

# COMU Journal of Marine Sciences and Fisheries

Journal Home-Page: <http://jmsf.dergi.comu.edu.tr> Online Submission: <http://dergipark.org.tr/jmsf>



## RESEARCH ARTICLE

### Effect of *Anisakis pegreffii* (Nematoda: Anisakidae) on Biochemical and Haematological Characteristics of Chub Mackerel (*Scomber japonicus* Houttuyn, 1782) Caught in the Dardanelles at Çanakkale, Türkiye

Ekrem Şanver Çelik<sup>1</sup>, Rıdvan Erdem Kanat<sup>2</sup>, Pinar Dermancı<sup>3</sup>, Dilek Kahraman Yılmaz<sup>4</sup>, Emre Turgay<sup>5</sup>, Süheyla Karataş<sup>6</sup>, Sevdan Yılmaz<sup>7\*</sup>

<sup>1</sup>Department of Marine Biology, Faculty of Marine Sciences and Technology, Çanakkale Onsekiz Mart University, Çanakkale 17100, Türkiye

<sup>2</sup>Department of Marine and Inland Water Sciences, Çanakkale Onsekiz Mart University School of Graduate Studies, Çanakkale 17100, Türkiye

<sup>3</sup>Department of Marine and Inland Water Sciences, Çanakkale Onsekiz Mart University School of Graduate Studies, Çanakkale 17100, Türkiye

<sup>4</sup>Department of Marine Biology, Faculty of Marine Sciences and Technology, Çanakkale Onsekiz Mart University, Çanakkale 17100, Türkiye

<sup>5</sup>Department of Aquaculture and Fish Diseases, Faculty of Aquatic Sciences, Istanbul University, 16 Mart Şehitleri Cad. No:2, 34134 Fatih, Istanbul, Türkiye

<sup>6</sup>Department of Aquaculture and Fish Diseases, Faculty of Aquatic Sciences, Istanbul University, 16 Mart Şehitleri Cad. No:2, 34134 Fatih, Istanbul, Türkiye

<sup>7</sup>Department of Aquaculture, Faculty of Marine Sciences and Technology, Çanakkale Onsekiz Mart University, Çanakkale 17100, Türkiye

<https://orcid.org/0000-0003-4514-457X>

<https://orcid.org/0000-0002-1452-8856>

<https://orcid.org/0000-0003-1981-3215>

<https://orcid.org/0000-0002-9626-5446>

<https://orcid.org/0000-0001-9964-3919>

<https://orcid.org/0000-0003-2006-7854>

<https://orcid.org/0000-0002-4809-5809>

Received: 18.07.2022 / Accepted: 23.09.2022 / Published online: 28.10.2022

#### Key words:

Worm  
Parasite  
Anisakiasis  
Health  
Biometric Indices  
18S rRNA

**Abstract:** Chub mackerel (*Scomber japonicus* Houttuyn, 1782) were obtained from the commercial fishermen in Çanakkale, Turkey, in July, 2017. We examined a total of 40 fish (20 non-infested and 20 infested) and assessed the biometric indices, haematological parameters and serum biochemical variables. The hepatosomatic index and gonadosomatic index of infested chub mackerel fish were lower than those of non-infested fish. Blood haematocrit ratio and haemoglobin concentration in naturally parasite-infested chub mackerel fish were significantly lower than those in non-infested fish. However, white blood cell counts of the parasite-infested chub mackerel fish were higher than those of healthy ones. Serum total protein, globulin, glucose, cholesterol, triglyceride, urea, chlorine and iron levels in naturally parasite-infested chub mackerel fish were significantly lower than those in non-infested fish. Moreover, serum lactate dehydrogenase, aspartate aminotransferase, alkaline phosphatase and alanine aminotransferase activities of the parasite-infested chub mackerel fish were higher than those in healthy ones. Therefore, observed variations in haematological parameters, serum biochemical variables and biometric indices influenced by the parasite, *A. pegreffii*, may potentially increase sensitivity of the chub mackerel, *Scomber japonicus*, to diseases and environmental conditions.

#### Anahtar kelimeler:

Kurt  
Parazit  
Anisakiasis  
Sağlık  
Biyometrik İndeksler  
18S rRNA

#### Çanakkale Boğazı'ndan Yakalanan Kolyoz'un (*Scomber japonicus* Houttuyn, 1782) Biyokimyasal ve Hematolojik Karakteristikleri Üzerine *Anisakis pegreffii* (Nematoda: Anisakidae)'nin Etkisi

**Öz:** Kolyoz (*Scomber japonicus* Houttuyn, 1782) Temmuz 2017'de Çanakkale, Türkiye'deki ticari balıkçılardan elde edildi. Toplam 40 balığın (20 parazitsiz, 20 parazitli) biyometrik indeksleri, hematolojik parametreleri ve serum biyokimyasal değişkenleri incelendi. Parazitli kolyoz balıklarının hepatosomatik indeksi ve gonadosomatik indeksi, parazitsiz balıklardan daha düşüktü. Doğal olarak parazitli istila edilmiş kolyoz balıklarında kan hematokrit oranı ve hemoglobin konsantrasyonu, parazitsiz balıklardan önemli ölçüde daha düşüktü. Bununla birlikte, parazitli kolyoz balıklarının beyaz kan hücresi sayıları sağlıklı olanlardan daha yüksekti. Doğal olarak parazitli kolyoz balıklarında serum total protein, globulin, glikoz, kolesterol, trigliserit, üre, klor ve demir seviyeleri, parazitsiz balıklardan önemli ölçüde daha düşüktü. Ayrıca, parazitli kolyoz balıklarının serum laktat dehidrojenaz, aspartat aminotransferaz, alkanin fosfataz ve alanin aminotransferaz aktiviteleri sağlıklı olanlardan daha yüksekti. Bu bulgular ışığında, parazitli kolyoz, *Scomber japonicus*'un hematolojik parametreler, serum biyokimyasal değişkenler ve biyometrik indeksler de meydana getirdiği değişimlerin balığı farklı hastalıklara ve/veya çevresel koşullara karşı daha duyarlı hale getirebileceği söylenebilir.

