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ORIGINAL RESEARCH

Evaluation of Aqueous Stem Bark Extract of Guiera senegalensis on Wistar Rats

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Abstract

Objective: Global herbal products however have enormous potential as natural drugs and are of vast commercial significance, are often processed and procured without being scientifically evaluated for their toxicity. This study evaluated the toxicological effects of *Guiera senegalensis* on wistar rats.

Material-Method: An acute toxicity evaluation was carried out to determine the LD_{50} of *Guiera senegalensis* stem bark extract where eleven (11) rats were used. Sub-acute toxicity was carried out to determine the effect of the plant extract on some liver and kidney function parameters and haematological parameters. For sub-acute toxicity studies, twenty (20) rats were randomly placed into 4 groups of 5 rats each. Group 2, 3 and 4 were orally treated with aqueous stem bark extract of *Guiera senegalensis* at a daily dose of 200, 400 and 800 mg/kg body weight (b.wt.) respectively for 28 days while group 1 served as a control group.

Results: Alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase and bilirubin levels increased significantly as dose increased. No significant increase in K^+ and urea was observed in group 2 and 3 treated with 200 and 400 mg/kg respectively. However, significant increase was observed in group 4 treated with 800 mg/kg. Na⁺ and Creatinine showed significant increase when compared with control group. Red blood cells, packed cell volume, and haemoglobin concentrations decreased significantly whereas a significant increase was observed in white blood cells with increase in dose respectively.

Conclusion: The aqueous stem bark extract of *Guiera senegalensis* have a dose-dependent toxic effect on liver, kidney and haematology of Wistar rats.

Keywords: Toxicological Evaluation, Guiera senegalensis, Liver, Kidney, Haematological Parameters

INTRODUCTION

Nearly all cultures use plants as a source of medicine from ancient times to present day, hence, a considerable percentage of people in both developing and developed countries use medicinal plants to remediate, alleviate or treat both human and animal diseases¹. As a consequence, medicinal plants are now of substantial significance due to the special characteristics they possess as a large source of therapeutic phytochemicals, that may lead to the discovery and development of novel drugs as most of the phytochemicals from natural sources such as phenolics and flavonoids have been reported to have positive impact on health and disease prevention². Ethnobotanical studies confirmed that indigenous plants are the main sources of traditional medicines³⁻⁴. Thus, there has been increase in herbal drugs usage globally with about 70% - 80% of the African population relying on nonconventional drugs which are predominantly of herbal sources both locally manufactured and imported⁴⁻⁵. However, most of these herbal drugs are often processed and procured without been scientifically evaluated for their safety.

The plant *Guiera senegalensis* a member of the family combretaceae is a tropical shrub widely distributed around the globe with high diversity in Africa and Asia growing on leached soils, fallows, and mostly on sandy soil and on very dry stations⁶ to a height of 3 to 5 m depending on the habitat⁷. *G. senegalensis* has been used to treat various illness where the same plant part is used to treat a number of different diseases and at the same time different parts of the plant used to treat the same disease⁷. It has been used to relief aches and pains and for treating fever and malaria^{8.9}. It has also been reported to possess healing properties against respiratory congestion and cough¹⁰, to ease breathe and treat bronchial disorder, severe diarrhea and

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dysentery^{9, 11}. It's safety has not been well documented. Hence this study evaluated the effects of aqueous stem bark extract of *G. senegalensis* on Wistar rats by determining the effects of the extract on haematological parameters, liver and kidney functions.

MATERIALS AND METHODS Plant material

Fresh stems of *G. senegalensis* were collected from Song Local Government Area, Adamawa state. The plant material was taxonomically identified by a Botanist in Botany Department of Adamawa State University, Mubi, Nigeria where the voucher specimen was deposited.

Experimental animals

Thirty-one (31) mature and healthy Wistar rats (both sexes) weighing about 180 g to 200 g obtained from the Animal Resource Unit, National Veterinary Research Institute (NVRI) VOM, Plateau State, Nigeria were used for the study. The animals were housed at room temperature in wired cages for one week acclimatization before the initiation of the experiment where a 12 h light/dark cycle was maintained. An ethical clearance for conducting the experiment on research animals was secured from the University Ethical Committee prior to the initiation of the experiment with an approval number of IACEC/ANP-A045/2021

Extraction of plant material

G. senegalensis stem bark was washed, peeled and dried for four weeks under shade and was then pulverized to powder form using mortar and pestle. Maceration method of extraction described by Abdullahi and Mainul¹² was used to extract the powdered *G. senegalensis* stem bark. The powdered plant material was soaked in distilled water and allowed to stand at room temperature where it was stirred every 24 h for a period of three (3) days in order to soften and break the cell wall of the plant to release the soluble phytoconstituents. After three days, the mixture was pressed and strained by filtration, and the solvent was evaporated using a water bath and crucible at 40° c until the extract was dried.

