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**THE ANALYSIS OF FATTY ACİDS LEVEL IN RAT HEARTH TISSUES PERFORMED
MYOCARDIAL ISCHEMIA-REPERFUSION INJURY AND MELATONIN**

ABSTRACT

Melatonin (N-acetyl-5-methoxytryptamine), the main secretory product of the pineal gland, is a free radical scavenger that has been found to protect against lipid peroxidation in many experimental models. In the present many study the effect of melatonin on lipid peroxidation of long chain polyunsaturated fatty acids located in rat. The most sensitive fatty acids in the heart were the monounsaturated eicosenoik acid (C20:1 n-6), polyunsaturated arachidonic acid (C20:4 n-6) docosapenthonoic acid (C22:5 n-3) and docosahexanoic acid (C22:6 n-3) which compared with the corresponding control values. I/R performed group showed remarkable differerences (p<0.05). In this study, myokardial- ischemia reperfusion is determined to cause increase or decrease in different types of fatty acids.

Keywords: Melatonin, Myocardial Ischemia-Reperfusion, Tissues, Fatty acids, Gas Chromatography

**SIÇANLARIN KALP DOKULARINDA YAĞ ASİTLERİ DÜZEYLERİNİN I/R VE MELATONİN
UYGULAMALARIYLA DEĞİŞİMLERİNİN İNCELENMESİ**

ÖZET

Melatonin- beyin epifizinin ana ürünü olan salgısı-birçok deneyde yağ peroksidasyonuna karşı savaşan serbest radikal bir temizleyici olduğu gözlemlendi. Mevcut birçok çalışmalarda-farede yapılan-melatoninin çoklu doymamış yağ asitlerine etkisi olduğu belirlendi. Araşidonik asit (C20:4 n-6) dokosapentaenoik asit (C22:5 n-3, tekli doymamış yağ asitleri, eicosenoik asit (C20:1 n-6) ve dokosaheksaenoik asit (C22:6 n-3) karşılık gelen kontrol değeri ile karşılaştırıldı. I/R çalışılan grup büyük farklılıklar gösterdi. Bu çalışmada miyokardiyal iskemi reperfüzyonunun farklı yağ asitleri türlerinde artmalara veya azalmalara sebep olduğu belirlendi.

Anahtar Kelimeler: Melatonin, Miyokardiyal İskemi Reperfüzyon, Doku, Yağ Asitleri, GC.



1. INTRODUCTION (GİRİŞ)

Melatonin (N-acetyl-5-metoxytryptamine) is an indole synthesized mainly by the the pineal gland of all mammals including humans (Reiter RJ 2003) and also, it is produced in a limited number of organs in tract (Tang PL 1997). Melatonin possesses free radical scavenging activity. Experimental studies have shown that melatonin directly scavenges the hydroxyl radical, peroxy radical, peroxy nitrite anion and singlet oxygen. Furthermore, this tryptophan derivative stimulates a number of antioxidant enzymes and stabilizes cell membranes (Allegra 2003). It is demonstrated that myocardial ischemia-reperfusion injury is related to increased free radical generated and intracellular calcium overload especially during the period of reperfusion (Sahna E, 2006). Moreover, melatonin has been shown to reduce the ischemia-reperfusion injury in various models of experimental I/R injuries (Deniz İ, 2006).

Several publications indicated that populations in Greenland and Japan exhibited a low incidence of sudden cardiac death (Bang, 1971). This was associated with a high n-3 PUFA intake. Since these early studies, numerous epidemiological investigations have been carried out. Most have reported the protective effect of n-3 PUFAs (Bang H. O 1981, Hodgson J., 1993, Osler M 2000) but others have not shown any correlation between the intake n-3 PUFAs and CHD mortality (Keys A 1986, Oomen C. M 2000, Marckmann P 1999). The first reference to the cardioprotective effects of fish oil (rich in n-3 PUFA) comes from the Greenland Eskimo population and dates back to 1976—the year when also the first statin was invented (H.O. Bang 1976, A. Endo 1976). Besides statins and life style changes, the n-3 PUFAs, especially eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), could play an important role in cardiovascular disease prevention (particularly in patients with heart failure R.B. Singh 2002). Lipid peroxidation is a free radical chain reaction (Kappus HA, (1987)) which causes the degeneration of cell membranes. Most products of lipid peroxidation are known to have mutagenic and/or carcinogenic properties (Fang, J.L. 1997). Free radical species affect all species affect all important components of cells such as lipids, proteins, carbohydrates and nucleic acids (Sarkar, 1997). Lipids are oxidized by free radical attack, and hence membranes are damaged (Cheesman, 1993).