\*Corresponding author: [sevdanyilmaz@comu.edu.tr](mailto:sevdanyilmaz@comu.edu.tr)

**How to cite this article:** Çelik, E.Ş., Kanat, R. E., Dermancı, P., Kahraman Yılmaz, D., Turgay, E., Karataş, S., Yılmaz, S. (2022). Effect of *Anisakis pegreffii* (nematoda: anisakidae) on biochemical and haematological characteristics of chub mackerel (*Scomber japonicus* Houttuyn, 1782) caught in the Dardanelles at Çanakkale, Türkiye. COMU J. Mar. Sci. Fish, 5 (Special Issue): 55-62.  
doi: 10.46384/jmsf.1140211

## Introduction

*Anisakis pegreffii*, is a nematode parasite belonging to Anisakidae family (Campana-Rouget and Biocca, 1955) with a complex life cycle that affects marine organisms at different trophic levels. In the life cycle of the *A. pegreffii*, small crustaceans are the intermediate host, fish and cephalopods are the intermediate and/or paratenic hosts while marine mammals are the definitive host (Mattiucci et al., 2008). The third-stage *A. pegreffii* larvae (L3) are infective to fish and cephalopods and are commonly found in the viscera and the musculature of these host species. Consuming infected animals raw or undercooked causes a disease called Anisakiasis in humans (Macchioni et al., 2021; Mattiucci et al., 2008; Mattiucci & D'Amelio, 2014; Mladineo & Poljak, 2014; Sakanari & McKerrow, 1989).

*Anisakis pegreffii* larvae have been reported in various fish species in many regions so far such as Africa, Australia, Oceania, Europe, South America and also in Turkish coastal waters (Aibinu et al., 2019; Özbakış Beceriklisoy et al., 2020; Pekmezci et al., 2014). *A. pegreffii* is the most important anisakid nematode larva in pelagic and demersal fish living in the Mediterranean waters, and life cycle of the parasite mostly includes fish species belonging to the families Salmonidae, Gadidae, Scombridae, Merlucciidae and Carangidae (Mattiucci et al., 2018; Menconi et al., 2022).

The relationship between parasites and hosts in both aquaculture and natural populations and their adverse effects on growth and survival have been demonstrated in parasite-host systems (Bichi & Dawaki, 2010). Damage on the host depends on the parasite species, the type of deterioration in the hosts tissue, the amount of parasite and the host's health condition (Tavares-Dias et al., 1999). Parasites can cause a decrease in haemoglobin concentrations, hematocrit levels and erythrocyte counts which called anemia (Martins et al., 2004). Also parasites affect the energy metabolism of their hosts. In many host-parasite relationships, parasites increase the metabolic rate of the host (Devevey et al., 2008). Therefore, changes associated with haematological and serum biochemical variables due to different parasites form a database that can be used in diagnosing diseases and preventive measures (Çelik & Aydin, 2006; Nnabuchi et al., 2015).

Chub mackerel, *Scomber japonicus* Houttuyn, (1782) is a pelagic fish species, which is distributed in the Atlantic, Black Sea, Mediterranean Sea, Indian and Pacific Oceans (Whitehead et al., 1984). Small pelagic fish such as anchovy, sardines, sprats, silversides and small invertebrates are the main food source of this species while *S. japonicus* is one of the most important element in the diet of other larger pelagic species and sea mammals (Collette & Nauen, 1983; Zardoya et al., 2004). However, there is no study on the impact of *A. pegreffii* on the biometric indices and blood variables of chub mackerel. Therefore, the present study was carried out to investigate the impact of *A. pegreffii* infestation on the biometric

indices, haematological and serum biochemical parameters of the chub mackerel, *S. japonicus*.

## Material and Methods

### Fish and parasite sampling

Chub mackerels (*Scomber japonicus* Houttuyn, 1782) were obtained from commercial fishermen in Çanakkale, Turkey in July, 2017. We examined a total of 40 fish (20 non-infested, 20 infested) and assessed the haematological and biochemical parameters. After recording wet body mass and body length, all individuals were dissected and the liver, spleen and gonad of each fish were weighed. Then, individuals were immediately examined for the parasites. For this purpose, the abdominal cavity was visually checked for the presence of nematodes and the fish with parasites were immediately examined. Nematode larvae were collected from the muscles, body cavum and visceral organs and fixed in 70% ethanol. The collected *Anisakis* larvae were morphologically identified according to Quiazon et al. (2009).

### Calculations of biometric indices

The condition factor (CF), splenosomatic index (SSI), hepatosomatic (HSI) and gonadosomatic index (GSI) were calculated according to the formulae proposed by Nikolsky (1963) and White and Fletcher (1985).

### Blood sampling

1-1.2 mL blood were taken from the tail vein through a plastic syringe, and 400-500 µL blood transferred to EDTA tubes for haematological analysis. 600-700 µL blood was harvested in serum separation tubes. Then, the serum samples were separated by centrifugation at 4000 ×g for 5 min and kept in a -80 °C freezer for analysis.

