

## The Possible Beneficial Effects of Lazaroid U-74389G on Ovarian Torsion Detorsion Injury

Mustafa Can GÜLER<sup>1</sup>, Ayhan TANYELİ<sup>1\*</sup>

<sup>1</sup> Department of Physiology, Faculty of Medicine, Atatürk University, Erzurum, Turkey,

[mcanguler@yahoo.com](mailto:mcanguler@yahoo.com)  
[ayhan.tanyeli@atauni.edu.tr](mailto:ayhan.tanyeli@atauni.edu.tr)

### Abstract

It was planned to search the possible beneficial effects of Lazaroid U-74389G (Laz) on ovarian tissue injury caused by bilateral ovarian torsion detorsion (T/D) in the experimental rat model. Wistar type female rats were randomly allocated to 3 groups. Groups of this research were designed as sham, T/D, and T/D+Laz groups. In sham group, the abdomen was incised and sutured but no intervention was performed. In T/D group, following the incision, ovarian T/D model was carried out and the incision was sutured. In treatment group, Laz was administered intraperitoneally at 20 mg/kg dose just before detorsion. After the detorsion period, rats were sacrificed and ovarian tissues were excised. Oxidant parameters elevated and antioxidant activity declined significantly in T/D group compared to sham group in ovarian tissues. Laz treatment reversed the oxidant and antioxidant parameters. Thereby, Laz protected against T/D-induced ovarian tissue injury in experimental rats.

**Keywords:** Lazaroid U-74389G, Ovarian Torsion Detorsion, Ovary, Rat..

### 1. Introduction

Ovarian torsion detorsion (T/D) is among gynecological emergencies and it is widely observed in reproductive age (1). Ischemia reperfusion (I/R) injury induces oxidative stress and inflammation leading to tissue injury (2; 3). I/R injury enhances the generation of reactive oxygen species (ROS) and malondialdehyde (MDA) production (4). ROS acts on membrane lipids and elevates MDA level (5). MDA is a lipid peroxidation metabolite and used for oxidative stress determination (6). During detorsion, ROS production increases and plays role in ovarian injury (7). Antioxidant enzymes including superoxide dismutase (SOD) prevent oxidative injury (8). The levels of ROS and antioxidant activity determine the rate of oxidative stress (9). Interleukin-1 $\beta$  (IL-1 $\beta$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) exist in the beginning of inflammatory respond and induce the release of free radicals (10; 11).

Different agents have been studied to alleviate or eliminate I/R injuries in various organs (12-16). Lazaroid U-74389G (Laz), a lazaroid family member, blocks lipid peroxidation through removing free radicals (17; 18). Laz has been examined in various I/R injury models including renal I/R injury (19). Laz eased I/R-induced intestinal injury in a previous study (20). Laz declined lipid peroxidation through reduction in MDA level (21).

Current study was planned to investigate the protective effect of Laz against ovarian oxidative damage induced by T/D.

### 2. Materials and Methods

#### 2.1. Experimental Animals and Ethical Approval

Animals were procured from Atatürk University Experimental Animal Research and Application Center and also experimental steps were carried out at same place. The rats were housed in laboratory conditions such as polypropylene cages, appropriate humidity, temperature, etc. They could reach both food

Received:20.05.2020

Accepted:21.06.2020

Published:24.06.2020

\*Corresponding author: Ayhan TANYELİ, PhD

Department of Physiology, Faculty of Medicine, Atatürk University, Erzurum, Turkey

E-mail: [ayhan.tanyeli@atauni.edu.tr](mailto:ayhan.tanyeli@atauni.edu.tr)

Cite this article as: M. C. Guler and A. Tanyeli, The Possible Beneficial Effects of Lazaroid U-74389G on Ovarian Torsion Detorsion Injury, *Eastern Anatolian Journal of Science*, Vol. 6, Issue 1, 46-51, 2020.

and water but fasted 12 hours prior to experiment. Atatürk University Experimental Animals Local Ethics Committee permitted the study (07.11.2019/203).

