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# The Effect of Seed Priming Applications on Germination Parameters of Red Clover (*Trifolium* pratense L.)

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**ABSTRACT:** This study aimed to determine the effects of silicon (Si) and salicylic acid (SA) seed priming applications on the germination parameters of red clover (*Trifolium pratense* L.). The study was carried out in Siirt University, Faculty of Agriculture, Field Crops Laboratory, under controlled conditions at  $24\pm1$  °C. The plant material of the study was Rajan red clover (*T. pratense* L.) cultivar. The laboratory study was carried out in Petri dishes according to the randomized plots trial design with 4 replications. The subject of the study consists of hydropriming application with 2 mM, 4 mM, and 6 mM priming doses of Si and 0.5 mM, 1.0 mM, and 1.5 mM priming doses of SA. The study also included the non-priming application as a control subject. The germination percentage, mean germination time, germination index, coefficient of uniformity of germination, and germination energy properties were examined in terms of the effect on germination development in different priming applications applied to red clover. Significant differences were found between the priming applications in terms of all germination rate can be achieved with priming applications in plants with germination problems such as red clover. In this respect, 2 mM Si priming application can be recommended.

Keywords: Red clover, silicon, salicylic acid, seed priming, germination index, germination energy

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#### **INTRODUCTION**

Seed priming is accepted as a simple, cost-effective, and efficient approach to increase seed germination, early seedling growth and development under both stress and non-stress conditions (Hameed et al., 2013; Maiti and Pramanik, 2013). To this end, different chemicals, ions, organic compounds, hormones, and some antioxidants are used, and it was observed that positive results were obtained in germination and seedling growth. For example, seed priming with acetylsalicylic acid was reported to provide uniform germination and seedling formation in hot pepper (*Capsicum annuum* L.) under normal and salty conditions (Khan et al., 2009), sodium silicate seed priming was reported to improve tolerance to water deficiency stress of wheat seeds and improve germination and seedling growth of wheat seeds (Abro et al., 2009; Hameed et al., 2013), seed priming with gibberellic acid (GA3) was reported to reduce salinity stress and increase germination and seedling growth parameters in alfalfa (Younesi and Moradi, 2014), and seed priming applications with KNO<sub>3</sub> and KH<sub>2</sub>PO<sub>4</sub> positively affected germination and seedling growth parameters of *Citrullus colocynthis* seeds (Ghiyasi et al., 2019). It is also a common practice to use silicon (Si) and salicylic acid (SA) in seed priming.

Silicon is the second most abundant element in the Earth's crust (Zhu and Gong, 2014) and is the only nutrient element that is not harmful when excessively accumulated in plants (Ma et al., 2001). Although not essential for plant growth (Zhang et al., 2017), Si was reported to be effective in reducing stresses related to salinity (Liang et al., 2003, 2005; Parveen and Ashraf, 2010; Malhotra et al., 2016) and drought (Maghsoudi et al., 2019) in plants. Furthermore, it was stated that Si increased the activity of antioxidant enzymes, thereby reducing stress and improving plant growth (Al-Aghabary et al., 2004).

Salicylic acid is a plant hormone that regulates physiological processes in plant growth and development and plays a role in the plant's resistance to biotic stress (Farhangi-Abriz and Ghassemi-Golezani, 2016; Dempsey and Klessig, 2017). Exogenous administration of SA improves growth and physiological processes and increases protection against abiotic stress (He et al., 2002; Kaya et al., 2002; Arfan et al., 2007; Eraslan et al., 2007; Noreen et al., 2012). Additionally, it was reported that SA had a very important function in stomatal regulation (Morris et al., 2000; Barros et al., 2019), mechanism of plant water stress tolerance (Ashraf and Foolad, 2007), seed germination in plants (Shakirova et al., 2003; Simaeia et al., 2011; Kim and Lee, 2013), increasing the dry weight of root and upper parts (Khodary, 2004; Stevens et al., 2006). Furthermore, Chakma et al. (2021) emphasized that it could be used as a seed priming material.

