



Synthesis and Comparative Antibacterial Studies of some Benzylidene Monosaccharide Benzoates

Mohammed M. Matin^{1*}, M.M.H. Bhuiyan¹, Md. Manir Hossain¹, and Mohammad Harun Or Roshid²

¹ Organic Research Laboratory, Department of Chemistry, University of Chittagong, Chittagong-4331, Bangladesh.

² Department of Anesthesia and Intensive Care, Chittagong Medical College, Chittagong-4203, Bangladesh.

E-mail: mahbubchem@cu.ac.bd, Tel. no.: 8801716839689

Abstract: Methyl a-D-glucopyranoside on reaction with benzaldehyde followed by direct dimolar benzoylation afforded 4,6-O-benzylidene-protected 2,3-di-O-benzoate **3** in good yield. Regioselective monobenzoylation of methyl a-D-mannopyranoside employing dibutyltin oxide method furnished 3-O-benzoate **5**. Compound **5**, on treatment with excess benzaldehyde followed by direct benzoylation, provided 2,3-di-O-benzoate **7** in high yield. *In vitro* antibacterial activity studies of the synthesized compounds along with the precursor materials (**1-7**) were evaluated against ten bacterial pathogens. The structure activity relationship study revealed that the benzoyl mannopyranosides (**5-7**) exhibited more antibacterial inhibitory property as compared to that of glucopyranosides (**2-3**).

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Introduction

Monosaccharides are wide spread in nature, being a component of some plant glycosides and bacterial polysaccharides of immunological importance [1-2]. Monosaccharide derivatives, especially acylated monosaccharides (esters), synthetic utility have as versatile intermediates in the syntheses of many natural products and their analogues which have great medicinal importance [3-4]. Various methods for acylation monosaccharides have so far been developed and employed successfully such as direct protection-deprotection [5-6], technique [7], organotin (bistributyltin oxide or dibutyltin oxide) mediated regioselective [8-10] *etc*. Yet, regioselective acylation (esterification) is a prominent challenge as monosaccharides contain several hydroxyl groups of similar reactivity. In this context, enzyme catalyzed acylation [11] and microwave assisted acylation [12] were also investigated in the past decade. In the present synthetic strategy, we employed acylation technique for both methyl a-Dglucopyranoside $(\mathbf{1})$ and methyl a-Dmannopyranoside (4). We have also used dibutyltin oxide method for the regioselective 3-O-benzoylation of mannopyranoside 4.

The emergence of multiple antibiotic resistant pathogenic bacteria represents a growing threat to human health worldwide. Thus, search for new antibacterial agents with novel mode of action represents a major target in chemotherapy [13]. Acylated sugars (esters) have been widely used as cosmetic and pharmaceutical industries for many years because they are considered biocompatible, biodegradable, and nontoxic [14-15].

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In ¹H NMR analysis of **1** in DMSO-d₆, aromatic protons appear at 8.58, 8.28, 8.02, 7.59, 7.11, 7.05 ppm as a doublet, singlet, multiple, triplet, doublet and doublet, respectively. The OH proton appear at 10.28 ppm as singlet. The ¹H NMR spectra of the compounds 2 and 3 are somewhat broader than corresponding signals the compound 1 due to aggregation the phthalocyanine isomers which is frequently encountered at the concentrations used for NMR spectroscopy. The inner NH protons of 2 were also identified in the ¹H NMR spectra with a broad chemical shift at -4.03 ppm.

In ¹³C NMR analysis of **1** in DMSO-d₆, aromatic

carbons appear at 158.53, 146.53, 135.11, 129.01, 130.37, 129.48, 128.38, 128.33, 127.87, 126.51, 126.11, 123.50, 123.16, 120.22, and 116.35 ppm, C-OH group appeared at 153.49 ppm, nitrile carbons appeared at 115.72 and 114.87 ppm, respectively.

In the EI⁺ mass spectrum of **1**, the presence of the characteristic molecular ion peak at m/z 298.0 [M]⁺ confirmed the proposed structure. In the case of **2**, the molecular ion peak was found at m/z 1233.5 [M+K]⁺ according to MALDI-TOF spectrum (Fig. 1). Also, the molecular ion peak for compound **3** found at m/z = 1260.7 [M+2]⁺. The elemental analyses were satisfactory.

