

Bacteria-to-bacteria communication, Signaling Molecules: AHLs, AIPs and AI-2, I can't talk now matey, gone to pathogenesis!

Nurdan Filik^{1,*}, Fethi Filik¹

¹Suleyman Demirel University, Department of Diseases, Isparta, Türkiye

Abstract: How do these primitive tiny organisms' hierarchy operate? The answer is simple: by communicating. Yes, bacteria talk, and it's called: Quorum Sensing System (QS). QS is a phenomenon where bacteria tynychat and arrange their moves via the accumulation of signaling molecules. The cells respond to this stimulus when the cut-off value concentration of molecules is achieved. This phenomenon is widely visible in the bacteria world. Bacteria never move solo and would need to gather up in critical mass because of secrete efficient toxins to be lethal. *N*-Acyl Homoserine Lactones (AHLs): Intercellular signaling molecules used by Gram-negative bacteria (Gram⁻) to monitor their critical mass density in QS controlling of critic gene expression. AHL signals are synthesized by LuxI proteins. AHLs are vital interbacterial signaling molecules used by bacteria to check their dependent bacteria density. Auto-inducing peptides (AIPs): signaling molecules involved in intercellular communication in Gram-positive bacteria (Gram⁺). Peptides are exported by dedicated systems, post-translationally modified and eventually sensed by other bacteria cells via membrane-located receptors that are part of a two-component system. AI-2 (Autoinducer-2): signaling molecule used by interspecies bacteria communication. This molecule chemically identified furanosylborate diester synthesized by LuxS proteins. Also, it's a universal signal because it carries out interspecies communication. The aim of this review is to summarize the AHL, AIPs, AI-2 bacterial signal molecules, QS systems target genes, effective in the prokaryotic world, and the micro-social lifestyle of bacteria.

ARTICLE HISTORY

Received: Feb. 23, 2023

Revised: Aug. 29, 2023

Accepted: Sep. 13, 2023

KEYWORDS

Signal Molecules: AHLs, AIPs AI-2, Quorum Sensing System, Bacterially Speaking, Bacteria-to-bacteria Communication, Intercellular Communication.

1. INTRODUCTION

Tynychat comes true via *N*-Acyl Homoserine Lactones (AHL, *N*-AHLs or AHLs) signal molecules managed by a Quorum Sensing System (QS). QS is the coordination of bacterial group movies based on cell density. Thanks to this attitude, bacterial tynychats are established. As a result of tynychat, serious big consequences such as diseases occur (Parveen & Cornell, 2011; Winsberg, 2022). The properties that a real QS molecule should have, unlike a metabolite, are summarized as follows: I. Production of the QS molecule occurs at different phases of reproduction, under special conditions changes. II. The QS molecule agglomerates extracellularly and is detected whereby specific receptors. III. Accumulation of the QS molecule elicits a planned response when it reaches a critical threshold. IV. The cellular

*CONTACT: Nurdan Filik ✉ nurdansal@hotmail.com 📍 Suleyman Demirel University, Isparta, Türkiye

response elicited by the QS molecule is much broader than metabolizing or detoxifying the QS molecule. While many metabolites show the first three of these four features, the fourth feature is a feature that a QS molecule must have (Winzer *et al.*, 2002).

Gram-negative bacteria (Gram⁻) use AHL as their command language to coordinate population behavior during invasion and colonization of higher organisms. AHL is synthesized from precursors by a synthase protein, “LuxI” and once they have reached a certain cut-off value concentration, they interact with a transcriptional activating “LuxR” protein to induce target gene expression (Fuqua *et al.*, 1996). So, QS operates that lets bacteria sense and answer target genes depending on their bacteria density. (Eberl *et al.*, 1996; Pütz *et al.*, 2022). In Gram-positive bacteria (Gram⁺), the instrument of the communication system functions via Autoinducing Peptide (AIPs) (Sahreen *et al.*, 2022). In interspecies, the instrument of communication is via Autoinducer-2 (AI-2) (Liu *et al.*, 2022). Bacteria become more prepotent by communicating. They act simultaneously, perform critical gene expressions such as virulence, and perform gene transfer. They coordinate such multiple social behaviors through signaling molecules. In fact, different types of bacteria can communicate with each other thanks to signal molecules in crosstalk.