The ability of melatonin to inhibit lipid peroxidation has been studied in various oxidative stress models of rat (Tang, 1997 and Deniz, 2006). Tang et al (3) reported that the direct effect of iron on lipid peroxidation in cell membranes from the brain, heart, kidney and liver of the male Sprague-Dawley rat was markedly reduced by melatonin in Tang 1997. The action of melatonin on the peroxidation of lipids in biological membranes has been studied in many laboratories (Garcia JJ 1997, Rudzite V 1999) but to a few studies have shown how long chain PUFA such as C20:4 n-6, C22:5 n-3 and C22:6 n-3 are protected by this indole or its derivatives during the lipid peroxidation process (Leadon, 2002 and Gavazza, 2004). Polyunsaturated fatty acids, specifically the n-3 series have been implicated in the prevention of various human diseases, including obesity, diabetes, coronary heart disease and stroke and inflammatory and neurologic diseases and regulate the expression of genes in various tissues, including the liver, heart, adipose tissue and brain (Sampath, 2004). The presented data offer the support to the hypothesis that pharmacological amounts of melatonin effectively reduce oxidative stress and display antihyperlipidemic activity. The decreased levels of thiobarbituric acid reactive substances found in the brain and liver tissues of melatonin-treated rats suggest that



melatonin may provide an effective protection against lipid peroxidation. In addition, melatonin reduces the free radical-induced alteration of microsomal membrane fluidity during induced lipid peroxidation (Garcia, 1997 and Longoni, 1998). A remarkable body of evidenc indicates that melatonin experts antioxidant protection in different experimental systems both in vitro and in vivo (Pandi-Perumal, 2006, Reiter, 2007 and Tan, 2007).

2. RESEARCH SIGNIFICANCE (ÇALIŞMANIN ÖNEMİ)

Melatonin (N-acetyl-5-methoxytryptamine), the main secretory product of the pineal gland, is a free radical scavenger that has been found to protect against lipid peroxidation in many experimental models. Cardiac-Ischemia reperfusion damage can appear in most clinic occassional; likes iscemic heart disease. We hope that this study will provide guidance to related with regulation of iscemic heart disease.

3. MATERIALS AND METHODS (MATERYAL VE METOT)

3.1. Experimental Groups (Deney Grubları)

Male Wistar rats weighing 200-250 g were placed in a temperature-(21±2°C) and humidty-(60 ±5%) controlled room in which a 12-12h light-dark cycle was maintained. Thirty rats were divided into six groups equally:-group1 (contro) received normal saline intraperitoneally(i.p) for 15 days-group 2 (L-NAME group) received non-specific NOS-N inhibitor LAME (40mg/kg, i.p.) for 15 days and melatonin's vehicle (ethanol) the last 5 days;-group 3 (melatonin group) received both L-NAME (40 mg/kg) for 15 days and melatonin (10 mg/kg, i.p) for the last 5 days of this time; and groups 4-6 were similar to groups 1-3, respectively, but at the end of 15 days, these groups were subjected to the myocardial IR.L-NAME (Fluka Chemie, Switzerland) was dissolved in normal saline (0.09% NaCl wt/vol). Melatonin (Sigma Chemical Co., St Louis, Missouri, USA) was dissolved in ethanol and Downloaded By:(TÜBTAK EKUAL) At: 14:01 15 October 2008 Hypertension, Reperfusion Injury, Myeloperoxidase 675 further diluted in salina(0.09% NaCl wt/vol) to give a final concentration of 1%. All experiments in this study were performed in accordancewith the guidelines for animal research from the National Institutes of Health and were approved by the Local Committee on Animal Research.

3.2. Ischemia-Reperfusion Procedure (İskemi-Reperfuzyon Uygulama)

Rats were anesthetized with urethane (1.2-1.4 g/kg) admimistered intraperitoneally. The trache was cannulated for artificial respiration. The chest was opened by a left thoracotomy. Positive-pressure artificial respiration was started immidiately with room air, using a volume of 1.5mL/100g body weight at a rate 60 beats/min to maintain normal pCO₂, pO₂, and pH parameters. A6/0 silk suture attached to a 10-mm micropoint reverse-cutting needle was quickly placed under the left main coronary artery. The artery was occluded for 30 min and then reperfused for 120 min (Sahna E, 2005).

