SERUM SELENIUM, ZINC, COPPER AND IRON CONCENTRATIONS AND SOME RELATED ANTIOXIDANT ENZYMES IN PATIENTS WITH CUTANEOUS LEISHMANIASIS

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ABSTRACT

Objective: This study has been carried out to measure the alterations of some trace element concentrations, their related antioxidant enzyme activities, and carrier proteins in patients with cutaneous leishmaniasis (CL).

Methods: Serum selenium, copper and zinc concentrations were determined by Atomic Absorption Spectrometer. Erythrocyte Cu-Zn superoxide dismutase (Cu-Zn SOD), glutathione peroxidase (GSH-Px) and catalase (CAT) activities and ceruloplasmin (Cp) levels were measured with enzymatic colorimetric methods and transferrin (Tf) levels were measured with turbidimetric method.

Results: Erythrocyte Cu-Zn SOD activities, serum copper concentrations and Cp levels were found to be significantly higher in patients group, than those of controls. However, GSH-Px and CAT activities, and selenium, zinc, iron and Tf levels were lower in patients according to the control subjects. There were positive important correlations between Cu-Zn SOD and Cp, Cu-Zn SOD and copper, Cp and copper, GSH-Px and selenium, and iron and CAT in patients group.

Conclusion: Our results showed that serum essential trace elements selenium, zinc, copper and iron concentrations and their related enzymes Cu-Zn SOD, GSH-Px and CAT activities change in CL patients. The changes may be a part of defense strategies of organism and are induced by the hormone-like substances.

Key Words: Cutaneous leishmaniasis, glutathione peroxidase, Cu-Zn superoxide dismutase, catalase, selenium, zinc, copper, iron.

INTRODUCTION

Leishmaniasis is initiated when sand flies inject the extracellular promastigote form of the parasite into the skin. Promastigotes are rapidly phagocytized, after which they transform into the intracellular amastigote form. Promastigotes are not seen in clinical lesions; the multiplication of amastigotes within macrophages leads to clinical disease (1).

Cutaneous leishmaniasis (CL), caused by L. Major and L. Tropica, is characterized by a skin ulcer which heals spontaneously, leaving an unsightly scar (2). It is widespread through the Southeastern Anatolia of Turkey. It mainly affects the low-income people or rural and suburban populations.

The mechanism(s) by which defense cells to kill microorganisms has been the subject of intense research in recent years (3). Although, reactive oxygen intermediates (ROI) and nitric oxide (NO) that are generated during the respiratory burst by macrophages and polymorphonulear leukocytes, are involved in intracellular killing it is now apparent that serum redistribution of essential trace elements also has an important role in cytotoxicity (4-6). The changes are part of defense strategies of organism and are induced by the hormone-like substances interleukin-1 (IL-1), tumor necrosis factor-alpha (TNF- α) and interleukin-6 (IL-6) (7, 8). These substances are cytokines liberated in a dose-dependent mode, mostly by activated macrophages, in response to several stimuli, including exercise, trauma, stress, or infection (9). In animals, many diseases are also caused by simultaneous deficiency of selenium. However, the exact mechanism causing this deficiency remains unknown (10).

Several enzymes, including SOD, CAT and GSH-Px are capable of scavenging ROI *in vivo*. However,

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these enzymes require mentioned trace elements for their activity. For example, Cu-Zn SOD is an important intracellular enzyme that requires both copper and zinc for normal enzymatic activity; zinc stabilizes the enzyme and copper is necessary for catalysis. Cu-Zn SOD eliminates O_2 ⁻ and hydroperoxides may oxidize cellular substrates, and they prevent propagation of free radical chain reaction (11). GSH-Px that is a glutathione recycling enzyme, catalyzes the oxidation of reduced glutathione by H_2O_2 and other hydroperoxides to form oxidized glutathione and require selenium for its activity (12). CAT is a tetrameric hemoprotein that catalyzes the dismutation of H_2O_2 to water and molecular oxygen (13).

The purpose of the present study was to measure the concentrations of the essential trace elements like selenium, zinc, copper and iron, their binding proteins, and some related antioxidant enzymes like Cu-Zn SOD, GSH-Px and CAT activities, together with the investigation of the relationship between these elements and their related enzyme activities in patients with CL.

