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Phytochemicals and Antioxidant Activities in Methanol Extracts of Endemic *Haplophyllum* Species from Türkiye

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Abstract

The aim of the current study is to determine the total flavonoids, phenolics and antioxidant activities of methanol extracts from aerial parts of four endemic Haplophyllum species to Türkiye (H. myrtifolium, H. vulcanicum, H. pumiliforme, and H. sahinii). There are two populations collected from different regions belonging to *H. myrtifolium* and *H. pumiliforme*. Antioxidant activities were measured by radical scavenging activity such as the 2, 2'-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) and 2, 2-diphenyl-1-picrylhydrazyl (DPPH), and compared with synthetic standards such as trolox, ascorbic acid and butylated hydroxytoluene (BHT). The highest total bioactive contents were recorded as H. sahinii for total phenolic content (66.4 mg GAE/g extract) and H. myrtifolium for total flavonoid content (40.8 mg QE/g extract), but H. vulcanicum displayed the lowest amount for both contents (38.0 mg GAE/g extract and 34.5 mg QE/g extract, respectively). Among the species, H. myrtifolium exhibited the strongest DPPH and ABTS radical scavenging activity followed by H. pumiliforme, H. sahinii and H. vulcanicum. All Haplophyllum species showed higher antioxidant activity for these two radical scavenging activities than trolox and BHT. The phytochemicals and antioxidant activities in methanol extracts from these endemic Haplophyllum species is reported for the first time. The diversity of the findings is to be discussed as likely consequence of the different species and regions.

Keywords: Haplophyllum L, phenolic, flavonoids, DPPH, ABTS

1. INTRODUCTION

Haplophyllum, belonging to Rutaceae family, herbaceous perennial and fragrant plants is a genus containing approximately 68 species, and has the most species diversity in the flora of Türkiye and Iran in terms of its spread over the world [1]. Türkiye is an important gene center for the Haplophyllum genus which is represented in the flora of Türkiye by 17 taxa belonging to 14 species with 52% endemism

Recently, [2]. two new species (Haplophyllum sahinii and H. ermenekense) was described by Tugay and Ulukuş, Ulukuş Türkiye has and Tugay [3, 4], 18 Haplophyllum taxa, 11 (58%) of which are endemic. This genus taxa are foetid perennial herbs, which grows mainly on rocky hills, steppes, slopes, rocky place on limestone, especially near pine forests, or sandy soils [2].

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Members of the Rutaceae family are of great economic importance, including wood, food, cosmetic and medicinal uses [5]. Haplophyllum species of this family are traditionally used actively in the treatment of different diseases in many countries. It is used in the treatment of malaria, rheumatoid arthritis and gynecological diseases in Saudi Arabia [6]. The herb part is used as an antispasmodic in the treatment of allergic rhinitis and gynecological diseases, asthma and respiratory distress in Sudan [7], while its leaves are used for skin infections in Oman [8]. Infusion samples of the herbal part are used to treat gynecological problems and digestive problems such as rheumatoid arthritis as well as constipation and diarrhea Moreover, Haplophyllum species. [9]. analyzed in previous studies, have been reported to exhibit incredible levels of biological activities including antimicrobial [10, 11], antioxidant [12, 13], antiinflammatory [14], and especieally anticancer [15-17]. The potential of these activities is based on phytochemicals in the plant. Many scientists reported that various Haplophyllum species contained important characteristic classes of phytochemical such phenolics, flavonoids, flavonols, as coumarins, alkaloids important and compounds of lignans [4, 15, 18].

Many compound classes or compounds with high antioxidant properties are preferred for preservative and additive purposes in food. industries such medicine, as pharmacology and cosmetics. These compounds are mostly synthetic products due to their cheapness and quick availability. As a result of the researches, it was revealed that synthetic compounds have toxic and carcinogenic effects, and instead, natural products/preparations with high antioxidant activity have become popular [19, 20]. Natural products of plant origin provide alternatives to synthetic antioxidants. Therefore, natural antioxidant products have been developed from aromatic plants, spices, and fruit powder and are still being developed [21, 22].