Materials and Methods

Chemicals and Apparatus

FT-IR spectra were recorded on an FT-IR spectrometer (Shimadzu, IR Prestige-21) using CHCl $_3$ mulls. Thin layer chromatography (TLC) was performed on Kieselgel GF $_{254}$ and visualization was accomplished by spraying the plates with 1% $\rm H_2SO_4$ followed by heating the plates at 150-200 $^{\rm 0}C$ until coloration took place. $^{\rm 1}H$ (400 MHz) and $^{\rm 13}C$ (100 MHz) NMR spectra were recorded using CDCl $_3$ as a solvent. Chemical shifts were reported in δ

unit (ppm) with reference to TMS as an internal standard and J values are given in Hz. Melting points (mp) were determined on an Electrothermal melting point apparatus and are uncorrected. Evaporations were performed under diminished pressure on a Büchi rotary evaporator. All reagents used were commercially available (Aldrich) and were used as received unless otherwise specified.

Synthesis

4,6-O-benzylidene-a-D-Methyl glucopyranoside (2): Benzaldehyde (3.0 g, 28.27 mmol) was added to methyl a-Dglucopyranoside (1) (0.4 g, 2.06 mmol) followed by addition of anhydrous zinc chloride (1.0 g, 7.337 mmol) at room temperature. The reaction mixture was stirred for 12 h and filtered through Celite® 545. Ice was added to the filtrate with constant shaking and the mixture was extracted several times with nhexane to remove unreacted benzaldehyde. The aqueous layer was then extracted with ethyl acetate (3×5 mL) with occasional The organic layer was warming. (MgSO₄), and concentrated under reduced pressure to yield a residue which, chromatographic purification with nhexane/ethyl acetate (1/2, v:v), yielded the

title compound $\mathbf{2}$ (0.401 g, 69%) as a white crystalline solid, mp 162-163 °C (lit. [22], mp 163-164 °C).

 $R_f = 0.48 \; (n\text{-hexane/ethyl acetate} = 1/2, \, \text{v/v}). \; \text{FT-IR} \; (\text{CHCl}_3, \, \text{v}, \, \text{cm}^{-1}): \; 3310\text{-}3400 \; (\text{OH}). \, \text{IH} \; \text{NMR} \; (400 \; \text{MHz}, \, \text{CDCl}_3, \, \delta, \, \text{ppm}): \; 7.53\text{-}7.47 \; (2\text{H}, \, \text{m}, \, \text{Ar-H}), \; 7.42\text{-}7.35 \; (3\text{H}, \, \text{m}, \, \text{Ar-H}), \; 5.54 \; (1\text{H}, \, \text{s}, \, \text{PhCH-}), \; 4.80 \; (1\text{H}, \, \text{d}, \, \textit{J}= 4.0 \; \text{Hz}, \; \text{H-1}), \; 4.30 \; (1\text{H}, \, \text{dd}, \, \textit{J}= 9.7 \; \text{and} \; 4.4 \; \text{Hz}, \; \text{H-6a}), \; 3.94 \; (1\text{H}, \, \text{dd}, \, \textit{J}= 9.3 \; \text{and} \; 9.2 \; \text{Hz}, \; \text{H-3}), \; 3.82 \; (1\text{H}, \, \text{ddd}, \, \textit{J}= 10.3, \; 9.2 \; \text{and} \; 4.4 \; \text{Hz}, \; \text{H-5}), \; 3.75 \; (1\text{H}, \, \text{ddd}, \, \textit{J}= 10.3 \; \text{and} \; 9.7 \; \text{Hz}, \; \text{H-6b}), \; 3.64 \; (1\text{H}, \, \text{dd}, \, \textit{J}= 9.2 \; \text{and} \; 4.0 \; \text{Hz}, \; \text{H-2}), \; 3.50 \; (1\text{H}, \; \text{dd}, \, \textit{J}= 9.3 \; \text{and} \; 9.2 \; \text{Hz}, \; \text{H-4}), \; 3.47 \; (3\text{H}, \, \text{s}, \; \text{OCH}_3). \; ^{13}\text{C} \; \text{NMR} \; (100 \; \text{MHz}, \; \text{CDCl}_3): \; \delta \; 137.1 \; \; (\text{Ar-C}), \; 129.3, \; 128.3, \; 126.3 \; (\text{Ar-CH}), \; 102.0 \; (\text{PhCH-}), \; 99.7 \; (\text{C-1}), \; 80.8 \; (\text{C-4}), \; 72.9 \; (\text{C-2}), \; 71.9 \; (\text{C-3}), \; 68.9 \; (\text{C-6}), \; 62.4 \; (\text{C-5}), \; 55.6 \; (\text{OCH}_3). \; \end{cases}$

