

In vitro Carbonic Anhydrase Inhibitory Effects of the Extracts of *Satureja cuneifolia*

Fatma Gülruy AYDIN¹, Emir Alper TÜRKÖĞLÜ^{2*}, Müslüm KUZU³, Turgut TAŞKIN⁴

¹Department of Pharmacognosy, Faculty of Pharmacy, University of Health Sciences Turkey, Turkey

²Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, University of Health Sciences Turkey, Turkey

³Department of Nutrition and Dietetics, Faculty of Health Sciences, Karabük University, Turkey

⁴Department of Pharmacognosy, Faculty of Pharmacy, Marmara University, Turkey

*Sorumlu Yazar: alper.turkoglu@sbu.edu.tr

Received: 09.08.2021 Received in revised: 10.09.2021 Accepted: 14.10.2021

Abstract

Carbonic anhydrases catalyze the interconversion between carbon dioxide and bicarbonate. Carbonic anhydrase inhibition has therapeutic importance and there are many drugs which use this mechanism. Carbonic anhydrase inhibitors are used as diuretic, antiglaucoma, antitumor and antiepileptic agents. As an alternative for synthetic chemicals with strong side effects, natural products have gained popularity in the recent years. In this study, *Satureja cuneifolia*, a medicinal plant used as folk medicine, was investigated for its inhibition effects on carbonic anhydrase I and II enzymes. Four different extracts were obtained with maceration method and three different solvents were used. The results have shown that methanol extracts of *Satureja cuneifolia* has the strongest inhibition activity on the enzymes with the IC₅₀ values of 31 µg/mL for hCA I and 12 µg/mL for hCA II. Further purification and analytical studies will be needed to obtain the active natural molecules and their potential for pharma and food industries.

Key words: Carbonic anhydrase, Inhibition, Extract, *Satureja cuneifolia*.

Satureja cuneifolia Ekstraktlarının *in vitro* Karbonik Anhidraz İnhibitör Etkileri

Öz

Karbonik anhidrazlar, karbon dioksit ve bikarbonat arasındaki karşılıklı dönüşümü katalize eder. Karbonik anhidraz inhibisyonunun terapötik önemi vardır ve bu mekanizmayı kullanan birçok ilaç vardır. Karbonik anhidraz inhibitörleri diüretik, antiglokom, antitümör ve antiepileptik ajanlar olarak kullanılmaktadır. Güçlü yan etkileri olan sentetik kimyasallara alternatif olarak son yıllarda doğal ürünler popülerlik kazanmıştır. Bu çalışmada, halk ilacı olarak kullanılan tıbbi bir bitki olan *Satureja cuneifolia*'nın karbonik anhidraz I ve II enzimleri üzerindeki inhibisyon etkileri araştırılmıştır. Maserasyon yöntemi ile dört farklı ekstrakt elde edilmiş ve üç farklı çözücü kullanılmıştır. Sonuçlar, *Satureja cuneifolia*'nın metanol ekstraktlerinin, hCA I için 31 µg/mL ve hCA II için 12 µg/mL IC₅₀ değerleri ile enzimler üzerinde en güçlü inhibisyon aktivitesine sahip olduğunu göstermiştir. Aktif doğal moleküllerin elde edilebilmesi ve bunların ilaç ve gıda endüstrileri için potansiyellerini değerlendirmek için daha fazla saflaştırma ve analitik çalışmalara ihtiyaç duyulmaktadır.

Anahtar Kelimeler: Karbonik anhidraz, İnhibisyon, Ekstrakt, *Satureja cuneifolia*.

Introduction

Carbonic anhydrases (CAs; EC 4.2.1.1) are widespread metalloenzymes catalyzing carbon dioxide hydration to bicarbonate and protons (Supuran, 2008). The enzymes are expressed in

organisms by eight gene families (α , β , γ , δ , ζ , η , θ and ι) (Güler et al., 2020). α -gene family secretes 16 CA isoforms with different locations (Nar et al., 2013; Orhan et al., 2016), molecular characteristics, kinetics to ligands and organization (Nocentini et al., 2016). hCA I, II, III, VII and XIII

isoenzymes are cytosolic isoforms, hCA IV, IX, XII, XIV and XV are membrane-bound forms, hCA VA and VB are present in mitochondria, hCA VI is secretory form (Almajan et al., 2008) and hCA VIII, X and XI are non-catalytic forms of the enzyme (Abdel-Aziz, et. al., 2014).