Phytochemical analysis

The qualitative phytochemical method described by Sofowara¹³ and Banu and Catherine¹⁴ was used to analyze the phytochemical constituents of aqueous *G. senegalensis* stem bark extract.

Acute toxicity

After one week of acclimatization according to criteria, the method described by Chinedu *et al.*¹⁵

was used for acute toxicity determination. The method is categorized into 3 separate stages with the result of each stage determining whether to further proceed to the next stage or end the process. The result of the final test was validated using a confidence (confirmatory) test. Eleven rats were used in total. At stage 1 four rats were divided into 4 groups of 1 rat each and were treated with 50 mg/kg, 200 mg/kg b.wt., 400 mg/kg b.wt., and 800 mg/kg of G. senegalensis stem bark extract. The rats were closely observed for 1 h post-administration and 10 min every 2 h space interval for 24 h. The testing proceeded to stage 2 if mortality was not recorded. In stage 2, three rats were divided into 3 groups of 1 rat each and were treated with 1000 mg/kg b.wt., 1500 mg/kg b.wt., and 2000 mg/kg b.wt. of the stem bark extract of G. senegalensis which were closely observed for 1 h postadministration and 10 min every 2 h space interval for 24 h. The testing proceeded to stage 3 if no mortality was recorded. In stage 3, three rats were divided into 3 groups of 1 rat each and were treated with 3000 mg/kg b.wt., 4000 mg/kg b.wt., and 5000 mg/kg b.wt. of G. senegalensis stem bark extract which were also observed closely for 1 h postadministration and 10 min every 2 h space interval for 24 h. Finally, 1 rat was used for the confirmatory stage and was treated with 5000 mg/kg b.wt. of the G. senegalensis stem bark extract and was observed closely for 1 h post-administration and 10 min every 2 hours space interval for 24 h.

Table 1. Doses for acute toxicity determination of aqueous G. senegalensis stem bark extract

Stages	Group 1	Group 2	Group 3	Group 4
1	50 mg/kg	200 mg/kg	400 mg/kg	800 mg/kg
2	1000 mg/kg	1500 mg/kg	2000 mg/kg	
3	3000 mg/kg	4000 mg/kg	5000 mg/kg	

Sub-acute toxicity

Three (3) doses were administered to rats for 28 days for sub-acute toxicity determination. The doses administered for sub-acute toxicity study were chosen based on the result acquired from the acute toxicity study. Twenty (20) rats were grouped into 4 groups of 5 rats each. The rats were grouped accordingly: group 1 received normal saline only, group 2 received 200 mg/kg b.wt. extract, group 3 received 400 mg/kg b.wt. extract and group 4 received 800 mg/kg b.wt. extract.

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Sample collection

After 28 days of daily administration, the rats were weighed and sacrificed by decapitation 24 h after the last dose was administered. The blood samples were collected and analyzed for haematological parameters based on the method described by Dacie and Lewis¹⁶, liver function parameters (ALT and AST) based on the method described by Rietman and frankel¹⁷, ALP based on the method described by Wright *et al.*¹⁸, while bilirubin based on the method described by Rifat²⁰. Cypress diagnostic kits were used for the analysis.

Statistical analysis

All data collected from the study were statistically recorded as mean \pm standard error of mean. Statistical differences between the mean values were determined using Analysis of Variance Test (ANOVA). Duncan's Multiple Range Test was used for comparison. IBM SPSS statistics software version 27 was used to analyze all data collected.

RESULTS

Phytochemical analysis

Phytochemical components of aqueous G. *senegalensis* stem bark extract are presented in Table 2. Phytochemicals tested included alkaloids, flavonoids, saponins, tannins, steroids and terpenoids. The result disclosed the presence of all the phytochemicals tested for.

Table 2. Phytochemical constituent of aqueous G.

 senegalensis stem bark extract.