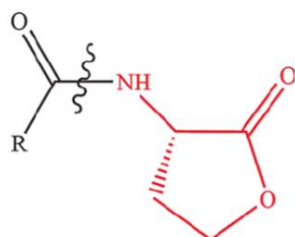
The aim of this review is to summarize the various AHL, AIPs, and AI-2 bacterial signal molecules currently available, QS system management prokaryotic world, and micro-social lifestyle of bacteria.

2. SIGNALING MOLECULES

2.1. AHL Molecules Structure

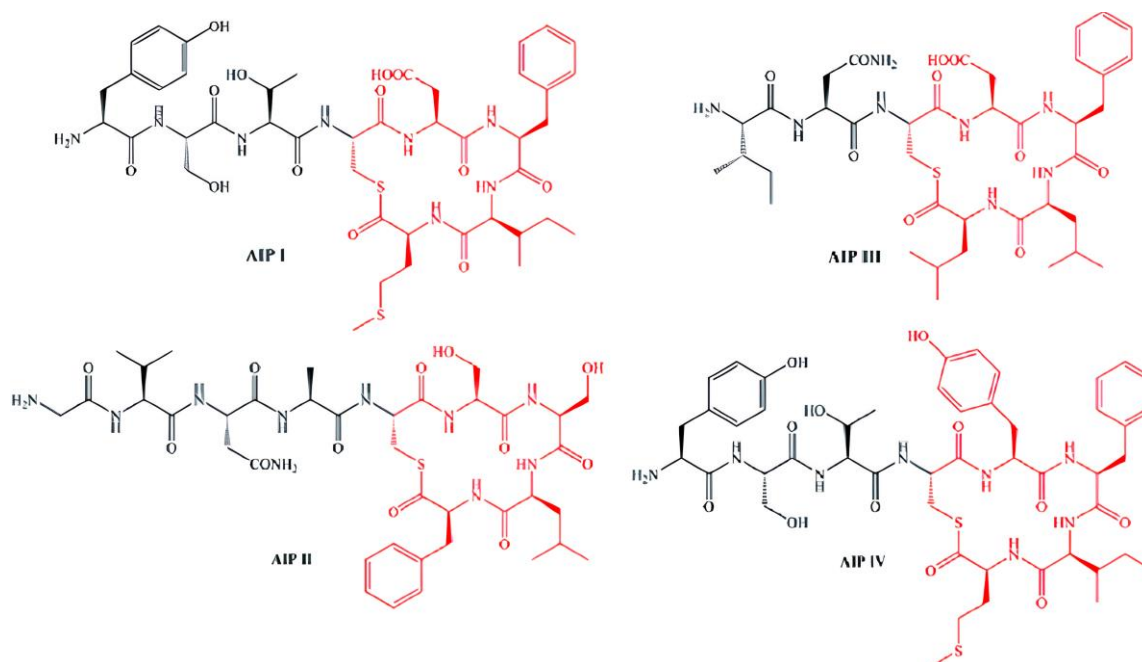
The main structural components of AHL (Figure 1) are hydrophilic sections of homoserine lactone ring (S-adenosylmethionine) and central amide group, as well as hydrophobic section strain-specific side hydrocarbon chain with varieties in length and level of oxygenation with a 3-oxo group. Acyl chain generally ranges from 4 to 18 carbons (Marin *et al.*, 2007). In bacterial signaling, AHL is produced and released by bacteria cells. AHLs produced by different bacteria differ in the length of the R-group sidechain (Kumari, 2006). In QS, LuxI protein is synthesized by AHL. AHL can pass through the cell membrane down a gradient to environmental space. When molecule concentration reaches the threshold value, cognate LuxR protein binds to AHL. So, AHL directs target gene transcription (von Rad *et al.*, 2008).

Figure 1. AHLs structure used by Gram⁻ (Konai *et al.*, 2018).



