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Proceeding Article

Cynarin, Chlorogenic and Caffeic Acid Flavonoids, Cyanidin, Peonidin Anthocyanidins in Head, Heart, Bractes of Artichokes as Antioxidative Quality Indicators: Alterations By Boiling, Steaming and Frying

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#### Abstract

In edible parts- bracte leaves, heart, head of artichoke varieties (Turkish *var*.Sakız, *var*.Bayrampaşa), proximate composition, antioxidative major phenolic acids [cyanarin-(1,3-dicaffeoylquinic acid), chlorogenic acid-(5-O-caffeoylquinic acid), caffeic acid], anthocyanidins (cyanidin, peonidin), total phenolic acids-TPA, total flavonoids-TFlav, total phenolics-TP, total antocyanins were detemined (p<0.01), the alterations on above-mentioned bioactive constituents through heat treatment effect (boiling, steaming, frying) were put forwarded. Cyanarin-(1,3-di-O- caffeoylquinic acid) was determined the major phenolic compound of head part of artichokes. It was determined about 6.2; 5.6; 3.48 fold increasing in TP content with boiling, steaming, frying, respectively respect to raw-form and total anthocyanins-TA rised as 1.93 fold with boiling whereas decreased as 1.04, 3.09 fold after steaming and frying, respectively. The important alterations were established in phenolic acids -PA, total phenolics-TP, cyanarin, chlorogenic acids in boiling (B) > steaming (S) > frying (F) towards whereas 1.36; 1.28; 2.59 fold decreasing in TFlav with B,S,F processing, respectively (p<0.01).

Keywords: Artichoke, Phenolic acid, Anthocyanin, Antioxidant, Heat treatment



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Artichoke (Cynara cardunculus L. scolymus) is an herbaceaus perennial plant belonging to Composite family (Asteraceae) and widely cultivated in the Mediterranean area and adjoining parts of Europe, which accounts for 85% of the world's production. Major producers of the globe artichoke are France, Italy, Spain, Egypt, Israel, Algeria, Morocco, and Turkey in Europe whereas California Castroville-Monterey County in USA (Tokusoğlu, 2018& Tokusoğlu and Basay, 2009).

Artichokes are consumed as fresh, traditional meal, and canned and are also traditionally used as a medicinal plant. The artichoke heads are edible and used worldwide. The leaves have beneficial effects against liver complaints and have strong antioxidant effects. Their leaves are brewed and consumed as teas and leaves are processed into pharmaceutical preparations such as capsules, tablets and juices. Artichokes contain bioactive compounds including phenolics which protect liver and have strong positive effects on several diseases and disorders. Also, inulin form carbohydrate in artichokes, stabilize blood sugar levels in diabetes (Tokuşoğlu & Başay,2009; Fratianni et al., 2007: Costabile et.al.2010; López-Molina et.al., 2005).

Majorly, such extracts from head and leaves of artichokes have been utilized for their hepatoprotective effects (Speroni et. al., 2003; Gebhardt and Fausel, 1997), their benefits on the liver and their protecting against toxins and infection are important (Adzet et.al., 1987). Artichoke head and leaves have antioxidative (Miccadei et.al,2008; Gebhardt and Fausel, 1997), anticarcinogenic (Agarwal andMukhtar, 1996) and hypocholesterolemic activity (Rondanelli et al.,2012). The artichoke has strong choleretic activity (promotes bile secretion in the liver), and choleretics increase the excretion of

cholesterol and decrease the manufacture of cholesterol in the liver (Bundy et.al.,2008). It is shown that artichoke leaf consuming improved the dyspeptic symptoms who suffer dyspepsia (digestive problems) and artichoke leaf extract has potential value in relieving irritable bowel syndrome symptoms (Bundy et.al.,2004).

These strong effects are attributed to the high polyphenolic content of artichokes including phenolic acids. majorly hydroxycinnamic acids, flavones and anthocyanins (Figure 1.) Artichoke have high proportion of phenolics (Fratianni et al., 2007; Llorach et al., 2002). The include cynarin (1,3-di-Ophenolics caffeoylquinic acid), luteolin, cynaroside (luteolin-7-O-glucoside), scolvmoside (luteolin-7-O-rutinoside); phenolic acids such caffeic. coumaric. as hydroxycinnamic, ferulic, caffeoylquinic acid derivatives: monoand dicaffeoylquinic acids. including chlorogenic; acid alcohols; flavonoid glycosides, among others (Tokuşoğlu & Basay,2009,2008; Fratianni et al.,2007; Sa'nchez Rabaneda et al., 2003).

Especially, the pleasant bitter taste of the artichoke is due mostly to a plant chemical cynarin called (1.5 dicaffeoylquinic acid), which is found in highest concentration in the leaves of the plant. It is known that extracts including cynarin have positive effects on liver health, hepatobiliary diseases, hyperlipidaemia and cholesterol metabolism (Tokuşoğlu & Başay,2009, Fratianni et al.,2007). Figure 2 shows two major compounds in globe chlorogenic artichoke are acid (5dicaffeoylquinic acid) and cyanarin (1,5dicaffeoylquinic acid), phenolic compounds that are derivatives of caffeic acid (Figure 2).

Anthocyanin pigments are responsible for most of the blue-purple and reddy colour intermediate hues of artichoke plant tissues and an increase in anthocyanin pigmentation is considered a positive attribute of plant. It is reported that the main major anthocyanins in artichoke heads were cyanidin aglycone (Figure 3) and cyanidin glycosides (cyanidin 3,5-diglucoside, cyanidin 3-O- $\beta$ -glucoside, cyanidin 3,5malonyldiglucoside, cyanidin 3-(3"malonyl)glucoside, and cyanidin 3-(6"malonyl) glucoside (Schütz et.al.,2006). Besides peonidin aglycon (Figure 3) and the two peonidin derivatives were identified as peonidin 3-O- $\beta$ -glucoside, peonidin 3-(6"malonyl) glycoside (Schütz et.al.,2006).

Currently, the data of vegetable composition includes are mainly determined regarding raw vegetable material. The limited data are reporting on cooking processes of vegetables. Cooking processes would bring about a number of changes in the chemical composition, antioxidant activities and physical properties and vegetables (Miglio et.al.,2008; Turkmen et.al.,2005; Zhang et.al.,2004; Sahlin et.al.,2004).

It is reported that there are only two studies concerning quality parameters and antioxidant activity of some cooked vegetables including artichoke (Pellegrini et.al.,2009; Jiménez-Monreal et.al.,2009) and only one study regarding antioxidant profiles and some physical properties of artichoke (Ferracane et.al.,2008) in the literature.

The present detailed study was undertaken to determine the antioxidant activity, total phenolics, the simultaneous quantitative determination of maiör flavonoids cynarin (1,5-dicaffeoylquinic acid), chlorogenic acid (5-dicaffeoylquinic acid) and caffeic acids and major anthocyanidins (cyanidin, peonidin) and major quality parameters; to investigate the influences of several heat treatments including boiling, steaming and frying on major phenolic these acids (cynarin,chlorogenic acid, caffeic acid), anthocyanidin phenolics (cyanidin, peonidin). antioxidant activity, total phenolics, and some quality indicators in head and bracte leaves of breaded artichoke varieties [Cynara Cardunculus L. Scolymus var. Sakız, BayramPasa] and to carry out the ratio of monitored phenolics in total phenolics of raw and cooked artichokes.

#### 2.Material and Methods 2.1.Research Material

The artichoke variety SAKIZ (Figure 4a.) was obtained from Çeşme-Karaburun via Ege University Horticultural Department, Agriculture Faculty, Izmir,Turkey. Artichoke variety BAYRAMPAŞA (Figure 4b.) was obtained from Atatürk Horticulture Institute, Yalova, Turkey.

For variety development using the clonal selection of the artichokes lineages, 2 developmental lineages and 2 control The lineages were used. material reproduction operations that were performed for the variety development experiment were set up in a randomised complete block design with 4 replications in 2 locations. In each plot containing 10 plants, 4 candidate varieties and 2 control varieties were used. Each of the experimental and the control plants had the stem weight, width, and the length, as well as the head weight, width, and height measured both at the beginning and at the end of the breeding experiment.

#### 2.2.Chemicals

Cynarin (1,5-Di-caffeoylquinic acid) (Cat.No:30964-13-7; 10 mg) from Carl Roth GmbH & Co. (Karlsruhe, Germany), chlorogenic acid (5-0-Caffeoylquinic acid) (Cat.No:327-97-9; 1 g) and caffeic acid (Cat.No.331-39-5; 1 g) from Sigma (Germany), cyanidin (Cat.No:528-58-5; 10 mg) and peonidin (Cat.No:134-01-0; 5 mg) from Extrasynthese, Genay (France). Cyanidin-3-O-glycoside chlorur (Cat.No: 7084-24-4; 5 mg) from Sigma (Germany), luteolin 7-Oglukozid (Cat.No:5373-11-5; 250 mg) from Extrasynthese, Genay (France), 1.1diphenyl-2-picryl-hydrazyl (DPPH) (Cat.No. D9132-1G;1g) from Sigma-Aldrich, Chemie Gmbh (Munich, Germany) were obtained. All HPLC grade solvents were purchased from Merck (Darmstadt, Germany).

### **2.3.Preparation of Artichokes to Analysis and Processing**

Prior to analysis and processing, artichokes were freshly transferred to laboratory (Product Chemistry and Quality Control Laboratory) at 4 1 C refrigerated conditions from breading areas and equilibrated to room temperature for about 2 h before treatments.