Red clover (*Trifolium pratense* L.) is one of the widely grown forage legumes in temperate regions (Boller et al., 2010). Red clover is a valuable feed source for ruminants as green forage, straw, silage or dried feed and is a nutrient-rich, nutritious plant that provides forage on the farm throughout the year (Przybylska et al., 2021). It is known that certified seed production of red clover, which has high forage yield especially under irrigable conditions, is insufficient (Zuk-Gołaszewska et al., 2019). Varieties of forage legume species usually produce seeds with low seed yield or low viability (Amdahl et al., 2017). Furthermore, hard seededness (Desai et al., 1997), a special type of dormancy found only in legumes, is a common feature in the cultivars of *Trifolium* species (Tomic et al., 2020). Tomic et al. (2020) reported that the important indicators of seed quality in legumes are germination, hard seededness, and seedling growth parameters.

Seed germination and seedling growth are two critical stages for the establishment of crops (Hubbard et al., 2012). Rapid and uniform seed germination and emergence are key factors in better and optimum crop formation (Hameed et al., 2013). To this end, seed priming, which initiates the germination process with some biochemical changes such as the activation of enzymes and dormancy

breaking, is important (Ajouri et al., 2004; Hameed et al., 2013). On the other hand, the response of plants to certain priming agents may differ, whether under stress or non-stress conditions. The concentration of the said substances, processing time, plant species, and the plant organ used are effective in this difference. In this sense, there is little information available for a large number of crops, and therefore further research is needed.

In this study, the effects of Si and SA seed priming applications on the germination parameters of red clover (T. pratense L.) were investigated.

#### **MATERIALS AND METHODS**

The study was carried out in Siirt University, Faculty of Agriculture, Department of Field Crops Laboratory. Rajan red clover (T. pratense L.) cultivar was used as a plant material. Sodium metasilicate pentahydrate (Na<sub>2</sub>SiO<sub>3</sub>.5H<sub>2</sub>O) was used as the Si source, and distilled water was used for hydropriming.

#### **Experimental Design and Treatment Details**

The subject of the study consists of hydropriming application with 2 mM, 4 mM, and 6 mM priming doses of Si and 0.5 mM, 1.0 mM, and 1.5 mM priming doses of SA. The study also included the non-priming application as a control subject.

A laboratory trial was set up according to a randomized plot trial design with 4 replications. Twenty-five seeds were used for each replication. Seeds were sterilized in 70% ethyl alcohol for 1 minute and rinsed 3 times with sterile water. Then, surface sterilization was carried out to cover the seeds with 10% sodium hypochlorite (NaOCl) for 5 minutes to deform the microorganisms on the seed surface. The sterilized seeds were placed between Whatman (Little Chalfont, Buckinghamshire, UK) Grade 2 filter paper in Petri dishes (90 mm x 15 mm). Si, SA and hydropriming solutions prepared according to the doses were applied to each Petri dish as 3 ml, and the seeds were kept in hydropriming for 18 hours (Farooq et al., 2017), in Si for 12 hours (Othmani et al., 2020), and in SA for 12 hours (Sundstrom et al., 1987; Ceritoğlu and Erman, 2020). The seed/solution ratio for each Petri dish was adjusted as 2:1 g ml<sup>-1</sup> (Johnson et al., 2005). After priming, the seeds were washed with distilled water to clean the entire surface and were first roughly dried in blotting paper, then again placed between dry filter paper and dried to the initial moisture  $(3\% \pm)$  (Jatana et al., 2020). The dried red clover seeds were placed in new Petri dishes, and 3 ml of distilled water was added to each Petri dish. Petri dishes were left to germinate in an oven set at 25±1 °C (BINDER, GmbH, Germany).

Germination controls were made every 24 hours during the test, and the germination test was completed on the 10<sup>th</sup> day. While germination was detected in seeds, at least 2 mm rootlet emergence was accepted as a germination criterion (Scott et al., 1984; Soleymani and Shahrajabian, 2018).

#### **Germination Measurements and Observations**

The germination percentage (GP), mean germination time (MGT), germination index (GI), coefficient of uniformity of germination (CUG), and germination energy (GE) properties were examined in terms of the effect on germination development in different priming applications applied to red clover.

The GP parameter was determined according to Equation 1 used by Scott et al. (1984) by counting the seeds that germinated every 12 hours. (1)

GP = (NGS/TS)x100

In the equation, NGS is the number of normally germinated seeds, and TS is the total number of seeds used.

The MGT is generally used to determine the germination day of seeds and was calculated according to Equation 2 (Ellis and Roberts, 1981).