### 2.2. Groups and Torsion/Detorsion Model

Prior to experiment, animals were fixed in supine position. Abdominal region was shaved, cleaned. Anesthesia was applied to animals before surgical process. Povidone-iodine was used for disinfection. 10 mg/kg i.p. xylazine hydrochloride (Rompun®, Bayer, Istanbul) and 60 mg/kg intraperitoneally (i.p.) ketamine (Ketalar®, Pfizer, Istanbul) were preferred for anesthesia as described before (22). Laz purchased from Sigma Aldrich Co.

18 Wistar Albino female rats, weighing 230-240 g, were randomly divided into 3 groups. Group I (sham group), the abdominal area applied 1-2 cm incision and repaired with 3/0 silk suture and no additional intervention was done. Group II (T/D group), following incision as described in group I, the ovaries, fallopian tubes, ovarian veins and arteria were rotated in clockwise 720 degrees and fixed with clamps for 3 hours. Then, clamps were removed and blood flow restarted for 3 hours (23; 24). Group III (T/D+Laz group), same steps in group II were done and 10 mg/kg Laz was administered i.p. to the rats just before detorsion. Finally, at the end of the experiment, the ovarian tissues were removed, cleaned and held frozen the analysis.

### 3. Biochemical assessments

Total antioxidant status (TAS) and total oxidant status (TOS) values were evaluated via appropriate kits (Rel Assay Diagnostics). TOS to TAS ratio, the oxidative stress index (OSI), was gauged as:  $OSI = [(TOS, \mu\text{mol H}_2\text{O}_2 \text{ equivalent L}) / (TAS, \text{mmol Trolox equivalent/L}) \times 10]$ . Evaluation of SOD activity depends on formazan dye level (25). Lipid peroxidation was measured by determining MDA level through thiobarbituric acid test (26). Myeloperoxidase (MPO) activity was gauged according to method described previously (27). IL-1 $\beta$  and TNF- $\alpha$  levels were evaluated via appropriate kits (Elabscience, Wuhan, China).

### 4. Statistical analysis

Data were analyzed with One-Way ANOVA and Tukey test using statistical package program, SPSS. All results were presented in table 1 and figure 1 as Mean $\pm$ SD. P value was considered significant when  $p < 0.05$ .

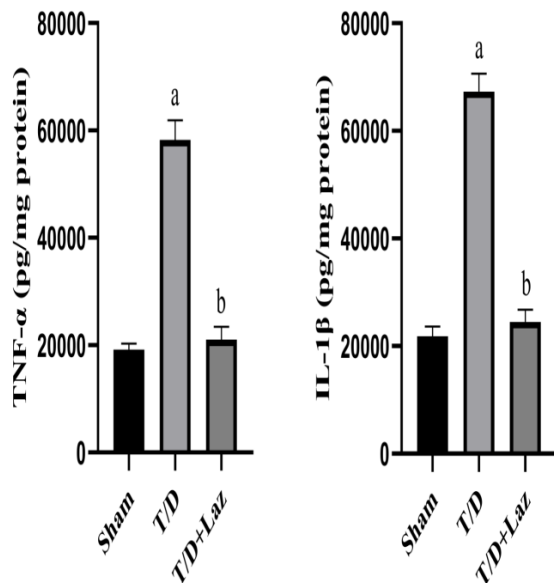
### 5. Results

TNF- $\alpha$  and IL-1 $\beta$  values, MDA, TAS, TOS, OSI levels, MPO and SOD activities in ovarian tissues were demonstrated in table 1 and figure 1. Oxidative parameters (MDA, TOS, OSI, MPO) and inflammatory mediators (TNF- $\alpha$ , IL-1 $\beta$ ) elevated significantly in T/D group compared to sham group. In Laz treatment group, these parameters declined significantly. Moreover, antioxidant activity (TAS and SOD) diminished in T/D group, whereas Laz treatment elevated TAS and SOD levels (Table 1; Figure 1,  $p < 0.001$ ).

