



## Use of Dietary *Pelargonium sidoides* Extract to Improve Growth and Body Composition of Narrow-Clawed Crayfish *Astacus leptodactylus* Eschscholtz, 1823 Juveniles

Funda Turan<sup>1\*</sup>, Yavuz Mazlum<sup>1</sup>, Yasemin Bircan Yıldırım<sup>1</sup>, Armağan Gezer<sup>1</sup>

<sup>1</sup> Mustafa Kemal University, Faculty of Fisheries and Aquaculture 31200 Iskenderun, Hatay.

\* Corresponding Author: Tel.: +90.326 6141866; Fax: +90.326 6141866;  
E-mail: turanfunda@yahoo.com

Received 11 May 2011  
Accepted 31 January 2012

### Abstract

The effects of the *Pelargonium sidoides* extract on survival, growth, and body composition of juvenile freshwater crayfish (*Astacus leptodactylus*) were investigated. Newly hatched third instars of *A. leptodactylus* were fed with experimental diets prepared by using supplementation of *P. sidoides* extract (0, 0.5, 1 and 2 ml 100 g<sup>-1</sup>) for 105 days. Growth rate was increased significantly in crayfish fed with 2 ml *P. sidoides* extract 100 g<sup>-1</sup> diet in comparison to the control groups (P<0.05). Feed conversion ratio, protein efficiency ratio and apparent net protein utilization were also significantly improved in group fed diet with 2 ml 100 g<sup>-1</sup> *P. sidoides* extract than that with control and the other groups. After 105 days of rearing the survival rate was found highest in 2 ml *P. sidoides* extract 100 g<sup>-1</sup> diet supplemented group (80.0%). Protein content (16.5%) of the 2 ml 100 g<sup>-1</sup> *P. sidoides* extract-supplemented group was significantly higher than the control and other dietary groups (P<0.05). The lipid contents were decreased in *P. sidoides* extract-supplementation. These results indicated that the *P. sidoides* extract is useful to improve growth, feed utilization and survival rate in freshwater crayfish, juvenile diets.

**Keywords:** *Pelargonium sidoides*, crayfish, *Astacus leptodactylus*, growth performance.

### Yavru Kerevitlerin (*Astacus leptodactylus* Eschscholtz, 1823) Büyümesi ve Vücut Kompozisyonlarının Geliştirilmesi Üzerine *Pelargonium sidoides* Ekstraktını Kullanımı

#### Özet

Bu çalışmada, yavru kerevitlerin (*Astacus leptodactylus* Eschscholtz, 1823) büyüme, yaşama oranı ve vücut kompozisyonları üzerine *Pelargonium sidoides* ekstraktının etkileri araştırıldı. Yeni çıkmış üçüncü aşamadaki *A. leptodactylus* yavruları 105 gün süre ile *P. sidoides* ekstraktı (0, 0.5, 1 ve 2 ml 100 g<sup>-1</sup>) destekli deneme diyetleri ile beslendi. Deneme sonunda; 2 ml 100 g<sup>-1</sup> *P. sidoides* ekstraktı ile beslenen grupta kontrol grubuna göre büyüme oranı istatistiksel anlamda önemli derecede artış gösterdi. Benzer şekilde; Yem değerlendirme oranı, Protein etkinlik oranı ve Görünür net protein kullanım oranlarının diğer gruplara göre 2 ml 100 g<sup>-1</sup> *P. sidoides* ekstraktı ile beslenen grupta daha iyi olduğu tespit edildi. Deneme sonunda, en yüksek hayatta kalma oranı %80 ile 2 ml 100 g<sup>-1</sup> *P. sidoides* ekstraktı ile beslenen grupta bulundu. Protein içeriği yönünden en iyi grup, %16,5 ile 2 ml 100 g<sup>-1</sup> *P. sidoides* ekstraktı ile beslenen grup olduğu belirlendi. Yağ içeriği ise diyetlerdeki *P. sidoides* ekstraktının kullanımına bağlı olarak azalma gösterdi. Bu çalışma; *P. sidoides* ekstraktını kullanarak hazırlanan diyetlerin, yavru kerevitlerde büyüme, yaşama oranı ve vücut kompozisyonları üzerine olumlu yönde etkili olduğu sonucunu göstermiştir.