3.3. Measurement of Fatty Acid Composition (Yağ Asitleri Ölçümü)

The lipids of the tissue samples were extracted with chlorophorm-metanol (2:1v/v) according to the method of Christie ve Folch et al. Tissue sample was homogenized with 10 ml hexane-isopropanol mixture. The homogenate was centrifuged at 5000 rpm for 5 min at 4°C and parts of tissue remnants were precipitated. Fatty acids in the lipid extracts were converted into methyl esters including 2% sulfuric acid (v/v) in methanol. The mixture was vortexed and then kept at 50°C for 12h.Then, after being cooled to room temperature, 5ml



of 5% sodium chloride was added and then it was vortexed. Fatty acid methyl esters were extracted with 2x5ml hexane. Fatty acid methyl esters were treated with 5 ml 2% KHCO₃ solution and then the hexane phase was evaporated by the nitrogen flow and then by dissolving in 0.5ml fresh hexane (Christie, Folch, Peter 1990). They were taken to auto sampler vials.

3.4. Gas Chromatographic Analysis of Fatty Acid Methyl Esters (Gas Kromatografisi İle Yağ Asitleri Tayini)

Methyl esters were analyzed with the Shimadzu GC-17 Ver. 3 gas chromatography (Kyoto, Japan). For this analysis, 25 m of long Permabond® FFAP-0.1 µm capillary column (Machery-Nagel, Germany) with an inner diameter of 0.25 µm and a thickness of 25 micron film was used. During the analysis, the column temperature was kept at 120-220 °C and the increment of temperature was 3°C/min, injection temperature was kept at 240°C and the detector temperature was kept at 280°C. The nitrogen carrier gas flow was 1 ml/min. The methyl esters of fatty acids were identified by comparison with authentic external standard mixtures analyzed under the same conditions. After this process, the necessary programming was made and the Class GC 10 software version 2.01 was used to process the data.

3.5. Statistical Analyses (İstatistiksel Analiz)

Data are expressed as the mean±SD. All statistical analyses were performed using SPSS 10.0 pack program (Chicago, IL, USA). Statistical comparisons were made with Mann-Whitney-U test determining differences among groups. A value of P<0.05 was considered significant.

Table 1. Chemical composition (relative % peak area) of the fatty acids in the Cardiac-Ischemia reperfusion, melatonin-Ischemia and control

Fatty Acids		Control	C-Ischemia	Mel-Ischemia	
		X ± Sx(n=4)	X ± Sx(n=4)	X ± Sx(n=4)	
Miristik acid	(C14:0)	0.56±0.29	0.59±0.20	0.37±0.04	b
Pentadecanoic	(C15:0)	0.20±0.02	0.22±0.01	0.19±0.03	
Palmitic acid	(C16:0)	16.30±1.29	15.80±1.74	14.77±0.15	ab
Palmitoleic acid	(C16:1)	1.50±0.27	1.36±0.19	0.94±0.27	ab
Heptadecanoic	(C17:0)	0.82±0.07	0.75±0.05	0.5 ±0.08	ab
Stearic acid	(C18:0)	17.63±1.65	16.49±0.87	18.70±0.76	b
Oleic acid	(C18:1)	12.58±1.09	12.73±1.84	9.52±0.74	ab
Linoleic acid	(C18:2)	26.33±1.79	26.96±1.15	24.32±1.06	b
Linolenic acid	(C18:3)	0.45±0.15	0.62±0.12	0.48 ±0.07	
Eicosenoic	(C20:1)	0.58±0.18	0.75±0.06	0.62±0.09	b
Eicosatetraenoic	(C20:4)	11.27±0.82	11.54±1.49	13.59±0.93	ab
Eicosapentaenoic	(C20:5)	0.44±0.08	0.42±0.08	0.43±0.10	
Docosenoic acid	(C22:1)	0.33±0.15	0.26±0.09	0.34±0.11	
Arachidonic acid	(C22:4)	0.15±0.04	0.13±0.04	0.12±0.02	
Docosapentaenoic	(C22:5)	1.65±0.16	1.63±0.15	1.94±0.17	ab
Docosahexaenoic	(C22:6)	9.20±1.14	9.63±0.81	12.90±0.84	ab
ΣSATURATE		35.38±0.31	33.86±1.22	34.63±0.81	
ΣSATURATE		64.61±0.31	66.15±1.22	65.35±0.80	
ΣMUFA (1)		15.10±1.02	15.35±2.33	11.46±1.10	ab
ΣPUFA (2, 3, 4, 5, 6)		49.51±1.92	50.98±3.20	53.92±0.94	ab

