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EFFECTS OF LIGHT AND SALINITY ON THE GERMINATION OF CLOSELY RELATED THREE SALSOLA TAXA

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ABSTRACT. Three closely related *Salsola* taxa (*Salsola boissieri* Botsch. subsp. serpentinicola (Freitag & Özhatay) Freitag & Uotila, Salsola boissieri Botsch. subsp. boissieri, Salsola turcica Yıldırımlı (halophytic ecotype), Salsola turcica Yıldırımlı (gypsicole ecotype)) from different edaphic conditions were studied according to changing light and salinity conditions. Seeds of target taxa were collected in 2017 and their weights were determined. The perianth segments were removed before the experimental trials and all the trials were conducted at 9°C/22°C which is the mean night and day temperatures of germination season. For the determination of the influence of light, one set of seeds for distilled water trial were kept at complete darkness. Different NaCl concentrations (distilled water, 100, 200, 300 mM NaCl) were used to evaluate the effects of salinity on germination. Viability of the seeds were determined by Triphenyl Tetrazolium Chloride (TTC) test which was applied to the seeds that did not germinate during the trials. As a result, it was found out that light stimulates germination of the taxa and Salsola seeds showed better germination ratio at light. The most tolerant taxa against salinity are the halophytic and gypsicole ecotypes of S. turcica, and the most susceptible one is S. boissieri subsp. serpentinicola. Both of the species show reduced germination ratios with increasing salinity. Salinity tolerance of S. boissieri subsp. serpentinicola and S. boissieri subsp. boissieri are very low, according to the Decreasing Germination Percentage (DGP) values. Although they show different germination response against increasing salinity, there is not any statistically meaningful difference between these three taxa according to germination percentages at different salinities, germination rates, last germination ratios and seed viabilities (F=1.818 p>0.05) (One Way ANOVA, SPSS 25).

Keyword and phrases. Amaranthaceae/Chenopodiaceae, germination, Gypsicole, halophyte, salinity, Salsola, serpentine

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İ.B. ÇINAR, G.N. TUĞ

1. INTRODUCTION

Soil salinity became an important problem as a result of global warming, lack of precipitation and also wrong irrigation policies, especially in arid and semi-arid areas increase [1] so halophytes became an important plant group whose biology and germination ecology attracts attention in last few decades [2-6]. Halophytes have some special adaptation mechanisms that make them germinate, survive and complete their life cycles at high salinities where the most of the plants can not survive [5, 7]. Although they have these adaptation they show better germination abilities at non-saline or less saline conditions [8-20], and increase in salinity cause decrease in their germination rate and ratio and/or retard germination [3, 12-19, 21-23].

The genus *Salsola* L. is represented by 18 species and 23 taxa in Turkey [24]. The members of the genus are generally adapted to saline or semi-saline areas of arid regions [25, 26]. Although *Salsola turcica* Yıldırımlı, *Salsola boissieri* Botsch. subsp. *boissieri* and *Salsola boissieri* Botsch. subsp. *boissieri* and *Salsola boissieri* Botsch. subsp. *serpentinicola* (Freitag & Özhatay) Freitag & Uotila are phylogenetically closely related, they are adapted to different edaphic conditions and became habitat specialists which make them more vulnerable against habitat loss and degradation [27].

S. turcica is endemic to Turkey and has two ecotypes spread over gypsaceous and saline steppes between altitudes 950 m and 1000 m [28]. The subspecies of *S. boissieri* are geographically separated from each other. *S. boissieri* subsp. *boissieri* prefers rocky and stony slopes at 900-2500 m altitudes around Kahramanmaraş and Sivas provinces [25]. *S. boissieri* subsp. *serpentinicola* is also endemic to Turkey and, prefers serpentine rocks and rock cracks between 1600 and 2000 m around Muğla and Burdur provinces [29]. If populations of a species is adapted to local conditions and have genetically preserved morphological and physiological differences, they are called as ecotypes [30]. *S. turcica* has a wide distribution area and populations from both halophytic and gypsicole areas were studied and because of this differentiation they may have different physiological adaptations. The germination trials of these ecotypes can provide some information about the differentiations of them.