Artichokes were washed, cleaned and blotted by blotting paper. Out bracte leaves and rude parts were seperated, stem parts were cutted by knife which cleaned with ethanole and awns of artichokes were discarded. Rude parts in nebs of bracte leaves were cleaned. Green parts were peeled as rolling by knife and accessed to head and heart, pileous parts were discarded by spoon. To prevent the browning of peeled parts, cutted and peeled artichokes were treated with lemon-water (For lemonwater content; 2 liter (10 glass-water) water and 3 lemon were used). Seperated green parts (bracte leaves) and white parts (headshearts) were homogenized at blender (*Waring*) apparatus. The homogenized bracte leaves and heart parts of artichokes were dried at N<sub>2</sub> atmosphere. Final samples were obtained for quality analyses, phenolic

#### 2.4. Extraction Methodology of Artichoke Phenolic Acids

Artichokes (*cynara cardunculus var. scolymus*) were extracted the method as shown below (n=2). Leave (or heart) part of artichoke sample was weighed (20 g)

Extraction with 70% ethanol and vortex during 8 min (*Nüve*, NM 110) Evaporation (79 °C) (Heidolp, *EssLab*, Essex, UK)

Acidification and pH regulation with formic acid to pH=2.4

Separation of lipid fraction (LF) with *n*-hexan and obtain the unlipidic fraction (UF)

Final extract (Extract of artichoke phenolics)

HPLC Analysis (20  $\lfloor L \rfloor$ 

### **2.5.High Performance Liquid Chromatographic (HPLC) Analysis Methodology** for Artichoke Phenolic Acids

Major artichoke phenolics cynarin, chlorogenic acid and caffeic acid were simultaneusly determined by modified isocratic HPLC based on the procedure from Sánchez-Rabaneda et.al.(2003) and Häusler et.al.(2002) and as shown below (n=2).

Column	: Hypersil-ODS
	[ $(250.4.6 \text{ mm} (5 \mid \text{m}) \text{ RP-18} (\text{Luna,Phenomenex,CAL})]$
Mobile Phase	: Acetonitrile/ phosphate buffer (25:75) (v/v) [pH=2.4]
Detection	: Fluorometric detection (254-370 nm) ((Shimadzu UV-1601)
Flow rate	: 1 ml/min
Sensitivity	: 0.05 A. U.F.

#### 2.6.Extraction Methodology of Artichoke Anthocyanidins

Artichoke (*Cynara cardunculus var. scolymus*) anthocyanidins were extracted the modified method from Goncalves et.al.(2007) and Schütz et.al.(2006) and as shown below (n=2). The pulp of leave (or heart) part of artichoke was weighed (5 g)

Extraction with 60% (v/v) aqueous methanol (MeOH) (50 ml) and vortex (*Nüve*, NM 110) during 10 min under the N<sub>2</sub> atmosphere at 25±1 °C

Filtration of extract mix with filtre paper (Whatman No. 1) (Main Extract)

Prior to HPLC injection, filtration with 0.45 m Durapore syringe (Millipore Corp., Bedford, MA)

Final Artichoke Anthocyanidin Extract (Extract of artichoke phenolics)

HPLC Analysis (20  $\lfloor L \rfloor$ 

2.7.High Performance Liquid Chromatographic (HPLC) Analysis Methodology for Artichoke Anthocyanidins

Major artichoke anthocyanidins cyanidin and peonidin were simultaneusly determined by modified HPLC method based on the procedure from Goncalves et.al.(2007) and as shown below (n=2).

**2.8.Total Anthocyanin Analyses of Artichokes** 

Total anthocyanins of artihockes were spectrophotometrically (*Optima SP 300*) determined based on the Estia et.al.(2002) and Prior et.al. (1998) (n=2). The pulp of leave (or heart) part of artichoke was weighed (5 g)

Extraction with HCl/ distilled water/ 80% (v/v) ethanol (EtOH) mixture (1/29/70) at ice-bath during 10 min

Extract was filtered

Absorbances were measured at 510 and 700 nm in buffers-pH 1.0 and pH 4.5

Maximum absorbance wave length of cyanidin-3-glycoside was determined as 534 nm

Total anthocyanin level was calculated according to equation as shown below

$$A = (A - A) - (A - A)$$
  
510 700 pH1.0 510 700 pH4.5

 $\mu$ g cyanidin-3-glycoside (C3G) /100 g extract was determined (MW = 445.2) (max. molar abs.  $\epsilon$ molar,  $\epsilon$  = 29600)

#### **2.9.Total Phenolic Analyses of Artichokes**

The level of phenolic compounds in artichoke samples was determined based on the Folin-Ciocalteu method(Singleton& Rossi,1965) and was expressed as gallic acid equivalents (n=2).

5 ml of 80% methanol including 1% HCl solution was added to 1 g artichoke leave (or heart)

The solution was mixed at  $4 \pm 1$  °C (ice-bath) during 2 h and centrifuged at 4000×g during 15 min.

The extract was filtered and clear part was separated for phenolic analyses. 2.5 ml of clear part (supernatant) was mixed with Folin-Ciocalteu reagent (2.5 ml) + distilled water (10 ml) [in ratio 0.5/0.5/10 (v/v/v)]

2 ml of 7% sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) was added to the mixture

The mixture was incubated for 2 h at room temperature and obtained blue-violet colour solution was measured at spectrophotometer (*Optima SP 300*) at 760 nm.

Total phenolics was expressed as chlorogenic acid equivalents (as  $\mu$ g Clg/ 100g fresh artichoke)

#### 2.10. Total Flavonoid Analyses in Artichokes

Total flavonoid analyses of artichokes were carried out spectrophotometrically(*Optima* SP 300) based on the aluminum chlorur chlorimetry method according to Singleton et.al. (1999) (n=2).

1 g artichoke leave (or heart) puree was treated with 5% sodium nitrite (NaNO<sub>2</sub>), 10% alüminum chlorure (AlCl<sub>3</sub>·6H<sub>2</sub>O) and 1 M sodium hydroxide (NaOH) mixture during 15 min

The absorbance of extract solution was measured at 510 nm and was expressed as luteolin 7-O-glukozid (luteolin-7-G) equivalent /100 g fresh artichoke

#### 2.11. Antioxidant Activity Analyses of Artichokes

Antioxidant activity analyses of artichokes were performed based on the 1,1-diphenyl-2-picryl-hydrazyl (DPPH) method described by Brand-Williams et.al.(1995) (n=2). 5 g of leave (or heart) part of artichoke was extracted with

80% (v/v) ethanol (EtOH) at ice-bath during 10 min

900  $\mu$ l of solutions in different concentration ranges (0.25–35  $\mu$ g/ml) were treated with 900  $\mu$ l 0.2 mM methanolic DPPH solution

The absorbance of each mixture was spectrophotometrically (*Optima SP 300*) measured at 517 nm,immediately.

After incubation at room temperature during 1,5,10,30 min, the absorbance of each mixture was again measured at 517 nm,immediately.

DPPH radical-scavenging activity was expressed as inhibition percent and calculated based on the equation as shown below.

DPPH radical-scavenging activity (%) =  $(1 - \text{antioxidant OD/ control OD}) \times 100$ The method was validated [ $y=12,33 \text{ x} - 3,87 (R^2=0.9997)$ ]

#### **2.12.Heat Treatments of Artichokes**

Three types of cooking methods (boiling, steaming, frying) were used. The optimized heat treatment conditions were applied for each artichoke. Testo 922 Dual Input Type K Thermocouple (Brandt Instruments, Inc., LA, USA) was used in temperature detection measurements for all heat treatments. In all cooking processes, same water with suitable pH (Erikli brand, pH 7.25) was used for the homogenize conditions and pH control of water was performed by Testo-206PH1 Tds (LA, USA) apparatus. All cooked samples with heat treatment were equilibrated to room temperature (25 $\pm$ 1 °C) under the ice-bath (4±1 °C) conditions via rapid cooling for prevent the artichoke antioxidants. All heating processes were done as twice replication.

#### 2.12.1. Boiling

Out bracte leaves of 6 of same calibre artichoke samples (*var*.Sakız 6 item; *var*.Bayrampaşa 6 item; as different experiment set) were peeled, washed and were transferred to stainless steel pan (Edition, *TEFAL*) including  $\frac{1}{2}$  pan boiled water and cooked at medium heat during 15 min. The excess water of boiled samples was decanted with colander about 40 sec, was equilibrated to room temperature (25±1 °C) and was prepared to analyses.

#### 2.12.2. Steaming

Out bracte leaves of 6 of same calibre artichoke samples (*var*.Sakız 6 item; *var*.Bayrampaşa 6 item; as different experiment set) were peeled, washed and transferred to steam cooker (VC 1002 Ultra Compact Buharlı Pişirici,*TEFAL*) and were cooked with <sup>3</sup>/<sub>4</sub> tea glass olive oil (*Tariş Naturel Sızma*).

Firstly, 1/3 proportion of water was transferred to water reservoir of steam cooker. Artichokes were placed to multilayered reservoirs of steam cooker, and olive oil was added, then cooked during 25 min and was equilibrated to room temperature ( $25\pm1$  °C) after cooking and was prepared to analyses.

#### 2.12.3. Frying

Out bracte leaves of 6 of same calibre artichoke samples (*var*.Sakız 6 item; *var*.Bayrampaşa 6 item; as different experiment set) were peeled, washed and were transferred to oiled oven tray (Oven:MF-26 GR Midi Oven, *VESTEL*) and were fried with <sup>3</sup>/<sub>4</sub> tea glass olive oil (*Tariş Naturel Sızma*). 5 min frying was performed in adjustable oven to 170 °C. After the frying, the excess frying oil was removed with blotting paper about 15-20 sec and fried artichokes were equilibrated to room temperature ( $25\pm1$  °C) and was prepared to analyses.

#### **3.Results and Discussion**

Figure 6 shows the standard and seperated sample HPLC chromatograms (Figure 6.). Average retention times (R.T.s) of cyanarin, caffeic acid and chlorogenic acid were 12.5 min, 15.7 min, 23.7 min, respectively in standard chromatogram whereas avg.retention times (R.T.s) of above mentioned compounds were 12.4 min, 15.6 min, 23.0 min, respectively in sample chromatogram (*var*.Bayrampaşa artichoke). It is shown that cyanarin, caffeic acid and chlorogenic acid were perfectly seperated by provided HPLC procedure (Figure 6.,chromatogram 1,2).

