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Effect of probiotic on total antioxidant (TAS) and total oxidant (TOS) in treatment of newborn calf diarrhea

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ABSTRACT

Objective: Newborn calf diarrhea causes significant economic losses. Therefore, this study was aimed to determine the effect of adding probiotics on Total Antioxidant (TAS) and Total Oxidant (TOS) levels in routine treatment of calf diarrhea.

Material-Method: The material of the study consisted of 12 calves, routine treatment (Group 1, n=6 calves) and probiotic group. (Group 2; n=6 calves).

Results: TAS values increased statistically after treatment (P < 0.05) in group 2 compared to pretreatment. Comparison of the groups after treatment revealed that TAS significantly increased (P < 0.05) in group 2. While there was no statistical difference in TOS values between the groups a statistically significant increase (P <0.05) was detected after treatment compared to pretreatment.

Conclusion: It was concluded that adding probiotics may be useful in addition to routine treatment in newborn calf diarrhea.

Keywords: Antioxidant, Diarrhea, TAS, TOS, Probiotic

INTRODUCTION

The neonatal period is the most critical period of calf rearing, covering the days between 0 and 28 following birth (Alkan, 1998). This time is the period when the diseases are most common and the mortality rates are the highest, especially in the first 15 days of calf life. Neonatal diseases in calves includes infectious (bacterial, viral, parasitic and mycotic) and noninfectious aetiolody (vitamin, mineral substance and, trace element deficiencies, congenital anomalies, etc.) (Ok et al., 2009; Aslan 1986; Burgu et al., 1986). Diarrhea is the most common problem. Treatment of diarrhea in calves involves; nutrition and prevention of bacteremia / septicemia and, it is important to correct dehydration, acid base and electrolyte balance (Şen

et al., 2013). Use of antibiotics may become a public health problem due to development of antibiotic resistance. To reduce the dose and side effects of antibiotic treatment applied in neonatal calf diarrhea and to increase the therapeutic effectiveness probiotic applications are of great importance. Probiotics used in single stomach animals aid in regulation of the normal digestive flora, reducing metabolic disorders thus increase the efficiency in a healthy way (Minssen and Nordberg, 2020).

Probiotics have been started to be used in ruminants (Nageshwar et al., 2016; Valencia et al., 2017). Probiotics protect the host's useful microorganisms (the natural microflora) which can positively effect on the health of the host, help in

food digestion and vitamin production, and inhibit the pathogenic bacteria with substances produced in the intestines (Oelschlaeger, 2011). Application of probiotics in the treatment of diarrhea, shows a progressive effect by strengthening the immune system, and it has also been observed that the gastrointestinal mucosa barrier function, which is impaired in both in vitro and in vivo models, is rearranged by probiotics (Yeoman et al., 2018; Garcia-Lafuente et al., 2001; Madsen et al., 2001). Probiotics, effects on the immune system include, antimicrobial (Lactobacillus acidophilus; Acidophilin, Lactobacillin) secretions, increasing defense release from cryptos in the small intestine, such as preventing the adhesion of pathogens (Salmonella typhimurium) and they also neutralize virulence factors by breaking down toxins (Clostridium difficile toxins). (Solis-Pereira and Lemonnir, 1993). In some studies, the use of probiotics, promotes cytokine production in blood cells and improves the activity of macrophages (Solis-Pereira and Lemonnir, 1993; Shah, 2001). In addition, the use of probiotics in ruminant's benefits from the increase in yield characteristics, it benefits from digestive regulatory effects in condition and rumen (Wallace and Newbold, 1992; Burçak and Yalçın, 2013; Yavuzarslan, 2018; Alıç Ural and Toplu, 2017).

In intestinal cells there are several enzymatic and non-enzymatic antioxidants, including superoxide dismutase (SOD), glutathione (GSH) and catalase (CAT) which plays a role in the antioxsidants system (Çadırcı et al., 2007). Reactive Oxygen Species (ROS) that actively contribute to the ongoing pathogenic cascade with oxidative stress increases with the release of different proinflammatory mediators and it connects to oxidative stress (Lewis et al., 2008; Moura et al., 2015). In infectious diarrhea intestinal permeability is impaired and Gram (-) bacterial agents cause sepsis which is an important complication and oxidative stress increases in sepsis (Winterbourn et al., 2000; Oldham et al., 2002). Many roles of probiotics in antioxidation have been identified, however, there is no research related to probiotic effects on antioxidant and oxidant system in ruminants and neonatal calves (Shen et al., 2011; Asemi et al., 2013; Kullisaar et al., 2002). Mechanisms of probiotics in antioxidation can be listed as: (1) metal ion chelating ability (Ahire et al., 2013), (2) antioxidant enzymes system capacity (LeBlanc et al., 2008), (3) reduction in antioxidant metabolites (Kullisaar et al., 2002; Endo et al., 2013) and (4) enhanced antioxidant effect by adjusting the microbiota composition (Doron & Gorbach, 2006)