Most of the *Salsola* taxa are salt tolerant during the germination phase of their life cycles, as supported by several studies on different *Salsola* species like: *S. kali* L. [21], *S. baryosma* (Schult.) Dandy [31], *S. villosa* Del. ex Roem. et Schult [32], *S. iberica* Sennen & Pau [3], *S. chandharyi* L. [33], *S. imbricata* Forssk. [34, 35],

S.affinis C.A.Mey. ex Schrenk [23], S. vermiculata L. [26], S. ferganica Drobow [36], S. grandis Freitag, Vural & N. Adıgüzel [37].

In this study, three closely related *Salsola* taxa adapted to different edaphic conditions were studied to understand the influence of their salinity tolerance on their distribution and also the germination characteritics of them under different NaCl concentrations. These taxa are *S. boissieri* subsp. *serpentinicola*, *S. boissieri* subsp. *boissieri*, *S. turcica* (halophytic ecotype), *S. turcica* (gypsicole ecotype).

2. MATERIALS AND METHODS

2.1 Collection of fruits and seeds

During vegetation period of 2017, populations were evaluated at their natural distribution areas and, fruits and seeds were collected according to the current population sizes of the taxa and at least 10 individuals were sampled. Until the start of experiments specimens were stored at 4°C. All the fruit and seed specimens were evaluated by BAB stereo binocular microscope and BAB image processing and analysis system (Bs200Pro). Mean weight of fruits and seeds were measured. The locations and collection numbers of specimens were given in Table 1.

Taxon	Collection number of specimen (IBÇınar)	Locality Burdur, Altınyayla, Dirmil pasture, serpentine soils, 1669 m		
S. boissieri subsp. serpentinicola (serpentinicole subspecies)	1139			
S. boissieri subsp. boissieri (glycophyte subspecies)	1141	Sivas, Yıldızeli, Yusufoğlan village, 1400 m		
<i>S. turcica</i> (halophytic ecotype)	1142	Konya, Cihanbeyli, Bolluk Lake, saline alkaline areas, 943 m		
<i>S. turcica</i> (gypsicole ecotype)	1143	Ankara, Beypazarı, about 14 km west of Beypazarı, on the right side of Beypazarı-Nallıhan highway, 948 m		

TABLE 1. Taxa with their subspecies and ecotype information, collection number of specimens and localities

2.2 Germination trials

Periant segments of seeds were removed before the trials and seeds were sterilized with 0,1% sodium hypochlorite. During the trials, the mean night and day temperatures of the germination period of the distribution areas were used, $9^{\circ}C / 22^{\circ}C$ with 12 h photoperiodism (light intensity 12000 lux ± %10). For the determination of the influence of salinity on germination distilled water and 100, 200, 300 mM NaCl solutions were used. At each trial, 25 seeds with 4 replicates were monitored for 10 days and the emergence of radicula was accepted as germination. Ungerminated seeds were taken into TTC test for detection of their viability under binocular microscope [38-39]. Stained red at TTC test is the indicator of viability which detects the cellular level respiratory activity [40]. Red stained seeds were accepted as unviable [15].

After the trials germination percentages, germination rates (Timson index) [41] and viability percentages were calculated.

Germination rate: $\Sigma G/t$ [41] Last germination: $(a / c) \times 100$ Seed viability: $[(a+d) / c] \times 100$ [15] ΣG : sum of the germination percentages of every 2 days t: total germinated seed number at the end of the trials (number of germinated seeds after recovery + number of germinated seeds at salinity) c: Total number of seeds d: seeds stained red after test- viable seeds

DGP: [(Germination percentage at distilled water – Germination percentage at salinity) / Germination percentage at distilled water] \times 100 [42].

2.3 Statistical Analysis

All the data were arcsin transformed and then evaluated by SPSS (IBM SPSS Statistics Version 25) with One Way ANOVA for the comparison of the influence of trials. T test was used for importance control (p < 0.05).

3. RESULTS AND DISCUSSION

Mean weight and sizes of all the taxa were given at Table 2 and the examples of fruits and seeds are provided in Figure 1 and 2.