**Anahtar Kelimeler:** *Pelargonium sidoides* kerevit, *Astacus leptodactylus*, büyüme.

#### Introduction

The narrow-clawed crayfish, *Astacus leptodactylus* Eschscholtz, 1823, is one of the most important crayfish species in Europe due to its aquaculture potential and wide consumer demand. It has been widely introduced into many countries, e.g. Poland, Italy, Germany, England and France, where it escaped into the wild and established large

populations in a number of water bodies (Skurdal and Taugbol, 2002). The only native crayfish species of Turkey, *A. leptodactylus* has a widespread distribution in lakes and ponds throughout the country (Harlioglu and Holdich, 2001; Skurdal and Taugbol, 2001; Mazlum, 2007; Harlioglu, 2009). At present it can be found in 30 countries, many of which it has been translocated to for stocking purposes (Souty-Grosset *et al.*, 2006). Until 1984, freshwater crayfish had an

important role as an export product, but after 1986 crayfish production declined dramatically (from 5000 tones to 200 tones) in most Turkish lakes, because of the crayfish plague (*Aphanomyces astaci*) and through over fishing, pollution and agriculture irrigation (Bolat, 2001; Mazlum, 2007; Harlioglu, 2004). Since the market value of narrow-clawed crayfish in Europe is higher than the one of imported species, it is more profitable to develop the farming of the former (Holdich, 1993). Crayfish farming is also played a significant role in the active protection of native species for restocking programmes. (Carral *et al.*, 2003; Smietana *et al.*, 2004). Therefore, there is a need to demand for crayfish reproduction under controlled conditions.

Medicinal herbs are efficacious for growth, health management, and for the immune systems of land mammals and humans. They originate mainly from vegetables are popular medicines in eastern Asia and Africa. Such treatment creates synergistic effects on various biological functions and mechanisms (Wu *et al.*, 2001; Park *et al.*, 2001; Lee *et al.*, 2003). One of the most important medicine herbs used in folk medicine by the Southern African native population is *Pelargonium sidoides* extract. The root of Umcka (*Pelargonium sidoides*) is used in traditional medicine to cure infectious respiratory diseases including tuberculosis (Haidvogel *et al.*, 1996). *Pelargonium* containing phytopharmaceuticals, elaborated from the traditional medicine, is nowadays successfully employed in modern phytotherapy in Europe (Haidvogel *et al.*, 1996). Polymeric polyphenols and coumarins have been identified as the principal ingredients (Koch and Biber, 2003). Most of the coumarins contain a methoxy function; functionality that is responsible for their antibacterial activity. Gallic acid and its methyl ester are present in large amounts. These were identified as the prominent immunomodulatory principle for this herbal medicine (Kayser and Kolodziej, 1998). In addition, *P. sidoides* is rich in phytochemical, vitamins, minerals and amino acids that enhance the body's functioning and protects it against diseases (Kolodziej *et al.*, 2003).

Some studies have been done in which herbs, as dietary additives, were fed to fish and shrimp. The focus of these studies includes their use as feeding attractants and their effects on growth, survival and immune system activity (Harada, 1991; Kwon *et al.*, 1999; Lee *et al.*, 2001; Jung *et al.*, 2002; Kim *et al.*, 2003; Immanuel *et al.*, 2004; Lee *et al.*, 2004; Ji *et al.*, 2007). Medicinal herbs promote lipid metabolism that catabolizes body fatty acids as a main energy expenditure, resulting in efficient protein accumulation and growth performance (Immanuel *et al.*, 2004). Also, Gezer (2009) has reported that *Pelargonium sidoides* extract -based diet improves growth and increase the level of protein in common carp. Although previous studies have mostly investigated on effects of herbal extracts on immunity and growth performance, there is a lack of data on the

use of *P. sidoides* extract in diets for crayfish growth.

In the present study we intended to ascertain whether *P. sidoides* extract included in the diet enhance the growth performance and survival rate of the narrow-clawed crayfish, juvenile stage under laboratory condition.