MATERIALS AND METHODS

Totally 80 subjects were enrolled in the study, 42 patients, and 38 healthy persons who were not exposed to CL. The study was conducted between November 1996 and March 1997 in Harrankapi Leishmaniasis Treatment Center, Sanliurfa which is an hyperendemic area for leishmaniasis in the Southeastern Anatolia of Turkey. Approval for the study was given by the Institution Ethical Committee. CL patients of any age and sex without regarding their lesion number were included in the study, while pregnants and those who were under antimonial or other treatments were excluded. If patients had 6 months or older lesions, they were excluded from study, because of spontaneous healing and immunity. Control group was selected among healthy parents or siblings which had not exposed to CL. Age, weight and height values were recorded. Additionally, size, localization, number and duration of lesions were recorded in the patient group. Diagnosis was confirmed clinically, as well as by laboratory demonstration of the parasite in the lesions by direct smears or cultures or both. Lesions were cleaned with ethanol, and punctured at the margins of the lesion with a sterile lancet. Smears were made from exudating materials, air dried, fixed methanol. The smears were stained with Giemsa's stain for examination by light microscopy. Microscopic diagnosis were made when amastigotes were identified in the smears. Also materials were cultured on Novy-Mac Neal-Nicolle (NNN) with rabbit blood agar medium for up to 3 weeks to detect the leishmanial promastigotes.

All of the materials (glass and plastic) employed were thoroughly cleaned with hot solution of nitric acid (20% v/v) for 48 hour and rinsed six times with demineralized water.

Total 10 mL venous blood was withdrawn. Of samples, 5 mL were transferred into tubes without any addition of anticoagulants and centrifuged for 15 min at a speed of 2000 x g. Sera were separated to determine selenium, copper, zinc and iron levels. Remaining 5mL of blood samples, 5 mL were transferred into heparinized tubes and erythrocyte Cu-Zn SOD, GSH-Px and CAT activities were measured immediately after preparation of erythrocyte hemolysate.

Serum selenium determination was performed by a SpectrAA 250 Plus Zeeman Atomic Absorption Spectrophotometer with a graphite furnace GTA-96 (Varian, Australia), with deuterium background correction. Varian hollow cathode lamps were employed at the 196-nm wavelength and 1.0-nm bandpass. Pyrolytically coated graphite tubes with pyrolytic graphite platforms (Varian, Australia) were used. Selenium concentration was determined by an internal standard addition method (14).

Serum copper and zinc were determined by a SpectrAA 250 Plus Zeeman Atomic Absorption Spectrometer (Varian, Australia) with a deuterium background correction. Serum copper and zinc values were expressed in mg/L.

Serum iron, Tf and albumin levels were determined with commercial kits (Boehringer Mannheim, Germany) by automatic analyser (Hithachi 911 Boehringer Mannheim, Germany) Serum ceruloplasmin (Cp)levels were determined by measurement of p-phenylenediamine oxidase activity according to the method of Sunderman and Nomoto (15).

In order to measure erythrocyte antioxidant enzyme activities, Erythrocytes were separated from heparinized blood by centrifugation at 2000 x g and washed with 0.9% isotonic NaCl solution repeatedly until a colorless supernatant was observed. Then, erythrocytes were lyzed by addition of 4 volumes of double distilled water. The resulting suspension was centrifuged twice to eliminate all of the cell membranes: first by 10 min in the tube centrifuge at 2000 x g, followed by centrifugation in an eppendorf centrifuge at 4500 x g for 5 min. Hemoglobin was determined by Drabkin's method in an 0.1/mL aliqout of hemolysate. Blood hematocrit (Htc) concentrations (%) were measured by automatic blood cell analyzer (STKS, Coulter, USA)

GSH-Px and Cu-Zn SOD activities were measured in erythrocyte by method of Paglia and Valentine (16),

and McCord and Fridovich (17), employing the RANSEL and RANSOD kits (RANDOX, Belfast). Measurements were carried out at 37°C. Internal quality control was maintained by use of a standardized control supplied by RANDOX. Measurements were performed in an autoanalyzer (Hithachi 911 Boehringer Mannheim, Germany) according to the Randox application procedure. Erythrocyte CAT activity was determined by a spectrophotometric method (18)