# $MGT = \sum (N_i T_i / N_i)$

Where  $N_i$  is the number of seeds germinated on day  $T_i$ , and  $T_i$  is the number of days counted from the beginning of germination.

The GI was calculated with the help of Equation 3 (Wang et al., 2004), CUG was calculated with the help of Equation 4 (Bewely and Black, 1994), and GE was calculated with the help of Equation 5 (Li et al., 2020).

## $GI = \sum (G_i/T_t)$

Where Gi is the germination percentage on the ith day, and Tt is the days of the germination test.  $CUG = \sum n / \sum [(MGT-t)^2 n]$ (4)

Where t is the time in days, starting from day 0, the day of sowing, and n is the number of seeds completing germination on day t.

# $GE = (T_1/N) \times 100$

Where  $T_1$  is the number of seeds germinated on the first day, and N is the total number of seeds.

# **Statistical Analysis**

The ArcSin transformation was applied to GP values before analysis of variance (Zar, 1996). The obtained data were subjected to analysis of variance according to the randomized plot trial design, and the differences between the means were checked by Tukey's multiple comparison test (Açıkgöz and Açıkgöz, 2001).

# **RESULTS AND DISCUSSION**

The GP results of the red clover plant in different priming applications are presented in Figure 1. The effect of different priming applications in terms of GP was found to be statistically significant at the p<0.01 level.



In the study, in terms of GP, the highest value was determined in 2 mM Si priming application with 40.0%, and the difference between all Si priming applications and hydropriming application was statistically insignificant. The lowest GP was determined in the control group (24.0%) without priming application and 0.5 mM SA priming application (25.3%). It was determined that Si priming showed more positive effects in terms of GP than SA priming (Figure 1).

When the effects of different priming applications were evaluated together in terms of GP, it was determined that all priming applications, except 0.5 mM SA priming, showed positive effects compared

40.0 a 36.0 ab 34.7 ab 36.0 ab 40.0 35.0 29.3 bc 28.0 bc 30.0 25.3 c 24.0 c 25.0 GP (%) 20.0 15.0 10.0 5.00.0 o. Hidrophning 2ml Si 0.5 mMSA 6mM Si AmMSi LODMASA 1.5 100 50 Priming Applications  $P = 0.0085^{**}$ 

(3)

(5)

(2)

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to the control, and especially Si priming applications promoted germination more. In studies conducted on many plant species, it was reported that hydropriming (Ghafoor et al., 2020; Açıkbaş and Özyazıcı, 2021; Ceritoglu and Erman, 2021), silicon priming (Janmohammadi et al., 2015; Khalaki et al., 2016; Biju et al., 2017; Bijanzadeh and Egan, 2018; Ivani et al., 2018; Ayed et al., 2021) and salicylic acid priming (Rajasekaran et al., 2002; Shakirova et al., 2003; Afzal et al., 2006; Farooq et al., 2008; Ceritoğlu and Erman, 2020; Ghafoor et al., 2020) had positive effects on the GP.

The findings of the MGT are shown in Figure 2. The effect of different priming applications in terms of MGT was found to be statistically significant at the p<0.05 level. When the MGT were examined, the latest germination was determined as 3.23 days in the control group. This was followed by hydropriming (2.60 days), 0.5 mM SA priming (2.50 days) and 1.5 mM SA priming applications (2.40 days), which were statistically in the same group. With the effect of priming applications, the earliest germination was detected in all Si priming applications (2, 4 and 6 mM) and 1.0 mM SA priming (Figure 2).



Figure 2. Effect of different priming applications on MGT in red clover

In general, it can be said that the MGT is shortened with priming applications. In this sense, it was observed that Si priming applications and 1.0 mM SA priming were more effective. As in this study, it was reported that hydropriming, Si and SA priming applications significantly affected the MGT and shortened the germination time in some other studies conducted on different plant species (Afzal et al., 2006; Bijanzadeh and Egan, 2018; Ivani et al., 2018; Ceritoğlu and Erman, 2020; Moghaddam et al., 2021).

Different priming applications affected the GI of red clover seeds in a statistically significant manner (p<0.01). The highest value in terms of the GI was determined in 2 mM Si priming with 5.67. The difference between this application, hydropriming and 4-6 mM Si priming applications was statistically insignificant. The lowest value was found in the control group with 2.03, in which no priming was applied (Figure 3).