Activation and/or inhibition of carbonic anhydrases have provided unique approaches in the treatment/prevention of many diseases (Scozzafava et al. 2004). For instance, carbonic anhydrase inhibitors (CAIs) have been utilized as potential pharmaceuticals in Alzheimer, glaucoma, epilepsy, cancer, osteoporosis and prevention of some infectious diseases (Supuran, 2008). Several synthetic compounds have been used as CAIs up to date (Türkoğlu et al. 2017; Ucar et al., 2021). Production procedures of these compounds are complex, and these molecules may show possible side effects (Akkemik et al., 2019). However, natural products have been important candidates in pharmaceutical industry and acted as vital position for the discovery of novel drugs. These kinds of products may possess the ability for binding to specific protein-based biomolecules whose functions are crucial in the treatment of many diseases (Türkoğlu et al., 2019).

Satureja L. is a genus that belongs to Lamiaceae family (Oke et al., 2009), the tribe Mentheae within sub-family Nepetoideae. The genus includes over thirty species. Five of them are endemic, 15 different species of *Satureja* L. are located in Turkey (Kan and Uçan, 2006). Various members of the genus *Satureja* have medicinal and aromatic properties (Eminagaoglu et al., 2007). *Satureja* L. can be found in different areas of the world such as Mediterranean Region, North Africa, North Asia, South America, and Canary Islands (Taslimi et al., 2020).

S. cuneifolia is an aromatic plant which is found in Mediterranean area of Turkey. It has characteristic taste and it is used as spice and tea (Milos et al., 2001). The aerial parts of the plant are used as traditional medicine. The tea prepared from *S. cuneifolia* is used for the treatment of infection, muscle pain, cramp, indigestion, diarrhea and nausea in folk medicine (Eminagaoglu et al., 2007).

In the view of therapeutic and medicinal potentials of CAs and natural products as CAIs, we aimed to determine the esterase activity of different extracts of *Satureja cuneifolia* on hCA I and II isoenzymes.

Materials and Methods

Plant material

Satureja cuneifolia was identified by Dr. İsmail Şenkardeş from Pharmaceutical Botany

Department of Marmara University and deposited at the Herbarium of Marmara University with the number of 18815 for future reference.

Chemicals and instruments

Human carbonic anhydrase I and II isoenzymes, 4-nitrophenylacetate as substrate and trizma base were purchased from Sigma-Aldrich (Saint Louis, MO, USA). Dimethyl sulfoxide was commercially supplied from Isolab (Turkey). Water for experimental purposes and buffer preparations was obtained Direct Q[®]3 UV water purification system (Millipore Corp., MA, USA). Ohaus PA224C (Ohaus Corp., USA) with the readability up to 0.1 mg was used to weigh all extracts and chemicals. Wtw - Inolab pH 730 pH-meter (WTW GmbH, Weilheim, Germany) was performed to arrange the pH values. For the mixing and stirring experimental processes, IKA RT10 magnetic stirrer (IKA-Werke GmbH & Co KG, Germany) and ZX3 Advanced Vortex Mixer (Velp Scientifica, Usmate, Italy) were operated. Bioactivities of plant extracts on the enzymes were performed by UV-1800 Spectrophotometer (Shimadzu, Kyoto, Japan) with the resolution of 1 nm.

Preparation of the extracts

As detailed in the previous study of one of the authors, the aerial parts of *S. cuneifolia* were dried at room temperature in the shade and then powdered. These powdered parts (50 g) were macerated with petroleum ether (SFPE), chloroform (SFC), methanol (SFM), respectively. In addition to this extraction process, another powdered parts of *S. cuneifolia* (20 g) were extracted directly with methanol and with the use of maceration method. This process was continued for the 24-hour cycles until colorless solution was obtained. Then filtration was performed for the extracts and these extracts were evaporated by rotary evaporator (Heidolph, Germany). The extracts were held in +4°C until the experimental use (Taşkın et al., 2020).

Esterase activity assay of human carbonic anhydrase I and II

The method described by Verpoorte et al. (1967) was used for the esterase activity assay hCA I and hCA II. Spectrophotometry-based detection in the conditions of 348 nm for 3 min. at 25°C was assayed in the absorbance changes of 4-nitrophenyl acetate (NPA) to 4-nitrophenylate ion (Ağgül et al., 2020). The reference analysis without enzyme was tested before kinetic analysis and then bioactivities of the extracts were carried out. The extracts for different concentration in the experiment were repeated in triplicate. Control

cuvette activity in the absence of inhibitor (extract) was acknowledged as 100%. Finally, the inhibition features for hCA I and II isoenzymes were determined from the activity (%)-[inhibitor] graph (Kuzu et al., 2016).