### Haematological analysis

Red blood cell (RBC,  $10^6 \text{ mm}^{-3}$ ) counts, haematocrit (Hct, %) ratios and haemoglobin (Hb, g/dL) concentrations were determined according to Blaxhall and Daisley (1973). The white blood cell (WBC,  $10^4 \text{ mm}^{-3}$ ) counts were estimated indirectly by the methods of Yilmaz et al. (2016).

### Biochemical analysis

Albumin (g/dL), globulin (g/dL), total protein (g/dL), urea (mg/dL), uric acid (mg/dL), creatinine (mg/dL), glucose (mg/dL), cholesterol (mg/dL), triglyceride (mg/dL), alkaline phosphatase (U/L), aspartate aminotransferase (U/L), lactate dehydrogenase (U/L), alanine aminotransferase (U/L), calcium (mg/dL), iron (µg/dL) and phosphorus (mg/dL) values were measured using commercial Bioanalytic kits in a spectrophotometer, as suggested by Yilmaz (2018), Yilmaz et al., (2012) and Yilmaz and Ergün (2012).

**DNA extraction, PCR amplification and DNA sequencing**

Nematode larvae were first homogenized with a bead-based homogenizer (Bullet Blender Storm - Next Advance Inc.) using glass beads (1.0 mm) for 5 min and then, genomic DNA was obtained using the commercial PureLink Genomic DNA Mini Kit (Catalog number: K182001, Invitrogen) following the manufacturer's instructions. To identify the parasites, partial 18S rRNA gene was amplified using a universal eukaryotic primer set F-566 and R-1200 (Hadziavdic et al., 2014). The PCR mix containing 0.4 µM of each primer, PCR Master Mix (2X) (Thermo Scientific), template DNA (approximately 50 ng) was used for amplifications using a thermal cycler (Biometra TAdvanced - Analytik Jena AG). The cycler was programmed as follows: initial denaturation 94°C for 5 min; 35 cycles of amplification (94°C for 30 s; 60°C for 30 s; 72°C for 1 min) and final extension at 72°C for 10 min. Agarose gel electrophoresis (1.6% wt/vol) was used for visualizing the PCR products in TAE buffer and electrophoresis was performed. All PCR products were purified and sequenced by a sequencing company (Medsantek Ltd., Istanbul/Turkey). Sequence editing and analyses were performed in Bioedit v7.0.0 (Hall, 1999) using the ClustalX 2.1 (Larkin et al., 2007) and BLASTN 2.2.20 algorithm (Zhang et al., 2000).

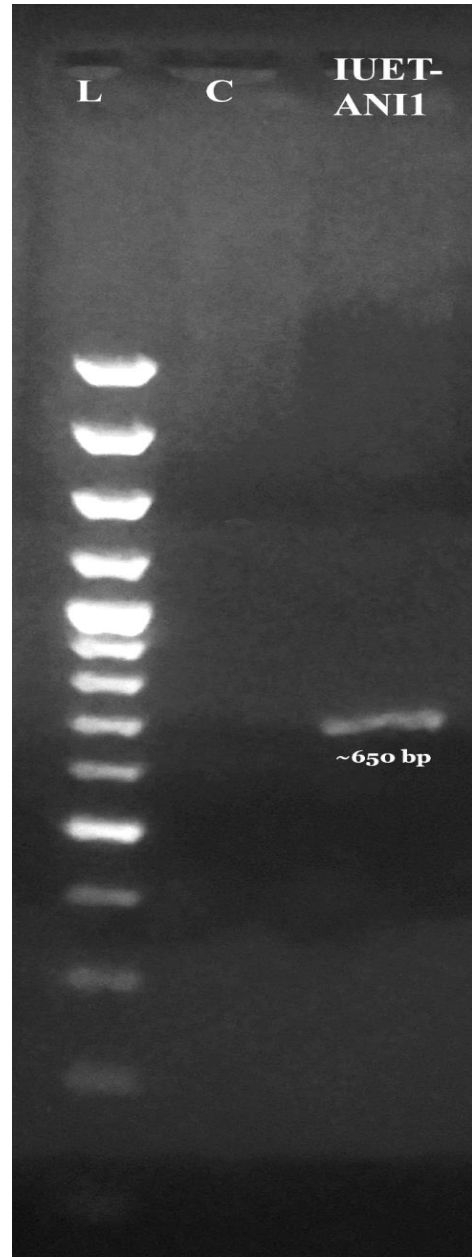
**Statistical analysis**

The statistical analysis was performed using SPSS 19.0 (SPSS Statistics) and the significance level was considered to be 0.05. All parameters were analyzed by T-test (Student's t-test).

**Results**

According to their morphological characteristics and partial 18S rRNA gene (Fig. 1) sequencing results with 100% similarity (acc. no. MF037865), the parasite was identified as *Anisakis pegreffii*.

The hepatosomatic index and gonadosomatic index of infested chub mackerel fish were significantly ( $p < 0.05$ ) lower than those of non-infested fish (Table 1). However, the means of body weight, body length, splenosomatic index and condition factor were not significantly different ( $p > 0.05$ ).



**Figure 1.** Agarose gel electrophoresis image of the PCR product Lane 1: (L) GeneRuler™ 100 bp DNA Ladder (Thermo Scientific), Lane 2: Negative control, Lane 3: Partially amplified 18S rRNA gene of the nematode

**Table 1.** The length, weight, hepatosomatic index (HSI), splenosomatic index (SSI), condition factor (CF), rate of gonadosomatic index (GSI) in the studied chub mackerel

Characteristic	Naturally parasite-infested fish mean ± SEM (n=20)	Healthy fish mean ± SEM (n=20)
Body weight (g)	126.12±2.15	138.15±3.11
Body length (cm)	24.62±0.21	24.98±0.41
HSI (%)	0.85±0.01 <sup>b</sup>	1.12±0.05 <sup>a</sup>
SSI (%)	0.15±0.02	0.16±0.02
CF (%)	0.79±0.10	0.82±0.05
GSI (%)	3.56±0.25 <sup>b</sup>	5.98±0.24 <sup>a</sup>

Values within a row denoted with different small letter is significantly different according to t-test

Blood haematocrit ratio and haemoglobin concentration in naturally parasite-infested chub mackerel fish were significantly ( $p<0.05$ ) lower than in non-infested fish (Table 2). However, white blood cell counts of the parasite-infested chub mackerel fish were higher than healthy ones ( $p<0.05$ ).