It has been understood that diarrhea and diarrhea complications cause an increase in oxidative stress, and antioxidant capacity in the intestines decreases in conditions and in this regard, no studies have been found on the effect of adding probiotics on TAS and TOS levels in neonatal calf diarrhea. This study was aimed to investigate the effect of probiotic use on the antioxidative mechanism in neonatal calf diarrhea treatments by using a probiotic combination of 2 bacteria (*Lactobacillus spp: L. plantarum, L. casei and Bacillus spp: B. subtilis*) and 1 yeast (*Saccharomyces spp: Saccharomyces cerevisiae*).

MATERIALS and METHODS

This study was carried out with the approval of Van Yüzüncü Yıl University Animal Experiments Local Ethics Committee (YÜHADYEK-code:03, 28.03.2019). The study included 12 calves with diarrhea from Van and its vicinity. The calves were those brought to the Department of Internal Medicine of the Faculty of Veterinary Medicine and / or private veterinary clinics for clinical examinations with the complaint of diarrhea. Diarrhoeic calves with moderate to severe dehydration, possible metabolic acidosis and hyperkalemia, hypothermia, arrhythmia and tachypnea were included in the study. All calves with diarrhea were divided into 2 groups.

I. Group animals (n=6); 1.3% NaHCO3, +0.9% NaCl + routine diarrhea treatment

II. Group animals (n=6); 1.3% NaHCO3 + 0.9% NaCl + Oral supplement (Probiotic) + routine diarrhea treatment

Blood samples were taken from the groups before treatment (0 hour) and on the third day after treatment,

Serum TAS was measured using the Commercial Kit (Product Code: RL0017, REL Assay Diagnostic, Mega T1p, Gaziantep, Turkey) developed by Erel (2004) on Biochrom Anthos Zenyth 200 rt Microplate Reader Device. Results were given as millimolar trolox equivalent per liter (mmol trolox equiv / L).

Serum TOS level was measured on Biochrom Anthos Zenyth 200 rt Microplate Reader using the Commercial Kit (Product Code: RL0017, REL Assay Diagnostic, Mega Tip, Gaziantep, Turkey) developed by Erel (2005). Statistical evaluation of data obtained in the research were paired t test to determine the statistical difference between the same parameter before and after treatment, variance analysis (ANOVA) to determine the significance of the difference between different groups before and after treatment and Duncan test to determine the difference between groups. SPSS 20.0 Statistical Package Program (SPSS Inc., Chicago, IL, USA) was used for this purpose. Statistical significance was determined as p < 0.05. All data are given as arithmetic mean \pm standard error of mean.

RESULTS

Clinically, there was dehydration and hypothermia along with diarrhoe and metabolic acidosis and hyperkalemia in sick animals. There was arrhythmia on the cardiac examination. Tachypnea was detected.

Serum TAS values before treatment in 2 groups significantly increased after treatment (P <0.05). When both groups were compared, a statistically significant increase (P <0.05) was observed in group 2 compared to group 1 after treatment (Table 1). While TOS values insignificantly decreased after treatment in group 2, the value significantly increased (P <0.05) after the treatment in group. TOS values significantly differed between the groups after treatment (P <0.05) (Table 1).

Groups	TAS (mmol Trolox Eq/L)		TOS (µmol H2O2 Eq/L)	
	Before Treatment	After Treatment	Before Treatment	After Treatment
I. Group	1.326±0.84	1.144±0.73ª	4.480±1.77*	7.784±1.42*
II. Group	1.116±0.59*	1.588±0.71 [*] a	5.492±1.68	4.178±1.89

*: The difference between the pre-treatment and post-treatment of the same group is statistically significant (p<0.05).

 $^{\rm a}$:The difference between groups is statistically significant (p<0.05).