Result and Discussion

Natural products are chemical molecules produced by all organisms of the three domains of life and have many drug-like bioactivity (Sorokina and Steinbeck, 2020). Plants as naturally gifted organisms (Huie, 2002) consist of many bioactive natural products commonly used in pharma, food and cosmetic industries. Extraction processes have been commonly used to separate significant natural molecules/products for the first step of sample preparation of plant materials (Wang and Weller, 2006). Extraction is an important sample preparation step in the studies on the bioactive components of the plant, and the results of the study depend on the selection of the appropriate extraction technique (Azmir et al., 2013). The selection of suitable solvent is one of the vital issues in the extraction process. Therefore, polar solvents extract polar substances while non-polar solvents extract non-polar substances. Solvent extraction is the most preferred extraction techniques for the preparation of extracts from plants (Gupta et al. 2012).

There are several *in vitro* bioactivity studies of plant extracts on carbonic anhydrase enzyme. Ahmad et al. (2019) reported bioactivity of *Cassia absus* L. seed extracts on CA. *In vitro* bioactivities of the fractions obtained from crude ethanol

extract were investigated in the study. Ethanol extract of the plant exhibited the best potential inhibitory features (IC_{50} : $1875 \pm 0.9 \mu\text{g/mL}$) against CA.

Kaya et al. (2019) have focused on the bioactivities of the extracts of *Alcea rosea*, *Foeniculum vulgare*, *Elettaria cardamomum*, *Laurus azorica* and *Lavandula stoechas* on hCA I and hCA II. In this study, the methanol extract of *Elettaria cardamomum* has possessed the highest inhibition profile (0.032 mg/mL) for hCA I. The methanol extract of *Lavandula stoechas* demonstrated the highest inhibitory characteristics (0.054 mg/mL) on hCA II.

Another study was conducted on the inhibitory features of *Cucumis melo* L. seed extracts against hCA I and hCA II. In this study, oil and methanol extracts of *Cucumis melo* L. seeds showed different bioactivities for hCA I and hCA II. While these extracts activated the hCA I, other isoenzyme, hCA II, was inhibited by oil extract with the IC_{50} value of 0.497 ng/mL and also methanol extract with the IC_{50} value of 10.98 $\mu\text{g/mL}$ (Akkemik et al., 2019).

Several bioactivity studies about the extracts of *Satureja cuneifolia* have been investigated in previous studies (Oke et al., 2009; Taslimi et al., 2020). To the best of our knowledge, the inhibitory features of the extracts of *Satureja cuneifolia* on hCA I and hCA II have been studied for the first time.

Table 1. Effect of *Satureja cuneifolia* extracts on hCA I and hCA II

Extracts of <i>S. cuneifolia</i>	hCA I		hCA II	
	IC_{50}	R^2	IC_{50}	R^2
Petroleum ether extract	44 $\mu\text{g/mL}$	0.9485	101 $\mu\text{g/mL}$	0.9569
Chloroform extract	161 $\mu\text{g/mL}$	0.9662	132 $\mu\text{g/mL}$	0.9799
Methanol extract	68 $\mu\text{g/mL}$	0.9511	12 $\mu\text{g/mL}$	0.9432
Direct methanol extract	31 $\mu\text{g/mL}$	0.9597	13.6 $\mu\text{g/mL}$	0.9261
Standard (acetazolamide)	6.07 nM (Taslimi et al., 2016)	0.9154 (Taslimi et al., 2016)	8.549 ng/mL	0.9891

Table 1 illustrates the effects of *Satureja cuneifolia* extracts on hCA I and II. According to the table, the direct methanol extract of *S. cuneifolia* has performed the best inhibitory characteristic among the evaluated extracts for hCA I isoenzyme with the IC_{50} value of 31 $\mu\text{g/mL}$. Direct methanol extract was followed by petroleum ether, methanol and chloroform extracts with the IC_{50} values 44 $\mu\text{g/mL}$, 68 $\mu\text{g/mL}$ and 161 $\mu\text{g/mL}$,

respectively. The methanol extract of *S. cuneifolia* possessed the highest inhibition profile with the IC_{50} value of 12 $\mu\text{g/mL}$ for hCA II among all tested extracts. Direct methanol extract also showed the potent inhibitory characteristic with the IC_{50} value of 13,6 $\mu\text{g/mL}$ against hCA II. Other extracts, petroleum ether and chloroform, showed the inhibition profiles against hCA II with the IC_{50} values of 101 $\mu\text{g/mL}$ and 132 $\mu\text{g/mL}$, respectively.