Figure 3. Effect of different priming applications on the mean GI in red clover

Since the GI parameter is affected by the germination ratio and calculated according to the number of germinated seeds on the germination days, the effect of different priming applications was clearly observed. Thus, many studies reported that the GI values of priming applications with Si and SA increased in plant species such as wheat (Hameed et al., 2013; Meena et al., 2014; Ayed et al., 2021), tomato (Shi et al., 2014), lentil (Biju et al., 2017), faba bean (Anaya et al., 2018), and bitter vetch (Moghaddam et al., 2021).

The values related to the CUG determined in red clover in different priming applications are given in Figure 4. There were statistically significant differences at the p<0.05 level between different priming applications in terms of the CUG. In terms of the CUG, the most uniform germination was revealed in 2 mM Si priming. In the study, in terms of the CUG, the difference between 0.5 mM SA priming and the other priming applications, excluding the control group, was statistically insignificant (Figure 4).



Figure 4. Effect of different priming applications on the CUG in red clover

The CUG is a parameter that expresses fluctuations and stability in germination times between the first and last germinated seeds. Higher values in this feature represent the stability of seeds, while lower values indicate variability in germination under the same conditions (Souhail and Chaabane, 2009). In a

study on chickpea regarding germination uniformity, it was reported that hydropriming and SA priming applications (0.1 mM, 0.2 mM, 0.3 mM) increased the CUG compared to the control group (Ceritoğlu and Erman, 2020).

The results of the GE values in the study are shown in Figure 5. Differences between different priming applications in terms of GE were found to be statistically significant at the p<0.01 level. The highest values in terms of GE were observed in 2 mM and 4 mM Si priming applications with 12.0. It was determined that no seeds germinated on the first day in the control group in which no priming was applied, and it constituted the lowest group (Figure 5).

Germination energy is generally a germination parameter related to how many seeds germinate on the first day. In terms of GE, 2 mM and 4 mM Si priming applications and 1.0 mM SA priming were found to be effective in seed germination on the first day. It was indicated that the effects of different priming applications on GE were different, and in a study conducted with *Triticum aestivum* L. in terms of GE, priming with sodium silicate had positive effects compared to hydropriming and control (Hameed et al., 2013). In another study, it was reported that different priming applications, including SA, had different effects on the GE of the maize plant (Kumari et al., 2017).



Figure 5. Effect of different priming applications on GE in red clover

## CONCLUSION

As a result of the study, it was observed that the effects of hydropriming, Si and SA priming applications on the germination characteristics of red clover showed significant differences. In particular, it was concluded that silicon priming and hydropriming were more effective than salicylic acid priming in terms of germination percentage and germination index, while in the other examined parameters, all priming applications had a positive effect compared to the control group in which no priming was applied. It is thought that early germination, more uniform germination, and higher germination percentage can be achieved with priming applications in plants with germination problems such as red clover. In this respect, 2 mM Si priming can be recommended.

## **Conflict of Interest**

The article authors declare that there is no conflict of interest between them.

#### **Author's Contributions**

The authors declare that they have contributed equally to the article.