Methyl 4,6-O-benzylidene-2,3-di-O-benzoyl-a-D-glucopyranoside(3): A solution of compound **2** (0.3 g, 1.281 mmol) in dry pyridine (1 mL) was cooled to 0 °C whereupon benzoyl chloride (0.396 g, 2.817 mmol) and DMAP (cat.) were added to it. The mixture was stirred overnight at room temperature. Usual work-up and silica gel column chromatography furnished the di-O-benzoyl derivative, 3 (0.521 g, 92%) as a crystalline solid. Recrystallization from ethyl acetate-*n*-hexane gave the analytically pure

Methyl 3-O-benzoyl-a-Dmannopyranoside (5): To a solution of methyl a-D-mannopyranoside (4) (0.3 g, 1.545 mmol) in dry methanol (10 mL) was added dibutyltin oxide (Bu₂SnO) (0.423 g, 1.699 mmol) and the mixture was heated under reflux. After 2 h the reaction mixture became homogeneous and clear. The mixture was then refluxed for an additional hour and the solvent was evaporated off in vacuo to leave a white solid. The solid tin complex was suspended in dry 1,4-dioxane (3 mL) and benzoyl chloride (0.239 g, 1.70 mmol) was slowly added to this solution, with stirring, at room temperature. The solution became clear upon addition of benzoyl chloride and stirring

Methyl 3-O-benzoyl-4,6-O-benzylidene-a-**D-mannopyranoside (6):** A solution of the 2,4,6-triol **5** (0.4 g, 1.341 mmol) benzaldehyde (3.0 g, 28.27 mmol) was treated with anhydrous zinc chloride (1.0 g, 7.337 mmol) at room temperature. The reaction mixture stirred was this temperature overnight and filtered through celite. Usual work-up as described for compound 2 and chromatographic purification (n-hexane/ethyl acetate = 1/2, v/v) yielded the title compound 6 (0.389 g, 75%) as white needles, mp 135-136 °C.

Methyl 4,6-O-benzylidene-2,3-di-O-benzoyl-α-D-mannopyranoside (7): The reaction of compound 6 (0.3 g, 0.776 mmol) and benzoyl chloride (0.396 g, 2.817 mmol) in anhydrous pyridine (1 mL) at 0 °C to room temperature for overnight followed by

sample as colorless needles, mp 146-148 °C. $R_f = 0.51$ (n-hexane/ethyl acetate = 4/1, v/v). FT-IR (CHCl₃, v, cm⁻¹): 1735, 1719 (CO). ¹H NMR (400 MHz, CDCl₃, δ , ppm): 7.81-8.05 (5H, m, Ar-H), 7.31-7.48 (6H, m, Ar-H), 7.21-7.30 (4H, m, Ar-H), 6.02 (1H, t, J= 9.8 Hz, H-3), 5.52 (1H, s, PhCH-), 5.21 (1H, dd, J= 9.8 and 3.6 Hz, H-2), 5.13 (1H, d, J= 3.6 Hz, H-1), 4.28-4.32 (1H, m, H-5), 4.03 (1H, t, J= 9.7 Hz, H-4), 3.78-3.87 (2H, m, H-6a and H-6b), 3.37 (3H, s, OCH₃).

was continued for 4 h. The solvent was evaporated off in vacuo to leave a syrupy mass, which was subjected to column chromatography (chloroform/methanol = 6/1, v/v) to give 3-O-benzoate **5** (0.313 g, 68%) as a thick syrup [11].

 $R_f = 0.48$ (chloroform/methanol = 4/1, v/v). FT-IR (CHCl₃, v, cm⁻¹): 3260-3350 (br OH), 1746 (CO). ¹H NMR (400 MHz, CDCl₃, δ , ppm): 7.55-7.73 (5H, m, Ar-H), 6.07 (1H, dd, J= 10.0 and 3.1 Hz, H-3), 4.62 (1H, d, J= 1.5 Hz, H-1), 4.22 (1H, dd, J= 3.1 and 1.5 Hz, H-2), 4.13 (1H, dd, J= 12.1 and 5.2 Hz, H-6a), 3.91-3.97 (1H, m, H-5), 3.88 (1H, app t, J= 9.8 Hz, H-4), 3.75 (1H, dd, J= 12.1 and 2.0 Hz, H-6b), 3.16 (3H, s, OCH₃).

