



Experimental Research

## Antibiotic susceptibilities of group B streptococci

Aziz Ramazan Dilek<sup>a\*</sup>, Hasan Kesbiç<sup>b</sup>, Nursel Dilek<sup>b</sup>

<sup>a</sup>Training and Research Hospital, Microbiology Laboratory, Rize, Turkey

<sup>b</sup>Dr. İ. Şevki Atasagun State Hospital, Nevşehir, Turkey

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#### \* Correspondence to

Aziz Ramazan Dilek

Eğitim Araştırma Hastanesi,

Mikrobiyoloji Laboratuvarı, Rize

e-mail:ar.dilek@hotmail.com

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### ABSTRACT

Streptococcus agalactiae is a frequent cause of serious infections in newborn babies. It has increasingly been recognized as a pathogen in nonpregnant adults. Penicillin and ampicillin are the drugs of choice for treatment of group B streptococcus (GBS) infections. Widespread use of these antibiotics has potentiated the emergence of antibiotic resistance. Total 56 isolates included in the study were recovered from vaginal swabs and urine samples. Isolates were tested for antimicrobial susceptibility by Kirby-Bauer disk diffusion susceptibility testing method. All of the 56 clinical isolates tested were fully susceptible to penicillin, ampicillin and cephalothin. Only 8 (14.2%) and 6 (10.7%) were resistant to erythromycin and clindamycin, respectively. All of the 56 clinical isolates were not susceptible to tetracycline.

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### 1. Introduction

Streptococcus agalactiae or group B streptococcus (GBS) is a frequent cause of serious infections often associated with mortality and morbidity in newborn babies (Al-Sweih et al., 2005). In the 1990s, 4 to 6 percent of affected newborns died from the infection (Schrag et al., 2000). It has increasingly been recognized as a pathogen in nonpregnant adults, especially among patients with underlying conditions (Betriu et al., 2003). The incidence of group B streptococcal disease is also high in pregnant women and the elderly (Schrag et al., 2000). Penicillin and ampicillin are the drugs of choice for treatment of GBS infections, and clindamycin and erythromycin are the recommended alternatives for patients who are allergic to  $\beta$ -lactam agents (Quiroga et al., 2008). Clindamycin and erythromycin resistance rates are 15-20 % and they are increasing worldwide (Ölçü and Eşel, 2007). Widespread use of these antibiotics in various clinical conditions has potentiated the emergence of antibiotic resistance (Quiroga et al., 2008).

This study was performed to determine the susceptibility patterns of the Streptococcus agalactiae isolates to a variety of antibiotics.

### 2. Material and Methods

The 56 isolates, recovered from vaginal swab and urine samples included in the study retrospectively. A total of 56 pa-

tients included the study were aged 22 to 66 years.

The arithmetic mean, median age of patients were 38.50 and 38.00, respectively. All isolates were identified by conventional methods as GBS. Isolates were tested for antimicrobial susceptibility to penicillin, ampicillin, cephalothin, clindamycin, ciprofloxacin, vancomycin, Trimethoprim/sulfamethoxazole, tetracycline, imipenem and erythromycin by Kirby-Bauer disk diffusion susceptibility testing method. Bacteria grown on blood agar (Oxoid, Basingstoke, UK) suspended in sterile saline to a 0.5 McFarland turbidity standard as an inoculum containing 105 cfu/ml. This was inoculated onto a Mueller-Hinton agar plate supplemented with 5% sheep blood, by streaking evenly with a swab. Then the plates were incubated in an aerobic incubator at 35 ° C for 24 h. The zone sizes were then recorded for each antimicrobial agent and interpreted according to the Clinical and Laboratory Standards Institute (CLSI).

### 3. Result

A total of 56 GBS isolates were collected. All of the 56 clinical isolates tested were fully susceptible to penicillin, ampicillin and cephalothin. All of the 56 clinical isolates were not susceptible to tetracycline. Only 8 (14.2%) and 6 (10.7%) were resistant to erythromycin and clindamycin, respectively. Ciprofloxacin resistance rate was 3 (5.3%). Results of antibiotic susceptibility test was demonstrated in Table 1.