#### REFERENCES

- Abro SA, Qureshi R, Soomro MF, Mirbahar AA, Jakhar GS, 2009. Effects of Silicon Levels on Growth and Yield of Wheat in Silty Loam Soil. Pakistan Journal of Botany, 41: 1385-1390.
- Açıkbaş S, Özyazıcı MA, 2021. The Effects of Silicon Priming Application on Germination in Common Grasspea (*Lathyrus sativus* L.). 3rd International African Conference on Current Studies, 27-28 February 2021, Abomey-Calavi, Benin, s. 404-412. (In Turkish).
- Açıkgöz N, Açıkgöz N, 2001. Common Mistakes in The Statistical Analyzes of Agricultural Experiments I. Single Factorials. ANADOLU Journal of the Aegean Agricultural Research Institute, 11 (1): 135-147. (In Turkish).
- Afzal I, Basra SM, Farooq M, Nawaz A, 2006. Alleviation of Salinity Stress in Spring Wheat by Hormonal Priming With ABA, Salicylic Acid and Ascorbic Acid. International Journal Of Agriculture & Biology, 8 (1): 23-28.
- Ajouri A, Haben A, Becker M, 2004. Seed Priming Enhances Germination and Seedling Growth of Barley Under Conditions of P and Zn Deficiency. Journal of Plant Nutrition and Soil Science, 167: 630-636.
- Al-Aghabary K, Zhu Z, Shi Q, 2004. Influence of Silicon Supply on Chlorophyll Content, Chlorophyll Fluorescence, and Antioxidative Enzyme Activities in Tomato Plants Under Salt Stress. Journal of Plant Nutrition, 27 (12): 2101-2115.
- Amdahl H, Aamlid TS, Marum P, Ergon Å, Alsheikh M, Rognli OA, 2017. Seed Yield Components in Single Plants of Diverse Scandinavian Tetraploid Red Clover Populations (*Trifolium pratense* L.). Crop Science, 57 (1): 108-117.
- Anaya F, Fghire R, Wahbi S, Loutfi K, 2018. Influence of Salicylic Acid on Seed Germination of *Vicia Faba* L. Under Salt Stress. Journal of the Saudi Society of Agricultural Sciences, 17 (1): 1-8.
- Arfan M, Athar HR, Ashraf M, 2007. Does Exogenous Application of Salicylic Acid Through The Rooting Medium Modulate Growth and Photosynthetic Capacity in Two Differently Adapted Spring Wheat Cultivars Under Salt Stress? Journal of Plant Physiology, 6: 685-694.
- Ashraf M, Foolad MR, 2007. Roles of Glycine Betaine and Proline in Improving Plant Abiotic Stress Resistance. Environmental and Experimental Botany, 59: 206-216.
- Ayed S, Othmani A, Bouhaouel I, Rasâa N, Othmani S, Amara HS, 2021. Effect of Silicon (Si) Seed Priming on Germination and Effectiveness of Its Foliar Supplies on Durum Wheat (*Triticum turgidum* L. ssp. *durum*) Genotypes under Semi-Arid Environment. Silicon, 1-11.
- Barros TC, de Mello Prado R, Roque CG, Arf MV, Vilela RG, 2019. Silicon and Salicylic Acid in The Physiology and Yield of Cotton. Journal of Plant Nutrition, 42 (5): 458-465.
- Bewely J, Black M, 1994. Seeds: Physiology of Development and Germination. New York.
- Bijanzadeh E, Egan TP, 2018. Silicon Priming Benefits Germination, Ion Balance, and Root Structure in Salt-Stressed Durum Wheat (*Triticum durum* desf.), Journal of Plant Nutrition, 41 (20): 2560-2571.
- Biju S, Fuentes S, Gupta D, 2017. Silicon Improves Seed Germination and Alleviates Drought Stress in Lentil Crops by Regulating Osmolytes, Hydrolytic Enzymes and Antioxidant Defense System. Plant Physiology and Biochemistry, 119: 250-264.
- Boller B, Schubiger FX, Kölliker R, 2010. Red Clover. In: B. Boller, Ed., Handbook of Plant Breeding. Springer, Vol. 5. pp. 439-455, Dordrecht, Netherlands.
- Ceritoglu M, Erman M, 2021. Effect of Silicon Priming Treatments on Germination and Some Agronomic Traits in Lentil. 3rd International African Conference on Current Studies, 27-28 February, Abomey-Calavi, Benin, pp. 436-444.
- Ceritoğlu M, Erman M, 2020. Mitigation of Salinity Stress on Chickpea Germination by Salicylic Acid Priming. International Journal of Agriculture and Wildlife Science, 6 (3): 582-591.
- Chakma R, Biswas A, Saekong P, Ullah H, Datta A, 2021. Foliar Application and Seed Priming of Salicylic Acid Affect Growth, Fruit Yield, and Quality of Grape Tomato Under Drought Stress. Scientia Horticulturae, 280: 109904.