Taxon Mean weight of a fruit (g)		Mean weight of a seed (g)	Mean diameter of a fruit (mm)	Mean diameter of a seed (mm)	
S. boissieri subsp. serpentinicola (serpentinicole subspecies)	0.615 ± 0.029	0.297 ± 0.008	5.53 ± 0.74	1.40 ± 0.18	
S. boissieri subsp. boissieri (glycophyte subspecies)	0.389 ± 0.019	0.225 ± 0.025	4.22 ± 0.64	1.23 ± 0.15	
S. turcica (halophytic ecotype)	0.862 ± 0.039	0.415 ± 0.016	4.34 ± 0.54	1.50 ± 0.15	
S. turcica (gypsicole ecotype)	0.583 ± 0.028	0.362 ± 0.035	4.63 ± 0.75	1.42 ± 0.17	

TABLE 2. Taxa, collection number of specimens and localities

Halophytic ecotype of S. turcica has the heaviest fruits and seeds according to the mean weights, and the lightest fruits and seeds belong to S. boissieri subsp. boissieri. S. boissieri subsp. serpentinicola has the largest mean fruit diameter and S. boissieri subsp. boissieri has the smallest. However, in general there is no big difference between the weights and diameters of fruits and seeds. Even though the values were close to each other; the largest seed diameter was measured at halophytic ecotype of S. turcica and the smallest at S. boissieri subsp. boissieri.

All the germination results, germination percentages, germination rates, last germination percentages, viability percentages after TTC test DGP ratios at changing salinities were given at Table 3 and Figure 2.

İ.B. ÇINAR, G.N. TUĞ
TABLE 3. Germination results of taxa

S. boissieri subsp. serpentinicola									
	Germination percentage	Germination rate	Last germination (%)	Viability (%)	DGP				
Darkness (Distilled water)	59	-	-	-	-				
Photoperiodism (Distilled water)	65	12.8	73	73	-				
100 mM NaCl	46	12.7	52	52	29.2				
200 mM NaCl	40	10.1	43	43	38.4				
300 mM NaCl	33	12.9	33	33	49.2				
S. boissieri subsp. boissieri									
	Germination percentage	Germination rate	Last germination (%)	Viability (%)	DGP				
Darkness (Distilled water)	76	-	-	-	-				
Photoperiodism (Distilled water)	100	24.9	100	100	-				
100 mM NaCl	59	17.3	60	60	41.0				
200 mM NaCl	57	16.2	65	65	43.0				
300 mM NaCl	55	13.8	59	59	45.0				
	S. turcio	ca (halophytic ed	cotype)						
	Germination percentage	Germination rate	Last germination (%)	Viability (%)	DGP				
Darkness (Distilled water)	69	-	-	-	-				
Photoperiodism (Distilled water)	77	22.9	80	80	-				
100 mM NaCl	74	22.5	82	82	3.8				
200 mM NaCl	70	21.6	84	84	9.1				
300 mM NaCl	45	12.5	55	55	41.5				
				22	41.5				
	S. turci	<i>ca</i> (gypsicole eco		55	41.3				
	S. turci Germination percentage			55 Viability (%)	DGP				
Darkness (Distilled water)	Germination	<i>ca</i> (gypsicole eco Germination	otype) Last germination	Viability					
Darkness (Distilled water) Photoperiodism (Distilled water)	Germination percentage	<i>ca</i> (gypsicole eco Germination	otype) Last germination	Viability (%)					
Darkness (Distilled water) Photoperiodism	Germination percentage 90	<i>ca</i> (gypsicole ecc Germination rate -	otype) Last germination (%) -	Viability (%)					
Darkness (Distilled water) Photoperiodism (Distilled water)	Germination percentage 90 96	ca (gypsicole eco Germination rate - 35.4	otype) Last germination (%) - 96	Viability (%) - 96	DGP -				



EFFECTS OF LIGHT AND SALINITY ON THE GERMINATION OF CLOSELY RELATED THREE SALSOLA TAXA

FIGURE 1. Fruits of analysed taxa (a. S. boissieri subsp. serpentinicola, b. S. boissieri subsp. boissieri, c. S. turcica (halophytic ecotype), d. S. turcica (gypsicole ecotype)) and seeds of taxa (A. S. boissieri subsp. serpentinicola, B. S. boissieri subsp. boissieri, C. S. turcica (halophytic ecotype), D. S. turcica (gypsicole ecotype))

All of the previous studies about halophyte germination showed that the seeds showed better germination ratios at distilled water [8-20]. Our results are consistent with the former results and we also found better germination ratios at distilled water (Figure 2, Table 3); and increasing NaCl concentrations inhibited the seed germination [18, 26, 37, 43-44].