DISCUSSION

The neonatal period covers the days 0 to 28 following birth and is the most critical time of calf breeding where, calf diseases especially are most common. Neonatal calf diseases and deaths cause serious economic losses (Alkan, 1998; Ok et al., 2009; Aslan, 1986). Recently, probiotic applications in addition to routine treatment are gaining importance in diarrhea treatments (Nageshwar et al., 2016; He et al., 2017; Le et al 2017). Diarrhea also triggers many local or systemic pathologies and causes an increase in oxidative stress (Lewis et al., 2008; Moura et al., 2015). Probiotics have been shown to reduce oxidative stress by many mechanisms (Ahire et al., 2013; LeBlanc et al., 2008; Endo et al., 2013; Kullisaar et al., 2002; Çatalkaya, 2020). Although there exist a few studies on the antioxidant and oxidant systems of calves. In a study conducted in healthy calves, TOS value was determined as 10.61 ± 3.82 (µmol H₂O₂ Eq/L) and TAS value was 1.72 ± 0.63 (mmol Trolox Eq/L) (Topraktaş, 2019). In another study, in calves with diarrhea, the value of TAS was 0.51 ± 0.02 (mmolTrolox Eq/L) before treatment and 0.55 ± 0.02 (mmolTrolox Eq/L) after treatment and the TOS value was 13.47 ± 0.81 (µmol), (H2O2 Eq / L) before treatment and 11.21 ± 0.26 (µmol H₂O₂ Eq / L) after treatment (Kabu et al., 2015). In a study conducted in calves with septicemia; TAS value was 0.50 ± 0.02 (mmolTrolox Eq/L) before treatment and 0.58 ± 0.03 (mmolTrolox Eq/L) after treatment. The TOS value was 5.30 ± 0.74 (µmol H₂O₂ Eq/L) before treatment and 2.13 ± 0.2 (µmol H₂O₂ Eq/L) after treatment in the study by Erkılıç et al., 2016. In this study, TAS value of 1,116 ± 0,59 (mmol Trolox Eq/L) before treatment increased to 1,588 ± 0,71 (mmol Trolox Eq/L) (P <0.05) after treatment in the group 2 while these values did not markedly differ in group did not receive probiotics. When both groups were compared, a marked increase (P <0.05) was observed in group 2 after treatment. This significant increase in TAS value might indicate that probiotics play a role in the rapid response in antioxidant capacity. Reactive Oxygen Species are 30% higher in calves on the first day of birth compared to their mothers and this is thought to cause increased oxidative stress, which is inevitably present in newborns after the first inhalation of atmospheric oxygen (Gaál et al., 2006). In our study, the value of TOS in Group 1 before treatment was $4,480 \pm 1,77$ (μ mol H₂O₂ Eq / L) and 7,784 ± 1,42 (μ mol H₂O₂ Eq /L) after treatment. This finding might suggest that probiotics were more effective along with antibiotic use when compared to the antibiotic use only.

In the group 2, the TOS value before the treatment $5,492 \pm 1,68 \ (\mu mol H_2O_2 Eq / L)$ decreased to $4,178 \pm 1,89 \ (\mu mol H_2O_2 Eq / L)$ after treatment. Although not statistically significant, it was concluded that this reduction may be related to increased capacity of antioxidant due to probiotic use. In many studies, including this study, differences in TAS and TOS values have been observed and this is thought to be due to measurement methods, kit differences or sensitivity of tests, but studies have reported that the TOS value increases and the TAS value decreases in disease condition (K121] et al., 2007; Topraktaş, 2019; Kabu et al., 2015; Erkılıç et al., 2016).

CONCLUSION

As a result, the use of additional probiotics in the treatment helps to reduce oxidative stress, if additional probiotics are not used an increase in oxidant capacity could be noted or might not cause a rapid decrease. Besides the local effects of probiotics on microbiota, they play a role in the antioxidant-oxidant mechanism systemically with symbiotic effects. It has been determined that adding the probiotics to neonatal calf diarrhea treatments protocol might cause an increase in antioxidant capacity. But there is a need for further investigation on the role probiotics in mechanism of antioxidant process.

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Author's Contributions: NY, EÇ and MY designed the study. EÇ and MY collected the materials, NY, EÇ, MY and YB performed the analysis. YB evaluated statistically. NY, EÇ, MY and YB wrote the study

REFERENCES

- Ahire JJ, Mokashe, NU, Patil HJ, Chaudhari BL. Antioxidative potential of folate producing probiotic *Lactobacillus helveticus* CD6. J Food Sci Technol. 2013; 50(1):26-34.
- Alıç Ural D, Toplu S. Sığırlarda süt verimini arttırmada probiyotiklerin kullanımı. MAE Vet Fak Derg. 2017; 2(2):153-162.
- Alkan F. Buzağı ishallerinde rotavirus ve corona virusların rolü. Ankara Üniv Vet Fak Derg. 1998; 45:29-37.