 $R_f=0.45~(n-hexane/ethyl acetate=3/1, v/v).$ FT-IR (CHCl $_3$, v, cm $^{-1}$): 3290-3500 (br OH), 1735 (CO). 1H NMR (400 MHz, CDCl $_3$, δ , ppm): 8.05 (2H, d, J=8.1 Hz, Ar-H), 8.05 (1H, t, J=7.6 Hz, Ar-H), 7.39-7.46 (5H, m, Ar-H), 7.27-7.35 (2H, m, Ar-H), 5.59 (1H, s, PhCH-), 5.54 (1H, dd, J=10.4 and 3.2 Hz, H-3), 4.78 (1H, d, J=1.2 Hz, H-1), 4.32 (1H, dd, J=3.2 and 1.2 Hz, H-2), 4.22-4.31 (2H, m, H-5 and H-6a), 3.95-4.03 (1H, m, H-6b), 3.92 (1H, t, J=10.1 Hz, H-4), 3.43 (3H, s, OCH $_3$).

chromatography (n-hexane/ethyl acetate =10/1, v/v) gave the di-O-benzoyl derivative, **7** (0.343 g, 90%) as a white crystalline solid. Recrystallization from ethyl acetate/n-hexane (1/1, v/v) gave the analytically pure sample as needles, mp 121-122 °C.

 $R_f = 0.51 \, (n\text{-hexane/ethyl acetate} = 4/1, v/v).$ FT-IR (CHCl₃, v, cm⁻¹): 1687, 1682 (CO). ¹H NMR (400 MHz, CDCl₃, δ , ppm): 8.12-8.19 (5H, m, Ar-H), 7.57-7.69 (4H, m, Ar-H), 7.41-7.54 (6H, m, Ar-H), 5.81 (1H, dd, J= 10.0

and 3.1 Hz, H-3), 5.79 (1H, dd, J= 3.1 and 1.2 Hz, H-2), 5.69 (1H, s, PhCH-), 4.93 (1H, d, J= 1.2 Hz, H-1), 4.32-4.41 (2H, m, H-5 and H-6a), 4.11-4.19 (1H, m, H-6b), 4.00 (1H, t, J= 10.0 Hz, H-4), 3.50 (3H, s, OCH₃).

Antibacterial screening tests

Four Gram-positive bacteria viz. Bacillus cereus BTCC 19, Bacillus megaterium BTCC 18, Bacillus subtilis BTCC 17 and Staphylococcus aureus ATCC 6538 and six Gram-negative bacteria viz. Escherichia coli ATCC 25922, INABAET (vibrio) AE 14748, Pseudomonas aeruginosa CRL (ICDDR,B), Salmonella paratyphi AE 14613, Salmonella typhi AE 14612, and Shigella dysenteriae AE 14369 were selected for antibacterial potentiality test. For the detection of antibacterial activities, the disc diffusion method described by Bauer et al.

[23] was followed. Mueller-Hinton (agar and broth) medium was used for the culture of bacteria. Dimethylformamide (DMF) was initially used as a solvent to prepare the desired solution (1%) of the compounds. The plates were incubated at 37 °C for 48 h. Proper control was maintained with DMF. Each experiment was carried out in triplicate. All the results were compared with the standard antibacterial antibiotic ampicillin [50 µg/disc, Beximco Pharmaceuticals Ltd., Bangladesh].

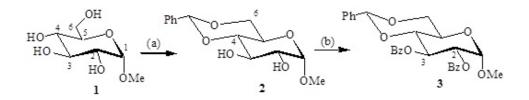
Results and Discussion

Our main aim was to synthesize 2,3-di-*O*-benzoyl derivatives (**3** and **7**) of methyl a-D-glucopyranoside (**1**) and methyl a-D-mannopyranoside (**4**), respectively, to study and compare their antibacterial properties.

Synthesis of methyl 4,6-O-benzylidene-2,3-di-O-benzoyl-a-D-glucopyranoside (3)

Initially we prepared methyl 4,6-*O*-benzylidene-a-D-glucopyranoside (**2**) from methyl a-D-glucopyranoside (**1**). The reaction of methyl a-D-glucopyranoside (**1**) with benzaldehyde in the presence of anhydrous zinc chloride for 12 h followed by work-up and chromatography gave a white crystalline solid,

mp 162-163 °C (Scheme 1). The FT-IR spectrum of this solid showed bands at 3310-3400 cm⁻¹ corresponding to hydroxyl stretching. In its ¹H NMR spectrum, a twoproton multiplet at δ 7.53-7.47, a three-proton multiplet at δ 7.42-7.35 and a one-proton singlet at δ 5.54 clearly indicated the formation of benzylidene acetal in the molecule. This was also confirmed by its 13C NMR spectrum where signals at δ 137.1 (Ar-C), 129.3, 128.3, 126.3 (Ar-CH) and 102.0 (PhCH-) were found for the benzylidene acetal. Complete analysis of its FT-IR, ¹H and ¹³C NMR spectra led us to assign the structure as methyl 4,6-O-benzylidene-a-Dglucopyranoside (2).