a: p<0.05,

b: p<0.001,

*: not detectable, Values are means ± SE



4. RESULTS (SONUÇ)

I/R performed group showed remarkable differences ($p < 0.05$). In this study, myocardial-ischemia reperfusion is determined to cause increase or decrease in different types of fatty acids. Table 1 summarizes the results obtained in the heart respectively. Melatonin at groups caused significant differences in the levels of Ischemia, free fatty acids in the heart when compared to controls. The most sensitive fatty acids in the heart were the monounsaturated eicosenoic acid (C20:1 n-6), polyunsaturated arachidonic acid (C20:4 n-6) docosapentenoic acid (C22:5 n-3) and docosahexanoic acid (C22:6 n-3) which compared with the corresponding control values, had increased. The addition of melatonin protected these PUFA especially arachidonic and docosahexanoic acids located in heart.

5. DISCUSSION (TARTIŞMA)

Cardiac-Ischemia reperfusion damage can appear in most clinical occasions; like ischemic heart disease, thrombotic treatment, percutaneous coronary interventions (balloon angioplasty), coronary artery bypass surgery, heart valve operations. (Sahna, Acet et al. 2002; Sahna, Parlakpınar et al. 2005). Most aerobic cells suffer oxidative damage as a consequence of the formation of reactive species that are generated as secondary products during respiration or from specific enzymatic reactions (Kohen, 2002). Damage to lipid deleteriously alters and modifies cellular membranes and, therefore, cellular function (Porter, 1984). Numerous studies have reported melatonin's protection against lipid peroxidation and DNA damage induced by ROS, both in vivo and in vitro (Karbownik, 2001 and Tan, 1993) but fewer workers have focused on melatonin's efficacy in reducing fatty acid damage analyzing the degradation of specific fatty acids located in selected membranes (Leadon, 2002 and Gavazza, 2004). Melatonin suppresses the ascorbate-Fe⁺⁺ induced lipid peroxidation process in rat liver, kidney and brain microsomal preparations and protects the most common PUFAs, arachidonic and docosahexanoic acid, from damage. Melatonin seems to be equally efficient in vitro in the protection of fatty acids that belong either to the n-6 and/or to the n-3 family. The physiological potential of melatonin in preventing lipid degradation of PUFAs in different organelles isolated from different tissues deserves further investigation (Patricio, 2005). The effect of a daily administration of melatonin for 45 days at two doses (0.5 and 1.0 mg/kg body wt.) on antioxidant status, lipid peroxidation and lipid profile in the brain and liver in rats. Both doses of melatonin caused a significant decrease in lipid peroxidation and the levels of cholesterol, phospholipids, triglycerides and free fatty acids in the examined tissues (Perumal Subramanian 2007). The decrease in the levels of cholesterol, phospholipids, triglycerides and free fatty acids found in the brain and liver of melatonin-treated rats underlines a significant antihyperlipidemic action of melatonin (Tunéz, 2002) which could be exerted by augmenting the clearance of endogenous cholesterol (Hoyos, 2000). Mortality and morbidity from coronary heart disease (CHD), diabetes mellitus (DM) and essential hypertension (HTN) are higher in people of South Asian descent than in other groups. There is evidence to believe that essential fatty acids and their metabolites may have a role in the pathobiology of CHD, DM and HTN. (Das, 1995).