Phytochemical	Inference
Alkaloid	+
Flavonoid	+
Saponin	+
Tannins	+
Steroid	+
Terpenoid	+

Key: + = present

Acute toxicity

Oral administration of aqueous *G. senegalensis* stem bark extract did not produce mortality up to a dose level of 5000 mg/kg b.wt. The bodyweight, food and water consumption of the rats did not show any significant difference when compared with the control group. Signs of toxicity such as aggression, depression, writhing, diarrhea and hypermotility were not recorded in rats used for oral LD_{50} determination when compared with control. **Sub-acute toxicity**

Table 3 shows the serum liver function parameters alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP) and bilirubin of rats treated with aqueous stem bark extract of *G. senegalensis* for 28 days. The result showed that the levels of liver parameters significantly (p<0.05) increased with an increase in dose of the extract when compared with the control.

Table 3. Effect of aqueous stem bark extract of G. senegalensis on liver function.

Parameters Groups	Alanine aminotranferase (IU/L)	Aspartate aminotransferase (IU/L)	Alkaline phosphatase (IU/L)	Total bilirubin (mmol/L)	Conjugated bilirubin (mmol/L)
1. Control	25.66 ± 1.20^{a}	$94.33{\pm}0.88^a$	37.33 ± 0.33^{a}	$7.63\pm0.15^{\text{a}}$	2.03 ± 0.03^{a}
2. 200 mg/kg b.wt	35.00 ± 0.58^{b}	103.33 ± 0.33^{b}	51.00 ± 1.00^{b}	8.63 ± 0.15^{b}	2.60 ± 0.12^{b}
3. 400 mg/kg b.wt	39.00 ± 0.88^{c}	121.66 ± 1.20 ^c	76.00 ± 1.16^{c}	12.33 ± 0.26^{c}	4.000 ± 0.58^{c}
4. 800 mg/kg b.wt	$41.00 \pm 1.15^{\rm c}$	141.33 ± 0.88^{d}	80.33 ± 1.45^{d}	13.50 ± 0.15^{d}	5.97 ± 0.09^{d}

All values are presented as mean \pm SEM. Different superscripts down the column indicates that they are significantly different at (p<0.05), n=5

Values for AST, ALP and bilirubin of all the groups vary significantly with each other. Table 4 shows the serum kidney function parameters (urea, creatinine, Na⁺ and K⁺) of rats treated with aqueous *G. senegalensis* stem bark extract. Values of Na⁺ and creatinine showed a significant (p<0.05) increase when compared with the control group whereas K^+ and urea values for group 2 and 3 did not differ significantly (p<0.05) when compared with the control group. However, group 4 that received 800 mg/kg b.wt. showed a significant increase when compared with the control.



Table 4.	Effect of ac	nueous stem	bark extract	of G .	senegalensis	on kidnev	function
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Parameters Groups	Na+ (mmol/L)	K ⁺ (mmol/L)	Creatinine (mmol/L)	Urea (mmol/L)
1. Control	137.67 ± 0.33^a	4.17 ± 0.07^{a}	25.00 ± 0.58^a	7.20 ± 0.12^{a}
2. 200 mg/kg b.wt	148.33 ± 0.67^{ab}	$4.40\pm0.06^{\rm a}$	36.33 ± 0.33^{b}	7.43 ± 0.09^{a}
3. 400 mg/kg b.wt	148.67 ± 0.33^{ab}	$4.20\pm0.12^{\rm a}$	45.66 ± 0.20^{c}	$7.60\pm0.12^{\rm a}$
4. 800 mg/kg b.wt	149.33 ± 0.33^b	$5.07\pm0.03^{\text{b}}$	54.00 ± 1.16^{d}	8.67 ± 0.15^{b}

All values are presented as mean \pm SEM. Different superscripts down the column indicates that they are significantly different at (p<0.05), n=5

Table 5 shows the haematological parameters, packed cell volume (PCV), haemoglobin (Hb), red blood cells (RBC) and white blood cells (WBC) of rats treated with aqueous *G. senegalensis* stem bark

extract. PCV, Hb and RBC significantly decreased with an increase in dose of the extract while WBC significantly increased with an increase in dose when compared with control.