Scheme 1. Reagents and conditions: (a) PhCHO, dry $ZnCl_2$, 25 ^{0}C , 12 h, 69%. (b) BzCl, dry pyridine, 0 ^{0}C -RT, 12 h, 92%.

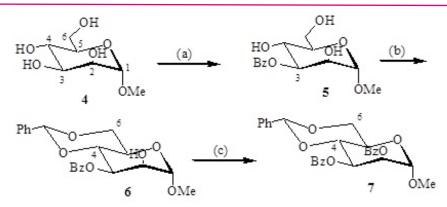
Having protected glucopyranoside **2** in hand, we carried out its dibenzoylation (Scheme 1). Thus, treatment of glucopyranoside **2** with dimolar benzoyl chloride (BzCl) in anhydrous pyridine for 12 h using a catalytic amount of DMAP gave a compound almost in quantitative yield (92%) as needles, mp 146-148 °C. In the FT-IR spectrum of this compound, bands at 1735 and 1719 cm⁻¹ were observed for carbonyl frequency and hence indicated the attachment of benzyloxy groups in the molecule. The FT-IR spectrum showed no band for hydroxyl stretching. In the ¹H NMR spectrum, the following peaks were observed

Synthesis of methyl 4,6-*O*-benzylidene-2,3-di-*O*-benzoyl-a-D-mannopyranoside,

Our next attempt was to synthesize methyl 4,6-*O*-benzylidene-2,3-di-*O*-benzoyl-a-D-mannopyranoside, (**7**). For this reason we prepared methyl 3-*O*-benzoyl-a-D-

in the aromatic region: δ 7.81-8.05 (5H, m), 7.31-7.48 (6H, m) and 7.21-7.30 (4H, m) and a one-proton singlet at δ 5.52 for PhCH-. The more ten aromatic protons in addition to benzylidene acetal protons indicated the attachment of two benzoyloxy groups in the molecule. Also, H-2 (at δ 5.21 as dd) and H-3 (at δ 6.02 as t) protons shifted down field as compared to its precursor compound **2**. This confirmed the attachment of benzoyloxy groups at position C-2 and C-3. So, the structure was established as methyl 4,6-*O*-benzylidene-2,3-di-*O*-benzoyl- α -D-glucopyranoside (**3**).

mannopyranoside (**5**) from methyl a-D-mannopyranoside (**4**) by dibutyltin oxide method using literature procedure [8, 24] (Scheme 2).



Scheme 2. Reagents and conditions: (a) Bu_2SnO , dry MeOH, reflux, 3h, BzCl, dioxane, RT, 4h, 68%. (b) PhCHO, dry $ZnCl_2$, 25 °C, 12 h, 65%. (c) BzCl, 0 °C-RT, 12 h, 90%.

In the dibutyltin oxide method initially a stannylene ring (intermediate tin complex) is formed between *cis*-vicinal glycol at C-2 and C-3 position, where the equatorial C-3 OH group is activated without exception (Figure

1). Hence the unimolecular benzoylation occurs at C-3 position only. Thus, the structure of this compound was established as methyl 3-*O*-benzoyl-a-D-mannopyranoside (**5**) by analyzing its FT-IR and ¹H NMR spectra.

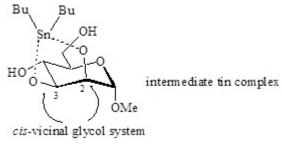


Figure 1. Formation of the intermediate stannylene ring.

The 3-O-benzoate 5 was then subjected for 4,6-*O*-benzylidene protection. Thus, the reaction of compound 5 with benzaldehyde in the presence of anhydrous zinc chloride for 12 h followed by work-up and chromatography gave as white needles, mp 135-136 °C. The FT-IR spectrum of this solid showed bands at 3290-3500 (br) and 1735 cm⁻¹ corresponding hydroxyl and carbonyl stretchings, respectively. In its ¹H NMR spectrum, a twoproton doublet at δ 8.05, a one-proton triplet at δ 8.05, a five-proton multiplet at δ 7.39-7.46, a two-proton multiplet at δ 7.27-7.35 and a one-proton singlet at δ 5.59 (PhCH-) clearly indicated the formation of benzylidene acetal and presence of a benzoyl group in the molecule. Complete analysis of its FT-IR and ¹H NMR spectra led us to assign the structure as 3-O-benzoyl-4,6-O-benzylidene-a-Dmethyl mannopyranoside (6).