The biosynthesis of these phytochemicals in plant and therefore their biological activities are affected by external factors such as environmental factors (soil, light intensity and climatic conditions) [23], as well as by internal factors such as the biotype and chemo-type of the plant [24], physiological and genetic aspects [25]. Therefore, it is an important first step to identify the preparation/extract or a specific compound that can be used for commercial purposes from nature.

of The high pharmaceutical effects *Haplophyllum* species suggest that there may be species with high antioxidant activity among the species. In this context, this study aims to investigate the antioxidant activities of endemic Haplophyllum species to Türkiye, of which there are very few studies, and to compare them with synthetic antioxidant standards. The results of this study are important in terms of the use of Haplophyllum species as a potential source of natural antioxidants in food and pharmaceutical products.

2. MATERIALS AND METHODS

2.1. Material

The erial parts of Haplophyllum species (H. myrtifolium, H. vulcanicum, H.pumiliforme and H. sahinii) respresenting a total of 40 shoots were collected according to completely randomized design at full flowering period (Figure 1). The species were identified by Prof. Dr. Osman Tugay, Faculty of Farmacy, Department of Pharmaceutical Botany, Selçuk University, Konya, Türkiye. The locations belonging to each taxon were recorded as following (Table 1).

2.2. Extraction

The dried and finely ground samples (about 4g) of the aerial parts of *Haplophyllum* species were extracted in methanol at 40°C for 24 h. The resulting solutions were filtered through whatman paper and solvent was

separated with a rotary evaporator (Heidolph, laborota 4000), and extract yields were calculated as %. Then, extracts were

dissolved in methanol. Each extraction process was repeated three times.



Figure 1 General view of habit and flowers of endemik *Haplophyllum* species (*H. myrtifolium* (A&B), *H. vulcanicum* (C&D), *H. pumiliforme* (E&F) and *H. sahinii* (G&H))

Plant Name	Abbreviated Names	Hazard Category ¹	Altitude (m)	Collection Site	Collector Number
Haplophyllum myrtifolium Boiss.	HM1	EN	1070	C4 ² Konya; Çumra, Apasaraycık Köyü, taşlı yerler	OT-9264- DU ³
	HM2	EN	1090	C4 Konya; Çumra, Apasaraycık Köyü	OT-7392-DU
<i>Haplophyllum vulcanicum</i> Boiss. & Heldr.	HV2	VU	1200	C4 Karaman; Karadağ	OT-9614-DU
Haplophyllum pumiliforme HubMor. & Reese	HP1	VU	1450	C3 Konya; Derebucak, Soğukoluk yolu	OT-7495-DU
	HP2		1470	C3 Konya; Derebucak	OT-7481-DU
Haplophyllum sahinii Tugay & Ulukuş	HS	EN	1090	C4 Konya; Çumra, Apasaraycık-Apa köyü, kayalık alan	OT-7410-DU

Table 1 Habitats of H. myrtifolium, H. vulcanicum, H. pumiliforme and H. sahinii from Türkiye

2.3. Phytochemical Contents

2.3.1. Total phenolic

Total phenolic content in the methanol extracts of *Haplophyllum* species will be measure using the Folin–Ciocalteu reagent method as described by Yaman et al. [13].

The total phenolic contents of the samples were expressed as mg gallic acid equivalent (GAE) / g extract according to the equation obtained from the standard gallic acid graph. The experiment was done in triplicates with two replicates.

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2.3.2. Total flavonoid

Total flavonoid content in the methanol extracts of *Haplophyllum* species will be determine using colorimetric method as described by Yaman et al. [13]. The total flavonoid contents will be calculate from the calibration curve and express as mg quercetin equivalent (QE) / g extract according to the equation obtained from the standard quercetin graph. The experiment will be done in triplicates with two replicates.