2.2. AIPs Peptide Signaling Molecules Structure

AIPs are characterized as AIP I-II-III-IV (Figure 2). Peptides may have differences in amino acid sequence; however, all of them possess an increased hydrophobicity from their N- to C-terminal end in peptide structure. At the end of the sequence to the C-terminal positions, AIPs are found to have amino acids with hydrophobic side chains (Mayville *et al.* 1999; MDowell *et al.* 2001; Yang *et al.* 2016). AIP linear peptide analogs or hydrolysis of the thioester moiety inactivates its functions molecule. Thioester macrocyclic ring is an ester moiety that was found to inactivate the signaling. Upon removal of N-terminal exocyclic structures, AIP loses signaling (Konai *et al.*, 2018).

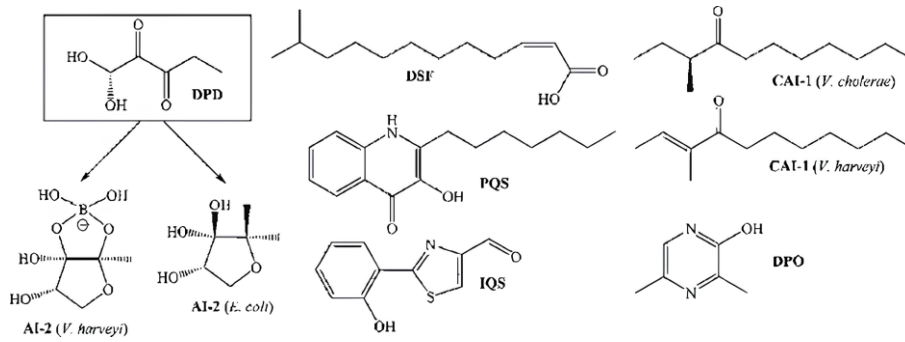
Figure 2. AIPs structure used by Gram⁺ (Konai *et al.*, 2018).

2.3. AI-2 Molecules Structure

AI-2 is a set of interconverting molecules derived from 4,5-dihydroxy 2,3-pentanedione (DPD) (Figure 3) (Qiao *et al.*, 2022). LuxS is a synthase enzyme of DPD (Pereira *et al.* 2013). AI-2 synthesis starts with SAM (Federle & Bassler 2003). Different forms of DPD act as AI-2 for interbacteria. AI-2's chemical nature structure is the same for many bacterial species. This suggests that AI-2 is used for inter-species communication (crosstalk) to detect other bacteria (Miller *et al.* 2004). In AI-2-based QS, signal receive, and transduction are conducted by a two-component pathway. The first component consists of LuxP and LuxQ while the second component constitutes phosphotransferase LuxU and cytoplasmic response LuxO. There exist LuxPQ complexes with symmetric heterotetramer in AI-2. Upon binding with AI-2, the tetramer undergoes a change. This change prevents the phosphorylation of the cytoplasmic proteins, LuxU and LuxO, which terminates the expression of genes. Finally, LuxR is produced (Konai *et al.*, 2018).

Significant works on the three-dimensional structures of proteins involved in QS first came into light in 2001 by determining the crystal structures of three LuxS orthologs with the help of X-ray crystallography. This was closely followed by determination of the crystal structure of the receptor LuxP of *Vibrio harveyi* with its inducer AI-2 (which is one of the few biomolecules containing boron) bound to it. AI-2 signalling is conserved among many bacterial species, including *Escherichia coli*, the model organism (Lewis *et al.*, 2001; Chen *et al.*, 2002). A database of QS peptides is existing “Quorumpeps” (Wynendaele *et al.*, 2013; Wynendaele *et al.*, 2015).