Having protected mannopyranoside **6** in hand, we carried out its 2-*O*-benzoylation. Thus, unimolar benzoylation of **6** with benzoyl

Antibacterial activities of the synthesized compounds

The results of the *in vitro* inhibition zone against the selected Gram-positive bacteria due to the effect of the chemicals (**1-7**) are mentioned in Table 1. It was observed from Table 1 that the tested chemicals were less

chloride in pyridine and chromatography gave a crystalline solid. Recrystallization from ethyl acetate/*n*-hexane (1:1,v:v) gave analytically pure sample as needles, mp 121-122 °C (Scheme 2). The FT-IR spectrum showed no band for hydroxyl stretching. In the FT-IR spectrum of this compound, bands at 1687 and 1682 cm⁻¹ were observed for carbonyl frequency. In the ¹H NMR spectrum of this compound the following peaks were observed in the aromatic region: δ 8.12-8.19 (5H, m), 7.57-7.69 (4H, m), 7.41-7.54 (6H, m) and a one-proton singlet at δ 5.69 (PhCH-). The additional five aromatic protons than that of 3-O-benzoate 6 indicated the attachment of one more benzoyloxy groups in the molecule. Also, H-2 (at δ 5.79 as dd) protons shifted down field as compared to its precursor compound 6. Thus, the structure of the compound was assigned as methyl 4,6-0benzylidene-2,3-di-O-benzoyl-a-Dmannopyranoside (7).

effective against these Gram-positive organisms. Only methyl 4,6-*O*-benzylidene-2,3-di-*O*-benzoyl-a-D-mannopyranoside (**7**) exhibited considerable inhibition (*20 mm) against *Bacillus cereus* which was comparable to that of the standard antibiotic, ampicillin (*22 mm).

Table 1. Inhibition against Gram-positive organisms by the tested chemicals (1-7).

Compound no.	Diameter of zone of inhibition, in mm (50 μg, dw / disc)								
	Bacillus cereus	Bacillus megaterium	Bacillus subtilis	Staphylococcus aureus					
1	N/A	N/A	N/A	N/A					
2	08	N/A	N/A	N/A					
3	08	N/A	N/A	N/A					
4	N/A	N/A	N/A	N/A					
5	08	N/A	11	N/A					
6	N/A	N/A	N/A	N/A					
7	*20	N/A	N/A	N/A					
**Ampicillin	*22	19	*25	*21					

NB. "N/A" indicates no inhibition, dw = dry weight, "**" indicates standard antibiotic, "*" shows good inhibition.

Inhibition zone against the selected Gramnegative bacteria due to the effect of the monosaccharides (1-7) are mentioned in Table 2. The test chemicals showed better activity against these organisms as compared to that of Gram-positive organisms. The compounds

(1-7) did not show considerable inhibition against *E. col*i and INABAET (vibrio). The study revealed that the tested chemicals were more effective against *Salmonella paratyphi* and *Salmonella typhi*.

Table 2. Inhibition of the tested chemicals (1-7) against Gram-negative organisms.

	Diameter of zone of inhibition in mm (50 μg dw / disc)							
Compound	E. coli	INABAET (vibrio)	Pseudomonas aeruainosa	Salmonella paratyphi	Salmonella tvphi	Shigella dysenteriae		
1	N/A	N/A	N/A	08	N/A	N/A		
2	N/A	07	N/A	06	07	N/A		
3	06	12	08	09	14	N/A		
4	N/A	N/A	N/A	*21	*20	N/A		
5	08	N/A	14	*20	*20	N/A		
6	N/A	09	12	*21	*22	N/A		
7	15	N/A	14	17	18	13		
**Ampicillin	*25	*24	17	*35	13	*35		

NB. N/A indicates no zone of inhibition. dw=dry weight. ** indicates standard antibiotic, * shows good inhibition.