Also, when the results of complete darkness were evaluated, it was found that light stimulates the germination and for this reason 12/12 h photoperiodism was used during the germination trials. The comparisons of the germination percentages at light and complete darkness and also changing NaCl conditions can be seen in Figure 2.



FIGURE 2. Germination percentages of each taxa at distilled water and at changing NaCl concentrations

The presence of halophyte taxa at their habitats is shaped by their salinity tolerance [45] and germination phase of their life cycles defines edaphic conditions that they are facing with throughout their life span [46] so salinity, light, photoperiodism etc. influence germination ability at time and space [6].

According to our results, not only the germination percentages and rates but also the seed viabilities of all taxa decreased with increasing salinity (Table 3). Likewise, there is a negative correlation between last germinations and increasing salinities.

Halophytes show species specific salinity tolerance at germination and seedling phases [42] but for our study this specificity is not so obvious which may be caused by very close relations of the studied taxa. Formulize DGP (decreasing germination percentage) to quantify the negative correlation between salinity tolerance and germination percentages, higher the DGP lower the salinity tolerance.

According to the statistical analysis, even though these closely related taxa prefer different edaphic conditions, there is not any statistically important difference between germination percentages at different salt concentrations, last germination percentages, germination rates and seed viabilities (F=1.818 p>0.05)(One Way ANOVA, SPSS 25).

Seeds did not germinate after salinity treatments were taken into viability test. At the end of TTC test colour change denotes viability as can be seen from Figure 3. According to the classification used by [47-51] seeds stained totally red are accepted as viable.



FIGURE 3. Un-viable (partially stained) and viable seeds (completely satined red) after $$\mathrm{TTC}$$

According to our results, we concluded that light stimulates germination better than complete darkness at both of the taxa. The highest salinity tolerance among these three taxa was observed at both halophytic and gypsicole ecotypes of *S. turcica* and according to salinity tolerance the most vulnerable one is the *S. boissieri* subsp. *serpentinicola*. Both of the examined taxa showed better germination at distilled water which is consistent with the general knowledge about salinity tolerance mechanism of halophytes [12, 52-53]. Even though these three taxa are closely

İ.B. ÇINAR, G.N. TUĞ

related, their salinity tolerances are different from each other, especially according to calculated DGP values. The studied taxa are phylogenetically close to each other [25, 28-29] but because of their habitat specifications they evolved some physiological adaptations at germination phase and ecotypes of *S. turcica* are more tolerant than the subspecies of *S. boissieri* are glycophytes.

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Conflict of interest The authors declare that they have no conflict of interest.