Statistics were calculated with the SPSS for Windows program Version 6.0. The mean values obtained in the different groups were compared by Student's t-test. The Pearson's correlation test was used to evaluate the correlation between two variables. All results were expressed as mean values \pm SD; significance was defined as p<0.05.

patients with CL had significantly lower levels of serum selenium, zinc and iron (p<0.001, p<0.01, p<0.05, respectively) (Table II). There were also significant decreases in serum Tf levels and GSH-Px and CAT activities in patients group (p<0.05, p<0.001, p<0.01 respectively). However, serum copper concentrations, Cp levels, and Cu-Zn SOD activities were higher in CL patients according to the control subjects (p<0.01, p<0.001 and p<0.01 respectively). Blood Htc and albumin levels were lower in patients group according to the controls, but it was not statistically significant (Table II).

There was a positive important correlation between selenium concentration and GSH-Px activity in total group (r=0.703, p<0.0001), as seen from Table III. In addition, positive correlations were noted between selenium and albumin (r=0.344, p<0.05), and GSH-Px and Htc (r=0. 482, p<0.01), Htc and selenium (r=0.289, p<0.05) in patients. There was also positive relationships between Cu-Zn SOD and Cp (r=0.365, p<0.01). Cu-Zn SOD and Cu (r=0.255, p<0.05), Cp and Cu (r=0.67, p<0.001), and CAT and iron (r=0.43, p<0.01) in patients group (Table III).

RESULTS

As seen from Table I, the cases and unmatched controls were similar in age, height, body weight, body mass index (BMI). When compared to controls,

Table I.	Physical	Characteristics	of CL	Patients	and Healthy	Subjects

	Patients (n= 42)	Healthy Subjects (n= 38)	р
Age (year)	29.2 ± 4.3	28.3 ± 4.5	>0.05
Height (cm)	168 ± 6	169 ± 5	>0.05
Weight (kg)	65.1 ± 8.9	66 ± 12.4	>0.05
BMI (kg/m ²)	23.4 ± 3.6	24.7 ± 4.4	>0.05

 Table II. Comparison of Parameters in Patients and Healthy Subjects

Parameters	Patients (n= 42)	Healthy Subjects (n= 38)	p
Erythrocyte Cu-Zn SOD (U/g Hb)	570.96 ± 77.04	425.4 ± 64.26	<0.01
Erythrocyte GSH-Px (U/g Hb)	315.08 ± 34.3	362.10 ± 33.28	<0.001
Erythrocyte CAT (K act./g Hb)	99 ± 0.57	153 ± 0.54	<0.01
Serum Zn (mg/L)	0.74 ± 0.112	0.847 ± 0.111	<0.01
Serum Cu (mg/L)	1.44 ± 0.298	1.227 ± 0.415	<0.01
Serum Fe (mg/L)	0.722 ± 0.183	0.924 ± 0.1765	<0.05
Serum Se (µg/L)	75 ± 20.37	106 ± 26.28	<0. 001
Serum Cp(U/L)	1110 ± 85	790 ± 42	<0.001
Serum Tf (mg/dL)	263 ± 28	296 ± 35	<0.05
Serum albumin (g/dL)	4.3 ± 0.3	4.5 ± 0.2	>0.05
Hematocrit (%)	39.5 ± 2.48	41.3 ± 3.23	>0.05

Parameters	r	р
GSH-Px- Se	0.703	<0.0001
GSH-Px-Htc	0. 482	<0.01
Albumin- Selenium	0.344	<0.05
Htc - Selenium	0.289	<0.05
Copper-Cp	0.67	<0.001
Cu-Zn SOD - Cp	0.356	<0.01
CAT-iron	0.43	<0.01

Table III. Some Significant Correlations in Patients Group

DISCUSSION

The serum redistribution of the essential trace elements, iron, copper and zinc together with the increase in synthesis of acute-phase proteins like ceruloplasmin and transferrin, which takes place during the course of most infections, is well established. The changes consist of a decrease in serum iron and zinc concentrations together with an increase in copper, and their magnitude depends on the intensity of the stimulus (6). In the present study we found that serum selenium, zinc and iron concentrations were significantly lower and copper concentrations were higher in patients with CL than those of control subjects. In addition, their related antioxidant enzymes GSH-Px and CAT activities were lower and Cu-Zn SOD activities were higher in CL patients.