- Dempsey DA, Klessig DF, 2017. How Does the Multifaceted Plant Hormone Salicylic Acid Combat Disease in Plants and Are Similar Mechanisms Utilized in Humans? BMC Biology, 15: 23.
- Desai BB, Kotecha PM, Salunkhe DK, 1997. Seeds Handbook Biology, Production, Processing, and Storage. Marcel Dekker Inc., pp. 627, New York, USA.
- Ellis RH, Roberts EH, 1981. The Quantification of Ageing and Survival in Orthodox Seeds. Seed Science and Technology, 9: 373-409.
- Eraslan F, Inal A, Gunes A, Alpaslan M, 2007. Impact of Exogenous Salicylic Acid on The Growth, Antioxidant Activity and Physiology of Carrot Plants Subjected to Combined Salinity and Boron Toxicity. Scientia Horticulturae, 113 (2): 120-128.
- Farhangi-Abriz S, Ghassemi-Golezani K, 2016. Improving Amino Acid Composition of Soybean Under Salt Stress By Salicylic Acid and Jasmonic Acid. Journal of Applied Botany and Food Quality, 89: 243-248.
- Farooq M, Aziz T, Basra SMA, Cheema MA, Rehman H, 2008. Chilling Tolerance in Hybrid Maize Induced By Seed Priming With Salicylic Acid. Journal of Agronomy and Crop Science, 194 (2): 161-168.
- Farooq M, Hussain M, Nawaz A, Lee D, Alghamdi SS, Siddique KHM, 2017. Seed Priming Improves Chilling Tolerance in Chickpea By Modulating Germination Metabolism, Trehalose Accumulation and Carbon Assimilation. Plant Physiology and Biochemistry, 111: 274-283.
- Ghafoor MF, Ali Q, Malik A, 2020. Effects of Salicylic Acid Priming for Salt Stress Tolerance in Wheat. Biological and Clinical Sciences Research Journal, 1: e024-e024.
- Ghiyasi M, Amirnia R, Rahimi A, Özyazıcı G, 2019. The Effects of Different Methods to Break Seed Dormancy and Seed Priming on Germination and Seedling Growth of *Citrullus colocynthis*. EJONS International Journal on Mathematic, Engineering and Natural Sciences, 11: 21-32. (In Turkish).
- Hameed A, Sheikh MA, Jamil A, Basra SMA, 2013. Seed Priming with Sodium Silicate Enhances Seed Germination and Seedling Growth in Wheat (*Triticum aestivum* L.) Under Water Deficit Stress Induced By Polyethylene Glycol. Pakistan Journal of Life and Social Sciences, 11 (1): 19-24
- He YL, Liu YL, Chen Q, Bian AH, 2002. Thermo-Tolerance Related to Antioxidation Induced By Salicylic Acid and Heat Hardening in Tall Fescue Seedlings. Journal Plant Physiology Molecular Biology, 28: 89-95.
- Hubbard M, Germida J, Vujanovic V, 2012. Fungal Endophytes Improve Wheat Seed Germination Under Heat and Drought Stress. Botany, 90: 137-149.
- Ivani R, Sanaei Nejad SH, Ghahraman B, Astaraei AR, Feizi H, 2018. Role of Bulk and Nanosized SiO<sub>2</sub> to Overcome Salt Stress During Fenugreek Germination (*Trigonella foenum-graceum* L.). Plant Signaling & Behavior, 13 (7): e1044190.
- Janmohammadi M, Sabaghnia N, Ahadnezhad A, 2015. Impact of Silicon Dioxide Nanoparticles on Seedling Early Growth of Lentil (*Lens culinaris* Medik.) Genotypes With Various Origins. Agriculture and Forestry, 61 (3): 19-33.
- Jatana BS, Ram H, Gupta N, 2020. Application of Seed and Foliar Priming Strategies to Improve The Growth and Productivity of Late Sown Wheat (*Triticum aestivum* L.). Cereal Research Communications, 48: 383-390.
- Johnson SE, Lauren JG, Welch RM, Duxbury JM, 2005. A Comparison of The Effects of Micronutrient Seed Priming and Soil Fertilization on The Mineral Nutrition of Chickpea (*Cicer arietinum*), Lentil (Lens *culinaris*), Rice (*Oryza sativa*) and Wheat (*Triticum aestivum*) in Nepal. Experimental Agriculture, 4: 427-448.
- Kaya C, Kirnak H, Higgs D, Saltali K, 2002. Supplementary Calcium Enhances Plant Growth and Fruit Yield in Strawberry Cultivars Grown at High Salinity. Scientia Horticulturae, 93: 65-74.
- Khalaki MA, Ghorbani A, Moameri M, 2016. Effects of Silica and Silver Nanoparticles on Seed Germination Traits of *Thymus kotschyanus* in Laboratory Conditions. Journal of Rangeland Science, 6 (3): 221-231.
- Khan HA, Pervez MA, Ayub CM, Ziaf K, Bilal RM, Shahid MA, Akhtar N, 2009. Hormonal Priming Alleviates Salt Stress in Hot Pepper (*Capsicum annuum* L.). Soil and Environment, 28 (2): 130-135.
- Khodary SEA, 2004. Effect of Salicylic Acid on The Growth, Photosynthesis and Carbohydrate Metabolism in Salt-Stressed Maize Plants. International Journal of Agriculture & Biology, 6 (1): 5-8.