Table 5. Effect of aqueous stem bark extract of G. senegatensis on macmatological multiple	Table 5. Effect of	of aqueous stem	bark extract of	f G. senegalensis	on haematological indice
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Parameters Groups	Packed cell volume (%)	Haemoglobin (g/dL)	White blood cells (×10 ⁹ /L)	Red blood cells (×10 ² /L)
1. Control	$36.00\pm1.15^{\rm c}$	$12.43\pm0.80^{\rm c}$	$9.80 \pm 1.17^{\rm a}$	$6.37\pm0.26^{\rm c}$
2. 200 mg/kg b.wt	32.66 ± 2.03^{ab}	11.43 ± 0.38^{bc}	12.93 ± 0.49^{b}	5.87 ± 0.26^{b}
3. 400 mg/kg b.wt	32.33 ± 1.20^{ab}	10.53 ± 0.30^{ab}	13.77 ± 0.19^{bc}	4.90 ± 0.46^{ab}
4. 800 mg/kg b.wt	30.33 ± 0.33^a	9.57 ± 0.37^{a}	$14.47\pm0.32^{\rm c}$	4.12 ± 0.28^{a}

All values are presented as mean \pm SEM. Different superscripts down the column indicates that they are significantly different at (p<0.05), n=5

DISCUSSION

Acute toxicity

The determination of LD₅₀ is usually the first step in evaluating and screening novel drugs. It is an initial assessment and evaluation of toxic characteristics and manifestation of test substance²¹. Oral LD₅₀ of aqueous stem bark extract of G. senegalensis was indeterminable up to a dose level of 5000 mg/ kg, thus the LD_{50} is greater than 5000 mg/kg. This may indicate that the aqueous G. senegalensis stem bark extract of is safe via the oral route. This is similar to the report of Moo et al.²² in which no mortality was recorded up to a dose level of 5000 mg/kg of N-hexane extract of Leptadenia hastata.

Effect of aqueous stem bark extract of *G.* senegalensis on liver function

Liver is an important organ that performs different varieties of biochemical activities such as synthetic and excretory functions. Therefore, no single biochemical test is capable of identifying or detecting the general function of the liver²³. The serum ALT and AST reflect the hepatocellular injury, serum ALP reflects the impaired bile excretion and bile flow, while the serum total and conjugated bilirubin represent the metabolic functions of the liver²⁴.

The observed significant increase in the liver function indices of the rats treated with aqueous *G*. *senegalensis* stem bark extract may indicate that the aqueous stem bark extract of *G*. *senegalensis* is dose dependently toxic to the liver. The increased levels of ALT may indicate damage to hepatocytes that results to the release of the enzymes into the circulation, increased ALP may indicate the effect of the stem bark extract that leads to both

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intrahepatic and extrahepatic obstruction of bile flow and the increased bilirubin level may indicate haemolysis and overproduction of bilirubin induced by the stem bark extract of *G. senegalensis*. This result is consistent with the report of Ashafa²⁵ in which the aqueous leaves extract of *Felicia muricata* Thunb. affected the liver function parameters of Wistar rats.

Effect of aqueous stem bark extract of G. senegalensis on kidney

Serum creatinine concentration has been used for assessment and evaluation of kidney function which begins to rise only when the glomerular filtration rate (GFR) has by one-half diminished and thereafter the rise of creatinine is exponential to decline in GFR²⁶. Urea, the by-product of protein and amino acid breakdown produced by the liver which is distributed throughout the intracellular and extracellular fluid is filtered out from the blood by the glomeruli in the kidney and is partially being reabsorbed with water. It is another frequent biochemical parameter for estimating kidney function which is useful in the differential diagnosis of acute and pre-renal conditions²⁷. Electrolytes are negatively and positively charged ions that are found within extracellular fluids and cells, blood and plasma. Test for serum electrolytes such as measurement of sodium, and potassium is used to evaluate renal functions and comprehensive metabolic biochemistry profiles.

The increased Na⁺ may indicate that the aqueous G. senegalensis stem bark extract causes alteration in osmotic pressure that leads to dehydration and a consequent increase in Na⁺. The increased K⁺ may indicate the poor function of the kidney thus may lead to abnormal or sometimes fatal cardiac arrhythmias²⁸. The increase in the K^+ level may indicate that the membrane channels were affected by the stem bark extract or the plant stem bark may have a hyperkalemic effect. Hyperkalemia occurs most often in renal failures leading to declined potassium excretion²⁹⁻³⁰. The increased serum creatinine and urea concentration may indicate that there is decreased GFR by the kidney due to the effect of the aqueous G. senegalensis stem bark extract. This result is not in agreement with the reports of Unuofin³¹ in which the whole-plant aqueous extract of Vernonia mespilifolia Less. did not affect the biochemical parameters of Wistar rats.

