




Chemical composition and antioxidant activities of essential oils and extracts from cones of *Tetraclinis articulata* (Vahl) Masters

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Abstract: This study was carried out to evaluate the in vitro antioxidant activity and the chemical composition of essential oils and organic extracts of Moroccan *Tetraclinis articulata* cones (Khemisset region). The GC–MS analysis of essential oils identified a total 23 volatile components. The major constituents of the oil were α -pinene (18.33%), cis-verbenone (10.02%), and L-pinocarveol (8.32%). For phytosterols analysis of hexane extract, β -sitosterol constitutes the majority with a percentage of 77.74%. The amount of total phenolic and flavonoid contents was high in the methanol extract (78.54 \pm 2.8 mg GAE / g and 41.11 \pm 4.5 mg QE/g, respectively) and the Antioxidant capacity determined by DPPH method showed a strongest radical scavenging activity exhibition by the methanol extract (IC₅₀=0.038 \pm 0.006 mg/mL). The results indicated that *T. articulata* contains bioactive compounds which are responsible for its antioxidant activity. Therefore, this plant could be potential candidates for the preparation of a natural antioxidant drug or an additive preparation.

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1. INTRODUCTION

Reactive oxygen species (ROS) include species, such as the hydroxyl radical (\cdot OH), hydrogen peroxide (H₂O₂), and superoxide (O₂⁻) readily react with most biological macromolecules and leads to their oxidative modification and consequently resulting in the loss of their activities (Kapoor *et al.*, 2019). They have the ability, to damage all biomolecules, causing peroxidation of lipids, oxidation of proteins, and damage to nucleic acids, enzyme inhibition (Madkour, 2020) which causes chemical alterations of these molecules (Mitra, 2020) leading to many chronic diseases such as pulmonary diseases (Park *et al.*, 2006), cancer (Sosa *et al.*, 2013), renal

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diseases (Massy & Nguyen-Khoa, 2002), neurodegenerative disorders (Uttara *et al.*, 2009), metabolic and cardiovascular diseases (Incalza *et al.*, 2018).

Synthetic phenolic antioxidants (SPAs) such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), tertiary butyl hydroquinone (TBHQ), and propyl gallate (PG) are closely correlated with human life due to their extensive usages, and increasing concerns have been raised on their biosafety (Liu & Mabury, 2020). Previous studies have found that some synthetic antioxidants and related transformation products showed toxicity effects including hepatic toxicity, endocrine disrupting effects, and even carcinogenicity (Yang *et al.*, 2018; Dassarma *et al.*, 2018). Recently there has been a considerable interest in finding natural antioxidants from natural resources such as medicinal plants to replace synthetic ones and to find more information concerning the antioxidant potential of plant (Jamshidi-kia *et al.*, 2020).

In searching for novel natural antioxidants, The Thuya of Berber, *Tetraclinis articulata* (Vahl) Masters, known by its name "*El ârâr*" in Arabic, belongs to Cupressaceae family, is an endemic species to North Africa and in particular to the Maghreb countries (Morocco, Algeria and Tunisia). It is also found in some very specific areas, in the south-east of Spain (region of Almeria) and on the island of Malta (Kouider & Assia, 2017). In addition, this forest species has an important sociological and economical role (Tsouli Faroukh *et al.*, 2017). It is also widely used in traditional medicine because of its multiple therapeutic effects. Indeed, different parts of Thuya are recommended in the treatment of intestinal infections, gastric pain, respiratory diseases, diabetes, severe diarrhea, hypertension, and fever (Teixidor-Toneu *et al.*, 2017; Hind *et al.*, 2017). These uses reveal that extracts and essential oils of *T. articulata* contain compounds which exert numerous biological activities such as antibacterial, antioxidant, immunostimulatory, antifungal, anti-inflammatory, cytotoxic properties, anticholinesterase, anti-tyrosinase, antidiarrheal, and anti-ulcerative activities (Sadiki *et al.*, 2019; Rabib *et al.*, 2020; El-Shemy, 2020; Rached *et al.*, 2018; Fidah *et al.*, 2016; Jlizi *et al.*, 2018; Calderón-Montaña *et al.*, 2018; Daoudi *et al.*, 2013; Ababsa *et al.*, 2019; Saber *et al.*, 2021). The objective of this work is the evaluation of antioxidant activity and chemical characteristics of the essential oil and the extracts of the cones of *Tetraclinis articulata* from Khemisset.