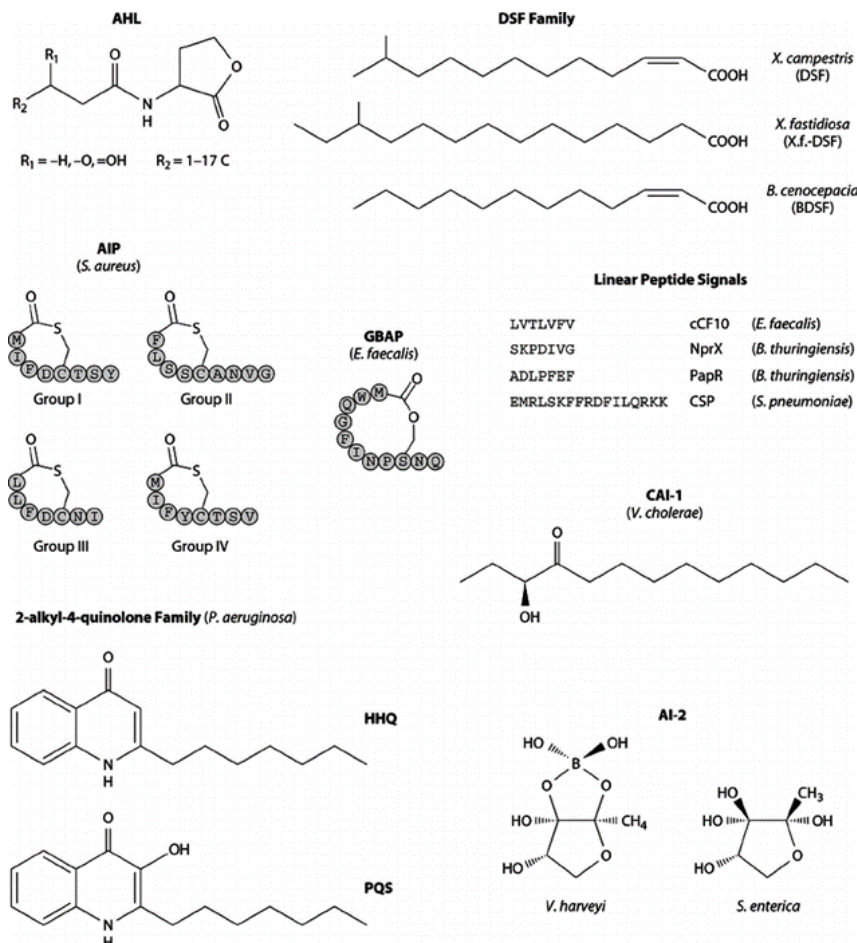
Figure 3. AI-2 and other signal molecules used by bacteria (Qiao *et al.*, 2022).



3. COMMUNICATION INSTRUMENTS IN BACTERIA

Briefly, AHL is produced by LuxI-type and by LuxR-type in Gram. The second mechanism involves AIPs in Gram⁺ which are sensed by two-component systems. The final system is the AI-2 system which is common in both Gram⁻ and Gram⁺ (Figure 4) (Brackman & Coeny, 2015). AIPs are often integral elements of a histidine kinase two-component signal transduction system. In this system, extracellular proteases process the secreted precursor-AIP into mature AIP. Kinase phosphorylates regulate gene transcription. This is called a two-component system (Rutherford & Bassler, 2012). AHLs are important “messenger” molecules involved in cellular communication of Gram⁻ (Rutherford & Bassler, 2012). Usually, AHL does not need additional processing and binds directly to transcription factors to regulate gene expression (Bassler, 1999). Communication instruments in bacteria are shown in Table 1.

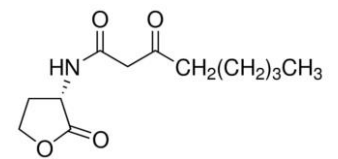
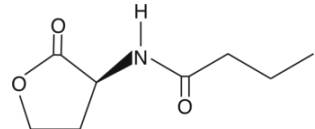
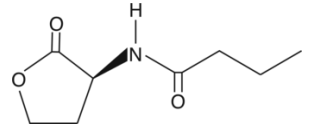
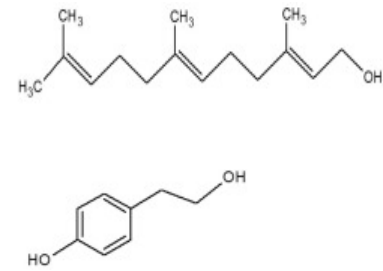
Figure 4. General structures of the selected QS signal molecules (LaSarre & Federle, 2013).