Structure activity relationship (SAR)

In vitro antibacterial study revealed that the monosaccharide derivatives (1-7) were more active against some Gram-negative organisms than that of Gram-positive organisms. An important observation was that the benzoylated mannopyranosides (5-7) were more active than that of the glucopyranosides (2-3). Again, compounds 1, 2 and 4 showed poor toxicity than that of compounds 3, 6, and 7 against these pathogens. This is probably due to the presence of more hydroxyl groups in 1, 2 and 4. While compounds 3, 6 and 7 having fewer or no hydroxyl groups showed much better antibacterial potentiality (mannopyranoside 5 was found to exceptional). Here the hydrophobicity of the molecules increased gradually from compounds 1, 2, and 4 to 3, 6, and 7. The hydrophobicity of compounds is an important parameter for bioactivity and is directly related to membrane permeation [25]. We believe that similar hydrophobic interaction might occur between the benzoyl groups of monosaccharides (3 and 7) accumulated in the lipid membranes of bacteria. consequence their hydrophobic interaction, bacteria lose their membrane permeability [26], ultimately causing death of the bacteria.

Conclusion

We have synthesized methyl 4,6-*0*benzylidene-2,3-di-O-benzoyl-a-Dglucopyranoside (3) and methyl 4,6-0benzylidene-2,3-di-O-benzoyl-a-Dmannopyranoside (7) from methyl a-Dglucopyranoside **(1)** and methyl a-D-(4),mannopyranoside respectively in reasonably good yields. A comparative in vitro

antibacterial study of these compounds was carried out successfully employing ten bacterial pathogens. The structure activity relationship (SAR) study revealed that the benzoylated mannopyranosides (5-7) were more active against the tested organisms than that of the glucopyranosides (1-3).

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References

- [1] Schaffer R. The Carbohydrates: Chemistry and Biochemistry. Pigman W, Horton D. (eds.), 2nd ed: Academic Press: New York; 1972. ISBN: 978-0318502380.
- [2] Kochetkov NK, Dmitrive BA, Backinowsky LV. New sugars from antigenic lipopolysaccharides of bacteria: identification and synthesis of 3-*O*-[(*R*)-1-carboxyethyl]-L-rhamnose, an acidic component of *Shigella dysenteriae* type 5 lipopolysaccharide. Carbohydrate Research 1976;51/2:229-237. DOI: 10.1016/S0008-6215(00)83330-3.
- [3] Dhavale DD, Matin MM. Selective sulfonylation of 4-C-hydroxymethyl- β -L-threo-pento-1,4-furanose: synthesis of bicyclic diazasugars. Tetrahedron 2004;60/19,4275-4281. DOI: 10.1016/j.tet.2004.03.034.
- [4] Dhavale DD, Matin MM. Piperidine homoazasugars: natural occurrence, synthetic aspects and biological activity study. Arkivoc 2005;iii:110-132. url: http://www.arkatusa.org/get-file/18916/
- [5] Matin MM, Bhuiyan MMH, Azad AKMS, Bhattacharjee SC, Harun-Or-Rashid M. Synthesis and antimicrobial studies of 6-*O*-lauroyl-1,2-*O*-isopropylidene-α-D-gluco-furanose derivatives. Chemistry & Biology Interface 2014;4/4:223-231. url: http://www.cbijournal.com/paper-archive/july-august-2014-vol-4/

- [6] Matin MM, Bhuiyan MMH, Debnath DC, Manchur MA. Synthesis and comparative antimicrobial studies of some acylated D-glucofuranose and D-glucopyranose derivatives. International Journal Biosciences 2013;3/8:279-287. DOI: 10.12692/ijb/3.8.279-287.
- [7] Matin MM. Synthesis and antimicrobial study of some methyl 4-O-palmitoyl- α -L-rhamnopyranoside derivatives. Orbital: The Electronic Journal of Chemistry 2014;6/1:20-28. url: http://www.orbital.ufms.br/index.php/Chemistry/art icle/view/553/pdf
- [8] Tusda Y, Haque ME, Yosimoto K. Regioselective monoacylation of some glycopyranosides via cyclic tin intermediates. Chemical and Pharmaceutical Bulletin 1983;31/5:1612-1624. DOI: 10.1248/cpb.31.1612.
- [9] Kabir AKMS, Matin MM, Ali M, Anwar MN. Comparative studies on selective acylation and antimicrobial activities of some D-glucofuranose derivatives. Journal of Bangladesh Academy of Sciences 2003;27/1):43-50. url: https://www.researchgate.net/publication/27274719 9.

[10] Kabir AKMS, Matin MM, Bhuiyan MMR, Rahim MA. Synthesis and characterization of some acylated derivatives of D-mannose. The Chittagong University Journal of Science 2001;25/1:65-73. url: https://www.researchgate.net/publication/2727918 94.