Serum iron, total protein, globulin, cholesterol, urea, triglyceride and glucose levels in naturally parasite-infested chub mackerel fish were significantly ( $p<0.05$ ) lower than in non-infested fish (Table 3). Moreover, serum aspartate aminotransferase, alkaline phosphatase, lactate dehydrogenase and alanine aminotransferase activities of the parasite-infested chub mackerel fish were higher than healthy ones ( $p<0.05$ ).

**Table 2.** Haematological parameters of naturally and healthy parasite-infested chub mackerel

Haematological parameters	Naturally parasite-infested fish mean $\pm$ SEM (n = 20)	Healthy fish mean $\pm$ SEM (n=20)
<i>Haematocrit</i> (%)	31.09 $\pm$ 1.56 <sup>b</sup>	36.54 $\pm$ 1.85 <sup>a</sup>
<i>Haemoglobin</i> (g/dL <sup>-1</sup> )	11.12 $\pm$ 0.28 <sup>b</sup>	13.16 $\pm$ 0.91 <sup>a</sup>
<i>Red blood cell</i> (10 <sup>6</sup> mm <sup>-3</sup> )	2.98 $\pm$ 0.47	3.02 $\pm$ 0.56
<i>White blood cell</i> (10 <sup>4</sup> mm <sup>-3</sup> )	2.56 $\pm$ 0.18 <sup>a</sup>	1.73 $\pm$ 0.34 <sup>b</sup>

Values within a row denoted with different small letter is significantly different according to t-test

**Table 3.** Serum biochemical characteristics of naturally parasite-infested and healthy chub mackerel

Serum Biochemical Parameters	Naturally parasite-infested fish mean $\pm$ SEM (n = 20)	Healthy fish mean $\pm$ SEM (n=20)
<i>Total protein</i> (g/dL)	2.56 $\pm$ 0.01 <sup>b</sup>	3.84 $\pm$ 0.21 <sup>a</sup>
<i>Albumin</i> (g/dL)	1.11 $\pm$ 0.12	1.42 $\pm$ 0.13
<i>Globulin</i> (g/dL)	1.45 $\pm$ 0.06 <sup>b</sup>	2.42 $\pm$ 0.12 <sup>a</sup>
<i>Creatinine</i> (mg/dL)	0.14 $\pm$ 0.01	0.15 $\pm$ 0.02
<i>Urea</i> (mg/dL)	1.32 $\pm$ 0.08 <sup>b</sup>	2.96 $\pm$ 0.18 <sup>a</sup>
<i>Uric acid</i> (mg/dL)	7.76 $\pm$ 0.12	7.55 $\pm$ 0.34
<i>Glucose</i> (mg/dL)	25.85 $\pm$ 0.96 <sup>b</sup>	56.47 $\pm$ 1.25 <sup>a</sup>
<i>Cholesterol</i> (mg/dL)	85.26 $\pm$ 2.55 <sup>b</sup>	108.22 $\pm$ 3.12 <sup>a</sup>
<i>Triglyceride</i> (mg/dL)	46.28 $\pm$ 2.16 <sup>b</sup>	81.22 $\pm$ 1.98 <sup>a</sup>
<i>Aspartate aminotransferase</i> (U/L)	95.21 $\pm$ 10.08 <sup>a</sup>	46.22 $\pm$ 10.17 <sup>b</sup>
<i>Alanine aminotransferase</i> (U/L)	25.16 $\pm$ 1.13 <sup>a</sup>	11.21 $\pm$ 0.96 <sup>b</sup>
<i>Alkaline phosphatase</i> (U/L)	59.14 $\pm$ 0.67 <sup>a</sup>	32.21 $\pm$ 1.18 <sup>b</sup>
<i>Lactate dehydrogenase</i> (U/L)	1268.16 $\pm$ 21.58 <sup>a</sup>	854.21 $\pm$ 62.13 <sup>b</sup>
<i>Calcium</i> (mg/dL)	14.44 $\pm$ 1.16	15.84 $\pm$ 1.51
<i>Phosphorus</i> (mg/dL)	16.31 $\pm$ 0.76	16.09 $\pm$ 0.85
<i>Iron</i> ( $\mu$ g/dL)	101.87 $\pm$ 10.19 <sup>b</sup>	155.85 $\pm$ 10.27 <sup>a</sup>

Values within a row denoted with different small letter is significantly different according to t-test

## Discussion

In the present study, *Anisakis pegreffii* were recovered within the body cavity, liver, gonad and muscles of chub mackerel (*S. japonicus*) caught in the Dardanelles, Çanakkale. Similarly, *Trachurus trachurus* (Mattiucci et al., 2008; Utuk et al., 2012), *S. japonicus*, *Scomber scombrus*, *Micromesistius poutassou*, *Merluccius merluccius*, *Trachurus mediterraneus*, *Mullus barbatus* (Pekmezci et al., 2014) and *Zeus faber* (Yardimci et al., 2014) were the fish intermediate hosts for *A. pegreffii* in the Turkish seas.