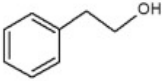
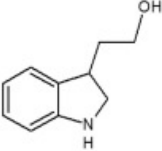
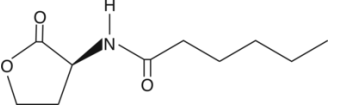
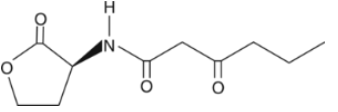
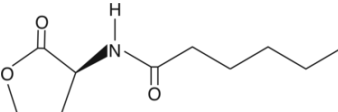
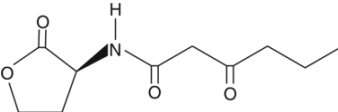


4. DISCOVERY of AHL, AIPs, AI-2

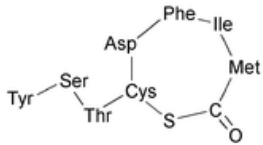
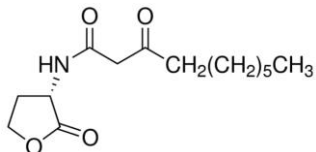
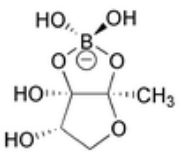
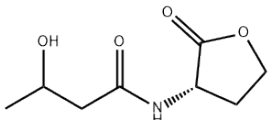
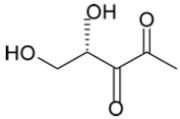
Previously pathogenic bacteria were tested against 2 AHL monitor bacteria (*Agrobacterium tumefaciens* NT1 and *Chromobacterium violaceum* CV026) in a well-diffusion assay (Bruhn *et al.*, 2005). However, currently for AHL extraction problem is components present in bacterial culture supernatants. Components are cell growth media and extracellular products produced by bacteria. To reduce extracellular products, a stationary bacteria growth phase is recommended at extraction. Liquid-liquid extraction (LLE) and solid-phase extraction (SPE) are commonly used methods to isolate AHL (Huang *et al.*, 2020). Several methods have been developed for purification, detection and quantification of the AHLs. Methods have been developed for the detection of tiny chat molecules. These methods are biosensors of HSLs, Thin-layer chromatography (TLC) (Laj *et al.*, 2022), Radiolabeled assay (Schaefer *et al.*, 2018), High-performance liquid chromatography (HPLC) (Bao *et al.*, 2022), Colorimetry (Jin *et al.*, 2020). For identification of AHL spectroscopic properties have been widely used to the characterization of QS molecule structures. Spectroscopic include Mass Spectrometry (MS) (Rosario *et al.*, 2022), MS/MS, nuclear magnetic resonance spectroscopy (NMR) (Sundar *et al.*, 2022), and infrared spectroscopy (IR) (Deepa *et al.*, 2022). These methods are HPLC-MS (High-performance liquid chromatography-Mass spectrometry) and GC-MS (Gas chromatography-Mass spectrometry), Nano-LC-MS/MS (Frommberger *et al.*, 2004), At-line coupling of UPLC (Ultra-high-pressure liquid chromatography) to chip-electrospray-FTICR-MS (Fourier-transform ion-cyclotron resonance mass spectrometry) (Li *et al.*, 2007a), Capillary zone electrophoresis mass spectrometry (CZE-MS), Nuclear magnetic resonance (NMR) (Bainton *et al.*, 1992), IR spectrometry (IR) (Wang *et al.*, 2011).

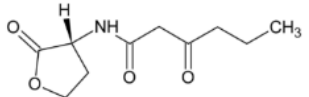
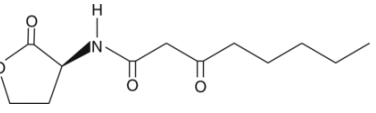
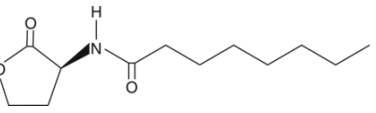
Table 1. Bacteria possessing QS system target genes, signal molecules, structure, and function in talking via chemical languages.