- Asemi Z, Zare Z, Shakeri H, Sabihi SS, Esmaillzadeh A. Effect of multispecies probiotic supplements on metabolic profiles, hs-CRP, and oxidative stress in patients with type 2 diabetes. Ann Nutr Metab. 2013; 63(1-2):1-9.
- Aslan V. Buzağı ishalleri ve tedavileri, Neonatal Buzağı Kayıpları Sempozyumu. SÜ Veteriner Fakültesi. Konya, 1986; 59-69.
- Burçak E, Yalçın S. Buzağı beslemede probiyotiklerin kullanımı. Lalahan Hay Araşt Enst Derg. 2013, 53:101-114.
- **Burgu İ, Öztürk F.** Neonatal dönemdeki buzağıların viral hastalıkları, Neonatal Buzağı Kayıpları Sempozyumu, S.Ü. Veteriner Fakültesi. Konya, 1986; 50-59.
- Çadırcı E, Süleyman H, Aksoy H, et al. Effects of Onosma Armeniacum root extract on ethanol-induced oxidative stress in stomach tissue of rats. Chemico Biol Interact. 2007; 170(1):40-48.
- **Çatalkaya E.** Probiotics and oxidant-antioxidant relationship. Journal of Istanbul Veterinary Sciences. 2020; 31:101.
- **Doron S, Gorbach SL**. Probiotics: Their role in the treatment and prevention of disease. Expert Rev Anti-Infect Ther. 2006; 4(2):261-275.
- Endo H, Niioka M, Kobayashi N, Tanaka M, Watanabe T. Butyrate-producing probiotics reduce nonalcoholic fatty liver disease progression in rats: New insight into the probiotics for the gut-liver axis. PLoS ONE. 2013; 8(5):1-19.
- **Erel O.** A new automated colorimetric method for measuring total oxidant status. Clin Biochem. 2005; 38:1103-111.
- **Erel O.** A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. Clin Biochem. 2004; 37:277-85.
- Erkılıç EE, Erdoğan HM, Olgun M, et al. Relationsheep between hepcidin and oxidant/antioxidant status in calves with suspected neonatal septicemia. Vet World. 2016; 9:1238-1241.
- Gaál T, Ribiczeyné-Szabó P, Stadler K, et al. Free radicals, lipid peroxidation and the antioxidant system in the blood of cows and newborn calves around calving. Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 2006; 143(4):391-396.
- García-Lafuente A, Antolín M, Guarner F, Crespo E, Malagelada JR. Modulation of colonic barrier function by the composition of the commensal flora in the rat. Gut. 2001; 48(4):503-507.
- He ZX, Ferlisi B, Eckert E, Brown HE, Aguilar A, Steele MA. Supplementing a yeast probiotic to pre-weaning Holstein calves: Feed intake, growth and fecal biomarkers of gut health. Anim Feed Sci and Technol. 2017; 226:81-87.
- Kabu M, Cigerci IH, Uyarlar C, Celik HA. Determination of pre and post treatment oxidative status and oxidative DNA damage in diarrheic calves. Indian J Anim Res. 2015; 49(6):830-833.
- Kızıl Ö, Özdemir H, Karahan M, Kızıl M. Oxidative stres and alterations of antioxidant status in goats naturally infected with *Mycoplasma agalactia*. Rev Med Vet. 2007; 158(6):326-330.
- Kullisaar T, Zilmer M, Mikelsaar M, *et al.* Two antioxidative lactobacilli strains as promising probiotics. Int J Food Microbial. 2002; 72(3):215-224.
- Le OT, Dart PJ, Harper K, *et al.* Effect of probiotic Bacillus amyloliquefaciens strain H57 on productivity and the incidence of diarrhoea in dairy calves. Animal Production Science. 2017; 57(5):912-919.

