

GENETIC CONTROL OF PURPLE PLANT COLOR IN SESAME

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ABSTRACT

The exploitation of exotic genetic resources in plant breeding is necessary to enhance diversity of cultivars. Purple color is an exceptional character in sesame and monitored rarely in stems, capsules and leaves. The purple sesame is suitable for commercial production with high antioxidant capacity. For understanding of its genetic behaviour, inheritance study was carried out for four years (2008-2011) by crossing Munganli-57 (♀) and ACS 70 (♂). Munganli-57 parent had green color in canopy while ACS 70 was purple. All the plants in the F₁ generation were purple colored. In F₂ population, 3:1 segregation ratio showed that purple color character was controlled by a single dominant gene. F₂ progenies were sown to single rows separately, either purple or green plants. The green plants in F₂ were also green colored in F₃ with no segregation while the purple plants obtained from F₂ indicated purple and green colors in F₃ with a segregation ratio of 3:1. These segregations demonstrated that purple color in sesame was under the control of a single gene and this unique color was dominant over traditional green color. This accession (ACS 70) is a valuable genetic resource by providing a unique color character in sesame. The result presenting in this study would be of importance for further improvement of high antioxidant capacity in sesame.

Keywords: Antioxidants, genetic control, purple capsule, *Sesamum indicum* L.

INTRODUCTION

Sesame (*Sesamum indicum* L.) is one of the world's important oilseed crops and has long been used as ingredient in human foods and animal feeding in the form of seed, seed oil and meal (Hahm et al., 2009). Sesame has also medicinal and pharmaceutical value owing to high quality oil (Uzun et al., 2008) and antioxidant lignans such as sesamin and sesamol (Peschel et al., 2007; Erbas et al., 2009). Recently, its antioxidants have been used exclusively for anti-aging processes.

Sesame is a plant breeder's dream because of its great variability (Langham and Wiemers, 2002). Hundreds of characters have been studied with respect to range of variability and many of them were classified by several researchers (Bisht et al., 1998; Furat and Uzun, 2010; Yol et al., 2010; Yol and Uzun, 2012). Color establishment in sesame also contributes to this variability with different cotyledon, petiole, corolla, capsule, leaf, stem, and seed coat color. Seed coat color has been studied intensively as it was related with amino acid composition (Lee et al., 1990), oil content (Namiki, 1995; Were et al., 2001; Zhang et al., 2004) and antioxidant capacity (Lee et al., 1999; Lee et al., 2002; Moazzami and Kamal-Eldin, 2006; Shahidi et al., 2006) whilst there has been limited investigation about plant color because of low variability.

In sesame, during the vegetative and reproductive phases the color is usually a shade of green, and then as the plants mature and begin to drop their leaves, the color will turn to many shades of yellow/green, and rarely purple (Langham, 2007). When purple lines reach late bloom stage until the drying phase, plants indicated purple color especially in stems and capsules on the contrary of usual sesame plants. Previously, purple plants were observed in Indian genotypes by Bedigian et al. (1986) who stated that purple form had dominancy however inheritance mechanism was not exhibited. Erbas et al. (2009) studied chemical composition of purple plant in sesame and they observed high antioxidant capacity in purple sesame compared with normal sesame plants. Purple forms should be assessed as commercial types due to their being a source of high antioxidant capacity. The aim of the present study is to determine genetic behaviour of purple sesame accession for understanding the gene system operating in the expression of purple color in sesame.

MATERIALS AND METHODS

In 2008 growing season, the accession with purple color, ACS 70 (♂) was crossed with Munganli-57 (♀) which is registered cultivar (green), at the West Mediterranean Agricultural Research Institute's fields of

Antalya (36°52'N, 30°50'E., 15 m elevation). F₁s were seld in the growing season of 2009 and F₂ populations were developed in 2010. All the plants in F₂ were scored either purple or green colored. F₂ progenies showing both colors were sown in single rows separately in 2011. Purple and green plants were also scored either purple or green in F₃ generation.

The crosses between Muganlı-57 (green color) and ACS 70 (purple color) were made using flower buds emasculated just before anthesis and pollinated the second day with pollen grains from freshly dehisced anthers of the male parents (Falusi and Salako, 2003). Parents, F₁s, F₂s and F₃s were grown in 70 cm row and 10 cm plant spacing in all the experiments. Standard cultural practices like regular weeding, irrigation and fertilization were followed. Fertilizer was applied at rates of 60 kg N, 60 kg P₂O₅ and 60 kg K₂O per hectare just prior to sowing. F₁, F₂ and F₃ generations were grown at the same location.

