



## Relationship Between Spermatozoa Motility, Egg Size, Fecundity and Fertilization Success in *Salmo trutta abanticus*

Yusuf BOZKURT<sup>1</sup> Selçuk SEÇER<sup>2</sup> Süleyman BEKCAN<sup>2</sup>

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**Abstract:** Sperm and eggs were collected without anesthesia by abdominal massage from *Salmo trutta abanticus*. The relationship between spermatozoa motility, egg size, fecundity and fertilization rates were investigated. Fertilization rates ranged from 72% to 80% and correlated positively with spermatozoa motility ( $r=0.111$ ,  $p>0.01$ ) and egg size ( $r=0.459$ ,  $p>0.01$ ). On the other hand, negative correlation ( $r=-0.535$ ,  $p>0.01$ ) was evaluated between fertilization rates and fecundity. Consequently it was determined that increase in spermatozoa motility and egg sizes effect the fertilization success positively in *Salmo trutta abanticus*.

**Key Words:** *Salmo trutta abanticus*, fertilization, motility, egg size, fecundity

### *Salmo trutta abanticus*'larda Fertilizasyon Başarısı ile Spermatozoa Motilitesi, Yumurta Büyüklüğü ve Yumurta Verimi Arasındaki İlişki

**Öz:** Abant alabalıklarından (*Salmo trutta abanticus*) sperma ve yumurta abdominal masaj yoluyla alınmıştır. Fertilizasyon oranları ile spermatozoa motilitesi, yumurta büyüklüğü ve yumurta verimi arasındaki ilişki araştırıldı. Fertilizasyon oranı ile spermatozoa motilitesi ( $r=0.111$ ,  $p>0.01$ ) ve fertilizasyon oranı ile yumurta büyüklüğü ( $r=0.459$ ,  $p>0.01$ ) arasında pozitif yönde korelasyon belirlenirken fertilizasyon oranı ile yumurta verimi ( $r=-0.535$ ,  $p>0.01$ ) arasında ise negatif yönde bir korelasyon belirlenmiştir. Sonuç olarak, spermatozoa motilitesi ve yumurta büyüklüğündeki artışların Abant alabalık (*Salmo trutta abanticus*)'larında fertilizasyon başarısını olumlu yönde etkilediği belirlenmiştir.

**Anahtar Kelimeler:** *Salmo trutta abanticus*, fertilizasyon, motilite, yumurta büyüklüğü, yumurta verimi

### Introduction

*Salmo trutta abanticus* is native to lake Abant and the nearest rivers and creeks and is known to be endemic to Lake Abant, Turkey (Geldiay and Balik 1988) being reared in natural conditions as well. Moreover, *Salmo trutta abanticus* is important as a biological gene source for restocking for Turkey. Nowadays, *Salmo trutta abanticus* populations are badly affected by natural phenomena (e.g., droughts and climate change) or human activities (e.g., overfishing, tourism, pollution and industry).

The use of high quality gametes from captive fish broodstock is of great importance for ensuring the production of viable larvae (Kjorsvik et al. 1990). The fish farming industry has been more focused on the quality of eggs or larvae rather than that of sperm, even though the quality of both gametes may affect

fertilization success and larval survival (Bozkurt et al. 2006). It is well known that the size of eggs of fish shows considerable intra- and inter- specific variation. Even parental fish of the same strain, weight and length have eggs that in different size (Bagenal 1971). Egg size is primarily determined by the genotype of parents of fish and it is also known to be affected by other factors including age and size of the female parent. Gall (1974) have shown in studies of hatchery-reared trout that older and heavier females produce larger eggs than younger and smaller fish. The availability of food also affects egg size (Springate et al. 1985). Alterations in egg size also occur in batch-spawning fish as the season progresses (Bagenal 1971) and in synchronous spawners as a result of photoperiodic modifications of maturation (Bromage et al. 1984).

<sup>1</sup>Mustafa Kemal Univ. Fac. of Fisheries-Hatay

<sup>2</sup>Ankara Univ. Fac. of Agriculture Department of Fisheries and Aquaculture-Ankara

On the other hand, sperm quality data are required for successful fertilization. In addition, sperm quality is a measure of the ability of sperm to successfully fertilize an egg. Any quantifiable physical parameter that directly correlates with the fertilization capacity of sperm could be potentially used as a measure of sperm quality. Optimal sperm quality is important for effective broodstock management and should be a criterion in the selection of male broodstock (Bozkurt et al. 2006).