Effect of aqueous stem bark extract of *G.* senegalensis on haematological parameters

Haematological parameters valuable are in observing and monitoring the toxicity of substances. RBCs are involved in the transport and distribution of oxygen in the body and transports CO_2 to the lungs; WBCs functions to defend the body by phagocytosis against foreign invaders, fight infections and to produce, transport and distribute antibodies throughout the body during immune response³².

The decrease in PCV and Red blood cells count may indicate that the stem bark extract of the plant may have haemolytic effect as reflected by the increased total bilirubin. This may lead to hyperkalemia and consequently disturbances in acid-base balance when administered at higher doses. The decrease in the level of haemoglobin may indicate anaemia as a result of erythrocyte haemolysis caused by the aqueous G. senegalensis stem bark extract which may lead to decreased oxygen transport to tissues as well as transport of CO₂ back to the lungs. The increased WBC may indicate that the plant may have some toxic compounds which trigger the immune response leading to the production of more immune cells. This study supports the report of Ilham *et al.*³³ in which the leaves of Ambrosia maritima affected the haematological parameters of Nubian goats.

This study is limited to four functional parameters of liver and kidney and haematological parameters. **CONCLUSION**

In conclusion, considering the serum level of biochemical parameters and the haematological parameters of the experimental rats treated with different oral doses of aqueous *Guiera senegalensis* stem bark extract, it suggests that the stem bark extract of the plant may be toxic especially at higher doses and longtime exposure.

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REFERENCES

- 1. Akerele O, Heywood V, Synge H. *Conservation of medicinal plants*. United States of Amarica Cambridge University Press, New York. 2009.
- 2. Azwanida N. N. A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation. *Medicinal and Aromatic Plants*. 2015;6.
- 3. Qureshi R, Ghazanfar SA, Obied H, Vasileva V, Tariq MA. Ethnobotany: A Living Science for Alleviating Human Suffering. *Evidence-Based Complementary and Alternative Medicine*. 2016;1-3.
- Buwa-Komoreng LV, Mayekiso B, Mhinana Z, Adeniran AL. An Ethnobotanical and Ethnomedicinal Survey of Traditionally Used Medicinal Plants in Seymour, South Africa: An Attempt toward Digitization and Preservation of Ethnic Knowledge. *Pharmacognosy Magazine*. 2019;15(60), 115-123.
- 5. Edebi NV, Gideon OA. Evaluation of pharmacognostical parameters and heavy metals in some locally manufactured herbal drugs. *Journal of Chemical and Pharmaceutical Research*.2011;3(2), 10.
- 6. Mubarak SH, Hassan E, Ahmed S, Eltayeb F, Reem HAA. Review on the Taxonomy, Ethnobotany, Phytochemistry and Pharmacology of *Guriea senegalensis* J.F.Gmel. (Combretaceae). *Medicinal & Aromatic Plants*. 2017;5.
- 7. Aimé AS, Kirti P, Drissa D, Lassine S, Jean CC, Gilles F, Pierre C. An ethnobotanical and phytochemical study of the African medicinal plant *Guiera senegalensis* J. F. Gmel. *Journal of Medicinal Plants Research*. 2011;5(9), 1-13.
- 8. Fiot J, Ollivier E, Timon-David P, Balansard. G. *Guiera senegalensis* J.F Gmel. (combrataceae) (S. G. Pandalai, Ed.) *Trivandrum India*: Research Signpost. 2004;2
- Zakawa NN, Akesa TM, Timon D, Yusuf CS, Magga B, Jacob GF. Ethnobotanical survey and phytochemical studies of Guiera senegalensis lam. in Mubi Local Government of Adamawa State. World Journal of Pharmaceutical Research. 2018;7(14), 12.
- 10. Alshafei NK, Ahmed SA. Nour. Preliminary Observations on the Uses of *Guiera Senegalensis* as a Traditional Medicinal Plants in Western Kordufan, Sudan. *International Journal of Applied and Pure Science and Agriculture*. 2016;2, 42-48.
- 11. Diatta W, Fall AD, Dieye AM, Faty S, Bassene E, Faye B. Experimental Evidence of Against Cough Activity of Total Alkaloids from *Guiera Senegalensis* Lam. in Guinea Pig. *Darkar Medical*. 2007;52(2), 130-134.
- 12. Abdullahi RA, Mainul H. preparation of medicinal plants. Basic extraction and fractionation procedures for experimental purposes. *Journal of Pharmacy and BioAllied Sciences*. 2020; 12(1), 1-16.
- 13. Sofowora EA. Medicinal plants and traditional use in Africa. Ibadan, Nigeria: Spectrum books Ltd. 2006
- 14. Banu KS, Catherine L. General Techniques Involved in Phytochemical Analysis. *International Journal of Advanced Research in Chemical Science*. 2015;2(4), 25-32.
- 15. Chinedu E, Arome D, Ameh FS. A New Method for Determining Acute Toxicity in Animal Models. *Toxicology International*. 2013;20(3), 1-6.
- 16. Dacie JV, Lewis SM. practical haematology (12th ed.). (C. Livingston, Ed.) London: Elsevier. 2016
- 17. Rietman S, Frankel SA. (1957) colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *American Journal of Clinical Pathology*. 1957;28. 56–63
- 18. Wright PJ, Leathwood PD, Plummer DT. Enzymes in rat's urine: Alkaline phosphatase. Enzymology. 1972; 42. 317-327
- 19. Penhaker M, Kasik V, Hrvolova B. Advanced Bilirubin Measurement by a Photometric Method. *Elektronika Ir Elektrotechnika*. 2013; 19(3). 47-50.
- 20. Rifai, N. Tietz Textbook of Clinical Chemistry and Molecular Diagnostics. 2018; 6th Edition
- 21. Akhila JS, Deepa S, Alwar MC. Acute Toxicity Studies and Determination of Median Lethal Dose. *Current Science*. 2007;93(7), 917-920.
- 22. Moo A, Jacks TW, Garba SH, Dibal N, Ojo P. Evaluation of Acute Oral Toxicity Induced by N-hexane Extract of Leptadenia hastata Leaves in Wistar Rats. *International Journal of Veterinary Sciences and Animal Husbandry*. 2019; 4(1): 40-44
- 23. Thapa B, Walia A. Liver Function Tests and their Interpretation. Indian Journal of Pediatrics. (2007;74, 1-9.
- 24. Hasan FA, Owyed S. Interpretation of liver chemistry tests. *Bulletin of the Kuwait Institute for Medical Specialization*. 2003;2, 27-31.
- 25. Ashafa AOT, Yakubu MT, Grierson DS, Afolayan AJ. Toxicological evaluation of the aqueous extract of *Felicia muricata* Thunb. leaves in Wistar rats. *African Journal of Biotechnology*. 2019; 8(6), 949-954.
- 26. Korhonen PE. How to Assess Kidney Function in Outpatient Clinics. *The International Journal of Clinical Practice*. 2014;69(2), 156–161.
- Gowda S, Desai PB, Kulkarni SS, Hull VV, Math AA, Vernekar SN. Markers of Renal Function Tests. North American Journal of Medical Sciences. 2010;2(4), 170-173.
- 28. Mushiyakh Y, Dangaria H, Qavi S, Ali N, Pannone J, Tompkins J. Treatment and pathogenesis of acute hyperkalemia. *Journal of Community Hospital Internal Medicine Perspectives*. 2012; 1(4), 7372.