A chi-square (χ^2) goodness of fit test was performed on the segregating populations against a possible theoretical segregation ratio using the formula:

$\chi^2 = \sum (O - E)^2 / E$, where O and E are the observed and expected values, respectively (Steel and Torrie, 1980).

RESULTS

Following to Muganlı-57 (♀) x ACS 70 (♂) cross, all the F₁ filials had purple color on their stems, capsules and leaves. In F₂ progeny, the ratio of purple color to green color was 65 to 22 (Table 1). Phenotypically, 65 plants indicated purple form (Figure 1a) and 22 plants indicated green form (Figure 1b). Chi-square values obtained from this cross showed a good fit for a monogenic inheritance with the F₂ phenotypic ratio of 3:1 (Table 1).

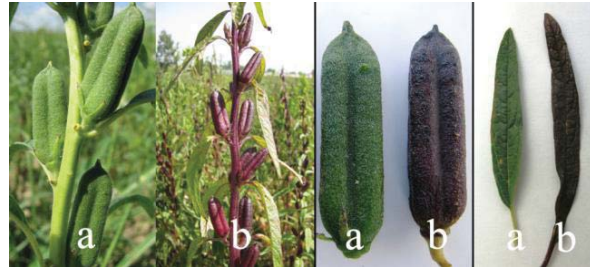


Figure 1. Green and purple sesame plants a - green stem, capsule and leaf, respectively. b - purple stem, capsule and leaf, respectively.

Table 1. Chi-square values of the cross between Muganlı-57 and ACS 70

Cross	Experimental		Theoretical		χ^2	p	Ratio
	Purple Plants	Normal Plants	Purple Plants	Normal Plants			
Muganlı-57 x ACS 70	65	22	65.25	21.75	0.0038	0.95	3:1

The plants with purple and green color in F₂ were separately advanced to F₃ generation in single rows. Totally, 106 offsprings in F₃ were obtained from four F₂

plants with green color. All these offsprings had green color and showed no segregation (Table 2).

Table 2. Segregation ratios of purple and green colored sesame in F₃

Cross	Number of plants		Ratio	Symbol
	Purple plants	Normal (green) plants		
Muganlı-57 x ACS 70				
F ₂ progeny with purple color				
Offspring 1	43	0	1:0	P/P
Offspring 2	37	0	1:0	P/P
Offspring 3	39	0	1:0	P/P
Offspring 4	32	0	1:0	P/P
Offspring 5	45	0	1:0	P/P
Offspring 6	41	0	1:0	P/P
Offspring 7	30	8	3:1 (P=0.57)	P/p
Offspring 8	51	0	1:0	P/P
Offspring 9	12	5	3:1 (P=0.67)	P/p
F ₂ progeny with normal (green) color				
Offspring 1	0	45	0:1	p/p
Offspring 2	0	12	0:1	p/p
Offspring 3	0	30	0:1	p/p
Offspring 4	0	19	0:1	p/p

Totally, 343 offsprings in F₃ were obtained from nine F₂ plants with purple color. Those of 288 offsprings coming from seven F₂ plants had purple color with no

segregation. The other two F₂ progeny (offspring 7 and 9) consisted of 42 purple and 13 green color plants. This result supported that purple and green color plants fitted

the expected 3:1 ratio (Table 2). These segregations proved that purple color in sesame was under the control of a single gene and it was dominant over traditional green color.

DISCUSSION

Monogenic inheritance for different colors in sesame was identified for petiole, nectar (Van Rheenen, 1970) and flower (Khidir, 1973) however there was no previous information about the inheritance of stem, leaf and capsule color. Same color characteristics were investigated for different crop plants. Red/purple color had dominancy over green stem color in *Ricinus communis* L. (Lavanya and Gopinath, 2008), in groundnut (Jadhav and Shinde, 1979), in pigeonpea (Narkhada et al., 1980) and purple plant color had dominancy over green plant color in *Vigna radiata* (L.) Wilczek (Khattak et al., 2000).

More knowledge on the inheritance mechanisms of desirable traits will indeed contribute much to varietal improvement in sesame (Ashri, 2007). Recently, antioxidant capacity has been taken into consideration as desirable character in sesame. Researchers use plant genetic resources to produce new crop varieties with specific characteristics like disease resistance, drought tolerance, or color; develop pharmaceutical or medical products; and determine the origins of a particular species (Delheimer, 2013). In particular, an adequate knowledge of genetic structure of plant genetic resources provides basic science and applied aspects like the efficient management of crop genetic diversity (Mondini et al., 2009). From this point of view, identifying inheritance mechanism of new traits would be beneficial for trait utilization. Purple sesame as a valuable genetic resource offers exploiting antioxidant capacity. Genetic behaviour of this unique character was explored and monogenic inheritance was established in this investigation.