А mixture of cynarin (1.5dicaffeoylquinic acid), chlorogenic acid (5dicaffeoylquinic acid) and caffeic acid were perfectly simultaneously separated by an isocratic HPLC and a baseline resolutions was obtained as shown in Figure 6 first and second chromatogram (Figure 6.). Figure 6, second chromatogram shows that in artichoke sample [var.Bayrampaşa heart], (1,5-dicaffeoylquinic cynarin acid). chlorogenic acid (5-dicaffeoylquinic acid) and caffeic acid gave good base-line seperation and were determined. simultaneously (n=30) (Figure 6 second chromatogram). The quantification and

concentration determination of individual artichoke phenolic acids (cynarin, caffeic acid, chlorogenic acid) were obtained through calibration curves of standards by HPLC software. Our utilized extraction methodology and chromatographic seperation was shorter than that of studies and total elution time for three compounds was about 25 min. Not only was time saving but also economical determination regarding solvent consuming.

#### 3.1. Phenolic Acids (Cynarin, Chlorogenic Acid, Caffeic Acid) Quantities

Cynarin, chlorogenic acid and caffeic acid quantities of heart, bracte leaves and head of artichoke samples as shown in Table 1.

In our study, major phenolic acid was found as cynarin (Cyn) in heart parts of artichokes and its level was found as 29483  $\pm$  201 mg kg<sup>-1</sup> in heart of artichoke *var*.Sakız and

was found as  $18087 \pm 21 \text{ mg kg}^{-1}$  in heart of artichoke *var*.Bayrampaşa (p < 0.01) (Table 1).

Cyn levels were  $1512 \pm 2 \text{ mg kg}^{-1}$  ve 1058  $\pm$  5 mg kg<sup>-1</sup> in bracte leaves of *var*.Sakız and *var*.Bayrampaşa, respectively. It is shown that cynarin was the main compound of artichoke hearts and was found as 1.6 fold high in *var*.Sakız than var.Bayrampaşa (p<0.01) (Table 1.).

Figure 7 shows the phenolic acid amounts in heart and bracte leave parts of *var*.Sakız and *var*.Bayrampaşa in our study (Figure 7.).

Head part of artichokes includes heart and bracte leaves of artichokes. The present results demonstrated that both heart and bracte leaves and also sum total of heart and leaves, head part data were in our study and it has been put forwarded the importance of our study. In the literature, detailed study on phenolic compounds and polyphenols in all artihoche edible parts (head, heart and bracte leaves) are limited.

Romani et.al.(2006) were found polyphenol levels in different parts of typical Italian artichokes (Cynara scolymus L.) var. Violetto di Toscana and var. Terom. acid (Clg) Chlorogenic level had determined in bracte leaves of var. Violetto de Toscana and *var*. Terom, avg.  $8.72 \pm 6$ mg kg<sup>-1</sup> and avg.  $2.53 \pm 2$  mg kg<sup>-1</sup> by Romani et.al (2006). In our study, chlorogenic acid (Clg) levels were found as  $3197 \pm 27 \text{ mg kg}^{-1}$  and  $2379 \pm 43 \text{ mg kg}^{-1}$  in of artichokes var.Sakız heart and var.Bayrampaşa, respectively. Clg levels were found as 569 $\pm$  3 mg kg<sup>-1</sup> and 1263  $\pm$ 11 mg kg<sup>-1</sup> in bracte leaves of artichokes var.Sakız and var.Bayrampaşa, respectively and it is seen that bracte leaves of artichokes in our study have very high Clg existence in comparison with Italian varieties (Table 1.) (Figure 7.). Romani et.al. (2006) showed that Clg amount had determined in head of var.Violetto de Toscana and var.Terom, avg.  $30.51 \pm 20 \text{ mg kg}^{-1}$  and avg.  $14.25 \pm 10$ mg kg<sup>-1</sup> (Table 1.) while  $3766 \pm 30 \text{ mg kg}^{-1}$ and  $3642 \pm 54 \text{ mg kg}^{-1}$  were found in heads of our artichokes var.Sakız and *var*.Bayrampaşa, respectively and Clg level of our artichoke heads were very high (Table 1.) (Figure 7.).

Caffeic acid (Caf) concentration was found as  $452\pm2$  mg kg<sup>-1</sup> ve  $688\pm7$  mg kg<sup>-1</sup> in artichoke var.Sakız heart part and bracte leaves part, respectively whereas found as  $106 \pm 5 \text{ mg kg}^{-1}$  ve  $881 \pm 3 \text{ mg kg}^{-1}$  in artichoke var.Sakız heart part and bracte leaves part, respectively. It is determined that bracte leaves part included more caffeic acid than heart of artichokes in both varieties and it is seen that Caf levels of Sakız variety-heart was 4.2 fold higher than that of Bayrampaşa variety-heart (p < 0.01) (Tablo 1.). Detailed data regarding caffeic which deproteinized acid form of chlorogenic acid in artichokes was not found in literature.

Romani et.al.(2006) stated that 63.57 $\pm$ 48 mg kg<sup>-1</sup>cynarin and 27.54 $\pm$ 21 mg kg<sup>-1</sup> cynarin in bracte leaves of artihocke *var*.Violetto di Toscana and *var*.Terom,respectively.In our artichokes 1512  $\pm$  2 mg kg<sup>-1</sup>cynarin and 1058  $\pm$  5 mg kg<sup>-1</sup> cynarin were detected in Sakız and Bayrampaşa artichokes (Table 1.) (Figure 7.) and higher than literature data by Romani et.al.(2006).

It was reported the  $253.35 \pm 244$  mg cynarin kg<sup>-1</sup> in head of Violetto di Toscana artichoke and 95.02 cynarin  $\pm$  91 mg kg<sup>-1</sup>in head of Terom artichoke by Romani et.al.(2006). In artichoke head *var*.Sakız and in artichoke head *var*.Bayrampaşa, cynarin concentration was

extremely high and 30995  $\pm$  203 mg kg<sup>-1</sup> and 19145  $\pm$  26 mg kg<sup>-1</sup> of cynarin were determined, respectively (Table 1.) (Figure 8.). Figure 8 shows the phenolic acid levels in head of var. Sakız and var. Bayrampaşa in our study (Figure 8.). It has also been revealed that cyanidin was major compound in studied artichoke varieties. Especially heart part of studied artichokes were rich in cynarin and thereby cynarin profile has been determined extremely high in heads of artichokes, var.Sakız Turkish and var.Bayrampaşa. As overall, owing to the richness of their phenolic acid compositions, especially sources of cynarin compounds of Turkish artichokes, it has been stated their wealthiness of liverhepatic functions and antioxidative availability.

## **3.2. Total Phenolic Acid, Total Flavonoid and Total Phenolics in Artichokes**

In studied artichokes, total phenolic acid, total flavonoid and total phenolics were determined as shown in Table 2.

Total phenolic acids were determined as  $33325.12 \pm 85 \text{ mg kg}^{-1}$  in heart part of artichoke var.Sakız and while 20992.25  $\pm 23$  mg kg<sup>-1</sup> in heart part of artichoke var.Bayrampasa and 1.59 fold differency was found in each other (Table 2.). Total flavonoid levels was higher in artichoke var.Bayrampasa (3302.78 ± 17 mg kg<sup>-1</sup>) and was found 1.84 fold higher than artichoke var.Sakız (p < 0.01) (Table 2.). Figure 9 shows total phenolic acids, total flavonoids and total phenolics in parts of studied artichokes (Figure 9.).

Total phenolic amounts was found as  $35482,64 \pm 77$  mg kg<sup>-1</sup> in artichoke

*var*.Sakız whereas 24438,14 ±38 mg phenolics kg<sup>-1</sup> in *var*.Bayrampaşa and was 1.45 fold higher in Sakız artichokes (p<0.01). It is shown that both artichokes were good sources of phenolics (Table 2.).

Total phenolic acids (TPA) were found 36097.31± 90 mg kg<sup>-1</sup> in head of as artichoke var.Sakız and while 24208.47  $\pm$ 26 mg kg<sup>-1</sup> in heart part of artichoke var.Bayrampasa. Sakız head was rich as 1.49 fold in phenolic acids (Table 2.). Romani et.al.(2006) stated that 287.92 mg kg<sup>-1</sup> and 109.83 mg kg<sup>-1</sup> of TPA in head of artichoke var. Violetto di Toscana and var.Terom, respectively. TPA in Turkish artihocke heads were higher about 84-125 fold than the study reported by Romani et.al.(2006). Romani et.al.(2006) found that 72.99 mg kg<sup>-1</sup> and 30.46 mg kg<sup>-1</sup> of TPA in bracte leaves of var. Violetto di Toscana and var.Terom, respectively. Based on our data, hovewer, total phenolic acids were lower than another parts in var.Sakız and var.Bayrampaşa, Turkish artichokes have very rich in phenolic acids. TPA in bracte leaves were  $2772.19 \pm 5 \text{ mg kg}^{-1}$  and 3216.22mg kg<sup>-1</sup> for *var*.Sakız and  $\pm$ 3 var.Bayrampaşa, respectively and at least 44 fold higher than the literature data. Detailed data concerning chlorogenic acid in artichokes was not found in literature.

Due to extremely Turkish artichokeheart part have very high total phenolics, the total phenolic (TP) concentration of head parts have been extremely high (Table 2.) (p < 0.01). Total phenolics were found as  $40784.96 \pm 83 \text{ mg kg}^{-1}$  in head part of Sakız and 29952.6 ± 54 mg kg<sup>-1</sup> in head part 2.). of Bayrampasa (Table Romani et.al.(2006) reported 297.50 mg kg<sup>-1</sup> and 111.81 mg kg<sup>-1</sup> of total phenolics in Violette Toscana and Terom artichokes, di respectively. Romani et.al.(2006) also reported that total polyphenols level was 74.65 mg kg<sup>-1</sup> and 32.09 mg kg<sup>-1</sup> in bracte leaves of Violette di Toscana and Terom artichockes, respectively. In our study, total phenolics of bracte leaves were found as  $5302.32 \pm 6 \text{ mg kg}^{-1}$  in Sakız artichokes and  $5514.46 \pm 16 \text{ mg kg}^{-1}$  in Bayrampaşa variety (p<0.01) (Table 2.), this bracte leaves data very high than Violette di Toscana and Terom. TP in heart parts of our artichokes were 35482.64 ±77 mg kg<sup>-1</sup> for Sakız and 24438.14 ±38 mg kg<sup>-1</sup> for Bayrampaşa artichokes. Detailed data study regarding TP in heart part of artichoke was not found in literature.