Strain	Gram* staining	Signal molecules	Structure	QS system target genes	Function	Reference
<i>Agrobacterium tumefaciens</i>	Gram ⁻	<i>N</i> -(3-oxooctanoyl)-L-homoserine lactone (OOHL)		<i>traR-traI</i>	Factors for conjugal Ti plasmids transfer	Zavilgelsky & Manukhov, (2001)
<i>Aeromonas hydrophila</i>	Gram ⁻	<i>N</i> -butanoyl-L-homoserine lactone (C ₄ -HSL, BHL)		<i>luxI-luxR</i> <i>AhyI-AhyR</i>	Virulence, biofilm, hemolysis	Filik, 2020
<i>Aeromonas salmonicidae</i>	Gram ⁻	C ₄ -HSL, BHL		<i>luxI-luxR</i> <i>AsaI-AsaR</i>	Exoprotease production	Miller & Bassler, (2001)
<i>Bacillus subtilis</i>	Gram ⁺	Competence and Sporulation Stimulating Factor (CSF)	Glu-Arg-Gly-Met-Thr		CSF adsorption, serious membrane damage, gene expression	Huang <i>et al.</i> (2021)
<i>Candida albicans</i>	Yeast**	Farnesol Tyrosol		<i>LuxS</i>	Dimorphism, biofilm	Kim & Yeon, (2018)

		Phenylethanol				
		Tryptophol				
<i>Chromobacterium violaceum</i>	Gram ⁻	<i>N</i> -hexanoyl-L-homoserine lactone (C ₆ -HSL, HHL)		<i>luxI-luxR</i> <i>CviI/CviR</i>	Violacein pigment production	McClellan <i>et al.</i> , (1997)
<i>Enterobacter agglomerans</i>	Gram ⁻	3-oxo-C ₆ -HSL		<i>luxI-luxR</i> <i>EagI/EagR</i>	Pheromone production	Swift <i>et al.</i> , 1993
<i>Erwinia carotovora</i>	Gram ⁻	C ₆ -HSL, HHL		<i>luxI-luxR</i>	Carbapenem antibiotic production	Zavilgelsky & Manukhov, 2001
		<i>N</i> -(β-ketocaproyl)-L-Homoserine lactone (3-Oxo-C ₆ -HSL)			Virulence	Burr <i>et al.</i> , 2006

<i>E. coli</i>	Gram ⁻	AI-2		<i>LuxS</i>	Cell division, chromosome replication, cell signaling, translational modification, pathogenesis mechanismsprotein	Soni <i>et al.</i> , 2007
		3OC ₈ HSL		NA (Not applicable) - SdiA	Motility, acid resistance	Soares & Ahmer 2011
<i>Pseudomonas aeruginosa</i>	Gram ⁻	C ₄ -HSL, BHL		<i>rhlI-rhlR</i>	Virulence, secondary metabolites (rhamnolipids)	Zavilgelsky & Manukhov, 2001
		<i>N</i> -(3-oxododecanoyl)-L-homoserine lactone (OdDHL, 3OC ₁₂ -HSL)		<i>lasI-lasR</i>	Virulence (toxin A, elastase)	
<i>Pseudomonas aureofaciens</i>	Gram ⁻	C ₆ -HSL, HHL		<i>LuxI-LuxR PhzI-PhzR</i>	Produces of exoproducts, including protease, phenazines, phenazine antibiotic biosynthesis	Zhang & Pierson, 2001

<i>Staphylococcus aureus</i>	Gram ⁺	Oligopeptide Autoinducers Peptides (AIPs)		<i>Agr</i>	Virulence	Malone <i>et al.</i> , 2007
<i>Vibrio anguillarum</i>	Gram ⁻	3-oxo-C ₁₀ -HSL (ODHL)		<i>LuxI-LuxR</i> <i>VanI/VanR</i>	Bioluminescence Violacein pigment production	Milton <i>et al.</i> 1997
<i>Vibrio harveyi</i>	Gram ⁻	<i>N</i> -octanoyl-L-homoserine lactone (OHL), Autoinducer-2 (AI-2)		<i>ainS-ainR</i>		
		<i>N</i> -(3-hydroxybutanoyl)-L-homoserine lactone (HBHL), AI-1		<i>luxL-luxM</i> <i>luxR-luxN</i>	Bioluminescence	Geske, 2008
		4,5-dihydroxypentane-2,3-dione (DPD)		<i>luxS-luxQ</i>		