It was demonstrated that the alterations of serum zinc, iron and copper concentrations particularly depend on cytokines specially IL-1 and TNF- α (9). Activated phagocytic cells release endogenous mediators that reduce plasma zinc concentrations in experimental animals by redistribution of zinc from plasma to the liver (19). Decreasing serum zinc levels apparently result from the synthesis of methallothionein (MT) in liver and other tissues. Both IL-1 and TNF-α decrease plasma iron concentrations and the decreasing concentrations of iron induced by IL-1 is at last in part, due to the release of apolactoferrin by granulocytes and apolactoferrin can remove iron from Tf, thus iron is sequestered in compartments which are nutritionally unavailable to bacteria or parasites (20). Increased serum copper is associated with Cp and also induced by IL-1 (21). It was demonstrated that IL-1, but not TNF- α increase plasma copper content (6). Increased Cp and copper content may be attributable to inflammation associated with the disease.

As seen above, the changes of these trace metal contents in CL patients may depend on cytokines

specially IL-1 and TNF- α , although we could not determine these cytokines. However, some observations have shown that the production of IL-1 and TNF- α were induced by CL (22-24).

In animals, many diseases are caused by simultaneous deficiency of selenium. However, the exact mechanism causing this deficiency remains unknown. It has been demonstrated that GSH-Px enzyme activity was lower in the patients with Leishmania Donovani (25). However, it was not known whether the causes of this depletion depended on selenium content or the other factors. Also, we could not found any report about selenium status and relationship between selenium and GSH-Px activity in patients with CL. Dworkin (26).demonstrated that both selenium concentrations, GSH-Px activities and serum albumin levels were significantly lower in patients and there was an important correlation between albumin and GSH-Px in acquired immune deficiency syndrome (AIDS). He has claimed that selenium deficiency seems to be a part of the protein-calorie malnutrition and impaired intestinal absorption common in AIDS. However, Dworkin's patients were predominantly malnourished drug users. Olmsted et al. (27), have also found that selenium concentrations were significantly lower in AIDS patients. Whereas, they were mainly homosexuals from higher socioeconomic classes where malnutrition is usually not a problem and intestinal selenium absorption were not impaired in these patients. We also found that there was a good correlation between blood selenium content and GSH-Px activity. However, there were weak correlations between serum albumin and selenium, and Htc and GSH-Px in these groups. We assessed albumin levels because they depend on serum protein intake and it is frequently used to assess nutritional status (28). Also, we have measured Htc levels, BMI, weight and height of the patients and controls. These findings demonstrated that in our patients malnutrition is not a problem. Whereas, blood selenium concentrations were significantly

lower in this group. The decreased levels of selenium in our patients may be attributed to redistribution from the plasma pool into the tissues as a defense mechanism modulated by the effect of infection. The decrease in the activity of GSH-Px may also be related to decreased selenium levels in those patients.

Numerous studies have demonstrated for a long time that ROI such as O₂.⁻ and H₂O₂ peroxide, which are products of the macrophage respiratory burst, were the major killing mechanisms (29). A number of groups reported evidences that ROI's were involved in macrophages leishmanicidal activity, and H₂O₂ is an effector molecule against these parasites (30). It would be expected that, increased amounts of hydroperoxides might be generated to kill protozoa as host defense strategies. It was known that there are strong relationships between antioxidant enzymes and their cofactors (31). We also found significant correlations between GSH-Px and selenium (r= 0.703, p<0.0001), CAT and iron (r=0.430, p<0.01). Consequently, lack of selenium and iron leads to a and CAT enzyme deficiency of the GSH-Px activities and, consequently, to a decreased ability to degrade H₂O₂ (32).

