide and MCP-1 regulation in LPS activated rat Kupffer cells. *Mol Cell Biochem* 2008; 319(1-2): 91-98.
- Wang D, Yin Y, Yao Y. Advances in sepsisassociated liver dysfunction. *Burns Trauma* 2014; 2(3): 97-105.
- **15.** Xu Y, Jagannath C, Liu XD, Sharafkhaneh A, Kolodziejska KE, Eissa NT. Toll-like receptor 4 is a sensor for autophagy associated with innate immunity. *Immunity* **2007**; 27(1): 135-144.
- **16.** Fujita K, Maeda D, Xiao Q, Srinivasula SM. Nrf2mediated induction of p62 controls Toll-like receptor-4-driven aggresome-like induced structure formation and autophagic degradation. *Proc Natl Acad Sci USA* **2011**; 108(4): 1427-132.
- **17.** Klionsky DJ, Abdelmohsen K, Abe A, et al. Guidelines for the use and interpretation of assays for monitoring autophagy (3rd edition). *Autophagy* **2016**; 12(1): 1-222.
- Yu L, Chen Y, Tooze SA. Autophagy pathway: Cellular and molecular mechanisms. *Autophagy* 2018; 14(2): 207-215.
- 19. Hu W, Chan H, Lu L, et al. Autophagy in intracellular bacterial infection. *Semin Cell Dev Biol* 2020; 101: 41-50.
- **20.** Feng Y, Liu B, Zheng X, Chen L, Chen W, Fang Z. The protective role of autophagy in sepsis. *Microb Pathog* **2019**; 131: 106-111.
- **21.** Yin X, Xin H, Mao S, Wu G, Guo L. The Role of Autophagy in Sepsis: Protection and Injury to Organs. *Front Physiol* **2019**; 10: 1071.
- **22.** Oami T, Watanabe E, Hatano M, et al. Suppression of T Cell Autophagy Results in Decreased Viability and Function of T Cells Through Accelerated Apoptosis in a Murine Sepsis Model. *Crit Care Med* **2017**; 45(1): 77-85.

- 23. Ding WX, Yin XM. Mitophagy: mechanisms, pathophysiological roles, and analysis. *Biol Chem* 2012; 393(7): 547-564.
- 24. Chang AL, Ulrich A, Suliman HB, Piantadosi CA. Redox regulation of mitophagy in the lung during murine Staphylococcus aureus sepsis. *Free Radic Biol Med* 2015; 78: 179-189.
- **25.** Baechler BL, Bloemberg D, Quadrilatero J. Mitophagy regulates mitochondrial network signaling, oxidative stress, and apoptosis during myoblast differentiation. *Autophagy* **2019**; 15(9): 1606-1619.
- 26. Aki T, Unuma K, Uemura K. Emerging roles of mitochondria and autophagy in liver injury during sepsis. *Cell Stress* 2017; 1(2):79-89.
- 27. Akçiçek E, Yücel B, Apiterapi'de Apilarnil. Arı Ürünleri ve Sağlık (Apiterapi). İzmir: Sidas Yayınevi; 2015.
- 28. Isidorov VA, Bakier S, Stocki M. GC-MS investigation of the chemical composition of honeybee drone and queen larva homogenate. J Apic Res 2016; 60(1): 111-120.
- **29.** Silici S. Honeybee Products and Apitherapy. *TURJAF* **2019** 7(9): 1249-1262.
- **30.** Silici S. Chemical Content and Bioactive Properties of Drone Larvae (Apilarnil). *Mellifera* **2019** 19(2):14-22.
- **31.** Meda A, Lamien CE, Millogo J, Romito M, Nacoulma OG. Therapeutic uses of honey and honeybee larvae in central Burkina Faso. *J Ethnopharmacol* **2004**; 95(1): 103-107.
- **32.** Doganyigit Z, Kup FO, Silici S, Deniz K, Yakan B, Atayoglu T. Protective effects of propolis on female rats' histopathological, biochemical and genotoxic changes during LPS induced endotoxemia. *Phytomed* **2013**; 20(7): 632-639.
- **33.** Kanbur M, Eraslan G, Beyaz L, et al. The effects of royal jelly on liver damage induced by paracetamol in mice. *Exp Toxicol Pathol* **2009**; 61(2): 123-132.
- **34.** Doganyigit Z, Okan A, Kaymak E, Pandir D, Silici S. Investigation of protective effects of apilarnil against lipopolysaccharide induced liver injury in rats via TLR 4/HMGB-1/NF-kappaB pathway. *Biomed Pharmacother* **2020**; 125: 109967.
- **35.** Watanabe E, Muenzer JT, Hawkins WG, et al. Sepsis induces extensive autophagic vacuolization in hepatocytes: a clinical and laboratory-based study. *Lab Invest* **2009**; 89(5): 549-561.
- **36.** Sun Q, Gao W, Loughran P, et al. Caspase 1 activation is protective against hepatocyte cell death by up-regulating beclin 1 protein and mitochondrial autophagy in the setting of redox stress. *J Biol Chem* **2013**; 288(22): 15947-15958.
- **37.** Kroemer G, Marino G, Levine B. Autophagy and the integrated stress response. *Mol Cell* **2010**; 40(2): 280-293.