<i>Vibrio fischeri</i>	Gram ⁻	N-(3-oxohexanoyl)-L-homoserine lactone (OHHL), AI-1		<i>luxI-luxR</i>	Bioluminescence	Zavilgelsky & Manukhov, 2001
<i>Yersinia ruckeri</i>	Gram ⁻	OOHL		<i>luxI-luxR</i>	Virulence	Bruhn <i>et al.</i> , 2005
		OHL				

*Gram staining is an assay applied only to bacteria.

***C. albicans* is yeast and has nothing to do with gram stain.

5. DISCUSSION and CONCLUSION

Interbacterial communication systems based on signaling molecules have been determined in pathogenic Gram⁻ and Gram⁺ causing tough problems. The ability to mark bacterial communication at single-cell level in situ enables exploration of interbacterial communication, intercellular signal transduction, and QS, and it gives us a tremendous potential for studying the efficiency of communication systems in complex virulence systems scenarios (Andersen *et al.*, 2001). Many species of microorganisms exhibit a social behavior.

QS in prokaryotic biology refers to the ability of bacteria to sense information from other cells in the population when they reach a critical concentration. They communicate with each other through signal molecules they have produced, monitor whether they have reached a certain majority, and trigger critical gene expressions such as synthesis of virulence as soon as they reach a sufficient majority. Thus, by not stimulating the host's immune system prematurely, it creates a successful infection process (Thirunavukkarasu *et al.*, 2023).

Researchers have established “SigMol” (<http://bioinfo.imtech.res.in/manojk/sigmol>), a specialized repository of molecules in procaryotes. SigMol harbors information on QSSMs pertaining to different signaling systems namely AHL, AIPs, AI-2, and others. The database consists of 1382 entries of 182 peerless signal molecules from 215 microorganisms. SigMol encompasses biological (genes etc.) and chemical (IUPAC name, SMILES and structure etc.) properties of molecules (Rajput *et al.*, 2016).

By secreting chemical toxins, all bacteria by themselves would receive the signals and answer them by activating the transcription of virulent genes and changing their social behavior to become highly pathogenic. Bacteria possess an extraordinary repertoire for interbacterial communication and social movements.

The discovery of communication molecules in the bacterial world attracts attention to fish disease control. Destroying AHL signal molecules, which are communication instruments, before they can occur the disease, brings up the concept of early diagnosis and in this case, it is aimed to era in prophylaxis. So, explaining to molecules in all detail will be an important step in diagnosis.

The similarities between the signals used by bacteria and artificial neural networks are striking. Based on the finding that bacteria have many of the properties of a neural network, bacteria may have a low level of intelligence. Researchers have stated that billions of bacteria collectively carry out the same command in their experiment by putting bacteria into communication with each other with the program loaded in their DNA. Scientists state that billions of bacteria that can communicate with each other can be managed at the same time and directed to certain tasks. They point out that in the future, smart biological devices will also be reflected in daily life.

Declaration of Conflicting Interests and Ethics

The authors declare no conflict of interest. This research study complies with research and publishing ethics. The scientific and legal responsibility for manuscripts published in IJSM belongs to the authors.

Authorship Contribution Statement

Nurdan Filik: Investigation, conceptualized the review articles. Also, a literature review. **Fethi Filik:** The draft preparation and involved in the drafting and edition. Also, a literature review. The drafts were critically discussed and revised by all authors.