2. MATERIAL and METHODS

2.1. Plant collection and preparation

The cones were harvested in March 2017 from Khemisset region and dried in the open air, at room temperature and protected from humidity. All measurements are performed in triplicate.

2.1.1. Preparation of essential oil (EO)

The extraction of essential oils was carried out by hydrodistillation in a Clevenger type apparatus (Clevenger, 1928). Briefly, 100g of the dry plant is placed in a balloon filled to 2/3 with water; the whole is brought to the boil for 6 hours. The oil is recovered and then stored at a temperature of 4 °C for the tests.

2.1.2. Preparation of extracts

Firstly, 40 g of crushed Thuya cones are taken then put it in a cartridge before extracting it using a solvent (hexane, methanol) using the Soxhlet as an extraction material (Jensen, 2007). The system is heated at reflux for 6 hours, until discoloration. Samples are prepared by two different solvents used a successive extraction: hexane (for delipidation) then with methanol. The extracts obtained are evaporated to dryness using a rotary evaporator equipped with a vacuum pump and then stored at 4 °C for the tests.

2.2. Phytochemical screening

Chemical assays for screening and identification of bioactive constituents in *T. articulata* cones were performed with extracts prepared by qualitative characterization reactions.

2.2.1. Flavonoid detection

Briefly, 1mL of extract with a few drops of concentrated HCl, and then adds a few milligrams of magnesium turnings. The presence of flavonoids is confirmed by the appearance of a pink to red orange color (N'Guessan *et al.*, 2009).

2.2.2. Alkaloid detection

100 mg of extract with 3 ml H₂SO₄ (1%), the whole is brought to a boil in a water bath at 100 ° C for 5 min. After cooling and filtration, 5 drops of Mayer's reagent are added. The formation of a white precipitate indicates the presence of alkaloids (Mojab *et al.*, 2010).

2.2.3. Tannin detection

The extract dissolved in distilled water and added drops of a solution of FeCl₃ (1%). The appearance of a blue-black color indicates the presence of gallic tannins and the appearance of a green-blackish color indicates the presence of catechetical tannins (Y *et al.*, 2004).

2.2.4. Saponosides detection

Mix 1ml of the extract with 2 mL of hot distilled water and stir for 15 seconds then let stand for 10 min. A height of persistent foam, greater than 1 cm indicates the presence of Saponosides (Bekro *et al.*, 2016).

2.2.5. Sterols and polyterpenes detection

The extract is diluted in 2 mL of acetic anhydride. The addition of a few drops of concentrated H₂SO₄ allows the appearance of a violet color which indicates the presence of sterols and polyterpenes (N'Guessan *et al.*, 2009).

2.3. Determination of the total phenolic content

The determination of the total polyphenols of the methanol extract is carried out by the Folin-Ciocalteu method (Singleton *et al.*, 1999). Briefly, 200 µL of extract or reference (Gallic acid) with 800 µl of the Na₂CO₃ solution (7.5%), After stirring (5 min), 1 mL of the Folin-Ciocalteu solution (diluted with distilled water 1:10) is added to the whole, after 2 hours of incubation at room temperature, the absorbance is read at 765 nm against a blank without extract. The results are expressed in micrograms of Gallic acid equivalent per milligram of extract (µg EAG / mg of extract).

2.4. Determination of flavonoids content

The determination of flavonoids in our experiments is carried out by the method (Quettier-Deleu *et al.*, 2000). In test tubes 1 mL of methanol extract or standard (quercetin) and 1 mL of methanol solution of aluminum chloride (2%). After 15 min of incubation at room temperature, the absorbances are read using a UV spectrophotometer visible at 430 nm against a blank (methanol added to AlCl₃). The results are expressed in µg Quercetin equivalent per milligram of extract (µg EQ / mg of extract).