We found that Cu-Zn SOD activity were higher together with serum copper and Cp levels in patients, and there was a positive significant correlation between copper and Cp, and Cp and Cu-Zn SOD, and Cu-Zn SOD and copper in patients group. We suggest that, cytokines induce not only trace element metabolism, but also antioxidant enzyme activity. It was demonstrated that both manganese (Mn) SOD and Cu-Zn SOD activities were significantly induced by IL-1 and TNF- α (33). Disilvestro et al. (34), and Assreuy et al. (3), also demonstrated that addition of SOD to macrophages, enhanced leishmanicidal activity. Although there is no available report about CL and antioxidant activity, our results are supported by mentioned reports. However, it was found that Cu-Zn SOD activity was lower together with GSH-Px and CAT activities in Leishmania Donovani infection (25). This difference between visceral and CL may depend on a distinct pathology and immune response in two species of leishmania. For example, the high production of IL-1 in CL is in contrast to visceral leishmaniasis that resulted in suppression of the IL-1 response as seen above (35).

We conclude that serum essential trace elements selenium, zinc, copper and iron concentrations and their related enzymes Cu-Zn SOD, GSH-Px and CAT activities change probably by the effects of some cytokines as host defense strategies of organism during CL infection. However, there is no available report about selenium and cytokines. Further investigations will be needed to study the cytokines together with trace elements specially selenium and antioxidant enzyme activities.

REFERENCES

- 1. Berman JD, Waddel D, Hanson BD. Biochemical Mechanisms of the antileishmanial activity of sodium stibagluconate. Antimicrob Agents Chemother 1985; 27(6): 916-920.
- 2. Dowtlati Y. Cutaneous leishmaniasis. Int J Dermatol 1975; 18:362-368.
- Assreuy J, Cunha FQ, Epperlein M, Noronha-Dutra A, O'Donnell CA, Liew FY, Moncada S. Production of nitric oxide and superoxide by activated macrophages and killing of Leishmania major. Eur J Immunol 1994; 24: 672-676.
- 4. Haidaris CG, Bonventre PF. A role for oxygendependent mechanisms in killing of Leishmania donovani tissue forms by activated macrophages. J Immunol 1982; 129: 850-855.
- 5. Dockrell HM, Playfair JHL. Killing of Plasmodium yoelii by enzyme-induced products of the oxidative burst. Infect Immun 1984; 43:451-456.
- 6. Mastousek AJ, Burguera JL, Burguera M, Anez N. Changes in total content of iron, copper and zinc in serum, heart, liver, spleen and skeletal muscle tissues of rats infected with Trypanosoma cruzi. Biol Trace Elem Res 1993; 37: 51-69.
- 7. Barber EF, Cousins RJ. Interleukin-1-Stimulated induction of ceruloplasmin synthesis in normal and copper-deficient rats. J Nutr 1988; 118: 375-381.
- 8. Cousins RJ. Absorption, transport, and hepatic metabolism of copper and zinc: special reference to metallothionein and ceruloplasmin. Physiol Rev 1985; 65: 238-246.
- 9. Klassing KC. Nutritional aspects of leukocytic cytokins. J Nutr 1988; 118: 1435-1443.
- 10. Dworkin BM, Rosenthal WS, Wormser GP, et al. Abnormalities of blood selenium and gluthathione peroxidase activity in patients with acquired immunodeficiency syndrome and aids-related complex. Biol Trace Elem Res 1988;15:167.
- 11. Panemangalore M, Bebe FN. Effect of high dietary zinc on plasma ceruloplasmin and erythrocyte superoxide dismutase activities in Cu-depleted and repleted rats. Biol Trace Elem Res 1996; 55: 111-126.
- 12. Sodhi CP, Katyal R, Rana SV, Attri S, Singh V. Study of oxidative-stress in rotavirus infected infant mice. Indian J Med Res 1996; 104: 245-249.
- *13.* Kono Y, Fridowich I. Superoxide radical inhibits catalase. J Biol Chem 1982; 257:5751-5755.
- 14. Korunova V, Skodova Z, Dedina J. Serum selenium in adult Czechoslovak (Central Bohemia) population. Biol Trace Elem Res 1993; 37: 91-99.