## Materials and Methods

Forty-two ovigerous female *Astacus leptodactylus* were collected from experimental ponds at Eğirdir Fisheries Research Institute located in Eğirdir, Isparta, Turkey, on three occasions, all on 1 May 2007. After each collection, the crayfish were immediately packed in insulated live-fish shipping boxes and shipped by bus to Iskenderun, Turkey, where they were transported by car to the Aquaculture Research Facilities at Mustafa Kemal University where spawning took place, survival at arrival was 95.2%.

Prior to experimental use, the crayfish were kept in circular fiberglass tanks provided with shelter (e.g., onion sacks and pieces of polyvinylchloride pipe), and well-aerated water was used in the flow-through system. The animals were fed commercial carp diet to develop and shed eggs. Hatched larvae were reared until the third-instar stage (young-of-the-year, YOY). Total length (TL) of subsamples of the *A. leptodactylus* third instars was measured to the nearest millimeter, to estimate the size of the starting YOY crayfish.

Newly hatched third instars of *A. leptodactylus* with mean body weight  $42.4 \pm 1$  mg were fed with a commercial carp diet, and stocked into 60 L aquaria at a density of 10 crayfish per aquarium ( $50 \text{ crayfish/m}^2$ ) (Mazlum, 2007). Experiment was conducted in aquaria ( $n=12$ ) with the dimensions of  $0.8 \times 0.4 \times 0.25$  m (length x width x height; area:  $0.20 \text{ m}^2$ ). Each aquarium was supplied with continuous aeration. A static system was used and 20% of the water in each aquarium was changed with 7 days interval. The bottom of each tank was provided with a PVC pipe and shelter.

Carp diets (Aquamak, Turkey: 28% protein, 12% lipid (on wet basis)) were used to prepare experimental diets. *Pelargonium sidoides* extract (UMCA®) was supplied by Dr. Willmar Schwabe GmbH and Co. (Ettlingen, Germany). (UMCA® contents; coumarins, gallic acid, polychemical ingredients). In the preparation of experimental diet, liquid *P. sidoides* extract were mixed with a solid carp diet in which water ( $450 \text{ mL kg}^{-1}$ ) were added and pelletized through a food grinder with a 2 –mm diameter die plate. These pelleted diets were dried in the open air for 24 hours. The dry pellets were placed in covered plastic containers and stored in a refrigerator at  $+5^\circ\text{C}$  (Lee *et al.*, 2004). Three different dosages of *P. sidoides* extract (0.5, 1, 2 ml extract  $100\text{g}^{-1}$  diet) were used in the experiment. The control diet was also mixed with 450 ml water. The

extrusions were broken into small pieces and stored in freezer until feeding. Each of the three experimental diets was randomly assigned to triplicate groups. Feed was provided twice a day, at a quantity of 5% of their body weight, and uneaten food was removed before next feeding for 105 days (Mazlum *et al.*, 2011). The amounts of feeds were adjusted according to the crayfish weight calculated for each sampling periods.

Water temperature and oxygen content were measured daily with a thermometer and a model 55 YSI oxygen meter (Yellow Springs Instruments Cy. Ohio), respectively. Water samples were collected for analysis per week: pH was determined with Accumet pH meter (Model 915, Fisher Scientific, Pennsylvania), ammonia nitrogen N-NH<sub>4</sub> (Nessler method) and nitrite nitrogen N-NO<sub>2</sub> (sulphanil method) were determined colorimetrically on spectrophotometer (UV-160 1 PC, Shimadzu visible). The average water temperature was 25±1.5 °C, and the oxygen content of the water was 8.5±0.5. At pH 7.8±0.9, the ammonia nitrogen content did not exceed 0.1 mg N-NH<sub>4</sub>/l, and nitrite nitrogen was no greater than 0.04 mg N-NO<sub>2</sub>/l. The photoperiod was maintained on a 12-h light: 12-h dark schedule.