Total flavonoid (TF) content was also detected in bracte leaves of Sakız  $(2011.53 \pm 4 \text{ mg kg}^{-1})$  and Bayrampaşa  $(1697.57 \pm 8 \text{ mg kg}^{-1})$  and in our previous study,  $3805.35 \pm 6 \text{ mg kg}^{-1}$  and  $5000.35 \pm$ 25 mg kg<sup>-1</sup> of TF in head parts of Sakız and Bayrampasa.respectively (p < 0.01) (Table 2.). In the literature, avg.166 mg kg<sup>-1</sup> of TF in bracte leaves while 198-958 mg kg<sup>-1</sup> of in heads were reported in Italian TF artichokes (Romani et.al., 2006). As it is seen that our artichokes were rich in TF contents (Table 2). TF levels of heart part of Turkish artichokes were found as 1793.82 -3302.78 mg kg<sup>-1</sup> while detailed data study regarding TF in heart part of artichoke was not found in literature.

As overall evaluation, Turkish artihocke head>heads>bracte leaves were very rich in phenolic compounds (Table 1 and Table 2). Especially, alongside of normal consuming of heart and leaves (totally thereby head) as meal, heart parts can be used as canned food goods, bracte leaves powder can be used as food additive and nutraceutical food.

## **3.3. Major Anthocyanidins and Total Anthocyanin Quantities in Artichokes**

Figure 10. shows the standards and seperated sample HPLC chromatogram of majör anthocyanidins in studied artichoke *var*.Sakız (Figure 10.). Average retention times (R.T.s) of cyanidin and peonidin were 2.58 min [(R.T) peonidin = 2.58 min] for peonidin and 5.08 min for cyanidin [(R.T)  $_{cyanidin}$  = 5.08 min] in sample artichokes.

It is seen that cyanidin and peonidin anthocyanidins were perfectly simultaneously seperated by provided HPLCprocedure (Figure 10). As it is seen, cyanidin and peonidin were perfectly simultaneously determined with HPLC base-line seperation (n=30). The sugar moities containing glycosides were removed with the used method and were obtained aglycon forms as anthocyanidins. Total anthocyanidin content including glycosides was determined, spectrophotometrically (Table 3.).

In our study, majör anthocyanidin (aglycon form) was cyanidin in both artichoke varieties. In studied artichokes, cyanidin aglycon which gives orange-red colour, amount was found as  $92.73 \pm 3.1 \,\mu\text{g}^{-1}/100\text{g}$  ( $0.92 \pm 0.03 \,\text{mg kg}^{-1}$ ) in heart of artichoke var.Sakız while  $101.11 \pm 4.0 \,\mu\text{g}^{-1}/100\text{g}$  ( $1.01 \pm 0.04 \,\text{mg kg}^{-1}$ ) in heart of artichoke var.Bayrampaşa. It was detected that dominant aglycon form was cyanidin and its concentration was higher in heart part than that of in bracte leaves (p<0.01) (Table 3.) (Figure 11.).

Peonidin which give more colour, was found less in both varieties and was found higher concentration in bracte leaves of both artichokes (Table 3.) (Figure 11.).  $156.84 \pm 9.4 \ \mu g/100g \ (1.56 \pm 0.09 \ mg \ kg^{-1})$ and  $154.42 \pm 9.9 \ \mu g/100g \ (1.54 \pm 0.09 \ mg$ kg<sup>-1</sup>) of cyanidin aglycon in artichoke *var*.Sakız and *var*.Bayrampaşa,respectively and the levels of major aglycon cyanidin var.Sakız in accordance with var.Bayrampaşa. It is stated that individual aglycon levels in Turkish artichokes as shown in Table 3 and Figure 11 (p < 0.01). Detailed study regarding individual aglycons in bracte leaves, in heart parts and in head parts of artichokes were not found in literature.

Total anthocyanin (TA) content of artichoke *var*.Sakız was determined as  $912.28 \pm 9.4 \ \mu g/100g \ (9.12 \pm 0.09 \ mg \ kg^{-1})$  and  $2091.42 \pm 11.2 \ \mu g/100g \ (20.91 \pm 0.11 \ mg \ kg^{-1})$  in head parts of artichokes *var*.Sakız and *var*.Bayrampaşa, respectively. We reported that TA level was 2.3 fold higher in Bayrampaşa artichoke heart. TA amount was determined as  $528.46 \pm 1.2 \ \mu g/100g \ (5.28 \pm 0.01 \ mg \ kg^{-1})$ in bracte leaves of artichoke *var*.Sakız and there was no significant differences with TA levels in *var*.Bayrampaşa (p<0.01). In artichokes, total anthocyanin levels in head part (including heart and bracte leaves) was also detected as shown in Table 3 and Fig.12.

In head parts of artichokes *var*.Sakız and *var*.Bayrampaşa, total anthocyanins were found as 1440.74±10.6  $\mu$ g<sup>-1</sup>/100g (14.40 ± 0.10 mg kg<sup>-1</sup>) and 2589.78±13.5  $\mu$ g<sup>-1</sup>/100g (25.89 ±0.13 mg kg<sup>-1</sup>), respectively and it is seen that artichokes were rich in anthocyanins. With regards to colour intensity, anthocyanidin compounds in Bayrampaşa artichokes was 1.8 fold higher than that of Sakız artichokes (*p*<0.01) (Table 3.) (Figure 12.).

Schütz et.al.(2006) had carried out anthocyanin characterization and quantification in artichokes varieties by high performance liquid chromatographyelectrospray ionization mass spectrometry (HPLC-DAD-ESI-MS). Schüts et.al.(2006) reported that anthocyanin profilles (with glycoside compounds) in the heads of German artichokes (Cynara scolymus L.) var."Camus", var."Green Globe", var."Le Castel", var."Petit Violet" and the heads of French artichokes (Cynara scolymus L.) var."Buette" and var."Poivrade"(Schütz et.al.,2006). Total anthocyanin level were determined as  $8.4 - 1705.4 \text{ mg kg}^{-1}$  in the study reported by Schütz et.al. (2006) and it has been stated that major anthocyanin compound was cyanidin-3-(6"-malonyl) glycoside and delphinidin and two peonidin as the others.

In our present study, both individual anthocyanidins-cyanidin, peonidin as aglycons and also total anthocyanins were determined in bracte leaves, in hearts, in head part of artichokes and as it is seen that that is detailed research on artichoke parts. In the literature, total anthocyanins (TA) in head part of German were found artichoke *var*. "Petit Violet" as  $8.4 \pm 0.0$  mg kg<sup>-1</sup> and in head part of French artichoke *var.*" Poivrade" as  $20.8 \pm 0.2 \text{ mg kg}^{-1}$ (Schütz et.al., 2006). In our study, TA was determined as  $14.40 \pm 0.10 \text{ mg kg}^{-1}$  $(1440.74 \pm 10.6)$  $\mu g/100g$ ) in head of artichoke var.Sakız and as  $25.89 \pm 0.13$  mg kg<sup>-1</sup> (2589.78±13.5µg/100g) in head of artichoke var.Bayrampaşa (Table 3) (Figure 11,12); especially TA level in var.Bayrampaşa was very high than German and French varieties. TA levels were found as  $912.28 \pm 9.4 \ \mu g/100g \ (9.12 \pm$  $0.09 \text{ mg kg}^{-1}$ ) and  $528.46 \pm 1.2 \mu \text{g}/100 \text{g} (5.28 \pm$ 0.01 mg kg<sup>-1</sup>) in heart and in bracte leaves of var.Sakız while 2091.42 ±11.2 µg/100g  $(20.91\pm0.11 \text{ mg kg}^{-1})$  and  $498.36\pm2.3$  $\mu g/100g (4.98 \pm 0.02 \text{ mg kg}^{-1})$  in heart and in bracte leaves of var.Bayrampasa, data respectively. Detailed study concerning total anthocyanins in bracte leaves and heart parts of artichokes were not found in literature. As it is seen, the importance of our study were revealed.

## **3.4.** The Alterations in Artichokes by Heat Treatment Effects

In our study three different cooking methods were applied to samples (Part 2.12.) and monitored the alterations of phenolic profiles, selected parameters and antioxidant activities. The changes in phenolic profiles after boiling, steaming, frying were shown in Table 4 (p<0.01) (Table 4.).

## **3.4.1. The Alterations of Phenolic Acid Profiles After Boiling and Steaming**

It is determined that after the heat treatment, especially caffeoylquinic acids levels importantly increased. Cynarin (1,3di-O- caffeoylquinic acid) and chlorogenic acid (5-O- dicaffeoylquinic acid) of artichokes rised (p < 0.01) (Table 4.) and it has been considered that the increasing of caffeoylquinic acids were owing the formed different dicaffeoylquinic acid isomers after heat treatments. High total phenolic acid levels of studied artichokes after heat treatments have been verified our remarks on forming of different dicaffeoylquinic acid isomers (p < 0.01) (Table 4.). Especially via the boiling effect, phenolic acid levels of artichokes mostly increased by comparison steaming and frying, respectively to (p<0.01) (Table 4.) (Table 4.1.). Between the dicaffeoylquinic acid concentrations formed after steaming and frying were not found significant differency as statistically (p<0.01) (Table 4.) (Table 4.1.).

Chlorogenic acid (Clg) level in bracte leaves of var.Sakız was 1354.22±19 mg kg<sup>-1</sup> after boiling process and was found as 2.38 fold high in comparison to raw form while its level was  $6745.67 \pm 152 \text{ mg kg}^{-1}$ in heart part of Sakız and was detected as 2.11 fold increased in comparison with raw heart form. In boiled head part of artichoke *var*.Sakız, 8099.89 ±171 mg kg<sup>-1</sup> of chlorogenic acid (5-O-caffeoylquinic acid) was detected and was 2.15 fold higher than its raw head form (p < 0.01) (Table 4). After boiling process, in bracte leaves of Bayrampaşa,  $2867.01 \pm 8 \text{ mg kg}^{-1}$  of Clg was found and its level was 2.27 fold higher than its concentration in raw leaves whereas 6994.26 ±93 mg kg<sup>-1</sup> of Clg was determined in heart part of Bayrampaşa and it was 2.94 fold high from raw heart of Bayrampaşa (*p*<0.01) (Table 4).