**Table 1:** Results of biochemical parameters among all the experimental groups.

Experimental Group (n=6)	TAS (mmol/L)	TOS ( $\mu\text{mol/L}$ )	OSI (arbitrary unit)	SOD (U/mg protein)	MPO (U/g protein)	MDA ( $\mu\text{mol/g tissue}$ )
Sham	0.86 $\pm$ 0.04	6.6 $\pm$ 0.89	0.7 $\pm$ 0.10	437.02 $\pm$ 16.57	2594.71 $\pm$ 3024.66	64.6 $\pm$ 5.71
T/D	0.22 $\pm$ 0.03 <sup>a</sup>	11.6 $\pm$ 1.08 <sup>a</sup>	5.20 $\pm$ 0.97 <sup>a</sup>	174.32 $\pm$ 7.05 <sup>a</sup>	22.10 $\pm$ 4257.53 <sup>a</sup>	64 $\pm$ 11.83 <sup>a</sup>
T/D+Laz	0.82 $\pm$ 0.11 <sup>b</sup>	6.99 $\pm$ 0.37 <sup>b</sup>	0.84 $\pm$ 0.09 <sup>b</sup>	407.85 $\pm$ 20.51 <sup>b</sup>	13.46 $\pm$ 2458.82 <sup>b</sup>	68.6 $\pm$ 5.33 <sup>b</sup>

<sup>a</sup> $p < 0.001$  compared to sham group. <sup>b</sup> $p < 0.001$  compared to T/D group.



**Figure 1:** Results of IL-1 $\beta$  and TNF- $\alpha$  among all the experimental groups.

## 6. Discussion

Ovarian torsion (O/T) mostly occurs during reproductive period (28). Ischemic tissue injury is based on insufficient materials for energy supply. Ovarian detorsion means the recovery of blood flow. But replenishment of blood flow results in ovarian tissue injury (29). O/T is an emergency situation with a 3% prevalence (30; 31).

During ovarian T/D, blood reflow elevates in lactic acid, proinflammatory cytokine and lipid peroxide levels (7; 32; 33). Increased ROS damages cells via lipid peroxidation (34). MDA is a lipid peroxidation product and harmful for tissues. It reflects oxidative stress (6). It is created by ROS during I/R (35). It has been proven that oxidative stress causes tissue damage in various animal models (36-39). Neutrophil infiltration also leads to I/R injury besides oxidative stress. MPO activity represents neutrophil activation and infiltration (40). Neutrophil infiltration, TNF- $\alpha$ , IL-1 $\beta$  and several proinflammatory cytokine production accompany I/R (41). IL-1 $\beta$  plays role in apoptosis and inflammation (10; 42).

Besides ischemic injury, reperfusion also leads to injury in tissues (43). Antioxidant enzymes like SOD compose cellular defense system against oxidative injury (44). SOD is a crucial antioxidant enzyme and SOD activity declined during ovarian T/D in previous

studies (45; 46). Oxidative stress is the surpass of oxidant activity against antioxidant system. OSI is the ratio of TOS to TAS. It is preferred for the determination of oxidative stress (47; 48). TOS and TAS play role in I/R injury assessment (49).

Laz has been studied in various I/R injury models including renal I/R injury (19), intestinal I/R injury in rats (50). Different agents with feature anti-inflammatory, antioxidant and radical scavengers have beneficial effects have been reported in alleviation or elimination of I/R injuries (51-56). In the current study, we thought that Laz could minimize T/D damage with its antioxidant and anti-inflammatory effects. Therefore, current study was planned to investigate possible protective effects of Laz on ovarian tissue by using an ovarian T/D model.

Here, several inflammatory mediators and oxidative stress biomarkers were diminished and antioxidant activity was enhanced by Laz treatment in ovarian T/D injury model.

## 7. Conclusions

Laz alleviated T/D-induced ovarian injury in experimental rat model through declining oxidative mediators and elevating antioxidant parameters. This is a hope-inspiring result in order to evaluate for T/D pathologies.

## 8. Acknowledgement

None.

## 9. Conflict of interest statement

None.