REFERENCES

- [1] Greenway, H., Munns, R., Mechanisms of salt tolerance in nonhalophytes, *Ann. Rev. Plant Physiol*, 31 (1980), 149-190.
- [2] Mariko, S., Kachi, N. Ishikawa, S., Furukawa, A., Germination ecology of coastal plants in relation to salt environment. *Ecological Research*, 7 (1992), 225-233.
- [3] Khan, M.A., Gul, B., Weber, D.J, Seed germination in the Great Basin halophyte Salsola iberica. Canadian Journal of Botany, 80 (2002), 650-655. https://doi.org/10.1139/b02-046.
- [4] Parsons, R.F., Incidence and ecology of very fast germination, *Seed Science Research*, 22 (2012), 161-167. https://doi.org/10.1017/S0960258512000037.
- [5] Estrelles, E., Biondi, E., Galiè, M., Mainardi, F., Hurtado, A., Soriano, P, Aridity level, rainfall pattern and soil features as key factors in germination strategies in salt-affected plant communities, *J Arid Environ*, 117 (2015), 1-9. https://doi.org/10.1016/j.jaridenv.2 015.02.005.
- [6] Rasheed A., Ahmed M.Z., Gul B., Khan M.A. & Hameed A., Comparative Seed Germination Ecology of Sabkha and Playa Halophytes of Pakistan. In: Gul B., Böer B., Khan M., Clüsener-Godt M., Hameed A. (eds) Sabkha Ecosystems. Tasks for Vegetation Science, Vol 49 (2019), 41-54. Springer, Cham.
- [7] Gul, B., Ansari, R., Flowers, T. J., Khan, M.A., Germination strategies of halophyte seeds under salinity. *Environmental and Experimental Botany*, 92 (2013), 4-18. https://doi.org/10.1016/j.envexpbot.2012.11.006.
- [8] Seneca, E.D., Germination response to temperature and salinity of four dune grasses from the outer banks of North Carolina, *Ecology*, 50 (1969), 45-53.
- [9] Onnis, A., Bellettato, R, Dormienza e alotolleranza in due specie spontanee di *Hordeum* (*H. murinum* L. e *H. marinum* Huds), *G. Bot. Ital*, 106 (1972), 101-113. https://doi.org/10.1080/11263507209426542.

- [10] Breen, C.M., Everson, C., Rogers, K, Ecological studies on Sprorobolus virginicus (L.) Kunth with particular reference to salinity and inundation, Hydrobiologia, 54 (1977), 135-140. https://doi.org/10.1007/BF00034987.
- [11] Dietert, M.F., Shontz, J.P., Germination ecology of a Maryland population of saltmarsh bulrush (Scirpus robustus), Estuaries, 1 (1978), 164-170. https://doi.org/10.2307/13514 58.
- [12] Ungar, I. A., Halophyte seed germination, The Bot. Rev, 44 (1978), 233-264. https://doi. org/10.1007/BF02919080.
- [13] Huiskes, A.H.L., Stienstra, A.W., Koutataal, B.P., Markusse, M.M., Van Soelen, J., Germination ecology of Salicornia dolichostachya and S. brachystacya, Acta Bot. Neerl, 34 (4) (1985), 369-380. https://doi.org/10.1111/j.1438-8677.1985.tb01943.x.
- [14] Khan, M. A., Ungar, I.A., Effects of thermoperiod on recovery of seed germination of halophytes from saline conditions, Am. J. Bot, 84 (2) (1997), 279-283. https://doi.org/10 .2307/2446089.
- [15] Baskin, C.C., Baskin, J.M., Seeds: Ecology, Biogeography and Evolution of Dormancy and Germination, (1998), Academic Press. San Diego.
- [16] Khan, M.A., Gul, B., High salt tolerance in germinating dimorphic seeds of Arthrocnemum indicum, International Journal of Plant Sciences, 159 (1998), 826-832. https://doi.org/10.1086/297603.
- [17] Khan, M.A., Gul, B., Weber, D.J., Temperature and high salinity effects in germinating dimorphic seeds of Atriplex rosea, Western North American Naturalist, 164 (2) (2004), 193-20.
- [18] Sekmen, A.H., Ozdemir, F., Türkan, İ., Effects of salinity, light and temperature on seed germination in an Turkish endangered halophyte, Kalidiopsis wagenitzii (Chenopodiaceae), Israel Journal of Plant Sciences, 52 (2004), 21-30.
- [19] Wang, L., Huang, Z., Baskin, C.C., Baskin, J.M., Dong, M., Germination of dimorphic seeds of desert annual halophyte Suaeda aralocaspica (Chenopodiaceae), a C4 plant without kranz anatomy, Ann. Bot, 102 (2008), 757-769. https://doi.org/10.1093/aob/mc n158.
- [20] Terzi, H., Yıldız, M., Altuğ, Ü, Halofit Salsola crassa tohum çimlenmesi üzerine tuzluluk, sıcaklık ve ışığın etkileri, AKÜFEMÜBİD, 17 (2017), 1-9. Doi: 10.5578/fmbd.52763.
- [21] Woodell, S.R.J., Salinity and seed germination patterns in coastal plants, Vegetatio, 61 (1985), 223-229. https://doi.org/10.1007/978-94-009-5524-0_24.
- [22] Ungar, I.A., Seed germination and seed bank ecology in halophytes. In seed development and seed germination, Edited by J. Kigel and G. Galili, Marcel Dekker, (1995), 599-628, New York.
- [23] Wei, Y., Dong, M., Huang, Z.Y., Tan, D.Y., Factors influencing seed germination of Salsola affinis (Chenopodiaceae) a dominant annual halophyte inhabiting the deserts of Xinjiang, China, Flora, 203 (2008), 134-140. https://doi.org/10.1016/j.flora.2007.02. 003.
- [24] Yaprak, A.E., Chenopodiaceae. In: Güner, A., Aslan, S., Ekim, T., Vural, M. ve Babaç, M.T. (edr.) Türkiye Bitkileri Listesi (Damarlı Bitkiler) Nezahat Gökyiğit Botanik Bahçesi ve Flora Araştırmaları Derneği Yayını, (2012), İstanbul.