2.5. Analysis of Essential oil

The gas chromatography/mass spectrometry (GC/MS) device is made by Perkin Elmer ClarusTM GC-680 with Q-8 MS. It is equipped with an auto-sampler, which gives access to the automatic injection of samples into the injector and a capillary column type RxiR-5Sil MS traversed by Helium gas. The mass spectrometer is powered by a SMART electronic ionization source. This source can ionize and vaporize the different molecules as well as a quadrupole filter to separate the different ions in their m/z ratio. The GC/MS system is computer-controlled with Turbomass (TM) software, which allows programming of analytical methods as well as qualitative and quantitative identification of detected species. The analysis parameters are as follows: analysis time: 2 hours, vector gas flow rate: 1 mL/min, ionization energy: 70 eV,

injector temperature: 260 °C, oven temperature: 40 °C for 2 minutes, then rise from 10 °C / min to 290 °C, and the injected volume is 0.5 µL.

2.5. Determination of the composition of sterols

Sterol composition was quantified according to the (ISO 6799, 1991), standard method using capillary gas chromatography (CGC) on an apolar column (Chroma pack) (30m× 0.32 mm, DI: 0.25 µm, phase: CPSIL8CB). The Varian CP-3800 chromatograph is equipped with a divider injector type 1079 (T: 300°C) and an FID (T: 300°C) and using helium as carrier gas (flow: 1.5 mL/min).

2.6. Evaluation of antioxidant activity (DPPH test)

The scanning activity of the DPPH radical is measured according to the protocol described by (Lopes-Lutz *et al.*, 2008). A methanol solution of 0.3 mM of DPPH is mixed with different concentrations of the samples from the cones of *T. articulata* (0.0025; 0.005; 0.01; 0.025; 0.05; 0.1; 0.25; 0.5; 1; 2 mg / mL). In a test tube, 2.5 mL of samples and controls solution (Ascorbic acid, Gallic acid and Quercetin) are added to 1 mL of the methanol solution of DPPH, after incubation for 30 min in the dark and at room temperature, the absorbances are measured at 517 nm against a blank which contains pure methanol. The negative control is composed of 1 mL of the methanol solution with DPPH and 2.5 mL of methanol.

3. RESULTS / FINDINGS

3.1. Yield

Table 1 summarizes the results of the study of the yield of the essential oil and extracts of *T. articulata* cones collected in the Khemisset region.

Table 1. Yield obtained from the different samples.

Samples	Yields (%)
Essential Oil	0.33 ± 0.05
Hexane extract	2.46 ± 0.20
Methanol extract	13.22 ± 1.01

The yield of the methanol extract is higher than that of the hexane extract by the order of 13.22 ± 1.01% and 2.46± 0.20%, respectively (Table 1). The yield of essential oils calculated after 6 hours of extraction from the dry plant material is 0.33±0.05%. These results are close to 0.31% found by (Buhagiar *et al.*, 2000), have studied the essential oils cones of a cultivated stand of *T. articulata* growing in Malta (Spain). Different yields are present in the literature, especially in Algeria. It was reported that the EO yield extracted from fresh cones of *T. articulata* from the Hammam Melouane region was 0.80% and that of Tipazaregion was 1.60% (Chikhoun *et al.*, 2013). Also, the EO yield collected in Ain-Defla region was of the order of 0.52% (Djouahri *et al.*, 2014). This yield being lower than that obtained by the sawdust of our other study (2.19%) (Saber *et al.*, 2021).

3.2. Phytochemical screening

The flavonoids and saponosides test indicate the presence with a strongly positive amount, then the alkaloid test reveals the presence in methanol extract. For the sterols and polyterpenes test shows the absence of these in the methanol extract. Also, the tannins test shows the absence of catechetal tannins and gallic tannins as can be seen in Table 2.

Table 2. Results of the phytochemical screening of the methanol extract of the cones studied.