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- 29. Soar J, Perkins DG, Abbas G, Alfonzo A, Barelli A, Bierens JLM, Brugger JLMJ, Deakin HD, Dunning DC, Georgiou J, Handley M, Lockey JA, Paal JD, Sandroni P, Thies C, Zideman K, Nolan JP. European Resuscitation Council Guidelines for Resuscitation 2010 Section 8. Cardiac arrest in special circumstances: Electrolyte abnormalities, poisoning, drowning, accidental hypothermia, hyperthermia, asthma, anaphylaxis, cardiac surgery, trauma, pregnancy, electrocution. *European Resuscitation Council. Elsevier Ireland Ltd* 2010; 81, 1400–1433.
- 30. Palmer FB, Clegg JD. Diagnosis and treatment of hyperkalemia. *Cleveland Clinic Journal of Medicine*. 2017; 84(12), 193-194.
- 31. Etim NN, Williams ME, Akpabio U, Offiong EE. Haematological Parameters and Factors Affecting Their Values. *Science and Education Centre of North America*. 2014;1(2), 37-47.
- 32. Unuofin OJ, Otunola AG, Afolayan AJ. Evaluation of acute and subacute toxicity of whole-plant aqueous extract of *Vernonia mespilifolia* Less. in Wistar rats. *Journal of Integrated Medicine*. 2018.
- 33. Ilham MOA, Mohammed AS, Halima MO, Ibtehal MA. Ahmed. Toxicological effects of Ambrosia maritima in Nubian goats. *Journal of Plant and Environmental Research*. 2016;1(1):0001-0010