- Kim YH, Lee IJ, 2013. Influence of Plant Growth Regulator Application on Seed Germination of Dandelion (*Taraxacum officinale*). Weed & Turfgrass Science, 2 (2): 152-158.
- Kumari N, Rai PK, Bara BM, Singh I, Rai K, 2017. Effect of Halo Priming and Hormonal Priming on Seed Germination and Seedling Vigour in Maize (*Zea mays* L.) Seeds. Journal of Pharmacognosy and Phytochemistry, 6 (4): 27-30.
- Li W, Zhang H, Zeng Y, Xiang L, Lei Z, Huang Q, Li T, Shen F, Cheng Q, 2020. A Salt Tolerance Evaluation Method for Sunflower (*Helianthus annuus* L.) at the Seed Germination Stage. Scientific Reports, 10 (1): 1-9.
- Liang Y, Chen Q, Liu Q, Zha W, 2003. Exogenous Silicon (Si) Increases Antioxidant Enzyme Activity and Reduces Lipid Peroxidation in Roots of Salt-Stressed Barley (*Hordeum vulgare* L.). Journal Plant Physiology, 60: 1157-1164.
- Liang Y, Zhang W, Chenc Q, Ding R, 2005. Effects of Silicon on H<sup>+</sup>-ATPase and H<sup>+</sup>-PPase Activity, Fatty Acid Composition and Fluidity of Tonoplast Vesicles From Roots of Salt-Stressed Barley (*Hordeum vulgare* L.). Environmental and Experimental Botany, 53 (1): 29-37.
- Ma JF, Miyake Y, Takahashi E, 2001. Silicon as A Beneficial Element for Crop Plants. Studies in Plant Science, 8:17-39.
- Maghsoudi K, Emam Y, Ashraf M, Arvin MJ, 2019. Alleviation of Field Water Stress in Wheat Cultivars By Using Silicon and Salicylic Acid Applied Separately or in Combination. Crop and Pasture Science, 70 (1): 36-43.
- Maiti R, Pramanik K, 2013. Vegetable Seed Priming: A Low Cost, Simple and Powerful Techniques for Farmers' Livelihood. International Journal of Bio-resource and Stress Management, 4 (4): 475-481.
- Malhotra CCH, Kapoor R, Ganjewala D, 2016. Alleviation of Abiotic and Biotic Stresses in Plants by Silicon Supplementation. Scientia, 13 (2): 59-73.
- Meena VD, DotaniyaML, Coumar V, Rajendiran S, Kundu S, Rao AS, 2014. A Case for Silicon Fertilization to Improve Crop Yields in Tropical Soils. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences, 84 (3): 505-518.
- Moghaddam SS, Pourakbar L, Rahimi A, Jangjoo F, 2021. Mitigation of Salinity Effects by Salicylic Acid Priming on Germination and Physiological Characteristics of Bitter Vetch (*Vicia ervilia* L.). Yuzuncu Yıl University Journal of Agricultural Science, 31 (1): 98-110.
- Morris K, MacKerness SA, Page T, John CF, Murphy AM, Carr JP, Buchanan-Wollaston V, 2000. Salicylic Acid Has A Role in Regulating Gene Expression During Leaf Senescence. The Plant Journal, 23: 677-685.
- Noreen S, Ashraf M, Akram NA, 2012. Does Exogenous Application of Salicylic Acid Improve Growth and Some Key Physiological Attributes in Sunflower Plants Subjected to Salt Stress?. Journal of Applied Botany and Food Quality, 84 (2): 169-177.
- Othmani A, Ayed S, Bezzin O, Farooq M, Ayed-Slama O, Slim-Amara H, Younes MB, 2020. Effect of Silicon Supply Methods on Durum Wheat (*Triticum durum* Desf.) Response to Drought Stress. Silicon, 1-11.
- Parveen N, Ashraf M, 2010. Role of Silicon in Mitigating The Adverse Effects of Salt Stress on Growth and Photosynthetic Attributes of Two Maize (*Zea mays* L.) Cultivars Grown Hydroponically. Pakistan Journal of Botany, 42: 1675-1684.
- Przybylska A, Cwintal M, Pszczółkowski P, Sawicka B, 2021. Effect of Attractants and Micronutrient Biofortification on the Yield and Quality of Red Clover (*Trifolium pratense* L.) Seeds. Agronomy, 11 (1): 152.
- Rajasekaran LR, Stiles A, Caldwell CD, 2002. Stand Establishment in Processing Carrots: Effects of Various Temperature Regimes on Germination and The Role of Salicylates in Promoting Germination at Low Temperatures. Canadian Journal of Plant Science, 82 (2): 443-450.
- Scott SJ, Jones RA, Williams WA, 1984. Review of Data Analysis Methods for Seed Germination. Crop Science, 24: 1192-1199.