After 45, 75 and 105 days all the crayfish were collected and counted. The whole stock was individually weighed to the nearest 0.1 g. weight gain (Watanabe *et al.*, 1990), feed conversion ratio (Steffens, 1989), specific growth rate (Clark *et al.*, 1990), protein efficiency ratio (Steffens, 1989), apparent net protein utilization (Bender and Miller, 1953) and survival rate (Watanabe *et al.* 1990) were calculated at the end of the experiment. Then, subsample of ten crayfish were pooled and stored at – 20 °C for proximate analysis. Standard methods (AOAC, 1990) as described for the experimental diets were used to determine the initial and final whole body proximate composition.

In the experiment, the variables were tested for normality and homogeneity of variances using the UNIVARIATE procedure. The data were homogenous and showed normal distribution, and all

data were subjected to a one-way analysis of variance (ANOVA) to determine if there is a difference in weight gain and body composition among treatments. Duncan test was used to compare the means of the treatments when differences occurred (Norris, 1993).

## Results and Discussion

The effects of different concentrations of dietary *P. sidoides* extract on growth and survival of juvenile crayfish (*Astacus leptodactylus*) for 105 days are shown in Table 1. After 105 days of rearing the survival rate was highest in 2 ml *P. sidoides* extract 100g<sup>-1</sup> diet supplemented group (80.0%) and was lowest in control group (53.3%). Moreover, there was no adverse influence of *Pelargonium sidoides* extract on the health status of the crayfish larvae in the present study. During the study, there were not abnormal observation on the behavior and general activities of treated cray fish.

Growth rate and Specific growth rate (SGR) were significantly increased in crayfish fed with *P. sidoides* extract-supplemented diets in comparison to the control groups (P<0.05). Among the *P. sidoides* extract-supplemented groups, the crayfish fed diet with 2 ml 100 g<sup>-1</sup> *P. sidoides* extract exhibited significantly higher than crayfish fed diets with 0.5 and 1 ml 100 g<sup>-1</sup> *P. sidoides* extract (Table 1). Feed conversion ratio (FCR), protein efficiency ratio (PER) and apparent net protein utilization (ANPU) were also significantly improved in group fed diet with 2 ml 100 g<sup>-1</sup> *P. sidoides* extract than that with control, 0.5 and 1 ml 100 g<sup>-1</sup> *P. sidoides* extract groups (P<0.05, Table 1).

The effects of different concentrations of dietary *P. sidoides* extract on the proximate composition of the whole-body juvenile freshwater crayfish (*Astacus leptodactylus*) for 105 days are shown in Table 2. Carcass moisture at the end the rearing trial did not change with dietary *P. sidoides* extract concentrations. Whole- body ash was significantly lower in 2 ml 100 g<sup>-1</sup> *P. sidoides* extract group than in

**Table 1.** The effects of different concentrations of dietary *P. sidoides* extract on growth and survival of juvenile freshwater crayfish (*Astacus leptodactylus*) for 105 days\*

Parameters	<i>P. sidoides</i> (ml 100g <sup>-1</sup> )			
	0	0.5	1	2
Weight gain (g)	1.27±0.08 <sup>a</sup>	1.41±0.04 <sup>a</sup>	1.42±0.05 <sup>a</sup>	1.88±0.11 <sup>b</sup>
SGR	3.23±0.10 <sup>a</sup>	3.37±0.05 <sup>a</sup>	3.42±0.06 <sup>ab</sup>	3.61±0.02 <sup>b</sup>
FCR	2.19±0.24 <sup>b</sup>	1.92±0.02 <sup>b</sup>	2.03±0.06 <sup>b</sup>	1.45±0.07 <sup>a</sup>
PER	1.33±0.14 <sup>a</sup>	1.49±0.01 <sup>a</sup>	1.40±0.04 <sup>a</sup>	1.97±0.10 <sup>b</sup>
ANPU (%)	19.15±1.24 <sup>a</sup>	20.43±0.48 <sup>a</sup>	22.11±1.30 <sup>a</sup>	32.26±2.51 <sup>b</sup>
Survival rate (%)	53.33±3.34 <sup>a</sup>	60.00±5.77 <sup>a</sup>	63.33±3.33 <sup>a</sup>	80.00±5.78 <sup>b</sup>

\*Values (mean ± S.E. of triplicate) with different superscripts in each line indicate significant differences (P<0.05).