It was found that boiled head part of Bayrampaşa included 9861.27  $\pm$ 93 mg kg<sup>-1</sup> of Clg and this boiled form contained 2.7 fold high in Clg (*p*<0.01) (Table 4).

After boiling, cynarin (1,3-di-Ocaffeoylquinic acid) content was found as 8.02 and 8.42 fold high in boiled bracte leaves and boiled heart forms, respectively. With the boiling process, the raising of major artichoke phenolic substance cynarin was considerably high in comparing to that of chlorogenic acid (p<0.01) (Table 4). It is stated that due to the dicaffeoylquinic acid groups of cynarin, it has been formed more isomers, likewise chlorogenic acid (5-Ocaffeoylquinic acid) is monocaffeoylacid group. Figure 13 shows the cynarin (1,3-di-O- caffeoylquinic acid) level after boiling and steaming process (Figure 13).

After boiling process, total phenolic acids (TPA) in heart parts of Sakız and Bayrampaşa were found as  $63531.26 \pm 163$ ve  $58638.56 \pm 87$  mg kg<sup>-1</sup> and it was found that 1.76 and 2.4 fold increasings in their concentrations, respectively after the boiling (p<0.01) (Table 4.). With steaming process, TPA in head of *var*.Sakız and *var*.Bayrampaşa was found as 75602.19 ±66 and 70952.66 ± 61 mg kg<sup>-1</sup> and was detected 2.09 and 2.93 fold raising in their TPA amounts after steaming. In steaming forms, phenolic acid contents of *var*.Sakız and *var*.Bayrampaşa were 1.19 and 1.21 fold high, comparison to their boiled forms, respectively. (p<0.01) (Table 4.).

After the cooking at steam, cynarin (1,3-di-O-caffeoylquinic) level was 81219.44 ±89 mg kg<sup>-1</sup> in head of Bayrampaşa artichoke variety and was detected as 4.24 fold raising in cynarin concentration of steamed heads, as comparing to raw heads (p<0.01) (Table 4.).

In steamed forms, cynarin contents were lower in Sakız and Bayrampaşa, comparing to boiled forms and were detected 2.34 and 2.14 fold lower (p < 0.01) (Table 4.). From the point of total phenolic acid (TPA) content, in artichokes, hovewer TPA of steamed forms were high, individual cynarin contents were higher in boiled forms (p < 0.01) (Table 4.). The establishing of these findings were notable for our study. Due to it is known more strong antioxidant and liver protector (hepatoprotective), anti-LDL effects of cynarin which is the major phenolic compound of artichoke; regarding findings on mostly high concentration of cynarin in boiled artichokes and regarding findings on good levels of cynarin in steam cooking were notable for consumers (Table 4.).

After steaming process, chlorogenic acid was found in steamed head parts of var.Sakız and var. Bayrampaşa as 6439.86  $\pm 43$  and 6154.98  $\pm 24$  mg kg<sup>-1</sup> and 1.71 and 1.69 fold rising was found, respectively (Table 4.). As it is known, chlorogenic acid (Clg) is strong antioxidant, livergallbladder-friendly, anti-cancer and antimicrobial agent. It is shown that with boiled artichoke or steam cooked artichoke, high concentrations of Clg can be absorbed. Our study has been put forwarded the notable data. In steamed forms, cynarin contents were detected as 1.26 and 1.6 fold

lower than their boiled forms in *var*.Sakız and in *var*.Bayrampaşa, respectively, but both boiled and steamed forms can be fruitfull (p<0.01) (Table 4.).

# **3.4.2.** The Alterations of Total Flavonoids, Total Phenolics After Boiling and Steaming

In the head parts of *var*.Sakız, after the boiling, total flavonoid levels were found as  $2782.94 \pm 23 \text{ mg kg}^{-1}$  luteolin-7-*O*-glukozid equivalent (p < 0.01) (Table 4.). It was stated that total flavonoids level affected by boiling process and was detected 1.36 fold decreasing (Table 4.).

Total flavonoid (TF) levels of steamed var.Sakız was found as 2954.45  $\pm$ 34 mg kg<sup>-1</sup> luteolin-7-O-glukozid equivalent and was 1.28 fold lower than that of raw forms. TF levels in boiled head of var.Bayrampasa and in steamed head of same cultivar were  $4717.31 \pm 10 \text{ mg kg}^{-1}$ and  $4733.83 \pm 23 \text{ mg kg}^{-1}$  luteolin-7-Oglukozid equivalent and there was no significant difference regarding the cooking losses between two processing, statistically (p < 0.01) (Table 4.). As TF quantity, the cooking losses in both method were not high, thereof, the consuming of boiled or steamed artichokes can be healthy for consumers owing to they are also good flavonoid sources with antioxidative. auxiliary of anticancer, LDL cholesterol inhibition properties (Table 4.).

Total phenolic (TP) substances in boiled head of var.Sakız was 252866.75±127 mg Clg/kg and TP in steamed head of var.Sakız 228388.55±138 mg Clg/kg. In Bayrampaşa variety, TP level was 188400.43 ±92 and 186969.12  $\pm 110 \text{ mg Clg/kg}$ , after boiling and steaming processes, respectively. In both varieties, with boiling and steaming, about 6.2 and 5.6 fold increasing were determined, compared to their raw forms (p < 0.01) (Table 4.). Figure 14 shows the alterations in total phenolic acids, total flavonoids and total phenolics of studied artichokes after boiling and steaming (Figure 14).

In our study, it has been put forwarded that boiling or steaming were effective cooking methods for maximum phenolic availability from artichoke vegetable (p<0.01) (Tablo 4.).

# **3.4.3.** The Alterations of Anthocyanidins (Aglycons), Total Anthocyanins After Boiling and Steaming

After boiling, in head of var.Sakız, total anthocyanin (Tantho) quantities were found 2780.62  $\pm 105 \ \mu g / 100 g$  as cyanidin-3-glycoside (C3G) equivalent and was detected as 1.93 fold high total anthocyanin in comparing to raw form whereas Tantho quantities were 1383.11  $\pm$ 58 µg C3G /100g in head of var.Sakız with steam cooking application and was detected 1.04 fold decreasing than that of raw form (p < 0.01) (Tablo 4.). It is considered that the duration of boiling application was 15 min and the duration of steaming cooking was 25 min (parts 2.12.1. and 2.12.2.), due to the longer heating process duration in steaming, it may be ring opening in unstable anthocyanin compounds, so a far amount of anthocyanin loss may be in steam cooking of artichokes, comparing to raw form. It is stated that the proposed mechanism the convertion of cyanidin aglycon to cyanidin 3-glycosid and cyanidin 3,5-di-glycoside (Figure 15.) (Anonymous,2005).

Aglycon form (anthocyanidin) has been formed via attaching of sugar moities from 3. and 5. sites of the molecule, cyanidin-glycoside forms has been increased with the boiling process. It has been commented that sugar (glycoside) content of vegetable may complex to colour compounds, merely, within the longer heating time, it may be the openings from O<sup>+</sup> position of the ring or it may be rupture from sites attached of glycosides in the ring or it may be the conversions in the molecule (Anonymous,2005;Tokuşoğlu & Başay, 20 09).

With the boiling effect, in the head of Bayrampaşa, total anthocyanin (TA) quantity was found as 5904.69  $\pm$ 77 µg /100g (as cyanidin-3-glycoside,C3G,

equivalent) and was detected as 2.28 fold increasing in TA content. After the steam cooking, TA levels was determined as 2686.78 ±25 µg C3G /100g in head of var.Bayrampaşa, comparing to raw form. As it was seen that TA levels had not decreased steamed in *var*.Bayrampaşa,compared to steamed var.Sakız; this result may be interpreted that the sugar content of var. Bavrampasa was higher than that of var.Sakız (p < 0.01)(Table 4.).

Artichoke colour substances, anthocyanins are antioxidants, strong effective on specific cancer types and have positive effects on health of urinary system (urinary tract system and urologic system), constituents important on memory functions and eye health (Anonymous,2008; Tokuşoğlu & Basay,2009).

Cyanidin aglycon level was identified in boiled head of *var*.Sakız and in boiled head of *var*.Bayrampaşa as  $68.19\pm4$ µg/100g and  $65.97\pm5$  µg/100g, respectively was found as about 50% of decreasing in both varieties, comparing to raw forms (p<0.01) (Table 4.).

For effective availability of artichoke anthocyanins, boiling >steaming effective cooking were consuming methods, respectively (p < 0.01) (Table 4.). After steam cooking,  $35.78 \pm 12 \text{ µg}/100\text{ g of}$ cyanidin aglycon was detected in head of Bayrampaşa and was determined the 4.3 fold of decreasing in raw head form (p < 0.01) (Table 4.). The more cyanidin decreasing in var.Bayrampaşa may be due to the more dark green content of var.Bayrampaşa. It may be interpreted that the dark green colour content of artichoke may be more increase and also its in cyanidin content may be more decrease. For our hypothesis verification, L\*a\*b\* Colour-Hunter values were measured and it is (redness) determined the а values decreasing. In heart part of the artichokes, it was detected the same level of cyanin stability (*p*<0.01) (Table 4.).

## **3.4.4. The Alterations of Major Phenolic Profiles After Frying**

After frying (part 2.12.3.) process, cynarin was found in head of *var*.Sakız and in head of *var*.Bayrampaşa as 74078.21  $\pm 155$  ve 43650.52  $\pm 103\mu g/100g$  and it was detected as 2.39 and 2.28 fold increasing of cynarin for *var*.Sakız and *var*.Bayrampaşa, respectively, in comparing to raw forms (p<0.01) (Table 5.). With frying, chlorogenic acid quantity rised as 2.13 fold in *var*.Sakız and as 1.9 fold in *var*.Bayrampaşa (p<0.01) (Table 5.).