## References

- OELSNER G, COHEN SB, SORIANO D, ADMON D, MASHIACH S, CARP H. 2003. Minimal surgery for the twisted ischaemic adnexa can preserve ovarian function. *Hum Reprod* 18:2599-602
- VAGHASIYA J, SHETH N, BHALODIA Y, MANEK R. 2011. Sitagliptin protects renal ischemia reperfusion induced renal damage in diabetes. *Regulatory peptides* 166:48-54
- SEHIRLI AO, SENER G, SATIROGLU H, AYANOGLU-DULGER G. 2003. Protective effect of N-acetylcysteine on renal ischemia/reperfusion injury in the rat. *Journal of nephrology* 16:75-80
- KUROSE I, GRANGER DN. 1994. Evidence implicating xanthine oxidase and neutrophils in reperfusion-induced microvascular

- dysfunction. *Annals of the New York Academy of Sciences* 723:158-79
- SOZER S, DINIZ G, LERMIOGLU F. 2011. Effects of celecoxib in young rats: histopathological changes in tissues and alterations of oxidative stress/antioxidant defense system. *Archives of pharmacal research* 34:253-9
- DEL RIO D, STEWART AJ, PELLEGRINI N. 2005. A review of recent studies on malondialdehyde as toxic molecule and biological marker of oxidative stress. *Nutrition, metabolism, and cardiovascular diseases : NMCD* 15:316-28
- BOZKURT S, ARIKAN DC, KURUTAS EB, SAYAR H, OKUMUS M, et al. 2012. Selenium has a protective effect on ischemia/reperfusion injury in a rat ovary model: biochemical and histopathologic evaluation. *J Pediatr Surg* 47:1735-41
- YASAR M, ERDI I, KAYA B. 2018. The preventive effects of atorvastatin and N-acetyl cysteine in experimentally induced ischemia-reperfusion injury in rats. *Bratisl Lek Listy* 119:167-74
- MCCORD JM. 1985. Oxygen-derived free radicals in postischemic tissue injury. *N Engl J Med* 312:159-63
- ELTZSCHIG HK, COLLARD CD. 2004. Vascular ischaemia and reperfusion injury. *British medical bulletin* 70:71-86
- DINARELLO CA. 2000. Proinflammatory cytokines. *Chest* 118:503-8
- ERASLAN E, TANYELI A, POLAT E, POLAT E. 2019. 8-Br-cADPR, a TRPM2 ion channel antagonist, inhibits renal ischemia-reperfusion injury. *J Cell Physiol* 234:4572-81
- ERASLAN E, TANYELI A, POLAT E, YETIM Z. 2019. Evodiamine alleviates kidney ischemia reperfusion injury in rats: A biochemical and histopathological study. *J Cell Biochem* 120:17159-66
- OZTURK D, ERDOGAN DG, TANYELI A, ÇOMAKLI S, BAYLAN H, POLAT E. 2019. The protective effects of urapidil on lung tissue after intestinal ischemia-reperfusion injury. *Turkish Journal of Biochemistry* 44:539
- TANYELI A, GULER MC, ERASLAN E, EKİNCİ AKDEMİR FN, et al. 2020. Barbaloin attenuates ischemia reperfusion-induced oxidative renal injury via antioxidant and anti-inflammatory effects. *Medicine Sciences* 9:246-50
- DOGAN C, HALICI Z, TOPCU A, CADIRCI E, KARAKUS E, et al. 2016. Effects of amlodipine on ischaemia/reperfusion injury in the rat testis. *Andrologia* 48:441-52
- ALTAVILLA D, SQUADRITO F, CAMPO GM, SQUADRITO G, ARLOTTA M, et al. 1999. The lazaroid, U-74389G, inhibits inducible nitric oxide synthase activity, reverses vascular failure and protects against endotoxin shock. *European journal of pharmacology* 369:49-55
- SAIJA A, TOMAINO A, PELLEGRINO ML, GIUFFRIDA N, TROMBETTA D, CASTELLI F. 2001. In vitro evaluation of the antioxidant activity and biomembrane interaction of the lazaroid U-74389G. *Life sciences* 68:1351-66
- GARVIN PJ, NIEHOFF ML, ROBINSON SM, MISTRY B, ESTERL R, et al. 1997. Renoprotective effects of the 21-aminosteroid U74389G in ischemia-reperfusion injury and cold storage preservation. *Transplantation* 63:194-201
- de OCA J, CUADRADO S, VALLET J, BENASCO C, MARTÍN E, et al. 1998. Protective effects of lazaroid U74389G on intestinal graft after heterotopic small bowel transplantation in rats. *The Journal of surgical research* 75:18-23
- FUKUMA K, MARUBAYASHI S, OKADA K, YAMADA K, KIMURA A, DOHI K. 1999. Effect of lazaroid U-74389G and methylprednisolone on endotoxin-induced shock in mice. *Surgery* 125:421-30
- EKİNCİ AKDEMİR FN, YILDIRIM S, KANDEMİR FM, TANYELI A, KUCUKLER S, BAHAEDDIN DORTBUDAK M. 2019. Protective effects of gallic acid on doxorubicin-induced cardiotoxicity; an experimental study. *Archives of physiology and biochemistry*:1-8
- GÜLER MC, TANYELI A. 2020. Role of Hyperoside on Ovarian Tissue Damage Created by Ovarian Torsion Detorsion. *New Trends in Medicine Sciences* 1:1-5
- GÜLER MC, TANYELI A, ERASLAN E, EKİNCİ AKDEMİR FN. 2020. Role of 6-Shogaol Against Ovarian Torsion Detorsion-Induced Reproductive Organ Damage. *New Trends in Medicine Sciences* 1:29-34
- SUN Y, OBERLEY LW, LI Y. 1988. A Simple Method for Clinical Assay of Superoxide-Dismutase. *Clin Chem* 34:497-500
- OHKAWA H, OHISHI N, YAGI K. 1979. Assay for Lipid Peroxides in Animal-Tissues by Thiobarbituric Acid Reaction. *Anal Biochem* 95:351-8
- BRADLEY PP, PRIEBAT DA, CHRISTENSEN RD, ROTHSTEIN G. 1982. Measurement of cutaneous inflammation: estimation of neutrophil content with an enzyme marker. *J Invest Dermatol* 78:206-9
- HOURY D, ABBOTT JT. 2001. Ovarian torsion: a fifteen-year review. *Ann Emerg Med* 38:156-9
- JENNINGS RB, MURRY CE, STEENBERGEN C, JR., REIMER KA. 1990. Development of cell