Nevertheless, in commercial hatcheries, sperm is often inadequate both in terms of quantity and quality and it does not always give successful fertilization in the artificial insemination procedures commonly used for aquaculture species. Spermatozoa motility is the most commonly used criterion to evaluate semen quality (Bozkurt, 2006). However, investigations of the relationship between spermatozoa motility and fertility have given conflicting results or were of little value in predicting fertilization potential (Graham et al. 1978).

From this point of view, it seems that fecundity, egg size and spermatozoa motility should be considered together to estimate fertilization success. Therefore the aim of this study was to determine the relationship between motility, egg size, fecundity and fertilization rates in *Salmo trutta abanticus*.

## Materials and Methods

### Broodstock care and collection of sperm:

Fifteen mature *Salmo trutta abanticus* males (total weight  $1.452 \pm 1.86$  kg, total length  $45.39 \pm 4.25$  cm) and females (total weight  $1.310 \pm 480.83$  kg, total length  $44.5 \pm 3.53$  cm) were used as sperm and egg donors respectively. The broodstock were held in raceways under a natural photoperiod regime and fed with a commercial trout diet at 2% of their body weight per day. Water temperature varied between 7-8 °C during spawning season. Sperm from each fish was collected into 25 ml glass beakers by abdominal massage.

**Evaluation of spermatozoa motility:** A 10 µl sample was taken from each sperm batch and placed on a microscope slide and 100 µl activation solution (0.3% NaCl) was added to determine spermatozoa motility. The percentage of motility was defined as the percentage of progressively motile spermatozoa within each activated sample. Progressively motile spermatozoa were defined as actively swimming in a forward motion. Sperm cells that vibrated in place were not considered to be motile and observations were made within thirty minutes of sperm collection.

**Fecundity and egg size:** Fecundity and egg size was evaluated from fifteen females. Fecundity was calculated by volumetric method and egg size was determined by using a sensitive micrometer (at 0.01 mm sensitivity).

**Fertilization:** The fertilization capacity of sperm from each males resulting 45 sperm samples were tested with the same egg pool. All fertilization trials were done as 3 replicates in dishes with 10 ml of eggs ( $90 \pm 10$  eggs). The dry fertilization technique was used and the insemination dosage was  $2 \times 10^5$  spz/egg for each fertilization experiment. Sperm obtained from each fish was poured onto the eggs and gently mixed about 20 s respectively and one minute later 20 ml fertilization solution (3 g urea, 4 g NaCl and 1 l distilled water) was added. About 30 minutes later following the fertilization, the eggs were rinsed in hatchery water and incubated in a egg incubator. Fertilization rate was determined as the percentage of eyed eggs about forty days later following the fertilization.

**Statistical Analysis:** Results are expressed as mean  $\pm$  standart deviation. The arcsine transformation was used to the fertilization percentages before statistical analysis. Significant means were subjected to a multiple comparison test (Duncan) for post-hoc comparisons at  $\alpha=0.01$  level. All statistical analysis were carried out using SPSS 10 for Windows software package.

## Results and Discussion

Fertilization ratio ranged between 72% and 80% and was found as mean  $76.0 \pm 4.0\%$ . Statistical analysis shows positive allometry for the relationships between fertilization rates and spermatozoa motility and negative allometry for the relationship between fertilization rates and fecundity. Values related to fecundity, egg size, spermatozoa motility and fertilization rates were shown in Table 1. Correlations and relationships between spermatozoa motility, egg size, fecundity and fertilization rates are shown in Table 2 and Figure 1 (a-c).

Table 1. Fecundity, egg size, motility and fertilization rates in *Salmo trutta abanticus* (n=15).

	Total Fecundity (eggs/fish)	Egg size (mm)	Motility (%)	Fertilization rates (%)
Mean value ( $\pm$ sd)	1468 $\pm$ 180.21	5.42 $\pm$ 0.25	75.2 $\pm$ 3.24	76 $\pm$ 2.58
Range	1125-1800	5.0-5.8	60-85	72-80

Table 2. Correlations between spermatozoa motility, egg size, fecundity and fertilization rates in *Salmo trutta abanticus*

	Spermatozoa motility	Egg size	Fecundity
Egg size	-0.100		
Total Fecundity	0.055	-0.156	
Fertilization rates	0.111	0.459	-0.535

Correlation is not significant at the 0.01 level.

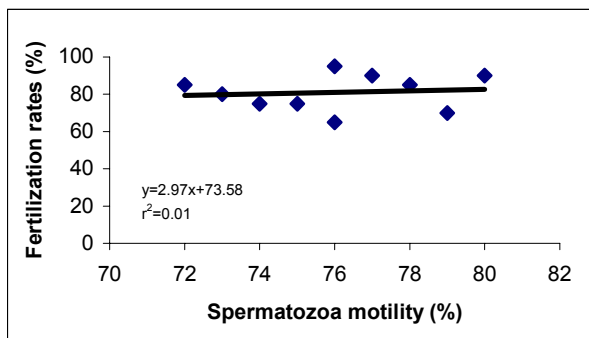


Figure 1a. Relationship between fertilization rates and spermatozoa motility.