WG (Weight Gain) (g) = Final weight-Initial weight.

SGR (Specific Growth Rate) (%) =  $[(\ln W^2 - \ln W^1) / (T^2 - T^1)] \times 100$ , where W<sup>1</sup>, and W<sup>2</sup> are mean body weight at times when the first and second samples were taken (T<sup>1</sup> and T<sup>2</sup>)

FCR (Food Conversion Ratio) = Feed consumption /weight gain

PER (Protein Efficiency Ratio) = Live body weight gained (g)/protein intake (g).

ANPU (Apparent Net Protein Utilization) (%) = [protein retained/unit of protein intake]x100.

**Table 2.** Proximate composition of the whole-body juvenile freshwater crayfish (*Astacus leptodactylus*) fed on diets containing different concentrations of dietary *P. sidoides* extract for 105 days\*

Parameters (%)	Initial	<i>P. sidoides</i> extract (ml 100g <sup>-1</sup> )			
		0	0.5	1	2
Moisture	76.60±0.57	76.68±0.38 <sup>a</sup>	77.43±0.46 <sup>a</sup>	77.01±0.41 <sup>a</sup>	76.80±0.52 <sup>a</sup>
Crude protein	14.66±0.84	14.30±0.64 <sup>a</sup>	14.93±0.27 <sup>a</sup>	15.78±0.71 <sup>ab</sup>	16.51±0.39 <sup>b</sup>
Crude lipid	1.76±0.03	1.71±0.04 <sup>d</sup>	1.51±0.02 <sup>c</sup>	1.33±0.03 <sup>b</sup>	1.13±0.03 <sup>a</sup>
Ash	6.87±0.05	6.78±0.13 <sup>b</sup>	6.15±0.16 <sup>ab</sup>	6.57±0.31 <sup>b</sup>	5.45±0.25 <sup>a</sup>

\*Values (mean ± S.E. of triplicate) with different superscripts in each line indicate significant differences (P<0.05).

Body composition data presented on a wet basis.

other dietary groups (Table 2). The protein contents of the whole body (16.51 %) of the 2 ml 100 g<sup>-1</sup> *P. sidoides* extract-supplemented group was significantly higher than protein contents of the control and other dietary groups (P<0.05). The lipid contents in the crayfish were dramatically decreased with increasing rate of *P. sidoides* extract-supplementation, and the control group had content (1.71%) (Table 2).

This was the first attempt to investigate potential of *Pelargonium sidoides* extract as a feed additive in crayfish culture. *Pelargonium sidoides* extract have been reported to increase antimicrobial activity and immunomodulatory effect in some terrestrial vertebrate animals (Koch and Biber, 2007; Bademkiran et al., 2009). However, no detailed information was found on the effects of *Pelargonium sidoides* extract on crayfish survival and growth. Only a study (Gezer, 2009) has reported that *Pelargonium sidoides* extract-based diet improves growth and survival in common carp. Therefore, the therapeutic use of *P. sidoides* extract prompted our studies on the usage of *P. sidoides* extract as dietary additives in crayfish culture. The present investigation revealed that *Pelargonium sidoides* extract-based diet improve growth and increase the level of protein in freshwater crayfish.

Also, there was no adverse influence of diet with *P. sidoides* extract on survival, and feed intake of crayfish in the present study and, FCRs were significantly improved in experimental treatments. This finding indicated that the *P. sidoides* extract (2 ml 100 g<sup>-1</sup>) is a positive dietary additive to induce effective technical and economical propagations for cultured crayfish. Similarly, in abalone *Haliotis discus hannai* (Lee et al., 2001), olive flounder *Paralichthys olivaceus* (Jung et al., 2002), shrimp *Penaeus indicus* (Immanuel et al., 2004), African catfish *Clarias gariepinus* (Turan and Akyurt, 2005), tilapia *Oreochromis aureus* (Turan, 2006), and common carp *Cyprinus carpio* (Turan et al., 2007) herbs in diets promoted growth and feed efficiency. Kim et al. (1998), suggested that unknown factors in various medicinal herbs led to favorable results in fish and shrimp trials. Sivaram et al. (2004) have been reported that medicinal herbs promote lipid metabolism, protein accumulation and growth performance. *P. sidoides* is rich in photochemical, vitamins, minerals and amino acids that enhance the