Secondary metabolites	Flavonoids	Saponosides	Alkaloids	Sterols and polyterpenes	Catechetical tannins	Gallic tannins
Methanol extract	+++	+++	++	-	-	-

(+++): Strongly positive test

(++): Positive average test

(+): Low positive test

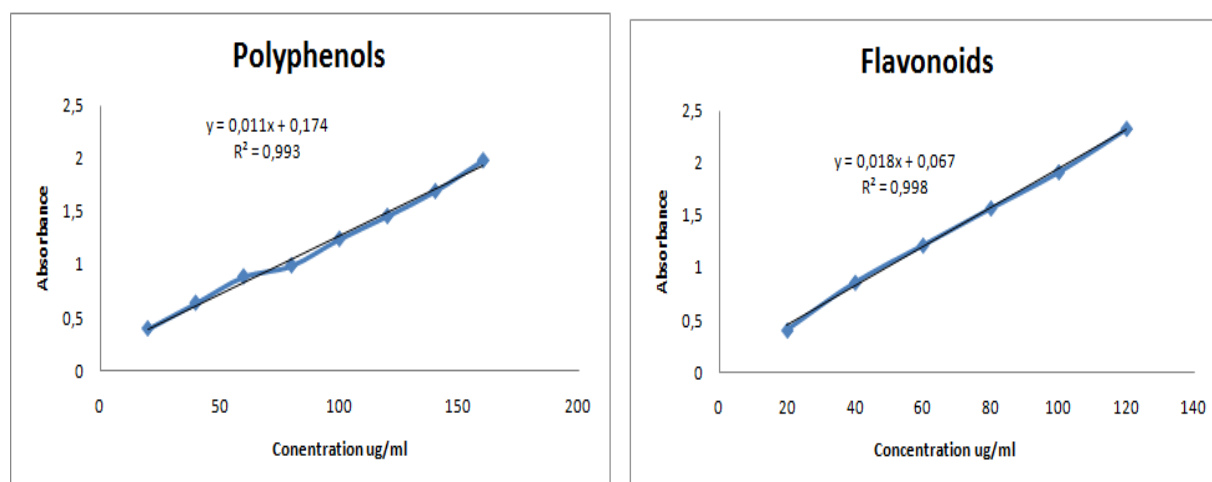
(-): Negative test

The presence of flavonoids and saponosides in our extract is probably responsible for the free radical scavenging effects observed. Indeed, flavonoids are phenolic compounds in plant medicinal which recognized by their antioxidant potentials (Ait Lahcen *et al.*, 2020). Also, saponosides have a wide range of pharmacological activities, including expectorant, anti-inflammatory, vasoprotective, gastroprotective and antimicrobial properties (Koczurkiewicz *et al.*, 2015). For Alkaloids they have shown to exert a broad spectrum of antimicrobial, anticancerogenic, and antimutagenic activity (Račková *et al.*, 2004).

3.3. Total phenolic and flavonoid content

The content of phenolic compounds in two extracts was determined from the calibration curve for gallic acid and the results are expressed in μg gallic acid equivalent per mg of extract (μg EAG/mg). On the other hand, the flavonoid content is expressed in μg quercetin equivalent per milligram of extract (μg EQ / mg of extract). The calibration curves are shown in Figure 1. Based on these results, the methanol extract showed high phenolic and flavonoid compounds ($78.54 \pm 2.8 \mu\text{g}$ GAE/mg and $41.11 \pm 4.5 \mu\text{g}$ QE/mg, respectively).

Figure 1. Gallic acid (phenolic) and quercetin (flavonoid) calibration curves.



2.4. Chemical compounds of Essential oils

The GC-MS analysis of cones TAE0 revealed the presence of 23 volatile compounds, the compounds identified are reported in Table 3. These compounds are belonging to different classes, including oxygenated monoterpenes (52.44%) and hydrocarbons monoterpene (27.78%). From these results, we note that this EO is mainly made up of α -pinene (18.33%), cis-Verbenone (10.02 %), L-pinocarveol (8.32%), bicyclo[4.1.0]hept-2-ene (6.76%), α -campholenal (6.10%), and D-limonene (5.75%). To our knowledge, this study is the first carried

out in Morocco, no study has been mentioned in the literature on the chemical composition of the Moroccan *T. articulata* cones.