Breeding objectives in sesame have focused many characters such as seed size, shape, flavor, coat color and oil content. Especially for physiological needs, color is an important argument because color production is an integral part of the development of various plant parts; type of color may adapt the plant part for a specific function (Padi, 2003). Therefore, color is one of the most important attributes of foods, being considered as a quality indicator and determining frequently their acceptance (Azerado, 2009). Particularly purple colored plants hold many healthy and agricultural advantages. Purple color can be due to anthocyanins which are phenolic compounds with high antioxidant activity that give many fruits and vegetables red or purple color (Wolfe and Liu, 2002). Purple sweet potato (*Ipomoea batatas*) has been concerned as a rich source of stable anthocyanins (Wu et al., 2008). Purple tomatillo (*Physalis ixocarpa*) genotypes appear to be good and safe source of antioxidants (Mendoza et al., 2010). Purple-pigmented rice bran includes high amounts of hydrophilic phenolic compounds and lipophilic antioxidants (Hu et al., 2003). Purple corn kernels are affluent in anthocyanins with well-established antioxidants (Adom and Liu, 2002). With this respect, the

introduction of purple color to sesame plants will satisfy new demands and markets as in other cultivated plants with purple color. The results of the study showed that purple color character is dominant over green color and information about inheritance of purple color character would be useful for the genetic improvement of sesame and improve new varieties with higher antioxidant capacity.

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LITERATURE CITED

- Adom, K.K. and R.H. Liu. 2002. Antioxidant activity of grains. *J. Agr. Food Chem.* 50:6182-6187.
- Ashri, A. 2007. Sesame (*Sesamum indicum* L.). In: Genetics Resources, Chromosome Engineering, and Crop improvement, Oilseed Crops, ed. Singh, R.J., 231-289, CRC Press, Florida.
- Azeredo, H.M.C. 2009. Betalains: properties, sources, applications, and stability – a review. *Int. J. Food Sci. Tech.* 44:2365-2376.
- Bedigian, D., C.A. Smyth. and J.R. Harlan. 1986. Patterns of morphological variation in sesame. *Econ. Bot.* 40:353-365.
- Bisht, I.S., R.K., Mahajan, T.R. Loknothan and R.C. Agrawal. 1998. Diversity in Indian sesame collection and stratification of germplasm accessions in different diversity groups. *Genet. Resour. Crop Evol.* 45:325-335.
- Delheimer, S. 2013. Conserving plant genetic resources. Fact sheet of the plant genetic resources conservation unit.
- Erbas, M., H. Sekerci, S. Gül, S. Furat, E. Yol, B. Uzun. 2009. Changes in total antioxidant capacity of sesame (*Sesamum* sp.) by variety. *Asian J. Chem.* 21:5549-5555.
- Falusi, O.A. and E.A. Salako. 2003. Inheritance studies in wild and cultivated 'Sesamum' L. species in Nigeria. *J. Sustain. Agr.* 22:75-80.
- Furat, S. and B. Uzun. 2010. The use of agro-morphological characters for the assessment of genetic diversity in sesame (*Sesamum indicum* L.). *Plant Omics J.* 3:85-91.
- Hahm, T.S., T.S.J. Park and Y.M. Lo. 2009. Effects of germination on chemical composition and functional properties of sesame (*Sesamum indicum* L.) seeds. *Bioresource Technol.* 100:1643-1647.
- Hu, C., J. Zawistowski, W. Ling and D.D. Kitts. 2003. Black rice (*Oryza sativa* L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems. *J. Agric. Food Chem.* 51:5271-5277.
- Jadhav, G.D. and N.N. Shinde. 1979. Genetic studies in groundnut (*Arachis hypogaea*). *Indian J. Agric. Res.* 13:93-96.
- Khattak, G.S.S., M.A. Haq, M. Ashraf and M.A. Awan. 2000. Inheritance of plant and flower bud color in mungbean (*Vigna radiata* (L.) Wilczek). *Songklanakarin J. Sci. Technol.* 22:249-251.
- Khidir, M.O. 1973. Genetic studies in sesame. II. Inheritance of flower color and number of locules per pod. *Exp. Agr.* 9:361-364.
- Langham, D.R. and T. Wiemers. 2002. Progress in mechanizing sesame in the US through breeding. In: Trends in New Crops