- Shakirova FM, Sakhabutdinova AR, Bezrukova MV, Fatkhutdinova RA, Fatkhutdinova DR, 2003. Changes in the Hormonal Status of Wheat Seedlings Induced By Salicylic Acid and Salinity. Plant Science, 164 (3): 317-322.
- Shi Y, Zhang Y, Yao H, Wu J, Sun H, Gong H, 2014. Silicon Improves Seed Germination and Alleviates Oxidative Stress of Bud Seedlings in Tomato Under Water Deficit Stress. Plant Physiology and Biochemistry, 78: 27-36.
- Simaeia M, Khavarinejada RA, Saadatmanda S, Bernardb F, Fahimia H, 2011. Interactive Effects of Salicylic Acid and Nitric Oxide on Soybean Plants Under NaCl Salinity. Russian Journal of Plant Physiology, 58 (5): 783-790.
- Soleymani A, Shahrajabian MH, 2018. Changes in Germination and Seedling Growth of Different Cultivars of Cumin to Drought Stress. Cercetări Agronomice în Moldova, 1 (173): 91-100.
- Souhail M, Chaabane R, 2009. Toxicity of The Salt and Pericarp Inhibition on The Germination of Some Atriplex Species. American-Eurasian Journal of Toxicological Sciences, 1 (2): 43-49.
- Stevens J, Senaratna T, Sivasithamparam K, 2006. Salicylic Acid Induces Salinity Tolerance in Tomato (*Lycopersicon esculentum* cv. Roma): associated Changes in Gas Exchange, Water Relations and Membrane Stabilization. Plant Growth Regulation, 49 (1): 77-83.
- Sundstrom FJ, Reader RB, Edwards RL, 1987. Effect of Seed Treatment and Planting Method on Tabasco Pepper. Journal of the American Society for Horticultural Science, 112 (4): 641-644.
- Tomić D, Stevović V, Đurović D, Stanisavljević R, Madić M, Petrović M, Lazarević D, Knežević J 2020. Seed Testing of Foliar-Fertilised Red Clover Crops After Various Periods of Storage. Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 48 (1): 284-293.
- Wang YR, Yu L, Nan ZB, Liu YL, 2004. Vigor Tests Used to Rank Seed Lot Quality and Predict Field Emergence in Four Forage Species. Crop Sciences, 44 (2): 535-541.
- Younesi O, Moradi A, 2014. Effect of Priming of Seeds of *Medicago sativa* 'bami' with Gibberellic Acid on Germination, Seedlings Growth and Antioxidant Enzymes Activity under Salinity Stress. Journal of Horticultural Research, 22 (2): 167-174.
- Zar JH, 1996. Biostatistical Analysis. 3rd ed. Prentice Hall, p. 662, New Jersey, USA.
- Zhang W, Xie Z, Wang L, Li M, Lang D, Zhang X, 2017. Silicon Alleviates Salt and Drought Stress of Glycyrrhiza Uralensis Seedling By Altering Antioxidant Metabolism and Osmotic Adjustment. Journal of Plant Research, 130: 611-624.
- Zhu Y, Gong H, 2014. Beneficial Effects of Silicon on Salt and Drought Tolerance in Plants. Agronomy for Sustainable Development, 34 (2): 455-472.
- Zuk-Gołaszewska K, Wanic M, Orzech K, 2019. The Role Of Catch Crops in The Field Plant Production-A Review. Journal of Elementology, 24 (2): 575-587.