2.4. Radical Scavenging Activity

2.4.1. DPPH free radical scavenging activity

Measurement of DPPH (2, 2-diphenyl-1picryl-hydrazyl) radical scavenging properties of the methanol extracts from Haplophyllum species will be carry out according to the method described by Yaman et al. [13] with some modifications. Trolox, Ascorbic acid (AA), butylated hydroxytoluene (BHT) will be used as positive control. The experiments will be doing in triplicates with two replicates. The results of the radical scavenging activity were calculated according to the following equation as % inhibition of the DPPH radical.

% inhibisyon =
$$\frac{(Abscontrol - Abssample)}{Abscontrol} \times 100$$

2.4.2. ABTS radical cation scavenging activity

Measurement of ABTS (2,2'-azino-bis-3ethylbenzothiazoline-6-sulfonic acid) radical scavenging properties of the methanol extracts from *Haplophyllum* species will be carry out according to the method described by Yaman et al. [13] with some modifications. Trolox and AA will be used as positive control. The experiments will be doing in triplicates with two replicates. Results of radical scavenging activity were denoted as % inhibition of ABTS radical. The % inhibition of ABTS radical cation scavenging activity was calculated according to the following equation:

$$\% inhibisyon = \frac{(Abscontrol - Abssample)}{Abscontrol} \times 100$$

2.5. Statistical Analysis

The findings were statistically analyzed using one-way ANOVA in SPSS statistical program, and comparison of the means was evaluated by Duncan's multiple range tests at a significance level of 0.05. Data were given as the mean \pm standard deviation.

3. RESULTS AND DISCUSSIONS

3.1. Extract Yield

The methanol extract yields of *Haplophyllum* species used in the project were analyzed and are given in Table 2 as %.

Table 2 The methanol extract yields of endemic
Haplophyllum species from Türkiye

Species	Extract Yield (%)	Standard Error
HM1	13.98 ^a	1.54
HM2	5.54 ^e	3.18
HV1	9.92 ^b	2.70
HP1	6.91 ^d	0.73
HP2	8.28 ^c	0.51
HS	9.32 ^b	2.43

HM, H. myrtifolium; HV, H. vulcanicum; HP, H.pumiliforme; HS, H. sahinii

Generally, in the extraction of plants, methanol solvent provides higher extract yield than other solvents [26, 27]. Yaman et al. [13] investigated ethanol extracts of similar species in their study, and reported their extract yields lower than the methanol extract yield in the current study.

HM1 gives the best extraction yield an average of 13.9%, while HM2 collected from a second region had the lowest yield (5.54 % on average). HV and HS had a statistically similar extract yield with 9.92% and 9.32% whereas HP from two regions was lower with 8.28% - 6.9%.

3.2. Phytochemical contents

The methanol of endemic extracts collected Haplophyllum species from different localities were investigated for their phytochemical contents such as total phenolic and flavonoid contents (Table 3). Results was calculated from the calibration curve (\mathbf{R}^2 = 0.999 for total phenolic content and $R^2 =$ 0.9997 for total flavonoid content). Differences for the species were showed in findings of this study. The great distinction between the same species collected from different regions appears due to different environmental and climatic conditions [13].

	Türkiye	
Species	Total phenolic content (mg GAE/g extract)	Total flavonoid content (mg QE/g extract)
HM1	48.0 ± 0.4^{d}	39.7 ± 0.2^{b}
HM2	56.1±0.8°	$40.8{\pm}0.2^{a}$
HV	$38.0{\pm}0.3^{\mathrm{f}}$	$34.5{\pm}0.1^{\rm f}$
HP1	40.1 ± 0.2^{e}	$35.3{\pm}0.1^{d}$
HP2	$60.1{\pm}1.0^{b}$	36.1±0.2°
HS	66.4±0.2ª	35.0±0.2 ^e
× × × × × ×		

HM, H. myrtifolium; HV, H. vulcanicum; HP, H.pumiliforme; HS, H. sahinii

The results revealed that the HS is very rich in phenolic compounds with 66.4 mg of GAE/g of extract, whereas HV is lowest with 38.0 mg of GAE/g of extract (Table 2). Interestingly, Yaman et al. [14] reported that ethanol extracts of the Haplophyllum vulcanicum (HV) species contained higher total phenolic than other Haplophyllum species. HV may differ from other species in phytochemical terms of content or biosynthesis ability, or affected by environmental factors.