Conclusion

The carbonic anhydrase inhibitory activities of *S. cuneifolia* have been investigated in this study and to best of our knowledge, it has been the first investigation performed on hCA I and II inhibitory features of *S. cuneifolia* extracts. In this study, direct methanol extract of *S. cuneifolia* has showed the best inhibitory potential against hCA I in all evaluated extracts. The extracts have potential inhibitory features for hCA II. However, methanol extract has performed the best inhibitory characteristics for hCA II in all studied extracts. According to the results, methanol and direct methanol extracts can be used as natural products/inhibitors because of their biological activities. Further purification and analytical studies for both extracts will be needed to obtain the active natural molecules and their potential for pharma and food industries.

Acknowledgement

We would like to thank Dr. İsmail Şenkardeş to identify the plant material.

Conflict of Interest Statement: The authors declare that the study was performed in the absence of any commercial and/or financial relationships that could be perceived as a conflict of interest.

Contribution Rate Statement Summary: The authors declare that they have contributed equally to the article. The authors have verified that all data in the manuscript have not been published before and have given approval for the final version of manuscript.

References

- Abdel-Aziz, A.A-M., El-Azab, A.S., Ceruso, M., Supuran, C.T. 2014. Carbonic anhydrase inhibitory activity of sulfonamides and carboxylic acids incorporating cyclic imide scaffolds, *Bioorganic and Medicinal Chemistry Letters*, 24: 5185-5189.
- Ağgöl, A.G., Kuzu, M., Kandemir, F.M., Küçükler, S., Çağlayan, C. 2020. Alterations in enzyme activity of carbonic anhydrase, 6-phosphogluconate dehydrogenase and thioredoxin reductase in rats exposed to doxorubicin and morin. *Clinical and Experimental Health Sciences*, 10(3): 228-234.
- Ahmad, S., Hassan, A., Rehman, T., Basit, A., Tahir, A., Adeel Arshad, M. 2019. In vitro bioactivity of extracts from seeds of *Cassia absus* l. growing in Pakistan. *Journal of Herbal Medicine*, 100258.
- Akkemik, E., Aybek, A., Felek, I. 2019. Effects of cefan melon (*Cucumis melo* l.) seed extracts on human erythrocyte carbonic anhydrase I-II enzymes, *Applied Ecology and Environmental Research*, 17(6): 14699-14713.
- Almajan, G.L., Barbuceanu, S-F., Innocenti, A., Scozzafava, A., Supuran, C.T. 2008. Carbonic anhydrase inhibitors. Inhibition of the cytosolic and tumor-associated carbonic anhydrase isozymes I, II and IX with some 1,3,4-oxadiazole- and 1,2,4-triazole-thiols, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 23: 101-107.
- Azmir, J., Zaidul, I.S.M., Rahman, M.M., Sharif, K.M., Mohamed, A., Sahena, F., Jahurul, M.H.A., Ghafoor, K., Norulaini, N.A.N., Omar, A.K.M. 2013. Techniques for extraction of bioactive compounds from plant materials: A review, *Journal of Food Engineering*, 117(4): 426-436.
- Eminagaoglu, O., Tepe, B., Yumrutas, O., Akpulat, H.A., Daferera, D., Polissiou, M., Sokmen, A. 2007. The in vitro antioxidative properties of the essential oils and methanol extracts of *Satureja spicigera* (K. Koch.) Boiss. and *Satureja cuneifolia* ten. *Food Chemistry*, 100(1): 339-343.
- Gupta, A., Naraniwal, M., Kothari, V. 2012. Modern extraction methods for preparation of bioactive plant extracts, *International Journal of Applied and Natural Sciences*, 1(1): 8-26.
- Güler, O.O., Supuran, C.T., Capasso, C. 2020. Carbonic anhydrase IX as a novel candidate in liquid biopsy, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 35(1): 255-260.
- Huie, C.W. 2002. A review of modern sample-preparation techniques for the extraction and analysis of medicinal plants, *Analytical and Bioanalytical Chemistry*, 373: 23-30.
- Kan, Y., Uçan, U. 2006. GC-MS analysis and antibacterial activity of cultivated *Satureja cuneifolia* Ten. essential oil. *Turkish Journal of Chemistry*, 30 (2): 253-259.
- Kaya, E., Erğün, B., Demir, Y., Alim, Z., Beydemir, Ş. 2019. The in vitro impacts of some plant extracts on carbonic anhydrase I, II and paraoxonase-1. *Hacettepe Journal of Biology and Chemistry*, 47(1): 51-59.
- Kuzu, M., Aslan, A., Ahmed, I., Comakli, V., Demirdag, R., Uzun, N. 2016. Purification of glucose-6-phosphate dehydrogenase and glutathione reductase enzymes from the gill tissue of Lake Van fish and analyzing the