İ.B. ÇINAR, G.N. TUĞ

- [25] Aellen, P., Salsola In: P.H. Davis [ed.], "Flora of Turkey and the East Aegean Islands", Vol. 2 (1967), 328-335. Edingburg University Press, Edinburgh.
- [26] Guma, I. R., Padrón-Mederos, M. A., Santos-Guerra, A., Reyes-Betancort, J.A., Effect of temperature and salinity on germination of *Salsola vermiculata* L. (Chenopodiaceae) from Canary Islands, *Journal of Arid Environments*, 74 (2010), 708-711. https://doi.org/ 10.1016/j.jaridenv.2009.10.001.
- [27] Pueyo, Y., Alados, C.L., Barrantes, O., Komac, B., Rietkerk, M., Differences in gypsum plant communities associated with habitat fragmentation and livestock grazing, *Ecological Applications*, 18 (4) (2008), 954-964. https://doi.org/10.1890/07-1770.1.
- [28] Yıldırımlı, Ş., Some new taxa, records and taxonomic treatments from Turkey, *Ot Sistematik Botanik Dergisi*, 17 (2) (2010), 64-68.
- [29] Güner, A., Ozhatay, N., Ekim, T., Baser, H.C., Flora of Turkey and East Aegean Islands, Vol. 11, (Supplement 2) (2000), Edinburgh, Edinburgh University Press.
- [30] Clausen, J., Keck, D. D. & Hiesey, William, M., Experimental studies on the nature of species. I. Publ. Carneg. Instn, (1940), no. 520.
- [31] Mohammad, S., Sen, D. N., Germination behavior of some halophytes in Indian desert, *Int. J. Exp. Biol*, 28 (1990), 545-549.
- [32] Assaeed, A., Effect of temperature and water potential on germination of *Salsola villosa* Del. ex. Roem. Et Schult. Assiut *Journal of Agricultural Science*, 32 (2001), 173-183.
- [33] Al-Khateeb, S.A., Differential response of Seidltizia rosmarnus L. and Salsola chandharyi L. to NaCl induced salinity under eastern Saudi Arabian conditions, Pakistan Journal of Agricultural Research, 17 (2002), 81-87.
- [34] El-Keblawy, A., Al-Ansari, F., Hassan, N., Al-Shamsi, N., Salinity, temperature and light affect germination of *Salsola imbricate*, *Seed Science and Technology*, 35 (2007), 272-281. https://doi.org/10.15258/sst.2007.35.2.03.
- [35] Mehrun-Nisa, Khan, M.A., Weber, D.J., Dormancy, germination and viability of Salsola imbricata seeds in relation to light, temperature and salinity, Seed Science and Technology, 35 (2007), 595-606. https://doi.org/10.15258/sst.2007.35.3.07.
- [36] Wang Y., Jiang G. Q., Han Y. N., Liu M. M., Effects of salt, alkali and salt-alkali mixed stresses on seed germination of the halophyte *Salsola ferganica* (Chenopodiaceae), *Acta Ecologica Sinica*, 33 (2013), 354-360. https://doi.org/10.1016/j.chnaes.2013.09.010.
- [37] Cinar, I. B., Ayyıldız G., Yaprak A. E., Tug G. N., Effect of salinity and light on germination of Salsola grandis Freitag, Vural & N. Adıgüzel (Chenopodiaceae). *Commun. Fac. Sci.Univ. Ank. Series C.*, 25 (2016), 25-32. https://doi.org/10.1501/Com muc_0000000184.
- [38] Grabe, D. F., Tetrazolium testing handbook for agricultural seeds. Association of Official Seed Analysts, Lan- sing, 62 (1970). Michigan, USA.
- [39] Williams, S., Reduced genetic diversity in eelgrass transplantations affects both population growth and individual fitness, *Ecol Appl*, 11 (2001), 1472-1488. https://doi. org/10.1890/1051-0761(2001)011[1472:RGDIET]2.0.CO;2.
- [40] França-Neto, J., Krzyzanowski, F., Tetrazolium: an important test for physiological seed quality evaluation, *Journal of Seed Science*, 41 (2019), 359-366. http://dx.doi.org/10.15 90/2317-1545v41n3223104.