NOTICE (NOT)

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REFERENCES (KAYNAKLAR)

- Reiter, R.J., Tan, D.X., Mayo, J.C., and et al., (2003). Melatonin as an antioxidant: Biochemical Mechanisms and Patho Physiological Implications in Humans. *Acta Biochim Pol*, 50:1129-46.
- Tang, P.L., Xu, M.F., and Qian, Z.M., (1997). Differential behaviour of cell membranes towards iron-induced oxidative damage and the effects of melatonin. *Biol Signals*, 6:291-300.
- Allegra, M., Reiter, R.J., Tan, D.X., and et al., (2003). The chemistry of melatonin 's interaction with reactive species. *J. Pineal Res*, 34:1-10.
- Sahna, E., Deniz, E., and Aksulu, H.E., (2006). Myocardial ischemia-reperfusion injury and melatonin. *Anadolu, Jun*, 6(2):163-8.
- Deniz, E., Colakoglu, N., Sari, A., Sonmez, M.F., Tugrul, I., Oktar, S., Ilhan, S., and Sahna, E., (2006). Melatonin attenuates renal ischemia-reperfusion injury in nitric oxide synthase inhibited rats. *Acta Histochem*, 108(4):303-9. Epub Jun 9.
- Bang, H.O., Dyerberg, J., and Nielsen, A.B., (1971). Plasma lipid and lipoprotein pattern in Greenlandic West-coast Eskimos. *Lancet* 1:1143-1145.
- Bang, H.O., and Dyerberg, J., (1981). Personal reflections on the incidence of ischaemic heart disease in Oslo during the Second World War. *Acta Med. Scand.* 210:245-248.
- Hodgson, J., Wahlqvist, M., Boxall, J., and Balazs, N., (1993). Can linoleic acid contribute to coronary artery disease? *Am. J. Clin. Nutr.* 58:228-234.
- Osler, M., Godtfredsen, J., Gronbaek, M.N., Marckmann, P., and Overvad, O.K., (2000). A quantitative assessment of the impact of diet on the mortality of heart disease in Denmark. Estimation of etiologic fraction. *Uges. Laeger* 162:4921-4925.
- Keys, A., Menotti, A., Karvonen, M., Aravanis, C., Blackburn, H., Buzina, R., and et al., (1986). The diet and 15-year death rate in the seven countries study. *Am. J. Epidemiol.* 124:903-915.
- Oomen, C.M., Feskens, E.J., Rasanen, L., Fidanza, F., Nissinen, A.M., Menotti, A., and et al., (2000). Fish consumption and coronary heart disease mortality in Finland, Italy, and The Netherlands. *Am. J. Epidemiol.* 151:999-1006.
- Marckmann, P., and Gronbaek, M., (1999). Fish consumption and coronary heart disease mortality. A systematic review of prospective cohort studies. *Eur. J. Clin. Nutr.* 53:585-590.
- Bang, H.O., Dyerberg, J., and Hjoorne, N., (1976). The composition of food consumed by Greenland Eskimos, *Acta Med. Scand.* 200:69-73.
- Endo, A., Kuroda, M., and Tanzawa, K., (1976). Competitive inhibition of 3- HMGCoA reductase by ML 236 A and ML 236 B, *FEBS Lett.* 72:323-326.
- Singh, R.B., Dubnov, G., NIaz, M.A., and et al., (2002). Effect of an Indo Mediterranean diet on progression of coronary artery disease in high risk patients (Indo-Mediterranean Diet Heart Study): a randomised single-blind trial, *Lancet* 360, 1455-1461.
- Kappus, H.A., (1987). A survey of chemicals inducing lipid peroxidation in biological systems. *Chem. Phys. Lipids.* 45:105-15.