To our knowledge, this is the first report to evaluate the biometric indices, haematological parameters and serum biochemical variables of chub mackerel naturally parasitized by *A. pegreffii*.

Biometric indices, haematological parameters and serum biochemical variables are important parameters of health status used in determining the effect of parasitic diseases in fish (Çelik & Aydın, 2006; Özdemir et al., 2016; Maqbool & Ahmed, 2016). In this study, the mean HSI and GSI levels of parasite-infested chub mackerel fish were significantly lower than that in healthy ones. Previous studies showed that high numbers of parasites caused destruction of organ structures and subsequent decrease in organ size (Ryberg et al., 2020; Ryberg et al., 2022). This result is in accordance with that of Ondračková et al. (2010) indicating that larval parasitic nematode *Raphidascaris acus* decreased GSI level in infested female *Neogobius kessleri*. Similarly, Çelik and Aydın (2006) declared that average HSI levels of the *Scorpaena porcus* fish naturally infested with *Trachelobdella lubrica* was lower than that of non-infested fish. However, *Anisakis* infections did not affect condition factor, total length, total weight, hepatosomatic index and gonadosomatic index of *Scomber colias* and *Scomber scombrus* (Santos et al., 2017). These different findings might be related with differences in season, geographic region, fish species and/or parasite species.

In this study, slight reduction of red blood cell count and a significant reduction in serum iron, Hb and Hct values were observed in *A. pegreffii*-infested fishes. Previous studies indicated that parasites like *T. lubrica* (Çelik & Aydın, 2006), *C. oestroides* (Özdemir et al., 2016), *Trypanosoma kashmirensis* (Maqbool and Ahmed, 2016), *Trypanosoma carassii* (McAllister et al., 2019), *Dactylogyrus* sp., *Trichodina* sp. and *Lernaea cyprinacea* (Panjvini et al., 2016) and *Trypanosoma* spp. (Sousa et al., 2020) often caused anemia which is characterized by decreased red blood cell count, Hb concentration and/or Hct ratio.

White blood cells, serum proteins, albumin and globulin play significant roles in the immune response in parasitic infections (Çelik & Aydın, 2006; Özdemir et al., 2016). The high number of leucocytes in infested chub mackerel can be a reaction of cellular immune system to *A. pegreffii* infection. Similar results were also observed in *Dactylogyrus* sp., *Trichodina* sp. and *L. cyprinacea* infested *Cyprinus carpio* (Panjvini et al., 2016),

*Thelohanellus mrigalae* infested *Cirrhinus mrigala* (Manna & Naskar, 2021) and *Argulus* sp. infested *Carassius auratus* (Shameena et al., 2021).

The decreased serum total protein and globulin concentrations may occur during infectious diseases and nutritional imbalances (Çelik & Aydın, 2006). The same results were also obtained in *C. oestroides* infested *Boops boops* (Özdemir et al., 2016). Ryberg et al. (2020) reported that the plasma protein level decreased while plasma globulin level increased significantly with increasing *Contracaecum osculatatum* density in *G. morhua*.

Previous studies demonstrated that creatinine, urea and uric acid levels could be interpreted as a biomarker for determining the damage of gill and kidney diseases (Campbell, 2004). Similar to our result, Çelik and Aydın (2006) who found that serum urea concentration in naturally infested black scorpion (*S. porcus*) were significantly lower than those in non-infested fish.

The low serum glucose, triglyceride and cholesterol levels in the infested fish group may be the result of undernourished condition, liver disorders and inflammation. Similarly, the decreased serum glucose, triglyceride and cholesterol levels were also observed in *S. porcus* infested with *T. lubrica* (Çelik & Aydın, 2006). Özdemir et al. (2016) reported that serum glucose and triglyceride levels were decreased in the *C. oestroides* infested fish.

Serum enzyme activities are accepted as important indices of tissue damages, for example aspartate aminotransferase and alanine aminotransferase are indices of liver damage, lactate dehydrogenase for liver or muscle damages, and alkaline phosphatase for biliary system damage in fish (Yilmaz et al., 2019). Therefore, the increase in these enzymes in *A. pegreffii* infested fish could be attributed to liver and muscle tissue damage. In agreement with our findings, earlier studies reported increases of lactate dehydrogenase, aspartate aminotransferase, alkaline phosphatase and/or alanine aminotransferase enzymes in response to parasitic infections in fish (Çelik & Aydın, 2006; Datta et al., 2022; Kundu et al., 2016; Nabi et al., 2022; Özdemir et al., 2016; Rastiannasab et al., 2016).

In conclusion, observed variations in haematological parameters, serum biochemical variables and biometric indices influenced by the parasite, *A. pegreffii*, may potentially increase sensitivity of the chub mackerel, *Scomber japonicus*, to diseases and environmental conditions.

## Acknowledgements

We would like to thank the commercial fishermen. Rıdvan Erdem Kanat and Pınar Dermancı are PhD students of YÖK 100/2000 project.

## Conflict of Interest

The authors declare that they have no conflict of interest.

## Author Contributions

E. Şanver Çelik: Study Idea; Investigation; Collect Data; Writing Original Draft, R. E. Kanat: Writing Original Draft, P. Dermanci: Writing Original Draft, D. Kahraman Yılmaz: Writing Original Draft; Review & Editing, E. Turgay: Writing Original Draft; Methodology; Review & Editing, S. Karataş: Writing Original Draft; Methodology, S. Yılmaz: Methodology; Statistical Analyses; Writing Original Draft; Review & Editing, All authors contributed to the study conception, design and writing.

## Ethics Approval

This study was carried out the approval of Canakkale Onsekiz Mart University Animal Experiments Local Ethics Committee (Protocol Number: 2017/05-05).