Total phenolic acids was determined as  $65697.23\pm42 \ \mu g/100g$  in fried head of *var*.Sakız and was detected 1.82 fold increasing, in comparison to raw head (p<0.01) (Table 5.). After frying, total flavonoid content was 1953.26 ± 23 and 1452.42 ±50  $\mu g/100g$  luteolin-7-*O*-glikozid in Bayrampaşa head and in Sakız head,respectively and was detected 2.56 and 2.62 fold of decreasing (p<0.01) (Table 5.).

After frying process, total phenolics (TP) was determined in fried head of *var*.Sakız and was detected as 3.34 fold high (136218.56  $\pm 97 \ \mu g/100g \ Clg \ equivalent)$ . In fried head of var.Bayrampaşa, TP levels was found as 104235.05  $\pm$ 111 µg/100g and was determined 3.48 fold increasing. After the frying, 3.14 and 3.05 fold decreasings were obtained in total anthocyanin (TA) content (p < 0.01) (Tablo 5.). Owing to the intensity of glycosid groups of *var*. Bayrampaşa, 849.108  $\pm$ 4 µg/100g cyanidin-3-*O*-glycosid equivalent was detected in head of Bayrampaşa artichokes and 1.85 fold higher than that of fried var.Sakız head (*p*<0.01) (Tablo 5.).

Figure 16 shows total anthocyanin levels of artichoke heads after frying process (Figure 16.) Cyanidin aglycon was determined in *var*.Sakız and *var*.Bayrampaşa as 29.89  $\pm$ 5 and 30.06  $\pm$ 2 µg/100g respectively and was found 5.25 and 5.14 fold of decreasings, comparing to their raw forms (*p*<0.01) (Tablo 5.).

After frying process, phenolics were also good levels in fried artichokes and were detected increasing levels in some phenolic profiles (p<0.01) (Tablo 5.). Artichoke frying is alternative consuming to boiling and steam cooking of artichokes, also aroma and flavor compounds has been formed in frying. In this point, frying time and frying temperature must be controlled and also frying oil quality must be considered for frying process.

Ferracane et.al. (2008) were determined that changing of phenolic caffeoylquinic acid isomers, apigenin derivatives of cooked Italian artichokes with various cooking procedures (Ferracane et.al.,2008). Ferracane et.al. (2008) reported that about 60% of increasings in total caffeoylquinic acids

in steamed Italian artichokes. It was no significant alteration in apigenin phenolics in Italian artichokes (Ferracane et.al.,2008).

#### 3.5. Antioxidant Activity in Artichokes

Antioxidant activity (AA) is a measurement of free radical distinctness ability of products. In our study, with DPPH method, antioxidant activities of raw, boiled, steamed, fried artichokes *var*.Sakız and *var*.Bayrampaşa were determined as Trolox equivalent (p<0.01) (Tablo 6.). Especially in boiled and steamed artichokes, it was determined the highest AA levels (p<0.01) (Table 6.).

In Turkish artichokes, the increasing of AA was about7 fold in boiled form, was about 11 fold in steamed form and also was about 5.5 fold in fried forms of artichokes (p<0.01) (Table 6.) (Figure 17.). Ferracane et.al.(2008) reported that the increasing of AA was 8 fold in boiled Italian artichokes whereas that of was about 15 fold in steam cooked Italian artichokes. Several research were found on antioxidant activity of artichokes (Fratianni et.al., 2007; Lattanzio Alamanni et.al.,2003) et.al.,2005; in literature but one research was found for comparable to our data. Our study data on AA was in accordance with that study by Ferracane et.al.(2008) however processing procedure differency was.

cooked Italian artichokes. Several research were found on antioxidant activity of artichokes (Fratianni et.al.,2007; Lattanzio et.al.,2005; Alamanni et.al.,2003) in literature but one research was found for comparable to our data. Our study data on AA was in accordance with that study by Ferracane et.al.(2008) however processing procedure differency was.

#### 4. Conclusion

In our performed study, various phenolic parameters were examined in two raw artichoke varieties and were also identified phenolic profiles after cooking processes including boiling, steaming and frying. It is seen thet our study was detailed on individual phenolic acids;caffeoylquinic acids, total flavonoids,total phenolics, anthocyanidins, total anthocyanins, antioxidant activity and their alterations on cooking processing effects. This previous study on phenolic profiles of processed artichokes has been given the strong data for food science and technology literature.

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#### Referance

Adzet T, Camarasa J, Laguna JC. **1987**. Hepatoprotective activity of polyphenolic compounds from Cynara scolymus against CCl4 toxicity in isolated rat hepatocytes. *J Nat Prod*. 50(4), 612-617.

Agarwal R, Mukhtar H. **1996.** Cancer chemoprevention by polyphenols in green tea and artichoke. Adv Exp Med Biol. 401, 35-50. Review.

Alamanni M.C., Cossu M. **2003**. Antioxidant Activity of the Extracts of the Edible Part of Artichoke (*Cynara Scolymus* L.) Var. Spinoso Sardo. *Ital.J.Food Sci.* n.2, vol.15, 187-195.

Anonim**2020**.Facts,comparisons.Artichoke .http://www.efactsonline.com (acces.2020 Jul)

Anonim**2020**.OceanmistFarm.<u>http://www.</u> <u>oceanmist.com</u> (accessed 2020 July).

Anonymous. **2012**. Oceanmist Farm. http://www.ocean.mist.com

Brand-Williams W., Cuvelier M.E., Berset C. **1995**. Use of free radical method to evaluate antioxidant activity, *Lebensmittel–Wissenschaft und Technologie* 28, 25–30.

Bundy R, Walker AF, Middleton RW, Wallis C, Simpson HC. **2008**. Artichoke leaf extract (Cynara scolymus) reduces plasma cholesterol in otherwise healthy hypercholesterolemic adults: a randomized, double blind placebo controlled trial. *Phytomedicine*. 15(9), 668-675.

Bundy R, Walker AF, Middleton RW, Marakis G, Booth JC. **2004**. Artichoke leaf extract reduces symptoms of irritable bowel syndrome and improves quality of life in otherwise healthy volunteers suffering from concomitant dyspepsia: a subset analysis. *J Altern Complement Med.* 10(4), 667-669.

Costabile A, Kolida S, Klinder A, Gietl E, Bäuerlein M, Frohberg C, Landschütze V,

Gibson GR. **2010**. A double-blind, placebocontrolled, cross-over study to establish the bifidogenic effect of a very-long-chain inulin extracted from globe artichoke (Cynara scolymus) in healthy human subjects. *Br J Nutr*. 104(7), 1007-1017.

Estia M., Cinquantaa L., Sinesiob F., Monetab E., Di Matteoc M. (2002). Physicochemical and sensory fruit characteristics of two sweet cherry cultivars after cool storage. *Food Chemistry* 76, 399– 405.

Ferracane R, Pellegrini N, Visconti A, Graziani G, Chiavaro E, Miglio C, Fogliano V. **2008**. Effects of different cooking methods on antioxidant profile, antioxidant capacity, and physical characteristics of artichoke. *J Agric Food Chem.* 56(18), 8601-8.

Fratianni, F., Tucci, M., De Palma, M., Pepe, R., Nazzaro, F., **2007.** Polyphenolic composition in different parts of some cultivars of globe artichoke (Cynara cardunculus L. var. scolymus (L.) Fiori). *Food Chemistry* 104, 1282–1286.

Gebhardt R., Fausel M. **1997.** Antioxidant and hepatoprotective effects of artichoke extracts and constituents in cultured rat hepatocytes. *Toxicol in Vitro.* 11(5), 669-672.

Goncalves B., Silva A.P., Moutinho-Pereira J., Bacelar E., Rosa E., Meyer A.S. **2007.** Effect of ripeness and postharvest storage on the evolution of colour and anthocyanins in cherries (Prunus avium L.) *Food Chemistry* 103, 976–984.

Häusler M., Ganzera M., Abel G., Popp M., Stuppner H. **2002**. Determination of Caffeoylquinic Acids and Flavonoids in *Cynara scolymus* L. by High Performance Liquid Chromatography.*Chromatographia*. 56(7-8), 407-411 Jiménez-Monreal AM, García-Diz L, Martínez-Tomé M, Mariscal M, Murcia MA. **2009**. Influence of cooking methods on antioxidant activity of vegetables. *J Food Sci.* 74(3), 97-103.

Lattanzio, V., Cicco, N. and Linsalata, V. **2005**. Antioxidant activities of artichoke phenolics. *Acta Hort*. 681:421-427.

Llorach R., Espin J.C., Tomas-Barberan, F.A., Ferreres F. **2002.** Artichoke (*Cynara scolymus* L.) by products as a potential source of health-promoting antioxidant phenolics. *J. Agric. Food Chem.* 50(12), 3458-3464.

López-Molina D, Navarro-Martínez MD, Rojas Melgarejo F, Hiner AN, Chazarra S, Rodríguez-López JN. **2005**. Molecular properties and prebiotic effect of inulin obtained from artichoke (*Cynara scolymus* L.). *Phytochemistry*. 66(12), 1476-1484.

Miccadei S, Di Venere D, Cardinali A, Romano F, Durazzo A, Foddai MS, Fraioli R, Mobarhan S, Maiani G.**2008.** Antioxidative and apoptotic properties of polyphenolic extracts from edible part of artichoke (Cynara scolymus L.) on cultured rat hepatocytes and on human hepatoma cells. *Nutr Cancer*. 60(2), 276-83.

Miglio, C.; Chiavaro, E.; Visconti, A.; Fogliano, V.; Pellegrini,N. Effects of different cooking methods on nutritional and physicochemical characteristics of selected vegetables. *J. Agric. Food Chem.* **2008**, *56*, 139–147.

Pellegrini N, Miglio C, Del Rio D, Salvatore S, Serafini M, Brighenti F. **2009**. Effect of domestic cooking methods on the total antioxidant capacity of vegetables. *Int J Food Sci Nutr.* 60 Suppl 2, 12-22.

Prior, R.L.; Cao, G.; Martin, A.; Sofic, E.; McEwen, J.; O'Brien, C.; Lischner, N.; Ehlenfeldt, M.; Kalt, W.; Krewer, G.; Mainland, C.M. **1998**. Antioxidant Capacity as Influensed by Total Phenolic and Anthocyanin Content, Maturity, and Variety of Vaccinium Species. J. Agric. Food Chem. **1998**, 46, 2686-2693.