When Table 3 was examined, the total flavonoid amounts of extracts varied from 34.5 to 40.8 mg QE/g extract. The highest levels of the total flavonoid were found in HM2 and HM1, respectively. Yaman et al.

[13] also reported a similar finding for ethanol extracts of *Haplophyllum myrtifolium* (HM).

The collection of species from different regions affected their phytochemical contents and compositions, especially total phenolic content (Table 2). Different results have been observed in populations at the different regions of one species. As a result of these populations belonging differences, to different regions of each species have ecological conditions (abiotic and biotic differentiators) and habitat (rocky, slopes etc. and diversity of flora). Such differences have also been identified by many researchers [28-31]. Rawat et al. [30] indicated that total phenol contents among the populations of Hedychium *spicatum* ranged were а significantly significant different. The present findings indicate to have a significant effect on different species and regions for total bioactive components.

3.3. Radical Scavenging Activity

Various phytochemicals (secondary metabolites) of plants such as flavonoids, polyphenols and other phenolics, tannins are the main group of components that serve as primary free radical scavengers [32, 33]. The obtained extracts from endemic Haplophyllum species collected from different localities were investigated for their two radical scavenging activities, namely ABTS and DPPH. Results were also expressed as % inhibition (Figure 2 and Figure 3).

The DPPH radical is largely used in the evaluation of free radical scavenger activity due to the ease of the reaction. When the DPPH radical is cleaned by a compound of antioxidant via hydrogen donation to form a stable DPPH-H molecule, the color of the solution is return from purple to yellow. All extracts in current study were determinate to reduce the stable violet DPPH radical to yellow. DPPH antioxidant activity ranged between 43.9-86.5% at a concentration of 100 μ g of all the assessed extracts and standards

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(Figure 2). Among extracts of *Haplophyllum* species, HP1, HP2 and HM1 exhibited maximum DPPH free radical scavenging activity and statistically the same DPPH activity with 85.0-85.1%, followed by HM2 (83.1%) and HS (71.7%) and HV (68.3%). The scavenging effect of different species on the DPPH radical for methanol solvent generally decreased in the order of HP \geq HM >HS >HV (Figure 2).



Figure 2 The DPPH free radical activities of methanol extracts of endemic *Haplophyllum* species from Türkiye (HM, *H. myrtifolium*; HV, *H. vulcanicum*; HP, *H.pumiliforme*; HS, *H. sahinii*)

The ABTS radical is blue, but transforms from blue to the colorless form through an antioxidant compound. As seen in Figure 3, ABTS radical scavenging activity ranged between 39.5%-89.9% at a concentration of 200 µg of all the assessed extracts and standards.

All values of methanol extracts from *Haplophyllum* species were statistically in different groups. Among *Haplophyllum* species, samples at both different regions of HM displayed the highest ABTS activity (HM1:76.5% and HM2:71.1%), followed by HP (HP2: 66.7%, HP1: 61.3%) and HS (56.5%) and HV (47.2%).



Figure 3 The ABTS free radical activities of methanol extracts of endemic *Haplophyllum* species from Türkiye (HM, *H. myrtifolium*; HV, *H. vulcanicum*; HP, *H.pumiliforme*; HS, *H. sahinii*)

Interestingly, DPPH and ABTS activities of all *Haplophyllum* species were lower than AA as the synthetic antioxidant standard, but stronger than the trolox standard. Also, DPPH activities of all *Haplophyllum* species had the higher than BHT standard. This finding indicates that the analyzed *Haplophyllum* species exhibit very powerful radical scavenging activity. In the other hand, the results between different localities of the species were close to each other.

Antioxidant activities of these *Haplophyllum* species have been reported very little in some previous studies [13, 34], but, for the first time, methanol extracts of endemic *Haplophyllum* species from Türkiye were compared among themselves and with synthetic standards in this study.