body's functioning and protects it against diseases (Kolodziej et al., 2003), and the presence of these phytochemical in the *P. sidoides* extract may stimulate growth in crayfish. From a proximate composition point of view, *P. sidoides* increased the level of protein in crayfish. Interestingly, we found that the survival of crayfish can be improved by *P. sidoides* extract-supplementation. Therefore, we expect that this result will stimulate a series of studies on the utilization of *P. sidoides* in diets for crayfish. Also *Pelargonium sidoides* extract is sold in local pharmacies in Turkey for about 3 US \$ per bottle (50 ml) for human medication. If it is supplied in bulk, it is expected that it would cost less.

In conclusion, the present findings suggested that disease and stress resistance.

## References

- Association of Official Analytical Chemists. 1990. Official Methods of Analysis. 15<sup>th</sup> edn. AOAC, Arlington, Virginia, USA.
- Bademkiran, S., Kurt, D., Yokus, B. and Celik, R. 2009. Comparison of *Pelargonium sidoides*, Placebo and Antibiotic Treatment of Chronic Endometritis in Dairy Cows: A Field Trial. *Journal of Animal and Veterinary Advances*, 8 (6): 1242-1247.
- Bender, A.E. and Miller, D.S. 1953. A new brief method of estimating protein value. *Journal of Biochemistry*, 53: 7-9.
- Bolat, Y. 2001. An estimation in the population density of freshwater crayfish (*Astacus leptodactylus salinus* Nordman, 1842) living in Hoyran Area of Eğirdir Lake, Isparta, Turkey. PhD thesis, Isparta: University of Süleyman Demirel, Turkey, 116 pp.
- Carral, J.M., Sáez-Royuela, M., Celada, J.D., Pérez, J.R., Melendre, P.M. and Aguilera, A. 2003. Advantages of Artificial Reproduction Techniques For White-Clawed Crayfish (*Austropotamobius Pallipes Lereboullet*). *Bulletin Francais de Pêche et de Pisciculture*, 370-371: 181-184. doi:10.1051/kmae:2003013
- Clark, A.E., Watanabe, W.O., Olla, B.L. and Wicklund, R.I. 1990. Growth, feed conversion and protein utilization of Florida red tilapia feed isocaloric diets with different protein levels in seawater pools. *Aquaculture*, 88: 75-85. doi:10.1016/0044-8486(90)90321-D
- Gezer, A. 2009. The effects of dietary *Sardinia* root extract (*Pelargonium sidoides*) on growth, whole-body composition and blood characteristics of the common