## References

- Aibinu, I. E., Smooker, P. M., & Lopata, A. L. (2019). Anisakis Nematodes in Fish and Shellfish- from infection to allergies. *International Journal for Parasitology: Parasites and Wildlife*, 9,384–393. <https://doi.org/10.1016/j.ijppaw.2019.04.007>
- Bichi, A., & Dawaki, S. (2010). A survey of ectoparasites on the gills, skin and fins of *Oreochromis niloticus* at Bagauda fish farm, Kano, Nigeria. *Bayero Journal of Pure and Applied Sciences*, 3(1), 83–86. doi: 10.4314/bajopas.v3i1.58720
- Blaxhall, P. C., & Daisley, K. W. (1973). Routine haematological methods for use with fish blood. *Journal of fish biology*, 5(6), 771-781. doi: 10.1111/j.1095-8649.1973.tb04510.x
- Campana-Rouget, Y., & Biocca, E. (1955). Une nouvelle espèce d'Anisakis chez un phoque méditerranéen. *Annales de Parasitologie Humaine et Comparée*, 30(5-6), 477-480.
- Campbell, T. W. 2004. Clinical chemistry of fish and amphibians. Pages 499–517 in M. A. Thrall, D. C. Baker, T.W. Campbell, D. DeNicola, M. J. Fettman, E. D. Lassen, A. Rebar, and G. Weiser, editors. *Veterinary hematology and clinical chemistry: text and clinical case presentations*. Lippincott Williams and Wilkins, Philadelphia.
- Collette, B. B., & Nauen, C. E. (1983). *Scombrids of the world: an annotated and illustrated catalogue of tunas, mackerels, bonitos, and related species known to date*. v. 2.
- Çelik, E. S., & Aydin, S. (2006). Effect of *Trachelobdella lubrica* (Hirudinea: Piscicolidae) on biochemical and haematological characteristics of black scorpion fish (*Scorpaena porcus*, Linnaeus 1758). *Fish Physiology and Biochemistry*, 32(3), 255–260. doi: 10.1007/s10695-006-9003-y
- Datta, N., Kar, P. K., & Saha, S. K. (2022). Primary stress response and biochemical profile of *Labeo rohita* (Hamilton, 1822) experimentally parasitized with *Argulus bengalensis* (Ramakrishna, 1951). *Journal of Fish Biology*.
- Devevey, G., Niculita-Hirzel, H., Biollaz, F., Yvon, C., Chapuisat, M., & Christe, P. (2008). Developmental, metabolic and immunological costs of flea infestation in the common vole. *Functional Ecology*, 22(6), 1091–1098. doi:10.1111/j.1365-2435.2008.01493.x
- Hadziavdic, K., Lekang, K., Lanzen, A., Jonassen, I., Thompson, E. M., & Troedsson, C. (2014). Characterization of the 18S rRNA gene for designing universal eukaryote specific primers. *PLoS one*, 9(2), e87624. doi: 10.1371/journal.pone.0087624
- Hall, T. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. In *Nucleic Acids Symp. Ser.* (Vol. 41, pp. 95-98).
- Kundu, I., Bandyopadhyay, P. K., Mandal, D. R., & Güreli, G. (2016). Study of pathophysiological effects of the nematode parasite *Eustrongylides* sp. on freshwater fish *Channa punctatus* by hematology, serum biochemical, and histological studies. *Türkiye Parazitoloji Dergisi*, 40(1), 42. doi: 10.5152/tpd.2016.4551
- Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., McGettigan, P. A., McWilliam, H., Valentin, F., Wallace, I.M., Wilm, A., Lopez, R., Thompson, J.D., Gibson T.J., & Higgins, D. G. (2007). Clustal W and Clustal X version 2.0. *Bioinformatics*, 23(21), 2947-2948. doi: 10.1093/bioinformatics/btm404
- Macchioni, F., Tedesco, P., Cocca, V., Massaro, A., Sartor, P., Ligas, A., Pretti, C., Monni, G., Cecchi, F., & Caffara, M. (2021). Anisakid and Raphidascaridid parasites in *Trachurus trachurus*: infection drivers and possible effects on the host's condition. *Parasitology Research*, 120(9), 3113–3122. doi:10.1007/s00436-021-07200-0
- Manna, S., & Naskar, S. (2021). Impact of *Thelohanellus mrigalae* tripathi, 1952 on *Cirrhinus mrigala*: prevalence, histopathological and haematological alterations. *Uttar Pradesh Journal of Zoology*, 42(9), 41-49. doi: 10.13140/RG.2.2.25174.57924
- Maqbool, A., & Ahmed, I. (2016). Haematological response of snow barbell, *Schizothorax plagiostomus* Heckel, naturally infected with a new Trypanosoma species. *Journal of Parasitic Diseases*, 40(3), 791-800. doi: 10.1007/s12639-014-0580-x
- Martins, M. L., Tavares-Dias, M., Fujimoto, R. Y., Onaka, E. M., & Nomura, D. T. (2004). Haematological alterations of *Leporinus macrocephalus* (Osteichthyes: Anostomidae) naturally infected by *Goezia leporini* (Nematoda: Anisakidae) in fish pond. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia*, 56(5), 640–646. doi:10.1590/s0102-09352004000500011