Bioactive compounds defined as secondary metabolites in plants have been taken the evidence as natural functional components, and various medicines have been developed all over the World [35, 36]. These compounds include total flavonoids, other phenolic compounds (phenolic acids, tocopherols, stilbenes, alcohols etc.) and polyphenolics (condensed and hvdrolvsable tannins. vitamins, saponin, lignin), terpenoids,

carotenoids, essential oils [37]. These compounds could use as a potential source of natural antioxidants [38]. Many researchers reported that found a correlation between the antioxidant activity and phenolics, in particular flavonoids [39-42]. Moreover, flavonoids are the most important and abundant polyphenols, more than 5000 reported up to today [43]. So, former studies have indicated that the amount of bioactive compounds in plants and their antioxidant activities depend on both biological factors (genetic, organ etc.) and environmental (precipitation, temperature, altitude, light intensity etc.) conditions [44]. Also, stress conditions may induce various flavonoid biosynthetic genes. The biotic and abiotic stresses such as drought, temperature, wounding, nutrient deprivation, metal toxicity and can increase the levels of flavonoids in the plants as a part of their defense strategy [45]. So, the antioxidant activity of flavonoids is principal gone on their ability to donate the electrons or hydrogen atoms [46]. The variety of phenolic and flavonoid compounds is as important as amount of the compounds, because effect of antioxidant activity of each compound is different.

4. CONCLUSIONS

Less than 10% of the world's biological diversity has been assessed for potential biological activity and there are many more natural compounds awaiting exploration to achieve this natural chemical variety. When both radical scavenging activities are taken into consideration, HM and HP have been exhibited the strongest antioxidant activity, especially the antioxidant activities of both species are very close to those of ascorbic acid. The findings suggest that these endemic species show much stronger antioxidant activities, involved significantly high levels of total phenolic and flavonoid contents, and could be a potential source of natural antioxidants. However, there is little research about antioxidant activities and total bioactive components of the species evaluated in this study. Further chemical investigations are

required to isolate the elements of active phenolic and flavonoid components of the plants that show a broad spectrum of pharmacological activity.

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The Declaration of Conflict of Interest/ Common Interest

No conflict of interest or common interest has been declared by the authors.

The Declaration of Ethics Committee Approval

This study does not require ethics committee permission or any special permission.

The Declaration of Research and Publication Ethics

The authors of the paper declare that they comply with the scientific, ethical and quotation rules of SAUJS in all processes of the paper and that they do not make any falsification on the data collected. In addition, they declare that Sakarya University Journal of Science and its editorial board have no responsibility for any ethical violations that may be encountered, and that this study has not been evaluated in any academic publication environment other than Sakarya University Journal of Science.

REFERENCES

- C. C. Townsend, "Haplophyllum A. Juss. In: Davis, P.H (Ed.) Flora of Türkiye and the Aegean Islands. vol 2," Edinburgh University Press, Edinburgh, pp. 496–506, 1967.
- [2] C. C. Townsend, "Taxonomic revision of the genus *Haplophyllum* (Rutaceae). In: Hooker's icones plantarumvol 40

parts 1," Bentham-Moxon Trustees Royal Botanical Gardens, Kew, 1986.