Romani A.,Pinelli P.,Cantini C.,Cimato A.,Heimler D. **2006**. Characterization of Violetto di Toscana, a typical Italian variety of artichoke (*Cynara scolymus* L.). *Food Chem* 95, 221-225.

Rondanelli M, Giacosa A, Opizzi A, Faliva MA, Sala P, Perna S, Riva A, Morazzoni P, Bombardelli E. **2012.** Beneficial effects of artichoke leaf extract supplementation on increasing HDL-cholesterol in subjects with primary mild hypercholesterolaemia: a double-blind, randomized, placebocontrolled trial *Int J Food Sci Nutr*. In Press

Sahlin, E., Savage, G. P.; Lister, C. E. **2004**. Investigation of the antioxidant properties of tomatoes after processing. *J. Food Compos. Anal.* 17, 635–647.

Sánchez-Rabaneda F.. Jáuregui O., Lamuela-Raventós R.M., Bastida J., Viladomat F., Codina C. 2003. Identification of phenolic compounds in artichoke waste by high - performance liquid chromatography - tandem mass spectrometry. Journal of Chromatography A, 1008: 57-72.

Singleton, V.L., Rossi, J.A.Jr. **1965.** Colou rimetry of total phenolics with phosphomolybdic-phosphotungustic acid reagents. Am. J. Enol. Viticult., 6;144-158.

Singleton, R. Orthofer and R.M. Lamuela-Raventos **1999.** Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin–Ciocalteu reagent, *Methods Enzymology* 299, 152– 178.

Speroni E, Cervellati R, Govoni P, Guizzardi S, Renzulli C, Guerra MC. **2003**. Efficacy of different *Cynara scolymus* preparations on liver complaints. *J Ethnopharmacol.* 86(2-3):203-211.

Schütz K., Persike M., Carle R., Schieber A. 2006. Characterization and quantification of anthocyanins in selected artichoke (*Cynara scolymus* L.) cultivars by HPLC – DAD–ESI–MS. *Analytical and Bioanalytical Chemistry*. 384, 1511–1517.

Tokuşoğlu Ö. 2018. Food By-Product BasedFunctionalFoodPowders,(TheNutraceuticals:BasicResearch/ClinicalApplication Series Book)CRC Press, Taylor& Francis Group, Boca Raton, Florida, USA.ISBN 9781482224375.

Tokuşoğlu Ö., Başay S. 2009. The Researches Determination on of the Antioxidant Phenolic Compounds and of Anthocyanins Turkish Artichoke (Cynara scolymus L.) varieties (Sakız, Bayrampaşa): The Researches on Monitoring of Alterations by heat process effects. Celal Bayar University Research Fund Project with Yalova Atatürk-CentralHorticultural Research Institute. Project No: 2005060

Tokuşoğlu Ö., Başay S. **2008.** The major functional phenolic constituents (cyanarin, chlorogenic acid, caffeic acid) in artichoke bracts and leaves. 6<sup>th</sup> Euro Fed Lipid Congress: Oils, Fats and Lipids in the 3rd Millennium: Challenges, Achievements and Perspectives, Book of Abstracts p.518, 7-10 September 2008, Athens, GREECE, 2008

Turkmen, N., Sari, F. and Velioglu, S. **2005**. The effect of cooking methods on total phenolics and antioxidant activity of selected green vegetables. *Food Chemistry* 93: 713-718.

Zhang, D.; Hamauzu, Y. Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking. *Food Chem.* **2004**, *88*, 503–509.

#### **FIGURES**

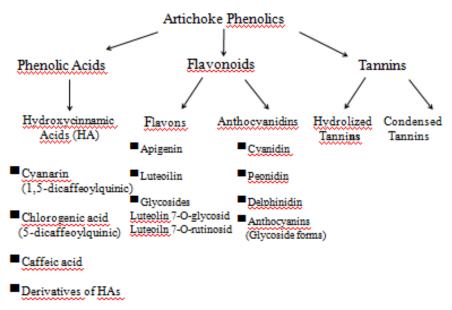
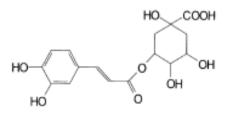
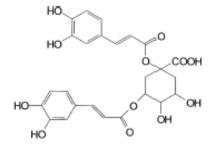
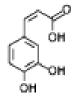


Figure 1.



Chlorogenic Acid (5-Dicaffeoylquinic Acid)





Cynarin (1,5-Dicaffeoylquinic Acid)

Caffeic Acid

Figure 2.

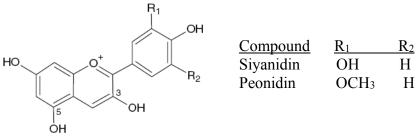


Figure 3.



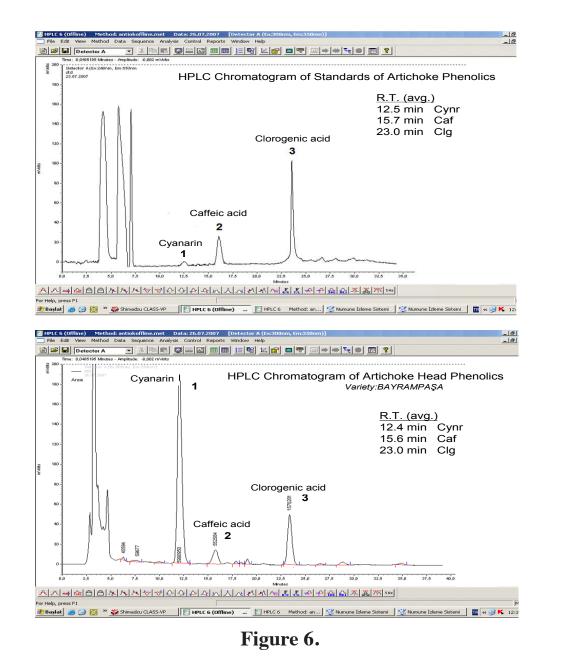
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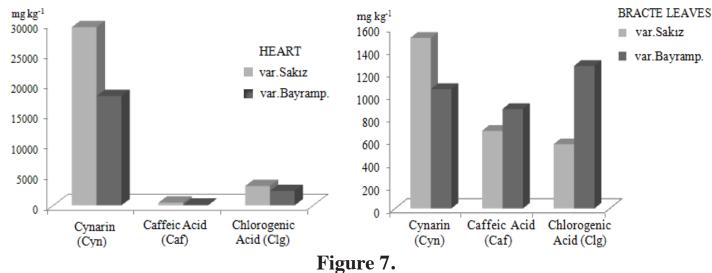


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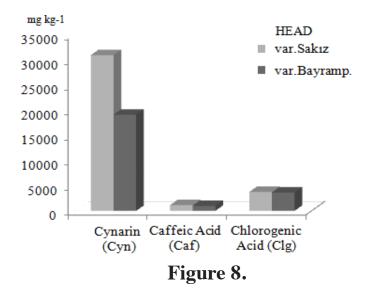


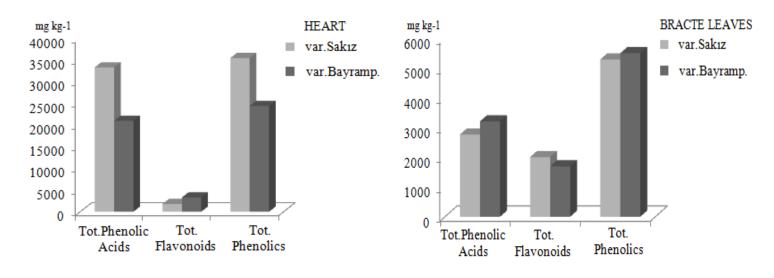
Figure 5.





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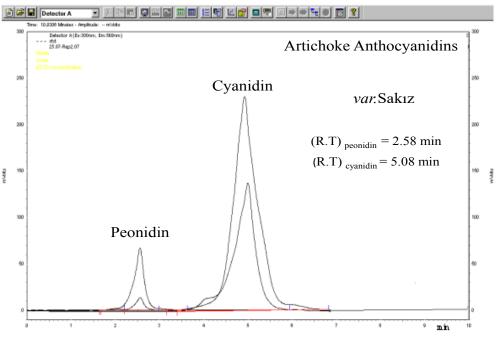


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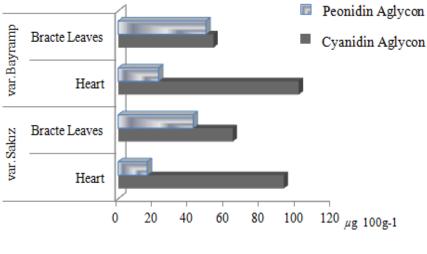


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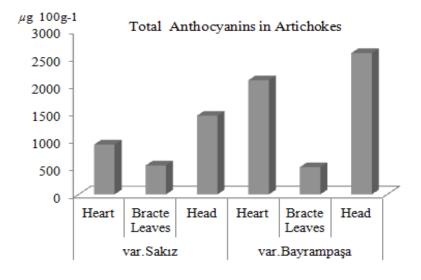


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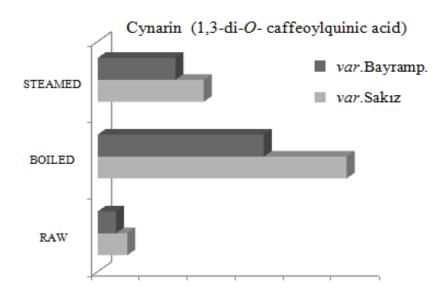
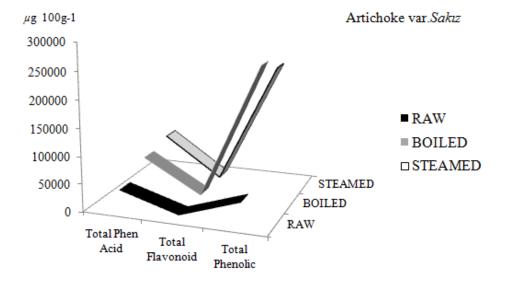


Figure 13.