- carp (*Cyprinus carpio* L. 1758). Master Thesis, Institute of Nature Sciences, Mustafa Kemal University, Hatay-Turkey.
- Haidvogel, M., Schuster, R. and Heger, M. 1996. Acute Bronchitis im Kindesalter-Multizenter-Studie zur Wirksamkeit und Vertraglichkeit des Phytotherapeutikums Umckaloabo Z. Phytotherapy, 17: 300-313.
- Harada, H. 1991. Attraction activities of herbal crude drugs for abalone, oriental weatherfish, and yellowtail. Nippon Suisan Gakkaishi, 57: 2083-2088. doi:10.2331/suisan.57.2083
- Harlioglu, M.M. and Holdich, D.M. 2001. Meat yields in the introduced crayfish, *Pacifastacus leniusculus* and *Astacus leptodactylus*, from British waters. Aquaculture Research, 32:411-417. doi:10.1046/j.1365-2109.2001.00577.x
- Harlioglu, M.M., 2004. The present situation of freshwater crayfish, *Astacus leptodactylus* (Eschscholtz, 1823) in Turkey. Aquaculture, 230: 181-187. doi:10.1016/S0044-8486(03)00429-0
- Harlioglu, M.M., 2009. A comparison of the growth and survival of two freshwater crayfish species, *Astacus leptodactylus* Eschscholtz and *Pacifastacus leniusculus* (Dana) under different temperature and density regimes. Aquaculture International, 17: 31-43. doi:10.1007/s10499-008-9177-7
- Holdich, D.M. 1993. A global review of astaciculture – freshwater crayfish farming. Aquatic Living Resources, 6: 307– 317. doi:10.1051/alr:1993032
- Immanuel, G, Vicnybai, V.C., Sivaram, V., Palavesam, A. and Marian, M.P. 2004. Effect of butanolic extracts from terrestrial herbs and seaweed on the survival, growth and pathogen (*Vibrio parahaemolyticus*) load on shrimp *Penaeus indicus* juveniles. Aquaculture, 236: 53–65. doi:10.1016/j.aquaculture.2003.11.033
- Ji, S.C., Takaoka, O., Jeong, G.S., Lee, S.W., Ishimaru, K., Seoka, M. and Takii, K. 2007. Dietary medicinal herbs improve growth and some non-specific immunity of red sea bream *Pagrus major*. Fisheries Science, 73: 63–69. doi:10.1111/j.1444-2906.2007.01302.x
- Jung, S.H., Lee, J.S., Han, H.K., Jun, C.Y. and Lee, H.Y. 2002. Effects of medicinal herbs extract on non-specific immune responses, hematology and disease resistance on olive flounder, *Paralichthys olivaceus* by oral administration. Journal of Fish Pathology, 15: 25–35.
- Kayser, O. and Kolodziej, H. 1998. Antibacterial activity of extracts and constituents of *Pelargonium sidoides* and *Pelargonium reniforme*. Planta Medica 63,508-510. doi:10.1055/s-2006-957752
- Kim, D.S., Noh, C.H., Jung, S.W. and Jo, J.Y. 1998. Effect of Obosan supplemented diet on growth, feed conversion ratio and body composition of Nile tilapia, *Oreochromis niloticus*. Aquaculture, 11:83-90.
- Kim, J.H., Lee, S.M., Baek, J.M., Cho, J.K., and Kim, D.S. 2003. Effect of dietary lipid level and herb mixture on growth of parrot fish, *Oplegnathus fasciatus*. Journal of Korean Fisheries Society, 36:113–119. doi:10.5657/kfas.2003.36.2.113
- Koch, E. and Biber, A. 2007. Treatment of rats with the *Pelargonium sidoides* extract EPs® 7630 has no effect on blood coagulation parameters or on the pharmacokinetics of warfarin. Phytomedicine, 14: 40-45. doi:10.1016/j.phymed.2006.11.026
- Kolodziej, H., Kayser, O., Radtke, O., Kiderlen, A. and Koch, E. 2003. Pharmacological profile of extracts of *Pelargonium sidoides* and their constituents. Phytomedicine, 10(4): 18-24. doi:10.1078/1433-187X-00307
- Kwon, M.G., Kim, Y.C., Shon, Y.C. and Park, S.I. 1999. The dietary supplementing effects of kugija, *Lycium chinese*, on immune responses of Nile tilapia, *Oreochromis niloticus* to *Edwardsiella tarda*. Journal of Fish Pathology, 1: 73–81.
- Lee, S.M., Park, C.S. and Kim, D.S. 2001. Effects of dietary herbs on growth and body composition of juvenile abalone, *Haliotis discus hannai*. Journal of Korean Fisheries Society, 34: 570–575.
- Lee, W.Y., Lee, S.D., Son, S.I., Chang, H.S., Kim, Y.H., Oh, T.H., Eom, K.D., Chang, K.H., Park, S.C., Yamamoto, O., Maeda, Y. and Lee, K.W. 2003. The effects of *Artemia capillaries* crude juice extract on CCl4 induced liver damage in dogs. Journal of Veterinary Clinics, 20: 389–395.
- Lee, K.J., Dabrowski, K., Rinchar, J., Gomez, C., Leszek, G. and Vilchez, C. 2004. Supplementation of Maca (*Lepidium meyenii*) Tuber Meal in Diets Improves Growth Rate and Survival of Rainbow Trout *Oncorhynchus mykiss* (Walbaum) Alevins and Juveniles. Aquaculture Research, 35: 215-223. doi:10.1111/j.1365-2109.2004.01022.x
- Mazlum, Y. 2007. Stocking density affects the growth, survival, and cheliped injuries of third instars of narrow-clawed crayfish, *Astacus leptodactylus* Eschscholtz, 1823 juveniles. Crustaceana, 80(7): 803-815. doi:10.1163/156854007781363114
- Mazlum, Y., Güner, Ö. and Sirin S. 2011. Effects of Feeding Interval on Growth, Survival and Body Composition of Narrow-Clawed Crayfish, *Astacus leptodactylus* Eschscholtz, 1823 Juveniles. Turkish Journal of Fisheries and Aquatic Sciences, 11: 283-289. doi:10.4194/trjfas.2011.0213
- Norusis, M.J. 1993. SPSS for Windows Advanced statistics release 6.0. SPSS Inc., USA, 578 pp.
- Park, R.J., Kim, N.J., Lee, K.T. and Seo, S.H. 2001. Comparative studies on concentration of decursinol in plasma after oral administration of *Angelicae gigantis radix* extract and combined use of decursin and *Cnidii rhizome* extract or *Bupleuri radix* extract in rats. Korean Journal of Pharmacology, 32: 72–78.
- Sivaram, V., Babua, M.M., Immanuela, G., Murugadassa, S., Citarasub, T. and Mariana, M.P. 2004. Growth and immune response of juvenile greasy groupers (*Epinephelus tauvina*) fed with herbal antibacterial active principle supplemented diets against *Vibrio harveyi* infections. Aquaculture, 237: 9-20.
- Skurdal, J. and Taugbol, T. 2002. *Astacus*. In: Holdich DM (Ed.), Biology of freshwater crayfish. Blackwell Science, Oxford: 467–510.
- Skurdal, J. and Taugbol, T. 2001. Crayfish of commercial importance-*Astacus*. In: D.M. Holdich (Ed.), Biology of Freshwater Crayfish. Blackwell Science, Oxford: 467–510.
- Smietana, P., Krzywosz, T. and Struzynski, W. 2004. Review of the National Restocking Program "Active Protection of Native Crayfish in Poland" 1999-2001. Francais de Pêche et de Pisciculture, 372-373(1-2): 289-299.
- Souty-Grosset, C., Holdich, D.M., Noel P.Y., Reynolds, J.D. and Haffner, P. 2006. Atlas of Crayfish in Europe. Museum national d'Histoire naturelle, Paris (Patrimoines naturels, 64), 187 pp.