- Mattiucci, S., & D'Amelio, S. (2014). Anisakiasis. In: Bruschi, F. (eds) *Helminth Infections and their Impact on Global Public Health* (pp. 325-365). Springer, Vienna.
- Mattiucci, S., Farina, V., Campbell, N., MacKenzie, K., Ramos, P., Pinto, A. L., Abaunza, P., & Nascetti, G. (2008). *Anisakis* spp. larvae (Nematoda: Anisakidae) from Atlantic horse mackerel: Their genetic identification and use as biological tags for host stock characterization. *Fisheries Research*, 89(2), 146–151. doi: 10.1016/j.fishres.2007.09.032
- Mattiucci, Simonetta, Cipriani, P., Levsen, A., Paoletti, M., & Nascetti, G. (2018). *Molecular Epidemiology of Anisakis and Anisakiasis: An Ecological and Evolutionary Road Map*. In *Advances in Parasitology* (1st ed., Vol. 99). Elsevier Ltd. doi: 10.1016/bs.apar.2017.12.001
- McAllister, M., Phillips, N., & Belosevic, M. (2019). *Trypanosoma carassii* infection in goldfish (*Carassius auratus* L.): changes in the expression of erythropoiesis and anemia regulatory genes. *Parasitology Research*, 118(4), 1147-1158. doi: 10.1007/s00436-019-06246-5
- Menconi, V., Pastorino, P., Canola, S., Pavoletti, E., Vitale, N., Scanzio, T., Righetti, M., Mugetti, D., Tomasoni, M., Bona, M. C., & Prearo, M. (2022). Occurrence and spatial variation of *Anisakis pegreffii* in the Atlantic horse mackerel *Trachurus trachurus* (Carangidae): A three-year monitoring survey in the western Ligurian Sea. *Food Control*, 131, 108423. doi: 10.1016/j.foodcont.2021.108423
- Mladineo, I., & Poljak, V. (2014). Ecology and genetic structure of zoonotic *Anisakis* spp. from Adriatic commercial fish species. *Applied and Environmental Microbiology*, 80(4), 1281-1290. doi:10.1128/AEM.03561-13
- Nabi, S., Tanveer, S., Ganie, S. A., & Bashir, K. (2022). Oxidative stress, serum biochemistry and DNA damage of *Cyprinus carpio* communis naturally infected with helminths. *International Aquatic Research*, 14(1), 13-22. doi: 10.22034/IAR.2022.1937373.1179
- Nikolsky, G.W. (1963). *The Ecology of Fishes*, Academic Press, London and New York, 352 p.
- Nnabuchi, O., Nwani, C. D., Ochang, S., & Somdare, P. (2015). Effect of parasites on the biochemical and haematological indices of some clariid (Siluriformes) catfishes from Anambra River, Nigeria. *International Journal of Fisheries and Aquatic Studies*, 3(2), 331–336.
- Ondračková, M., Francová, K., Dávidová, M., Polačik, M., & Jurajda, P. (2010). Condition status and parasite infection of *Neogobius kessleri* and *N. melanostomus* (Gobiidae) in their native and non-native area of distribution of the Danube River. *Ecological Research*, 25(4), 857-866. doi: 10.1007/s11284-010-0716-0
- Özbakiş Beceriklisoy, G., Aştı, C., & Gönenç, B. (2020). Marmara denizi Atlantik uskumrularındaki (*Scomber scombrus*, Linnaeus 1758) *Anisakis* spp. enfeksiyonu. *Veteriner Hekimler Derneği Dergisi*, 91(1), 80–85. doi:10.33188/vetheder.599455
- Özdemir, G., Çelik, E. Ş., Yılmaz, S., Gürkan, M., & Kaya, H. (2016). Histopathology and blood parameters of bogue fish (*Boops boops*, Linnaeus 1758) parasitized by *Ceratothoa oestroides* (Isopoda: Cymothoidae). *Turkish Journal of Fisheries and Aquatic Sciences*, 16(3), 579-590. doi: 10.4194/1303-2712-v16\_3\_28
- Panjvini, F., Abarghuei, S., Khara, H., & Parashkoh, H. M. (2016). Parasitic infection alters haematology and immunity parameters of common carp, *Cyprinus carpio*, Linnaeus, 1758. *Journal of Parasitic Diseases*, 40(4), 1540-1543. doi: 10.1007/s12639-015-0723-8
- Pekmezci, G. Z., Onuk, E. E., Bolukbas, C. S., Yardimci, B., Gurler, A. T., Acici, M., & Umur, S. (2014). Molecular identification of *Anisakis* species (Nematoda: Anisakidae) from marine fishes collected in Turkish waters. *Veterinary Parasitology*, 201(1–2), 82–94. doi: 10.1016/j.vetpar.2014.01.005
- Quiazon, K. M. A., Yoshinaga, T., Santos, M. D., & Ogawa, K. (2009). Identification of larval *Anisakis* spp. (Nematoda: Anisakidae) in Alaska pollock (*Theragra chalcogramma*) in northern Japan using morphological and molecular markers. *Journal of Parasitology*, 95(5), 1227-1232. doi: 10.1645/GE-1751.1
- Rastiannasab, A., Afsharmanesh, S., Rahimi, R., & Sharifian, I. (2016). Alternations in the liver enzymatic activity of Common carp, *Cyprinus carpio* in response to parasites, *Dactylogyrus* spp. and *Gyrodactylus* spp. *Journal of Parasitic Diseases*, 40(4), 1146-1149. doi: 10.1007/s12639-014-0638-9
- Ryberg, M. P., Huwer, B., Nielsen, A., Dierking, J., Buchmann, K., Sokolova, M., Krumme U., & Behrens, J. W. (2022). Parasite load of Atlantic cod *Gadus morhua* in the Baltic Sea assessed by the liver category method, and associations with infection density and critical condition. *Fisheries Management and Ecology*, 29(1), 88-99. doi: 10.1111/fme.12516
- Ryberg, M. P., Skov, P. V., Vendramin, N., Buchmann, K., Nielsen, A., & Behrens, J. W. (2020). Physiological condition of Eastern Baltic cod, *Gadus morhua*, infected with the parasitic nematode *Contracaecum osculatatum*. *Conservation Physiology*, 8(1), coaa093. doi: 10.1093/conphys/coaa093
- Sakanari, J. A., & McKerrow, J. H. (1989). Anisakiasis. *Clinical Microbiology Reviews*, 2(3), 278–284. doi:10.1128/CMR.2.3.278
- Santos, M. J., Castro, R., Cavaleiro, F., Rangel, L., & Palm, H. W. (2017). Comparison of anisakid infection levels between two species of Atlantic mackerel (*Scomber colias* and *S. scombrus*) off the Atlantic