- **38.** Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell* **2008**; 132(1): 27-42.
- 39. Betin VM, Lane JD. Caspase cleavage of Atg4D stimulates GABARAP-L1 processing and triggers mitochondrial targeting and apoptosis. *J Cell Sci* 2009; 122(Pt 14): 2554-66.
- **40.** Carchman EH, Rao J, Loughran PA, Rosengart MR, Zuckerbraun BS. Heme oxygenase-1-mediated autophagy protects against hepatocyte cell death and hepatic injury from infection/sepsis in mice. *Hepatology* **2011**; 53(6): 2053-2062.
- **41.** Puissant A, Fenouille N, Auberger P. When autophagy meets cancer through p62/SQSTM1. *Am J Cancer Res* **2012**; 2(4): 397-413.
- **42.** Lazova R, Camp RL, Klump V, Siddiqui SF, Amaravadi RK, Pawelek JM. Punctate LC3B expression is a common feature of solid tumors and associated with proliferation, metastasis, and poor outcome. *Clin Cancer Res* **2012**; 18(2): 370-379.
- **43.** Komatsu M, Kurokawa H, Waguri S, et al. The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1. *Nat Cell Biol* **2010**; 12(3): 213-223.
- **44.** Chen C, Deng M, Sun Q, Loughran P, Billiar TR, Scott MJ. Lipopolysaccharide stimulates p62dependent autophagy-like aggregate clearance in hepatocytes. *Biomed Res Int* **2014**; 2014: 267350.
- **45.** Waltz P, Carchman EH, Young AC, et al. Lipopolysaccaride induces autophagic signaling in macrophages via a TLR4, heme oxygenase-1 dependent pathway. *Autophagy* **2011**; 7(3): 315-320.
- **46.** Carchman EH, Whelan S, Loughran P, et al. Experimental sepsis-induced mitochondrial biogenesis is dependent on autophagy, TLR4, and TLR9 signaling in liver. *FASEB J* **2013**; 27(12): 4703-4711.

Authors' ORCID

Züleyha Doğanyiğit

http://orcid.org/0000-0002-6980-3384

Betül Köklü

http://orcid.org/0000-0003-3477-3290 Arda Üner

http://orcid.org/0000-0002-9657-7757 Aslı Okan

http://orcid.org/0000-0001-8152-7338 Emin Kaymak

http://orcid.org/0000-0002-3818-2693 Sibel Silici

http://orcid.org/0000-0003-2810-2917

https://dergipark.org.tr/tr/pub/ntms All Rights Reserved. © 2020 NTMS.