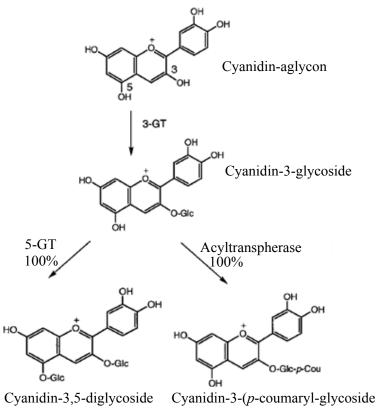
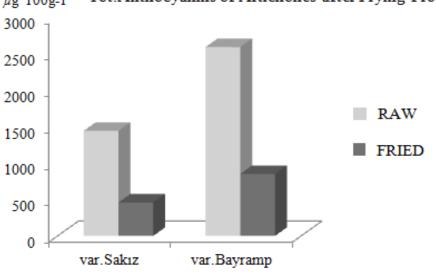


Figure 15.



μg 100g-1 Tot.Anthocyanins of Artichohes after Frying Process



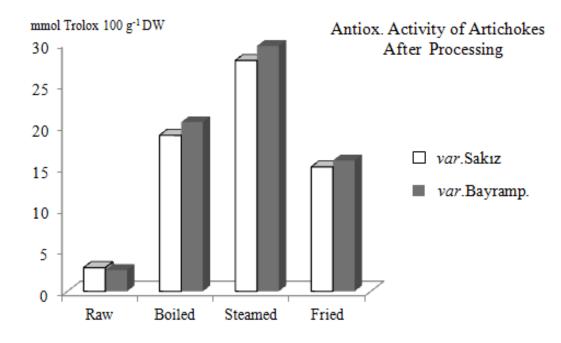


Figure 17.

Artihocke Varieties						
Parameters		var.Sakız		var.Bayrampaşa		
(mg kg <sup>-1</sup> )	Heart	Leave	Head	Heart	Leave	Head
Cynarin (Cyn)	29483 ±201	$1512 \pm 2$	$30995 \pm 203$	$18087 \pm 21$	$1058 \pm 5$	$19145 \pm 26$
Caffeic Acid (Caf)	452 ± 2	688 ± 7	1140±9	$106 \pm 5$	881 ± 3	987±8
Chlorogenic Acid (Clg)	3197 ± 27	569 ± 3	3766 ± 30	$2379 \pm 43$	$1263 \pm 11$	$3642 \pm 54$

#### Table 1. Phenolic Acid Levels of Artichokes\*

 $\Box(p < 0.01); n=30; \text{ as mg kg}^{-1} \text{ FW}$ 

	Artichoke Varieties						
Parameters	var.Sakız			<i>var</i> .Bayrampaşa			
(mg kg <sup>-1</sup> )	Heart	Bracte Leaves	Head	Heart	Bracte Leaves	Head	
Total Phenolic Acids	33325.12 ±85	2772.19 ± 5	36097.31±90	20992.25 ±23	$3216.22 \pm 3$	$24208.47 \pm 26$	
Total Flavonoids	1793.82 ± 2	$2011.53 \pm 4$	$3805.35 \pm 6$	3302.78 ± 17	1697.57±8	5000.35 ± 25	
Total Phenolics	35482.64 ±77	5302.32 ± 6	40784.96 ± 83	24438.14 ±38	5514.46 ± 16	29952.6 ± 54	

 $\Box(p < 0.01); n=30; \text{ as mg kg}^{-1} \text{ FW}$ 

	Artihoche Varieties					
Parameters	var.Sakız			l	<i>ar</i> .Bayrampaşa	
(µg 100 g <sup>-1</sup> )	Heart	Bracte Leaves	Head	Heart	Bracte Leaves	Head
Cyanidin Aglycon	92.73 ± 3.1	64.11±6.3	$156.84 \pm 9.4$	$101.11 \pm 4.0$	53.31±5.9	$154.42 \pm 9.9$
Peonidin Aglycon	$16.22 \pm 2.7$	41.95± 9.0	58.17±11.7	$22.55 \pm 2.2$	49.07± 3.6	$71.62 \pm 5.8$
Total Anthocyanins	912.28 ± 9.4	528.46± 1.2	1440.74±10.6	2091.42 ±11.2	498.36± 2.3	2589.78±13.5

 $\Box(p < 0.01); n=30;$  as  $\mu g 100 g^{-1}$  FW; In the text, the data was also compared as mg kg<sup>-1</sup> (convertion;  $\mu g g^{-1} = mg kg^{-1}$ )

							Heat Tr	eatments		
Compound	Variety	Raw		Boiled			Steamed			
		Bracte Leave	Heart	Head	Bracte Leave	Heart	Head	Bracte Leave	Heart	Head
Cynarin	Sakız	$1512 \pm 2$	29483 ± 201	30995 ± 203	12126.23±78	248246.86 ±185	260373.09 ±263	4868.64 ±21	106093.46 ±125	110962.1 ±146
	Bayrampaşa	1058 ±5	18087 ±21	$19145 \pm 26$	9978.55±113	163678.41 ±92	173656.96 ±205	3159.18 ±9	78060.26 ±80	81219.44 ±89
Chlorogenic Acid	Sakız	569 ± 3	3197 ± 27	3766 ± 30	1354.22 ± 19	6745.67 ±152	8099.89 ±171	938.85 ±12	5501.01 ±31	6439.86 ±43
	Bayrampaşa	1263 ± 11	2379 ± 43	3642 ± 54	2867.01 ± 8	6994.26 ±93	9861.27 ±93	2089.29 ±7	4065.69 ±17	6154.98 ±24
Total Phenolic Acid	Sakız	2772.19 ± 5	33325.12 ±85	36097.31±90	4684.68 ± 11	58846.58 ±56	63531.26 ± 67	5246.84 ±27	70355.35±39	75602.19 ±66
	Bayrampaşa	3216.22 ± 3	20992.25 ±23	$24208.47 \pm 26$	5628.22 ± 17	53010.34 ±70	58638.56 ± 87	6093.64±15	$64859.02 \pm 46$	70952.66 ± 61
Total Flavonoid	Sakız	2011.53 ± 4	$1793.82 \pm 2$	$3805.35\pm 6$	1399.18 ± 5	1383.76 ± 18	2782.94 ± 23	1418.2 ± 8	$1536.25\pm26$	$2954.45\pm34$
	Bayrampaşa	1697.57 ± 8	3302.78 ± 17	5000.35 ± 25	$1515.66 \pm 3$	3201.65 ±7	4717.31 ± 10	1610.63±14	3123.2 ±9	4733.83 ± 23
Total Phenolic	Sakız	5302.32±6	35482.64 ±77	40784.96 ± 83	32988.54 ±85	219878.21±42	252866.75±127	29145.79±51	199242.11±87	228388.55±138
	Bayrampaşa	5514.46 ± 16	24438.14 ±38	29952.6 ± 54	35123.31 ±23	153277.12 ±69	188400.43 ±92	30309.56 ±34	156659.56±76	186969.12 ±110
Tot.Anthocyanin	Sakız	528.46± 1.2	$912.28 \pm 9.4$	1440.74±10.6	1167.89 ± 19	1612.73 ±86	2780.62 ±105	493.27 ±40	889.84 ±18	1383.11 ±58
	Bayrampaşa	498.36± 2.3	2091.42±11.2	2589.78±13.5	1041.57 ±33	4863.12±44	5904.69 ±77	510.38±19	2176.4 ±6	2686.78 ±25
Cyanidin Aglycon	Sakız	64.11± 6.3	92.73 ± 3.1	$156.84 \pm 9.4$	22.10 ±2	46.09 ±2	68.19±4	18.94 ±4	23.67 ±3	42.61 ±7
	Bayrampaşa	53.31± 5.9	$101.11 \pm 4.0$	$154.42 \pm 9.9$	19.78 ±1	46.19 ±4	65.97±5	16.88 ±1	18.9 ±11	35.78 ±12

<b>Table 4.</b> The Alterations in Major Phenolic Profiles of Boiled and Steamed Processed Artichockes(as $\mu$ g 100g <sup>-1</sup> )	
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		Raw	Frying
		Head	Head
Cynarin (1,3-dicaffeoylquinic acid)	Sakız	30995 ± 203	74078.21 ±155
	Bayrampaşa	$19145\pm26$	43650.52 ±103
Chlorogenic Acid (5- <i>O</i> -caffeoylquinic)	Sakız	3766 ± 30	8021.58 ±73
	Bayrampaşa	$3642 \pm 54$	7243.20 ±54
Total Phenolic Acid	Sakız	36097.31±90	65697.23 ±42
	Bayrampaşa	$24208.47 \pm 26$	42848.77 ±66
Top.Flavonoids (as Lutein-7-G)	Sakız	$3805.35 \pm 6$	$1452.42 \pm 50$
	Bayrampaşa	$5000.35 \pm 25$	$1953.26\pm23$
Total Phenolic (as Clg)	Sakız	$40784.96 \pm 83$	136218.56 ±97
	Bayrampaşa	29952.6 ± 54	104235.05 ±111
Total Anthocyanin (as C3G)	Sakız	1440.74±10,6	458.834 ±15
	Bayrampaşa	2589.78±13,5	849.108 ±4
Cyanidin Aglycon	Sakız	156.84 ± 9,4	29.89 ±5
	Bayrampaşa	154.42 ± 9,9	30.06 ±2

**Table 5**. The Alterations in Phenolic Profiles in Artichokes After Frying (as  $\mu$ g 100g<sup>-1</sup>)

**Table 6.** Antioxidant Activity Levels of Sakız and Bayrampaşa Artichoke Heads

Antioxidant Activity (mmol Trolox 100 g <sup>-1</sup> DW)							
Variety Raw Boiled Steamed Fried							
Sakız	$2.87 \pm 0.03$	18.87± 0.22	$27.92 \pm 0.83$	$15.04 \pm 0.58$			
Bayrampaşa $2.58 \pm 0.02$ $20.46 \pm 0.64$ $29.68 \pm 0.80$ $15.82 \pm 0.58$							

*p*<0.01; *n*=30