Table 3. Chemical composition of the essential oil of *T. articulata* cones.

Compounds	RT	%
α -pinene	4.033	18.33
2,4-thujadiene	4.325	1.99
p-Xylene	5.045	0.66
o-cymene	5.347	1.71
D-limonene	5.410	5.75
Styrene, 2,5-dimethyl-	3.353	2.11
Linalool	6.474	0.85
α -campholenal	6.940	6.10
L-Pinocarveol	7.168	8.32
Cis-verbenol	7.232	2.96
3-Methylenecyclohexene	7.301	3.18
D-Pinocamphone	7.502	2.15
Bicyclo[4.1.0]hept-2-ene	7.571	6.76
Terpinen-4-ol	7.745	1.26
Thymol	7.846	2.53
α -terpineol	7.941	1.60
Myrtenal	8.052	5.22
cis-Verbenone	8.259	10.02
trans-2-Caren-4-ol	8.360	5.01
cis-Carveol	8.540	0.94
D-Carvone	8.757	2.62
Bornylacetate	9.387	2.18
Terpinylacetate	10.292	0.68
Monoterpenehydrocarbons		27.78
Oxygenatedmonoterpenes		52.44
Sesquiterpenehydrocarbons		-
Oxygenatedsesquiterpenes		-
Others		12.71
Total		92.93

The comparative study carried out in the literature on *T. articulata* cones EOs reveal a homogeneity of major compounds. Indeed, Buhagiar and collaborators identified α -pinene (68.2%), limonene (16.6%) (Buhagiar *et al.*, 2000). Djouahri, who worked on *T. articulata* as an antimicrobial and anti-inflammatory plant, cited α -campholenal (16.34%); trans-pinocarveol (15.45%); verbenone (13.36%); cis-verbenol (12.36%) (Djouahri *et al.*, 2014). In another study (Chikhoun *et al.*, 2013), Chikhoun *et al.* reported a chemical composition dominated by α -pinene; β -myrcene; limonene for two EOs of the cones studied. These results are close to the results found by Boussaïd *et al.* (Boussaïd *et al.*, 2016) who examined the chemical composition of six EO of *T. articulate* cones collected in six different regions of Algeria with a quantitative difference.

3.5. Sterols compounds

The sterols analyzed by CPG detected eight phytosterols for the hexane extract as summarized in Table 4. According to the results of Table 4, β -sitosterol constitutes the majority sterol with a percentage of 77.74%, followed by 10.55% of campesterol; brassicasterol (4.06%); stigmasterol (2.56%); cholesterol (0.48%); Δ -5-Avenosterol (0.37%); Δ -7-Avenosterol (0.27%) and Δ -7-stigmasterol (0.09%).

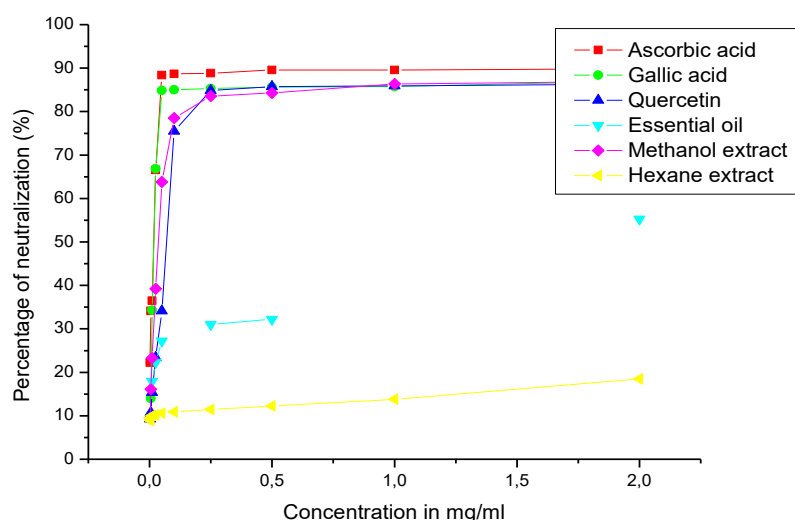
Table 4. Chemical composition in sterols of hexane extract of Thuya cones.