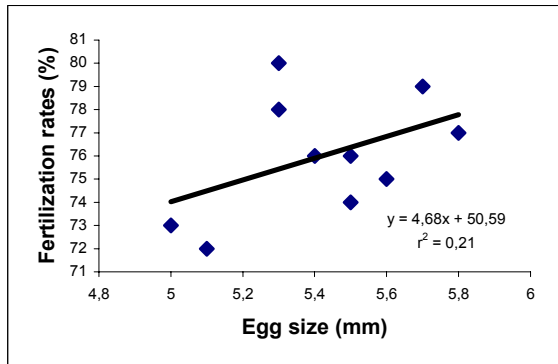


Figure 1b. Relationship between fertilization rates and egg size.

The global growth of intensive aquaculture has increased the need for efficient and effective means of conserving fish gametes. Viable sperm is an essential component in any successful animal production operation and the success of reproductive process is dependent on a supply of high quality gametes (Cruz-Casallas et al., 2005).

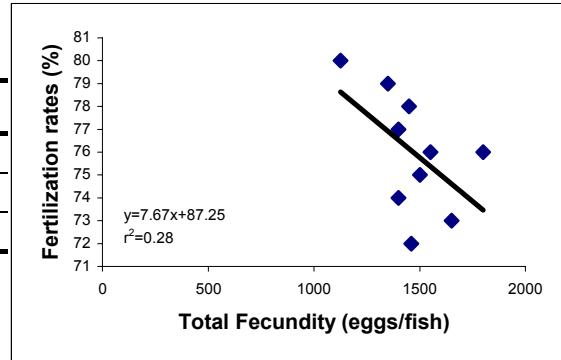


Figure 1c. Relationship between fertilization rates and fecundity.

The most reliable indicator of the sperm quality is spermatozoa motility. Subjective estimation of motility requires considerable experience and this indicator is being used in the selection of sperm for insemination and preservation. However, spermatozoa motility varies in vigor and duration not only among males but also within an individual male depending on its ripeness. The highest motility of the spermatozoa is observed at the peak of the breeding season (Terner 1986). Studies on most fish species recorded that motility of spermatozoa may show seasonal variation (Benau and Terner 1980).

Our results confirmed the data of Munkittrick and Moccia (1987) and of Ciereszko and Dabrowski (1994) who found a correlation between motility rate and fertilization capacity for the rainbow trout sperm using subjective estimation methods for motility determination. Magyary et al. (1996) concluded a strong, positive correlation ( $r=0.85$ ) of frozen/thawed carp spermatozoa motility and fertility at  $1 \times 10^5$ - $1.5 \times 10^5$ :1 spermatozoa/egg ratio. In addition, Linhart et al. (2000) also observed good correlation ( $r=0.53$ ) with fresh sperm between carp spermatozoa motility and fertilization at spermatozoa/egg ratios of around  $2 \times 10^5$ :1. The recommended sperm to egg ratio for commercial trout culture is based on the use of stripped milt (Scott and Baynes 1980). In this study, a positive relationship was determined between motility and fertilization rate ( $p>0.01$ ) at 200.000 spz:egg ratio.

In the case of fecundity, this parameter was found lower than values reported for rainbow trout (Bromage and Cumarantunge 1988, Estay et al. 1994). However values related to the egg size were found higher than that of rainbow trout (Springate et al. 1984, Estay et al. 1994). Most studies of reproduction tend to consider fecundity and egg size as separate indicators of reproductive performance. It is generally accepted

that there is an inverse relationship between fecundity and egg size in which fish produce either more eggs of a smaller size or fewer eggs of a larger size (Springate et al. 1985, Bromage et al. 1992). Similarly, a negative allometry was obtained between fecundity and egg size in this study. Also a negative relationship was determined between fecundity and fertilization rates. For this reason, it can be concluded that egg size is more important than fecundity for the fertilization success.

In conclusion, the information obtained from the present study can lead to more efficient gamete management and increased fry yields. On the other hand, further studies are needed to increase the viability, survival and development of larvae used in farming conditions.