- injury in sustained acute ischemia. *Circulation* 82:1i2-12
- HIBBARD LT. 1985. Adnexal torsion. *Am J Obstet Gynecol* 152:456-61
- MASHIACH S, BIDER D, MORAN O, GOLDENBERG M, BEN-RAFAEL Z. 1990. Adnexal torsion of hyperstimulated ovaries in pregnancies after gonadotropin therapy. *Fertil Steril* 53:76-80
- ERGUN Y, KOC A, DOLAPCIOGLU K, AKAYDIN Y, DOGRUER G, et al. 2010. The protective effect of erythropoietin and dimethylsulfoxide on ischemia-reperfusion injury in rat ovary. *Eur J Obstet Gynecol Reprod Biol* 152:186-90
- SAK ME, SOYDINC HE, SAK S, EVSEN MS, ALABALIK U, et al. 2013. The protective effect of curcumin on ischemia-reperfusion injury in rat ovary. *Int J Surg* 11:967-70
- GUTTERIDGE JM. 1993. Free radicals in disease processes: a compilation of cause and consequence. *Free Radic Res Commun* 19:141-58
- GIROTTI AW. 1998. Lipid hydroperoxide generation, turnover, and effector action in biological systems. *J Lipid Res* 39:1529-42
- TANYELI AYHAN, GÜZEL ERDOĞAN D. 2019. Investigation into the Biochemical Effects of Barbaloin on Renal Tissue in Cecal Ligation and Puncture-Induced Polymicrobial Sepsis Model in Rats. *Southern Clinics of Istanbul Eurasia* 30:285-9
- TANYELI A, GÜZEL ERDOĞAN D. 2019. Alliin mitigates Cecal Ligation Puncture (CLP)-induced lung injury through antioxidant and antiinflammatory effects. *Turkish Journal of Sciences* 4:46-59
- EKINCI AKDEMİR FN, TANYELI A. 2019. The Antioxidant Effect of Fraxin against Acute Organ Damage in Polymicrobial Sepsis Model induced by Cecal Ligation and Puncture. *Turkish Journal of Science* 4:22-9
- TANYELI A, ERASLAN E, GULER MC, OZBEK SEBIN S, CELEBI D, OZGERIS FB, TOKTAY E. 2019. Investigation of biochemical and histopathological effects of tarantula cubensis D6 on lung tissue in cecal ligation and puncture-induced polymicrobial sepsis model in rats. *Medicine Science International Medical Journal* 8:644-50
- ZHANG Y, ZHU J, GUO L, ZOU Y, WANG F, et al. 2017. Cholecystokinin protects mouse liver against ischemia and reperfusion injury. *International immunopharmacology* 48:180-6
- RAUP-KONSAVAGE WM, GAO T, COOPER TK, MORRIS SM, JR., REEVES WB, AWAD AS. 2017. Arginase-2 mediates renal ischemia-reperfusion injury. *American journal of physiology. Renal physiology* 313:F522-f34
- HASTURK A, ATALAY B, CALISANELLER T, OZDEMIR O, ORUCKAPTAN H, ALTINORS N. 2009. Analysis of serum pro-inflammatory cytokine levels after rat spinal cord ischemia/reperfusion injury and correlation with tissue damage. *Turkish neurosurgery* 19:353-9