- 15. Sunderman FW, Nomoto S. Measurement of human serum ceruloplasmin by its p-phenylenediamine oxidase activity. Clin Chem 1970; 16: 903-910.
- 16. Paglia DE, Valentine WN. Studies on quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J Lab Clin Med 1967; 70:158-159.
- 17. McCord JM, Fridovich I. Superoxide dismutase, an enzymatic function for erythrocuprein (hemocuprein). J Biol Chem 1969;. 244: 6049-6055
- 18. Mateo MCM, Martin B, Bene't MS, Rabadan J. Catalase activity in erythrocytes from colon and gastric cancer patients. Influence of nickel, lead, mercury and cadmium. Biol Trace Elem Res 1997; 57:79-90.
- 19. Svenson KLG, Hallgren R, Johansson E, Lindh U. Reduced zinc in peripheral blood cells from patients with inflammatory connective tissue disease. Inflammation 1985; 9 (2): 189-199.
- 20. Adam'k B, Wlaszczyk A. Lactoferrin-its role in defense against infection and immunotropic properties. Postepy Higieny i Meddycyny Doswiadczalnej 1996; 50(1): 33-41.
- 21. Klassing KC, Laurin DE, Penk RK, Fry DM. Immunological mediated growth depression in chicks: influence of feed intake, corticosterone and interleukin-1. J Nutr 1987; 117:1629-1637.
- 22. Pirmez C, Yamamura M, Uyemura K, Oliveria MP, Silva VC. Cytokine patterns in the pathogenesis of human leishmaniasis. Clin Invest 1993; 91:1390-1395.
- 23. Wagner HM, Buesher HU, Rollinghoff M, Solbach W. Interferon-gamma inhibits the efficacy of interleukin -1 to generate a Th2-cell based immune response induced by leishmania major. Immunobiol 1991; 182(3-4):292-306.
- 24. Cillarie E, Dieli M, Maltese E, Milano S, Salerno A, Liew FY. Enhancement of macrophage IL-1 production by leishmania major infection in

vitro and its inhibition by IFN-γ. J Immunol. 1989; 143(6): 2001-2005.

- 25. Biswas T, Ghosh DK, Mukherjee N, Ghosal J. Lipid peroxidation of erythrocytes in visceral leishmaniasis. J Parasitol 1997; 83(1): 151-152.
- 26. Dworkin BM. Se deficiency in HIV infection and the acquired immunodeficiency syndrome (AIDS). Chem Biol Interact 1994; 91:181-186.
- 27. Olmsted L, Schrauzer GN, Flores-Arce M, Dowd J. Selenium supplementation of symptomatic human immunodeficiency virus infected patients. Biol Trace Elem Res 1989; 20:59-65.
- 28. Apelgren KN, Rombeau JL, Twomey PL, Miller RA. Comparison of nutritional indices and outcome in critically ill patients. Crit Care Med 1982; 10: 305-312.
- 29. Hughes HPA. Oxidative killing of intracellular parasites mediated by macrophages. Parasitol Today 1988; 4: 340-347.
- 30. Ockenhouse CF, Shear HL. Oxidative killing of the intraerythrocytic malaria parasite Plasmodium yoelii by activated macrophages. J Immunol 1984; 132: 424-432.
- 31. Sies H. Antioxidant activity in cells and organs. Am Rev Respir Dis 1987;136: 478-480.
- 32. Burk RF. Biological activity in selenium. Annu Rev Nutr 1983; 3: 53-70.
- 33. Niwa J, Lizawa O, Ishimato K, Akamatsu H, Kanoh T. Age-dependent bazal level and induction capacity of Cu-Zn and Mn superoxide dismutase and other scavenging enzyme activities in leukocytes from young and elderly adults. Am J Pathol .1993;143(1): 312-320.
- 34. DiSilvestro RA, David EA, Collignon C. Interleukin-1 slowly increases lung fibroblast Cu-Zn superoxide dismutase activity levels. Proc Soc Exp Biol Med 1991;197:197-200.
- 35. Reiner NE. Parasite accessory cell interaction in murine leishmaniasis I. Evasion and stimulusdependent suppression of the macrophage interleukin-1 response by Leishmania Donovani. J Immunol 1987; 138: 1919-1925.