Phytosterols	Pourcentage (%)
Cholesterol	0.48
Brassicasterol	4.06
Compesterol	10.62
Stigmasterol	2.56
β -Sitosterol	77.21
Δ -5-Avenosterol	0.37
Δ -7-Stigmasterol	0.09
Δ -7-Avenosterol	0.27

Among these, the most abundant phytosterols is β -sitosterol, which has a broad range of *in vivo* biological functions including antitumor, anti-inflammatory, antidiabetic, and gallstone-reducing activities (Soleimanian *et al.*, 2020). To our knowledge, this study is the first carried out on the composition of phytosterols, no study has been mentioned in the literature on the composition of phytosterols in *T. articulata* cones.

3.5. Antioxidant activity of samples

From Figure 2, it is quite clear that the percentages of neutralization of DPPH radical increase with increasing concentration of samples and standards (ascorbic acid, gallic acid and quercetin). The antioxidant activity is expressed in IC_{50} , it defines the concentration of the extract or reference tested necessary to reduce 50% of the DPPH radical. In addition, the lower the IC_{50} values, the stronger the antioxidant power.

Figure 2. Evolution of the neutralization percentage (%) for samples and standards.

The concentrations which provide 50% inhibition (IC_{50}) are grouped together in Table 5, where it is found that the IC_{50} values calculated for the methanol extract and references confirmed the reactivity of these samples with respect to DPPH. The results obtained demonstrated that the methanol extract ($IC_{50} = 0.038 \pm 0.006$ mg/mL) has an antioxidant activity close to that of the standards and greater than the essential oils and hexane extract (absence) over the entire range of concentrations studied.

Table 5. Antioxidant activity expressed in IC₅₀.

	IC ₅₀ (mg/mL)
Ascorbic acid	0.018±0.001
Gallic acid	0.020±0.003
Quercetin	0.070±0.004
Essential oil	1.677±0.026
Methanol extract	0.038±0.006
Hexane extract	Absence

In comparison with the literature, the value of methanol extract obtained is higher than those obtained in the work carried out by Bensebia and collaborators (IC₅₀ = 0.14 mg/ml) were obtained from the *T. articulata* leaves extracted with 80 % ethanol (Bensebia, s. d.). In addition, our essential oil shows better activity than two samples of essential oils from cones in two different sites in Algeria which showed low activity (Chikhounne *et al.*, 2013).

4. DISCUSSION and CONCLUSION

The aromatic plants currently present a reliable source of active ingredients known for their therapeutic properties, in particular anti-oxidant activity. This work is interested in the study of the antioxidant activity of essential oil and organic extracts from the cones of *T. articulata*. The phytochemical tests carried out by the characterization reactions made it possible to highlight alkaloids, flavonoids and saponosides in the Thuya cones of the Khemisset region. The determination of phenolic compounds of methanol extract revealed considerable contents of polyphenols and flavonoids. Regarding antioxidant activity, we studied the antioxidant power through the capacity of DPPH radical scavenging, the methanol extract studied revealed a significant antioxidant potential. This study may find important application in the pharmaceutical and food industries.

Declaration of Conflicting Interests and Ethics

The authors declare no conflict of interest. This research study complies with research and publishing ethics. The scientific and legal responsibility for manuscripts published in IJSM belongs to the authors.

Authorship Contribution Statement

Mohammed Saber: Investigation, Resources, Visualization, Software, Formal Analysis, and Writing-original draft. **Hicham Harhar:** Methodology, Supervision, and Validation. **Mohamed Tabyaoui:** Methodology, Supervision, and Validation. **Latifa El Hattabi:** Visualization, Software and Formal Analysis. **Gokhan Zengin:** checked and corrected the final version. **Abdelhakim Bouyahya:** validation, checking and correction of the manuscript.

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