- Fang, J.L., Vaca, C.E., Valsta, L.M., and Mutanen, M., (1996). Determination of DNA adducts of malonaldehyde in humans: effects of dietary fatty acid composition. *Carcinogenesis* 17:1035-1040.
- Sarkar, S., Yadav, P., and Bhatnagar, D.J., (1997). Cadmium-induced lipid peroxidation and the antioxidant system in rat erythrocytes: the role of antioxidants. *Trace Elem. Med. Biol* 11:8-13.
- Cheesman, K.H., and Slater, T.F., (1993). Introduction to free radical biochemistry. *Br. Med. Bull.* 49:481-493.
- Garcia, J.J., Reiter, R.J., Guerrero, J.M., and et al., (1997). Melatonin prevents changes in microsomal membrane fluidity during induced lipid peroxidation. *FEBS Lett*, 408:297-300.
- Rudzite, V., Jurika, E., and Jirgensons, J., (1999). Changes in membrane fluidity induced by tryptophan and its metabolites. *Adv Exp Med Biol*, 467:353-367.
- Leaden, P., Barrionuevo, J., and Catala, A., (2002). The protection of long chain polyunsaturated fatty acids by melatonin during non enzymatic lipid peroxidation of rat liver microsomes. *J Pineal Res* 32:129-134
- Gavazza, M.B., and Catala, A., (2004). Protective effect of N-acetyl-serotonin on the non enzymatic lipid peroxidation in rat testicular microsomes and mitochondria. *J Pineal Res*, 37:153-160.
- Sampath, H., and Ntambi, J.M., (2004). Polyunsaturated fatty acid regulation of gene expression. *Nut Rev* 2004; 62:333-339.
- Garcia, J.J., Reiter, R.J., Guerrero, J.M., Escames, G., Yu, B.P., Oh, C.S., and Munoz-Hoyos, A., (1997). Melatonin prevents changes in microsomal membrane fluidity during induced lipid peroxidation. *FEBS Lett.* 408:297-300.
- Longoni, B., Salgo, M.G., Pryor, W.A., and Marchiafava, P.L., (1998). Effects of melatonin on lipid peroxidation induced by oxygen radicals. *Life Sci*, 62:853-859.
- Sahna, E., Parlakpınar, H., Turkoz, Y., and Acet, A., (2005). Effects of melatonin on myocardial ischemiareperfusion-induced infarct size and oxidative stress. *Physiol Res*, 54:491-495.
- Christie, W.W., (1990). *Gas Chromatography and Lipids*, The Oil Press, Glasgow, 302-320.
- Folch, W., Robert, P., Lees, M., and Sladane-Stanly, G.H.A., (1957). Simple Method for The Isolation and Purification of Total Lipids from Animals Tissue, *J. Biol. Chem.*, 226, 447-509.
- Peter, S., (1973). Extraction and Purification of Lipids, II Why is Chloroform-Methanol Such Good Lipid Solvent, *Physiol. Chem. And Physic.* 5, 141-149.
- Kohen, R., and Nyska, A., (2002). Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. *Toxicol Pathol*, 30:620-650.
- Porter, N.A., (1984). Chemistry of lipid peroxidation. *Methods Enzymol*, 105:273-282.
- Karbownik, M., Reiter, R.J., Burkhardt, S., and et al., (2001). Melatonin attenuates estradiol-induced oxidative damage to DNA: relevance for cancer prevention. *Exp Biol Med*, 226:707-712.
- Tan, D.X., Chen, L.D., Poeggeler, B., and et al., (1993). Melatonin: a potent, endogenous hydroxyl radical scavenger. *Endocrine J*, 1:57-60.
- Leaden, P., Barrionuevo, J., and Catala, A., (2002). The protection of long chain polyunsaturated fatty acids by



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- melatonin during non enzymatic lipid peroxidation of rat liver microsomes. *J Pineal Res*, 32:129-134.
- Gavazza, M.B., and Catala, A., (2004). Protective effect of N-acetyl-serotonin on the non enzymatic lipid peroxidation in rat testicular microsomes and mitochondria. *J Pineal Res*, 37:153-160.
 - Patricio, J., Leaden and Angel Catala', (2005). Protective effect of melatonin on ascorbate-Fe²⁺ lipid peroxidation of polyunsaturated fatty acids in rat liver, kidney and brain microsomes: a chemiluminescence study *J. Pineal Res.* 39:164-169.
 - Subramanian, P., Mirunalini, S., Seithikurippu, R., Pandi-Perumal., and Ilya Trakht , D.P., (2007). Cardinali Melatonin treatment improves the antioxidant status and decreases lipid content in brain and liver of rats, *European Journal of Pharmacology* 571:116 -119.
 - Tunez, L., Munoz, M.C., Feijoo-Lopez, A.L., Valdovira, E., Bujalance-Arenas, L., and Montilla, P., (2002). Effect of melatonin on hyperlipidemic nephropathy under constant light exposure. *J. Physiol. Biochem.* 58:109-114.
 - Hoyos, M., Guerrero, J.M., Perez-Cano, R., Oliván, J., Fabiani, F., Garcia-Perganeda, A., and Osuna, C., (2000). Serum cholesterol and lipid peroxidation are decreased by melatonin in diet-induced hypercholesterolemic rats. *J. Pineal Res.* 28, 150-155.
 - Das, U.N., (1995). Essential fatty acid metabolism in patients with essential hypertension, diabetes mellitus and coronary heart disease. *Prostaglandins Leukot Essent Fatty Acids.* Jun, 52(6):387-91.