#### References

- Bagenal, T.B. 1971. The interpretation of the size of fish eggs, the date of spawning and the production cycle. *J. Fish Biol.* 3:207-219.
- Benau D. and C. Ternier. 1980. Initiation, prolongation and reactivation of the motility of salmonid spermatozoa. *Gamete Res.* 3: 247-257.
- Bozkurt, Y. 2006. The relationship between body condition, sperm quality parameters and fertilization success in rainbow trout (*Oncorhynchus mykiss*). *Journal of Animal and Veterinary Advances* 5 (4): 284-288.
- Bozkurt, Y., S. Secer, N. Bukan, E. Akcay and N. Tekin. 2006. Relationship between body condition, physiological and biochemical parameters in brown trout (*Salmo trutta fario*) sperm. *Pakistan Journal of Biological Sciences.* 9 (5): 940-944.
- Bromage, N.R., J.A. Elliot, J.R.C. Springate. and C. Whitehead. 1984. The effects of constant photoperiods on the timing of spawning in the rainbow trout. *Aquaculture* 43:213-223.
- Bromage, N. and P.R.C. Cumarantunga. 1988. Egg Production in the Rainbow Trout, In: R.J. Roberts and J.F. Muir (Editors), *Recent Advances in Aquaculture* Vol. 3, London, Croom Helm, 63-138.
- Bromage, N.R., J. Jones, C. Randall, M. Thrush, B. Davies, J. Springate, J. Duston and G. Barker. 1992. Broodstock management, fecundity, egg quality and the timing of egg production in the rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 100: 141-166.
- Ciereszko, A. and K. Dabrowski. 1994. Relationship between biochemical constituents of fish semen and fertility. The effect of short term storage. *Fish Physiol. Biochem.* 12: 357-367.
- Cruz-Casallas P.E., D.A. Lombo-Rodriguez and Y.M. Velasco-Santamaria. 2005. Milt quality and spermatozoa morphology of captive *Brycon siebenthalae* (Eigenmann) broodstock. *Aquaculture Research.* 36: 682-686.
- Estay, F., N.F. Diaz, R. Neira and X. Fernandez. 1994. Analysis of Reproductive Performance of Rainbow Trout in a Hatchery in Chile. *The Progressive Fish Culturist* 56: 244-249.
- Gall, G.A.E. 1974. Influence of size of eggs and age of female on hatchability and growth of rainbow trout. *Calif. Fish Game* 60:26-35.
- Geldiay, R. ve S. Balık. 1988. Türkiye Tatlısu Balıkları. (Freshwater fishes of Turkey). Ege Üniversitesi Fen Fak. Kitaplar Serisi. 97, 519 s.
- Graham, E.F., M.K.L. Schmehl, B.K. Evenson and D.J. Nelson. 1978. Viability assays for frozen semen. *Cryobiology* 15: 242-244.
- Kjorsvik, E., A. Mangor-Jensen and I. Holmefjord. 1990. Egg quality in fishes. In: Blaxter, J.H.S., Southward, A.J. (Eds.), *Adv. Mar. Biol.*, 26: 71-113.
- Linhart, O., M. Rodina and J. Cosson. 2000. Cryopreservation of sperm in common carp *Cyprinus carpio*: sperm motility and hatching success of embryos. *Cryobiology* 41: 241-250.
- Magyary, I., B. Urbanyi and L. Horvath. 1996. Cryopreservation of common carp (*Cyprinus carpio* L.) sperm: II. Optimal conditions for fertilization. *Journal of Applied Ichthyology* 12: 117-119.
- Munkittrick, K.R., and R.D. Moccia. 1987. Seasonal changes in the quality of Rainbow trout semen: effect of a delay in stripping on spermatozoa, volume and seminal plasma constituents. *Aquaculture* 64: 147-156.
- Scott A.P. and S.M. Baynes. 1980. A review of the biology, handling and storage of salmonid spermatozoa. *Journal of Fish Biology* 17: 707-739.
- Springate, J., N. Bromage J.A.K. Elliot and D.L. Hudson. 1984. The Timing of Ovulation and Stripping and the Effect on the Rates of Fertilization and Survival to Eying, Hatch and Swim-up in the Rainbow Trout (*Salmo gairdneri*). *Aquaculture* 43: 313-322.
- Springate, J.R.C., Bromage, N.R. and P.R.T. Cumarantunga. 1985. The effects of different ration on fecundity and egg quality in the rainbow trout (*Salmo gairdneri*) pp. 371-391. Ed: C.B. Cowey, A.M. Mackie, J.G. Bell. *Nutrition and Feeding in Fish*. Academic Press, London, UK.
- Terner, C. 1986. Evaluation of salmonid sperm motility for cryopreservation. *The Progressive Fish Culturist* 48: 230-232.

#### Correspondence address:

Süleyman BEKCAN  
Ankara University, Faculty of Agriculture,  
Department of Fisheries and Aquaculture  
E-mail: sbekcan@agri.ankara.edu.tr