- Portuguese coast. *Scientia Marina*, 81(2), 179-185. doi:10.3989/scimar.04552.26A
- Shameena, S. S., Kumar, K., Kumar, S., Kumari, P., Krishnan, R., Karmakar, S., S., Kumar, H. S., Rajendran, K. V., & Raman, R. P. (2021). Dose-dependent co-infection of *Argulus* sp. and *Aeromonas hydrophila* in goldfish (*Carassius auratus*) modulates innate immune response and antioxidative stress enzymes. *Fish & Shellfish Immunology*, 114, 199-206. doi: 10.1016/j.fsi.2021.04.026
- Sousa, L. F., Souza, D. C., Coelho, T. A., Tavares-Dias, M., & Correa, L. L. (2020). Morphometric Characterization of *Trypanosoma* spp. and blood parameters in *Pterygoplichthys pardalis* (Pisces: Loricariidae) from the Brazilian Amazon. *Anais da Academia Brasileira de Ciências*, 92.
- Tavares-Dias, M., Martins, M. L., & Kronka, S. D. N. (1999). Evaluation of the haematological parameters in *Piaractus mesopotamicus* Holmberg (Osteichthyes, Characidae) with *Argulus* sp. (Crustacea, Branchiura) infestation and treatment with organophosphate. *Revista Brasileira de Zoologia*, 16(2), 553-555. doi:10.1590/s0101-81751999000200019
- Utuk, A.E., Pişkin, F.Ç. & Balkaya, I. 2012. Molecular detection of *Anisakis pegreffii* in horse mackerels (*Trachurus trachurus*) sold for human consumption in Erzurum Province of Turkey. *Kafkas Üniversitesi Veterinerlik Fakültesi Dergisi*, 18(2), 303-307. doi:10.9775/kvfd.2011.5466
- White, A., & Fletcher, T. C. (1985). Seasonal changes in serum glucose and condition of the plaice, *Pleuronectes platessa* L. *Journal of Fish Biology*, 26(6), 755-764. doi:10.1111/j.1095-8649.1985.tb04316.x
- Whitehead P.J.P., Bauchot M-L., Hureau J-C., Nielsen J., & Tortonese E. (1984). *Fishes of the North-Eastern Atlantic and the Mediterranean*. 1, Paris, UNESCO.
- Yardimci, B., Pekmezci, G. Z., & Onuk, E. E. (2014). Pathology and molecular identification of *Anisakis pegreffii* (Nematoda: Anisakidae) infection in the John Dory, *Zeus faber* (Linnaeus, 1758) caught in Mediterranean Sea. *Ankara Üniversitesi Veteriner Fakültesi Dergisi*, 61(3), 233-236. doi: 10.1501/Vetfak\_0000002635
- Yılmaz, S. (2018). *Balık İmmunolojisi Analiz Yöntemleri/Methods of Fish Immunology Analysis*. Paradigma Akademi Kitabevi Yayınları, İSTANBUL, 105 sayfa.
- Yılmaz, S., & Ergün, S. (2012). Effects of garlic and ginger oils on hematological and biochemical variables of sea bass *Dicentrarchus labrax*. *Journal of Aquatic Animal Health*, 24(4), 219-224. doi: 10.1080/08997659.2012.711266
- Yılmaz, S., Ergun, S., Şanver Çelik, E., Yigit, M., & Bayizit, C. (2019). Dietary trans-cinnamic acid application for rainbow trout (*Oncorhynchus mykiss*): II. Effect on antioxidant status, digestive enzyme, blood biochemistry and liver antioxidant gene expression responses. *Aquaculture Nutrition*, 25(6), 1207-1217. doi: 10.1111/anu.12935
- Yılmaz, S., Ergün, S., & Çelik, E. Ş. (2016). Effect of dietary spice supplementations on welfare status of sea bass, *Dicentrarchus labrax* L. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, 86(1), 229-237. doi: 10.1007/s40011-014-0444-2
- Yılmaz, S., Sebahattin, E., & Celik, E. (2012). Effects of herbal supplements on growth performance of sea bass (*Dicentrarchus labrax*): Change in body composition and some blood parameters. *Journal of BioScience and Biotechnology*, 1, 217-222,
- Zardoya, R., Castilho, R., Grande, C., Favre-Krey, L., Caetano, S., Marcato, S., Krey, G., & Patarnello, T. (2004). Differential population structuring of two closely related fish species, the mackerel (*Scomber scombrus*) and the chub mackerel (*Scomber japonicus*), in the Mediterranean Sea. *Molecular Ecology*, 13(7), 1785-1798. doi: 10.1111/j.1365-294X.2004.02198.x
- Zhang, Z., Schwartz, S., Wagner, L., & Miller, W. (2000). A greedy algorithm for aligning DNA sequences. *Journal of Computational Biology*, 7(1-2), 203-214. doi: 10.1089/10665270050081478