- [3] O. Tugay, D. Ulukuş, "*Haplophyllum sahinii* (Rutaceae), a new species from Central Anatolia (Turkey),"Phytotaxa, vol. 297, No. 3, pp. 265-272, 2017.
- [4] D. Ulukuş, O. Tugay. O. *"Haplophyllum ermenekense* (Rutaceae), a new species from Turkey," PhytoKeys, (111), 119, 2018.
- [5] M. Debouba, B. Khemakhem, S. Zouari, A. Meskine, H. Gouia, "Chemical and biological activities of *Haplophyllum tuberculatum* organic extracts and essential oil," Journal of essential oil bearing plants, vol. 17, no. 5, pp. 787-796, 2014.
- [6] M. A. Al-Yahya, A. J. Al-Rehaily, M. S. Ahmad, M. S. Al-Said, F. S. El-Feraly, C. D. Hufford, "New alkaloids from *Haplophyllum tuberculatum*," Journal of Natural Products, vol. 55, pp. 899-903, 1992.
- [7] A. H. Mohamed, M. B. Ali, A. K. Bashir, A. M. Salih, "Influence of *Haplophyllum tuberculatum* on the cardiovascular system," International Journal of Pharmacognosy, vol. 34, no. 3, pp. 213-217, 1996.
- [8] J. S. Mossa, M. A. Al-Yahya, I. A. MAl-Meshal, Medical Plants of Saudi Arabia. Vol. 1, Riyadh, King Saud University Libraries, 1987.
- [9] O. Said, K. Khalil, S. Fulder, H. Azaizeh, "Ethnopharmacological survey of medicinal herbs in Israel, the Golan Heights and the West Bank region," Journal of Ethnopharmacology, vol. 83, no. 3, pp. 251-265, 2002.
- [10] S. A. M Abdelgaleil, M. M. G. Saad, N. R. Ariefta, Y. Shiono, "Antimicrobial and phytotoxic activities of secondary

metabolites from Haplophyllum tuberculatum and *Chrysanthemum coronarium*," South African Journal of Botany, vol. 128, pp. 35-41, 2020.

- [11] A. Abdelkhalek, M. Z. Salem, E. Hafez, S. I. Behiry, S. H. Qari, "The phytochemical, antifungal, and first report of the antiviral properties of Egyptian *Haplophyllum tuberculatum* extract," Biology, vol. 9, no.9, pp. 248, 2020.
- [12] C. Yaman, D. Ulukuş, O. Tugay, "Haplophyllum suaveolens varyetelerinin antioksidan aktivitesi ve sekonder metabolitleri üzerine farklı çözücülerin etkisi,"Türkiye Tarımsal Araştırmalar Dergisi, vol. 6, no. 3, pp. 277-284, 2019.
- [13] C. Yaman, D. Ulukuş, O. Tugay, "Endemik Haplophyllum A. Juss. türlerinin antioksidan aktivitesi üzerine lokasyon ve tür farkının etkisi," Journal of the Institute of Science and Technology, vol. 10, no. 1, pp. 648-657, 2020.
- [14] A. Hamdi, K. Majouli, A. Abdelhamid, B. Marzouk, H. Belghith, I. Chraief, Y. Vander Heyden, "Pharmacological activities of the organic extracts and fatty acid composition of the petroleum ether extract from *Haplophyllum tuberculatum* leaves,"Journal of ethnopharmacology, vol. 216, pp. 97-103, 2018.
- [15] M. Mohammadhosseini, A. Venditti, C. Frezza, M. Serafini, A. Bianco, B. Mahdavi, "The Genus Haplophyllum Juss.: Phytochemistry and Bioactivities—A Review, "Molecules, vol. 26, no. 15, pp. 4664, 2021.
- [16] A. Hamdi, A. Halouani, I. Aouf, J. Viaene, B. Marzouk, J. Kraiem, Y. Vander Heyden, "Cytotoxicity and Antiviral Activities of Haplophyllum tuberculatum Essential Oils, Pure

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Compounds, and Their Combinations against Coxsackievirus B3 and B4, "Planta medica, vol. 87, no.10/11, pp. 827-835, 2021.