- Steffens, W. 1989. Principles of fish nutrition. Ellis Horwood Limited, Chichester, West Sussex, U.K.
- Turan, F. and Akyurt, I. 2005. Effects of Red Clover Extract on Growth Performance and Body Composition of African Catfish *Clarias gariepinus* (Burchell, 1822). Fisheries Science, 71: 618-620.  
doi:10.1111/j.1444-2906.2005.01006.x
- Turan, F., Gürlek, M. and Yağlıoğlu, D. 2007. Dietary Red clover (*Trifolium pratense*) on Growth Performance of Common Carp (*Cyprinus carpio*). Journal of Animal and Veterinary Advances, 6(12): 1429-1433.
- Turan, F. 2006. Improvement of Growth Performance in Tilapia (*Oreochromis aureus* Linnaeus) by Supplementation or Red Clover (*Trifolium pratense*) in Diets. The Israeli Journal of Aquaculture-Bamidgeh, 58(1): 34-38.
- Watanabe, W.O., Clark, J.H., Dunham, J.B., Wicklund, R.I. and Olla, B.L. 1990. Culture of Florida Red Tilapia in Marine Cage. The Effect of Stocking Density and Dietary Protein on Growth. Aquaculture, 90: 123-124.  
doi:10.1016/0044-8486(90)90336-L
- Wu, T.S., Tsang, Z.J., Wu, P.L., Lin, F.W., Li, C.Y., Teng, C.M. and Lee, K.H. 2001. New constituents and antiplatelet aggregation and anti- HIV principles of *Artemisia capillaries*. Bioorganic and Medicinal Chemistry, 9:77-83.  
doi:10.1016/S0968-0896(00)00225-X