- [41] Khan, M. A., Ungar, I.A., The effect of salinity and temperature on germination of polymorphic seeds and growth of Atriplex triangularis Wild, American Journal of Botany, 71 (1984), 481-489.
- [42] Zhang H., Zhang G., Lü X., Zhou D., Han, X., Salt tolerance during seed germination and early seedling stages of 12 halophytes, *Plant Soil*, 388 (2015), 229-241. https://doi. org/10.1007/s11104-014-2322-3.
- [43] Yücel E., Duman A., Türe C., Böcük H., Özaydın B., Effects of different salt (NaCl), nitrate (KNO3) and acid (HCl and H₂SO₄) concentrations on the germination of some Hesperis species seeds, Biological Diversity and Conservation, 1/2 (2008), 91-104.
- [44] Orlovsky, N. S., Japakova, U. N., Shulgina, I., Volis, S., Comparative study of seed germination and growth of Kochia prostrata and Kochia scoparia (Chenopodiaceae) under salinity, Journal of Arid Environments, 75 (2011), 532-537. https://doi.org/10.101 6/j.jaridenv.2011.01.014.
- [45] Soriano, P., Moruno, F., Boscaiu, M., Vicente, O., Hurdato, A., Llinares, J.V., Estrelles, E., Is salinity the main ecologic factor that shapes the distribution of two endemic Mediterranean plant species of the genus Gypsophila?, Plant Soil, 384 (2014), 363-379. https://doi.org/10.1007/s11104-014-2218-2.
- [46] Ungar I.A., Germination ecology of halophytes. In: Sen D.N., Rajpurohit K.S. (eds) Contributions to the ecology of halophytes. Tasks for vegetation science, Vol 2, (1982), 143-154, Springer, Dordrecht.
- [47] Moore, R. P., Interpretation of color differences in tetrazolium testing, Seed Technologist News, 4 (3) (1972), 22-24.
- [48] Delouche, J. C., Standardization of vigor tests, Journal of Seed Technology, 1 (2) (1976), 75-85.
- [49] Grabe, D. F., Manual do teste de tetrazólio, (1976), Brasília: AGIPLAN.
- [50] De Guzman, L. E. P., Zamora O.B., Borromeo, T. H., Sta. Cruz, P. C., Mendoza, T., Seed Viability and Vigor Testing of Jatropha curcas L. Philippine Journal of Crop Science, 36 (2011), 10-18.
- [51] Melo, L. D. F. A., Junior, J. L.A.M., Ferreira, V. M., Neto, J.C.A., Viability Mimosa bimucronata (DC.) O. Kuntze. seeds by the tetrazolium test and oil content, Diversitas Journal, 5 (2020), 1575-1587. https://doi.org/10.17648/diversitas-journal-v5i3-846.
- [52] Flowers, T.J., Hajibagheri, M.A., Clipson, N.J.W., Halophytes, Q Rev Biol, 61 (1986), 313-335.
- [53] Flowers, T.J., Colmer, T.D., Salinity tolerance in halophytes, New Phytol, 179 (2008), 945-963. https://doi.org/10.1111/j.1469-8137.2008.02531.x.