- GRANGER DN, KVIETYS PR. 2015. Reperfusion injury and reactive oxygen species: The evolution of a concept. *Redox Biol* 6:524-51
- HALLIWELL B, AESCHBACH R, LÖLIGER J, ARUOMA OI. 1995. The characterization of antioxidants. *Food Chem Toxicol* 33:601-17
- YIGITER M, HALICI Z, ODABASOGLU F, KELES ON, ATALAY F, et al. 2011. Growth hormone reduces tissue damage in rat ovaries subjected to torsion and detorsion: biochemical and histopathologic evaluation. *European journal of obstetrics, gynecology, and reproductive biology* 157:94-100
- BAKAN V, CIRALIK H, TOLUN FI, ATLI Y, MIL A, OZTURK S. 2009. Protective effect of erythropoietin on torsion/detorsion injury in rat model. *Journal of pediatric surgery* 44:1988-94
- RABUS M, DEMIRBAG R, SEZEN Y, KONUKOGLU O, YILDIZ A, et al. 2008. Plasma and tissue oxidative stress index in patients with rheumatic and degenerative heart valve disease. *Turk Kardiyoloji Dernegi arsivi : Turk Kardiyoloji Derneginin yayin organidir* 36:536-40
- EREL O. 2005. A new automated colorimetric method for measuring total oxidant status. *Clin Biochem* 38:1103-11
- YAZICI S, DEMIRTAS S, GUCLU O, KARAHAN O, YAVUZ C, et al. 2014. Using oxidant and antioxidant levels to predict the duration of both acute peripheral and mesenteric ischemia. *Perfusion* 29:450-5
- ANDREADOU I, POUSSIOS D, PAPALOIS A, GAVALAKIS N, ARONI K, et al. 2003. Effect of U-74389G (21-lazaroid) on intestinal recovery after acute mesenteric ischemia and reperfusion in rats. *In vivo (Athens, Greece)* 17:463-8
- TANYELI A, EKINCI AKDEMİR FN, ERASLAN E, GULER M, SEBIN OZBEK S, GÜLÇİN I. 2020. Role of p-Coumaric acid in Alleviating of the Intestinal Ischemia/Reperfusion Injury. *Kocaeli Medical Journal* 9:166-73
- TOPDAGI O, TANYELI A, EKINCI AKDEMİR FN, ERASLAN E, GULER MC, COMAKLI S. 2020. Preventive effects of fraxin on



- ischemia/reperfusion-induced acute kidney injury in rats. *Life sciences* 242:117217
- GUZEL ERDOĞAN, TANYELİ A. 2018. Investigation of Chlorogenic Acid (Cga) as An Antioxidant in Renal Ischemia-Reperfusion Injury: An Experimental Study. *Sakarya Medical Journal* 8:410-5
- GÜZEL ERDOĞAN D, TANYELİ A. 2018. Investigation of Oxidative Damage of Lung Tissue in Experimental Renal Ischemia Reperfusion Model and The Protective Effects of Chlorogenic Acid (CGA). *Sakarya Medical Journal* 8:260-5
- TOPDAĞI Ö, TANYELİ A, EKİNCİ AKDEMİR FN, GÜZEL ERDOĞAN D, GÜLER MC. 2020. Higenamine decreases testicular damage injured by ischemia reperfusion: A biochemical study *Turkish Journal of Science* 4