- [11] Kobayashi T. Lipase-catalyzed syntheses of sugar esters in non-aqueous media. Biotechnology Letters 2011;33/10:1911-1919. DOI: 10.1007/s10529-011-0663-z.
- [12] Microwave in Organic Synthesis. Loupy A. (ed), 2nd ed: Wiley-VCH Verlag GmbH & Co.: Weinheim; 2006. ISBN: 3-527-31452-0.
- [13] Perez-Tomas R. Multidrug resistance: retrospect and prospects in anti-cancer drug treatment. Current Medicinal Chemistry 2006;13/16:1859-1876. DOI: 10.2174/092986706777585077.
- [14] Ahsan F, Arnold JJ, Meezan E, Pillion DJ. Sucrose cocoate, a component of cosmetic preparations, enhances nasal and ocular peptide absorption. International Journal of Pharmaceutics 2003;251:195-203. DOI: 10.1016/S0378-5173(02)00597-5.
- [15] Csóka G, Marton S, Zelko R, Otomo N, Antal I. Application of sucrose fatty acid esters in transdermal therapeutic systems. European Journal of Pharmaceutics & Biopharmaceutics 2007, 65/2, 233-237. DOI: 10.1016/j.ejpb.2006.07.009.
- [16] Pouillart P, Douillet O, Scappini B, Gozzini A, Santini V, Grossi A, et al. Regioselective synthesis and biological profiling of butyric phenylalkylcarboxylic esters derivated from mannose and xylitol: influence of alkyl chain length on acute toxicity. European Journal of Pharmaceutical Sciences 1999;7/2:93-106. DOI: 10.1016/S0928-0987(98)00011-6.
- [17] Chortyk OT, Pomonis JG, Johnson AW. Synthesis and characterizations of some insecticidal sucrose esters. Journal of Agricultural & Food Chemistry, 1996;44/6:1551-1557. DOI: 10.1021/jf950615t.

- [18] Kabir AKMS, Matin MM, Sanaullah AFM, Sattar MA, Rahman MS. Antimicrobial activities of some lyxoside derivatives. Bangladesh Journal of Microbiology 2001;18/1:89-95. url: https://www.researchgate.net/publication/2726773 0.
- [19] Kabir AKMS, Matin MM, Bhuiyan MMR, Rahim MA, Rahman MS. Biological evaluation of some monosaccharide derivatives. International Journal of Agriculture & Biology 2005;7/2: 218-221. DOI: 1560-8530/2005/07-2-218-221.
- [20] Kabir AKMS, Matin MM, Mridha MAU, Shahed SM. Antifungal activities of some methyl 6-O-trityl-a-D-mannopyranosides. The Chittagong University Journal of Science 1998;22/1:41-46. url: https://www.researchgate.net/publication/2728336 48.
- [21] Kabir AKMS, Matin MM, Rahman MS. Antimicrobial activities of some rhamnoside derivatives. Chittagong University Journal of Science 2000;24/1:129-135. url: https://www.researchgate.net/publication/2727924 69.
- [22] Demchenko AV, Pornsuriyasak P, de Meo C. Acetal protecting groups in the organic laboratory: Synthesis of methyl 4,6-O-benzylidene-a-D-glucopyranoside. Journal of Chemical Education 2006;83/5:782-784. DOI: 10.1021/ed083p782.
- [23] Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. American Journal of Clinical Pathology 1966;45/4: 493-496. Pubmed: http://www.ncbi.nlm.nih.gov/pubmed/5325707.
- [24] Nashed MA, Anderson L. Organotin derivatives and the selective acylation and alkylation of the equatorial hydroxy group in a vicinal, equatorial-axial pair. Tetrahedron Letters 1976;17/39:3503-3506. DOI: 10.1016/S0040-4039(00)71342-6.

[25] Kim YM, Farrah S, Baney RH. Structure-antimicrobial activity relationship for silanols, a new class of disinfectants, compared with alcohols and phenols. International Journal of Antimicrobial Agents 2007;29/2:217-222. DOI: org/10.1016/j.ijantimicag.2006.08.036.

[26] Judge V, Narasimhan B, Ahuja M, Sriram D, Yogeeswari P, Clercq ED, Pannecouque C, Balzarini J. Synthesis, antimycobacterial, antiviral, antimicrobial activity and QSAR studies of N2-acyl isonicotinic acid hydrazide derivatives. Medicinal Chemistry 2013;9/1:53-76. DOI: 10.2174/157340613804488404.