- [17] P. Varamini, M. Doroudchi, A. Mohagheghzadeh, M. Soltani, A. Ghaderi, "Cytotoxic evaluation of four *Haplophyllum* species with various tumor cell lines, "Pharmaceutical Biology, vol. 45, no. 4, pp. 299-302, 2007.
- [18] J. M. Prieto, "Haplophyllum A. Juss, a rich source of bioactive natural principles. In: Bioactive Compounds: Types," Biological Activities and Health Effects, pp. 341-380, 2012.
- [19] A. Pasqualone, A. M. Bianco, V. M. Paradiso, C. Summo G. Gambacorta, F. Caponio, A. Blanco, "Production and characterization of functional biscuits obtained from purple wheat," Food Chemistry, vol. 180, pp. 64-70, 2015.
- [20] B. Narayanasamy, N. Jeyakumar, D. K. Manoharan, "Effect of natural antioxidants on the oxidation stability of methyl ester of rubber seed oil," Journal Energy Sources, Part A: Recovery, Utilization, and Environmental Effects, vol. 40, no. 6, pp. 680-687, 2018.
- [21] S, Bajaj, A, Urooj, P. Prabhasankar," Effect of incorporation of mint on texture, colour and sensory parameters of biscuits" International Journal of Food Properties, vol. 9, pp. 691-700, 2006.
- [22] M. A. Shah, S. J. Don Bosco, S. A. Mir, "Plant extracts as natural antioxidants in meat and meat products," Meat Science, vol. 98, pp. 21-33, 2014.
- [23] A. Russo, C. Formisano, D. Rigano, F. Senatore, S. Delfine, V. Cardile, S. Rosselli, M. Bruno, "Chemical composition and anticancer activity of

essential oils of Mediterranean sage (*Salvia officinalis* L.) grown in different environmental conditions," Food and Chemical Toxicology, vol. 55, pp. 42-47, 2013.

- [24] A. R. Duarte, R. R. Naves, S. C. Santos, J. C. Seraphinand, P. H. Ferri, "Genetic and environmental influence on essential oil composition of *Eugenia dysenterica*," Journal Brazilian Chemical Social, vol. 21, no.8, pp. 1459-1467, 2010.
- [25] B. Li, C. Zhang, L. Peng, Z. Liang, X. Yan, Y. Zhu, Y. Liu, "Comparison of essential oil composition and phenolic acid content of selected *Salvia* species measured by GC–MS and HPLC methods," Industrial Crops Products, vol. 69, pp. 329-334, 2015.
- [26] D. H. Truong, D. H. Nguyen, N. T. A. Ta, A. V. Bui, T. H. Do, H. C. Nguyen, "Evaluation of the use of different solvents for phytochemical constituents, antioxidants, and in vitro anti-inflammatory activities of *Severinia buxifolia*, "Journal of Food Quality, 2019.
- [27] M. Barbouchi, K. Elamrani, M. El Idrissi, "A comparative study on phytochemical screening, quantification of phenolic contents and antioxidant properties of different solvent extracts from various parts of *Pistacia lentiscus* L., "Journal of King Saud University-Science, vol. 32, no. 1, pp. 302-306, 2020.
- [28] J. S. Dambolena, M. P. Zunino, E. I. Lucini, R. Olmedo, E. Banchio, P. J. Bima, J. A. Zygadlo, "Total phenolic content, radical scavenging properties, and essential oil composition of *Origanum* species from different populations," Journal of Agricultural and Food Chemistry, vol. 58, pp. 1115-1120, 2010.

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- [29] R. Chirinos, R. Pedreschi, H. Rogez, Y., Larondelle, D. Campos, "Phenolic compound contents and antioxidant activity in plants with nutritional and/or medicinal properties from the Peruvian Andean region," Industrial Crops and Products, vol. 47, pp. 145-152, 2013.
- [30] S. Rawat, I. D. Bhatt, R. S. Rawal, "Total phenolic compounds and antioxidant potential of *Hedychium spicatum* Buch. Ham. ex D. Don in west Himalaya, India," Journal of Food Composition and Analysis, vol. 24, pp. 574-579, 2011.
- [31] C. Çırak, J. Radušienė, V. Janulis, L. Ivanauskas, N. Çamaş, A. K. Ayan, "Phenolic constituents of *Hypericum triquetrifolium* Turra (Guttiferae) growing in Turkey: variation among populations and plant parts," Turkish Journal of Biology, vol. 35, pp. 449-456, 2011.
- [32] T. Sharma, V. Khandelwal, S. Gupta, S. Singh, "Secondary Metabolites, Boon for Plants; Their Role in Defence Mechanism and Antioxidant Activity of *Anthocephalus cadamba*," In Antioxidants in Plant-Microbe Interaction, Springer, Singapore, pp. 413-424, 2021.
- [33] T. K. Patle, K. Shrivas, R. Kurrey, S. Upadhyay, R. Jangde, R. Chauhan, screening "Phytochemical and determination phenolics of and flavonoids in Dillenia pentagyna using UV-vis and FTIR spectroscopy, "Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, vol. 242, pp. 118717, 2020.
- [34] G. Zengin, C. Sarikurkcu, A. Aktumsek, R. Ceylan, O. Ceylan, "A comprehensive study on phytochemical characterization of *Haplophyllum myrtifolium* Boiss. endemic to Turkey and its inhibitory potential against key