- **LeBlanc AM, LeBlanc JG, Perdigon G**, *et al.* Oral administration of a catalase-producing *Lactococcus lactis* can prevent a chemically induced colon cancer in mice. J Med Microbiol. 2008; 57(1):100-105.
- Lewis K, Caldwell J, Phan V, *et al.* Decreased epithelial barrier function evoked by exposure to metabolic stress and nonpathogenic E. coli is enhanced by TNF-alpha. Am J Physiol Gastrointest Liver Physiol. 2008; 294(3):669-678.
- Madsen K, Cornish A, Soper P, *et al.* Probiotic bacteria enhance murine and human intestinal epithelial barrier function. Gastroenterology. 2001; 121(3):580-591.
- Minssen T, Nordberg A. Rules and Tools in the Battle against Superbugs-A call for integrated strategies and enhanced international collaboration to promote antimicrobial drug development. In: Eccleston-Turner M, Brassington I, eds. Infectious Diseases in the New Millennium: Legal and Ethical Challenges. Denmark: Springer; 2020. p.111-136.
- Moura FA, Queiroz de Andrade K, Dos Santos JCF, Araújo ORP, Goulart MOF. Antioxidant therapy for treatment of inflammatory bowel disease: Does it work? Redox Biol. 2015; 6:617-639.
- Nageshwar A, Raval A P, Bhagwat SR, Rajgor BB. Studies on growth, nutrient utilization, immune modulation, and economic return at different levels of probiotic feed supplementation in kankrej female calves. Animal Science. 2016; 10(2):55-62.
- Oelschlaeger TA. Definition of "good bacteria" origin, effects and effective components. Online:http://www.dgmim.de/fileadmin/CONTENT/OElsc hlaeger_good_bacteria_vortrag.pdf 2011, Erişim Tarihi: 04.03.2012
- **Ok M, Güler L, Turgut K**, *et al*. The studies on the aetiology of diarrhoea in neonatal calves and determination of virulence gene markers of *Escherichia coli* strains by multiplex PCR. Zoonoses Public Health. 2009; 56(2):94-101.
- Oldham KM, Wise SR, Chen L, Stacewicz-Sapuntzakis M, Burns J, Bowen PE. A longitudinal evaluation of oxidative stress in trauma patients. J Parenter Enteral Nutr. 2002; 26(3):189-197.

- Shah NP. Functional foods from probiotics and prebiotics. Food Technology. 2001; 55(11):46-53.
- Shen Q, Shang N, Li P. In vitro and in vivo antioxidant activity of Bifidobacterium animalis 01 isolated from centenarians. Curr Microbiol. 2011: 62(4): 1097-1103.
- Solis-Pereira B, Lemonnier, D. Induction of human cytokines by bacteria used in dairy foods. Nutrition Research. 1993; 13:1127-1240.
- Şen İ, Güzelbekteş H, Yıldız R. Neonatal buzağı ishalleri: patofizyoloji, epidemiyoloji, klinik, tedavi ve koruma. Türkiye Klinikleri J Vet Sci. 2013; 4(1):71-78.
- **Topaktaş B.** Sağlıklı Buzağılarda Doğumu Takip Eden İlk Bir Ay İçerisinde Total Oksidan Seviye ve Total Antioksidan Kapasitedeki Değişimlerin Araştırılması [Yüksek Lisans Tezi]. Kayseri: Erciyes Üniversitesi; 2019.
- Valencia GL, Zapata-Ramirez O, Nunez-Gonzalez L, et al. Effective use of probiotic-glyconutrient combination as an adjuvant to antibiotic therapy for diarrhea in rearing dairy calves. Turkish J Vet Anim Sci. 2017; 41(4):578-581.
- Wallace RJ, Newbold CJ. Probiotics for ruminants. In: Fuller R, ed. Probiotics The Scientific Basis. Netherlands: Springer; 1992. p.317-353.
- Winterbourn CC, Buss IH, Chan TP, Plank LD, Clark MA, Windsor JA. Protein carbonyl measurements show evidence of early oxidative stress in critically ill patients. Crit. Care Med. 2000; 28(1):143-149.
- Yavuzarslan E. Değişen Miktarlarda Süte Katılan Prebiyotiklerin Süt Emen Simental Buzağılarda Büyüme Performansı ve Sağlığı Üzerine Etkileri [Yüksek Lisans Tezi]. Kırıkkale: Kırıkkale Üniversitesi; 2018.
- Yeoman CJ, Ishaq SI, Bichi E, Olivo SK, Lowe J, Aldridge BM. Biogeographical differences in the influence of maternal microbial sources on the early successional development of the bovine neonatal gastrointestinal tract. Sci Rep. 2018; 8:1-14.