- effects of some chalcone derivatives on enzyme activities, *Fish Physiology and Biochemistry*, 42(2): 482-491.
- Milos, M., Radonic, A., Bezic, N., Dunkic, V. 2001. Localities and seasonal variations in the chemical composition of essential oils of *Satureja montana* L. and *S. cuneifolia* Ten. *Flavour and Fragrance Journal*, 16(3): 157-160.
- Nar, M., Çetinkaya, Y., Gülçin, İ., Menzek, A. 2013. (3,4-Dihydroxyphenyl) (2,3,4-trihydroxyphenyl) methanone and its derivatives as carbonic anhydrase isoenzymes inhibitors, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 28: 402-406.
- Nocentini, A., Vullo, D., Bartolucci, G., Supuran, C.T. 2016. N-Nitrosulfonamides: A new chemotype for carbonic anhydrase inhibition, *Bioorganic and Medicinal Chemistry*, 24: 3612-3617.
- Oke, F., Aslim, B., Ozturk, S., Altundag, S. 2009. Essential oil composition, antimicrobial and antioxidant activities of *Satureja cuneifolia* Ten. *Food Chemistry*, 112(4): 874-879.
- Orhan, F., Şentürk, M., Supuran, C.T. 2016. Interaction of anions with a newly characterized alpha carbonic anhydrase from *Halomonas* sp, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 31: 1119-1123.
- Scozzafava, A., Mastrolorenzo, A., Supuran C.T. 2004. Modulation of carbonic anhydrase activity and its applications in therapy, *Expert Opinion on Therapeutic Patents*, 14: 667-702.
- Sorokina, M., Steinbeck, C. 2020. Review on natural products databases: where to find data in 2020, *Journal of Cheminformatics*, 13: 20.
- Supuran, C.T. 2008. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators, *Nature Reviews Drug Discovery*, 7: 168-181.
- Taslimi, P., Gülçin, İ., Öztaşkın, N., Çetinkaya, Y., Göksu, S., Alwasel, S.H., Supuran, C.T. 2016. The effects of some bromophenols on human carbonic anhydrase isoenzymes. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 31(4): 603-607.
- Taslimi, P., Köksal, E., Gören, A.C., Bursal, E., Aras, A., Kılıç, Ö., Alwasel, S., Gülçin, İ. 2020. Anti-Alzheimer, antidiabetic and antioxidant potential of *Satureja cuneifolia* and analysis of its phenolic contents by LC-MS/MS. *Arabian Journal of Chemistry*, 13(3): 4528-4537.
- Taşkın, T., Doğan, M., Cam, M.E., Şahin, T., Şenkardeş, İ. 2020. In vitro anti-urease, antioxidant, anticholinesterase, cytotoxic and in vivo anti-inflammatory potential of *Satureja cuneifolia* Ten., *Natulae Scientia Biologicae*, 12(2): 222-232.
- Türkoğlu, E.A., Kuzu, M., Ayasan, T., İnci, H., Eratak, S.V. 2019. Inhibitory effects of some flavonoids on thioredoxin reductase purified from chicken liver, *Brazilian Journal of Poultry Science*, 21(2): 001-008.
- Türkoğlu, E.A., Şentürk, M., Supuran, C.T., Ekinci, D. 2017. Carbonic anhydrase inhibitory properties of some uracil derivatives, *Journal of Enzyme Inhibition and Medicinal Chemistry*, 32: 74-77.
- Ucar, A., Findik, M., Kuzu, M., Pehlivanoglu, S., Sayin, U., Sayin, Z., Akgemci, E.G. 2021. Cytotoxic effects, microbiological analysis and inhibitory properties on carbonic anhydrase isozyme activities of 2-hydroxy-5-methoxyacetophenone thiosemicarbazone and its Cu (II), Co (II), Zn (II) and Mn (II) complexes. *Research on Chemical Intermediates*, 47(2): 533-550.
- Verpoorte, J.A., Mehta, S., Edsall, J.T. 1967. Esterase activities of human carbonic anhydrases B and C, *The Journal of Biological Chemistry*, 242: 4221-4229.
- Wang, L., Weller, C.L. 2006. Recent advances in extraction of nutraceuticals from plants, *Trends in Food Science & Technology*, 17(6): 300-312.