enzymes involved in Alzheimer, skin diseases and type II diabetes, "Industrial Crops and Products, vol. 53, pp. 244-251, 2014.

- [35] M. S. Butler, "The role of natural product in chemistry in drug discovery," Journal of Natural Products, vol. 67, pp. 2141-2153, 2004.
- [36] B. B. Mishra, V. K. Tiwari, "Natural products: An evolving role in future drug discovery," European Journal of Medicinal Chemistry, vol. 46, pp. 4769-4807, 2011.
- [37] M. Mazid, T. A. Khan, F. Mohammad, "Role of secondary metabolites in defense mechanisms of plants," Biology and Medicine, vol. 3, pp. 232-249, 2011.
- [38] Y. Z. Cai, Q. Luo, M. Sun, H. Corke, "Antioxidant activity and phenolic compounds of 112 Chinese medicinal plants associated with anticancer," Life Sciences, vol. 74, pp. 2157-2184, 2004.
- [39] V. Katalinic, M. Milos, T. Kulisic, M. Jukic, "Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols," Food Chemistry, vol. 94, pp. 550-557, 2006.
- [40] S. M. Cottica, A. Sawaya, M. N. Eberlin, S. L. Franco, L. M. Zeoula, J. V. Visentainer, "Antioxidant activity and composition of propolis obtained by different methods of extraction," Journal of the Brazilian Chemical Society, vol. 22, pp. 929–935, 2011.
- [41] Z. A. Abbas, S. Saggu, M. I. Sakeran, N. Zidan, H. Rehman, A. A. Ansari, "Phytochemical, antioxidant and mineral composition of hydroalcoholic extract of chicory (*Cichorium intybus* L.) leaves," Saudi Journal of Biological Sciences, vol. 22, no. 3, pp. 322-326, 2015.

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- [42] M. J. Iqbal, S. Hanif, Z. Mahmood, F. Anwar, A. Jamil, "Antioxidant and antimicrobial activities of Chowlai (*Amaranthus viridis* L.) leaf and seed extracts," Journal of Medicinal Plants Research, vol. 6, no. 27, pp. 4450-4455, 2012.
- [43] J. Dai, R. J. "Mumper, Plant phenolics: extraction, analysis and their antioxidant and anticancer properties," Molecules, vol. 15, pp. 7313-7352, 2010.
- [44] M. J. Bano, J. Lorente, J. Casstillo, G. O. Benavente, J. A. Rio, A. Ortuno, K. W. Quirin, D. Gerard, "Phenolic diterpenes, flavones, and rosmarinic acid distribution during the development ofleaves, flowers, stems and roots of *Rosmarinus officinalis* antioxidant activity," Journal of Agricultural and Food Chemistry, Vol. 51, pp. 4247-4253, 2003.
- [45] B. Winkel-Shirley, "Biosynthesis of flavonoids and effects of stress," Current Opinion in Plant Biology, vol. 5, pp. 218-223, 2002.
- [46] X. C. Li, J. Lin, W. J. Han, W. Q. Mai, L. Wang, Q. Li, M. F. Lin, M. S. Bai, L. S. Zhang, D. F. Chen, "Antioxidant ability and mechanism of Rhizoma *Atractylodes macrocephala*. Molecules. Vol. 17, pp. 13457-13472, 2012.