- and New Uses, ed. Janickand, J. and Whipkey, A., 157-173, ASHS Press, Virginia.
- Langham, D.R. 2007. Phenology of sesame. In:), Issues in New Crops and New Uses, ed. Janickand, J. and Whipkey, A., 144-182, ASHS Press, Alexandria, Virginia.
- Lavanya, C. and V. Gopinath. 2008. Inheritance studies for morphological characters and sex expression in pistillate lines of castor (*Ricinus communis* L.). Indian J. Genet. 68:275-282.
- Lee, J.I., B.H. Lee and J.K. Bang. 1990. Breeding of sesame (*Sesamum indicum* L.) for oil quality improvement. V. Varietal differences of amino-acid composition in protein of sesame seed. Kor. J. Breed. 21:300-307.
- Lee, S.W., C.W. Kang, D.H. Kim, Y. Satoko and K. Masumi. 1999. Varietal variation of sesamin, sesamol and oil contents according to seed coat colors in sesame. Kor. J. Breed. 31:286-292.
- Lee, W.C., J.Y. Wen, C.H. Shiow and D.D. Pin. 2002. Antioxidant activity of sesame coat. Food Chem. 78:347-354.
- Mendoza, D.G., O.G. Juárez, R.S. Ortiz, F.E. Garcia and J.F.S. Hernández. 2010. Evaluation of total phenolics, anthocyanins and antioxidant capacity in purple tomatillo (*Physalis ixocarpa*) genotypes. Afr. J. Biotechnol. 9:5173-5176.
- Moazzami, A.A. and A. Kamal-Eldin. 2006. Sesame seed is a rich source of dietary lignans. J. Am. Oil Chem. Soc. 83:719-723.
- Mondini, L., A. Noorani and M.A. Pagnotta. 2009. Assessing plant genetic diversity by molecular tools. Diversity 1:19-35.
- Namiki, M. 1995. The chemistry and physiological functions of sesame. Food Rev. Int. 11:281-329.
- Narkhada, B.N., A.B. Daokar and R. Cruz. 1980. Inheritance of some characters in a cross of pigeonpea, *Cajanus cajan* (L.) Millsp. Journal of Maharashtra Agricultural Universities. 5:205-208.
- Padi, K.F. 2003. Genetic analyses of pigmentation in Cowpea. Pakistan J. Biol. Sci. 6:1655-1659.
- Peschel, W., W. Dieckmann, M. Sonnenschein and A. Plescher. 2007. High antioxidant potential of pressing residues from evening primrose in comparison to other oilseed cakes and plant antioxidants. Ind. Crop. Prod. 25:44-54.
- Shahidi, F., C.M.L. Pathirana and D.S. Wall. 2006. Antioxidant activity of white and black sesame seeds and their hull fractions. Food Chem. 99:478-483.
- Steel, R.G.D. and J.H. Torrie. 1980. Principles and Procedures of Statistics: A Biometrical Approach. 2. ed. New York: McGraw-Hill, 631p.
- Uzun, B., C. Arslan and S. Furat. 2008. Variation in fatty acid compositions, oil content and oil yield in a germplasm collection of sesame (*Sesamum indicum* L.). J. Am. Oil Chem. Soc. 85:1135-1142.
- Van Rheenen, H.A. 1970. Intergeneric hybridization between *Ceratotheca sesamoides*, Endl. and *Sesamum indicum* L. Nigerian J. Sci. 4:251-254.
- Were, B.A., M. Lee, and S. Stymne. 2001. Variation in seed oil content and fatty acid composition of *Sesamum indicum* L., and its wild relatives in Kenya. Sveriges Utsadesforenings Tidskrift. 111:178-193.
- Wolfe, K. and R.H. Liu. 2002. Apple peels are rich in phytochemicals and have high antioxidant activity. New York Fruit Quarterly. 10:9-11.
- Wu, D.M., J. Lu, Y.L. Zheng, Z. Zhou, Q. Shan and D.F. Ma. 2008. Purple sweet potato color repairs D-galactose-induced spatial learning and memory impairment by regulating the expression of synaptic proteins. Neurobiol. Learn. Mem. 90:19-27.
- Yol, E., E. Karaman, S. Furat and B. Uzun. 2010. Assessment of selection criteria in sesame by using correlation coefficients, path and factor analyses. Aust. J. Crop Sci. 4:598-602.
- Yol, E. and B. Uzun. 2012. Geographical patterns of sesame (*Sesamum indicum* L.) accessions grown under Mediterranean environmental conditions, and establishment of a core collection. Crop Sci. 52:2206-2214.
- Zhang, J.H., H. Guo, Y. Wang and D.C. Chen. 2004. Study on method of determination of the content of sesame oil by second derivative ultraviolet spectrophotometry. J. Hunan. Agric. Univ. 30:367-370.