**Table 1:** Antibiotic susceptibility profiles of group B streptococcus

Antimicrobial agent	Susceptible	Intermediate	Resistant
Penicillin	100	--	--
Ampicillin	100	--	--
Cephalothin	100	--	--
Imipenem	100	--	--
erythromycin	85.7	--	14.2
clindamycin	89.2	--	10.7
ciprofloxacin	85.7	8.9	5.3
vancomycin	100	--	--
Trimethoprim/sulfamethoxazole	92.8	1.7	5.3
tetracycline	--	--	100
<b>Total: 56 isolates</b>			

#### 4. Discussion

*Streptococcus agalactiae* is one of the pathogens responsible for peripartum maternal and neonatal infections (De Mouy et al., 2001). The source of the organism in most of these cases maternal genital or gastrointestinal tract or both (Kulkarni et al., 2001). Transmission from mother to child has been reported to be 29 times higher in GBS-colonized mothers than in noncolonized mothers (Elbaradie et al., 2009). While the colonization rates of group B streptococcus in pregnant women have been reported range from 4% to 18.6% in various studies, (Karakus et al., 2007) a carrier rate of 31.6% was found in pregnant women in one area of Zimbabwe (Moyo et al., 2002). In other study GBS colonization was found in 19 of 100 parturient women (Kraśnianin et al., 2009).

A Centers for Disease Control and Prevention (CDC) surveillance study estimated that the use of intrapartum chemoprophylaxis has prevented 4500 cases per year of GBS sepsis and 225 deaths per year in the United States (Money and Dobson, 2004). Penicillin is still the drug of choice for treatment of GBS infection and erythromycin is alternative to the penicillins in cases of intolerance. The increasing prevalence of macrolide resistance raises concern about the empirical use

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of these antibiotics for the prevention of GBS infections (Fluegge et al., 2004; Tünger et al., 2005). While aminopenicillins are still highly active against GBS in most susceptibility studies such as our study (Al-Sweih et al., 2005; Quiroga et al., 2008; De Mouy et al., 2001; Fluegge et al., 2004; Topkaya et al., 2003; Figueira-Coelho et al., 2004; Monique et al., 2006), resistance to erythromycin and clindamycin have been reported in many studies range from 0.7% to 21.4% for erythromycin and 1.7% to 17.5% for clindamycin (Betriu et al., 2003; Quiroga et al., 2008; Ölçü and Eşel, 2007; De Mouy et al., 2001; Fluegge et al., 2004; Topkaya et al., 2003; Figueira-Coelho et al., 2004). Compared to this studies which have been reported from our country and other countries, the rates of erythromycin and clindamycin resistance were concordant in our experience.

While Resistance to quinolones has been described recently for GBS, An increase in resistance of GBS to quinolones has been reported (Quiroga et al., 2008; Topkaya et al., 2003; Kawamura et al., 2003). Compared the study which has been reported from Argentina the rate of ciprofloxacin resistance is higher in our study (Quiroga et al., 2008).

While resistance to Trimethoprim/sulfamethoxazole have been reported range from 10.6% to 53.2%, the rate of Trimethoprim/sulfamethoxazole resistance is lower in our study (De Mouy et al., 2001; Quiroga et al., 2008).

In addition All of the 56 clinical isolates were Resistant to tetracycline. Reported rates of tetracycline resistance were range 29% to 92.6 (Quiroga et al., 2008; Ölçü and Eşel, 2007; De Mouy et al., 2001; Topkaya et al., 2003).

With more widespread use of antibiotics, selection of antibiotic-resistant GBS may occur. We recommended that routine reporting of GBS susceptibilities should be performed to help guide the clinician in the choice of antibiotic therapy for invasive GBS infections and eradication of colonization.

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