Orcid

Nurdan Filik  <https://orcid.org/0000-0003-4376-7298>

Fethi Filik  <https://orcid.org/0000-0003-3564-8782>

REFERENCES

- Andersen, J.B., Heydorn, A., Hentzer, M., Eberl, L.E.O., Geisenberger, O., Christensen, B.B., ... & Givskov, M. (2001). gfp-based *N*-acyl homoserine-lactone sensor systems for detection of bacterial communication. *Applied and Environmental Microbiology*, 67(2), 575-585.
- Bainton, N.J., Bycroft, B.W., Chhabra, S.R., Stead, P., Gledhill, L., Hill, P.J., ... & Williams, P. (1992). A general role for the lux autoinducer in bacterial cell signalling: control of antibiotic biosynthesis in *Erwinia*. *Gene*, 116(1), 87-91.
- Bao, J., Guo, D., Jin, L., Li, T., & Shi, H. (2022). Accurate Identification of Diverse *N*-acyl Homoserine Lactones in Marine *Vibrio fluvialis* by UHPLC-MS/MS. *Current Microbiology*, 79(6), 1-11.
- Bassler, B.L. (1999). How bacteria talk to each other: regulation of gene expression by quorum sensing. *Current Opinion in Microbiology*, 2(6), 582-587.
- Brackman, G., Coenye, T. (2015). Quorum sensing inhibitors as anti-biofilm agents. *Current Pharmaceutical Design*, 21(1), 5-11.
- Bruhn, J.B., Dalsgaard, I., Nielsen, K.F., Buchholtz, C., Larsen, J.L., & Gram, L. (2005). Quorum sensing signal molecules (acylated homoserine lactones) in gram-negative fish pathogenic bacteria. *Diseases of aquatic organisms*, 65(1), 43-52.
- Burr, T., Barnard, A.M., Corbett, M.J., Pemberton, C.L., Simpson, N.J., & Salmond, G.P. (2006). Identification of the central quorum sensing regulator of virulence in the enteric phytopathogen, *Erwinia carotovora*: the VirR repressor. *Molecular microbiology*, 59(1), 113-125.
- Chen, X., Schauder, S., Potier, N., Van Dorsselaer, A., Pelczar, I., Bassler, B.L., & Hughson, F.M. (2002). Structural identification of a bacterial quorum-sensing signal containing boron. *Nature*, 415(6871), 545-549.
- Deepa, N., Chauhan, S., Kumari, P., Rai, A.K., Tandon, S., & Singh, A. (2022). Linalool reduces the virulence of *Pseudomonas syringae* pv. *tomato* DC 3000 by modulating the PsyI/PsyR quorum-sensing system. *Microbial Pathogenesis*, 105884.
- Eberl, L., Winson, M.K., Sternberg, C., Stewart, G.S., Christiansen, G., Chhabra, S.R., ... & Givskov, M. (1996). Involvement of *N*-acyl-l-homoserine lactone autoinducers in controlling the multicellular behaviour of *Serratia liquefaciens*. *Molecular microbiology*, 20(1), 127-136.
- Frommberger, M., Schmitt-Kopplin, P., Ping, G., Frisch, H., Schmid, M., Zhang, Y., ... & Kettrup, A. (2004). A simple and robust set-up for on-column sample preconcentration–nano-liquid chromatography–electrospray ionization mass spectrometry for the analysis of *N*-acylhomoserine lactones. *Analytical and bioanalytical chemistry*, 378(4), 1014-1020.
- Fuqua, C., Winans, S.C., & Greenberg, E.P. (1996). Census and consensus in bacterial ecosystems: the LuxR-LuxI family of quorum-sensing transcriptional regulators. *Annual review of microbiology*, 50, 727-752.
- Geske, G.D., O'Neill, J.C., & Blackwell, H.E. (2008). Expanding dialogues: from natural autoinducers to non-natural analogues that modulate quorum sensing in Gram-negative bacteria. *Chemical Society Reviews*, 37(7), 1432-1447.
- Huang, S., Zhang, H., Ng, T.C.A., Xu, B., Shi, X., & Ng, H.Y. (2020). Analysis of *N*-Acy-L-homoserine lactones (AHLs) in wastewater treatment systems using SPE-LLE with LC-MS/MS. *Water Research*, 177, 115756.
- Huang, Q., Chen, J., Zhu, J., Hao, X., Dao, G., Chen, W., ... & Huang, Q. (2021). Divergent bacterial transformation exerted by soil minerals. *Science of The Total Environment*, 784, 147173.

- Jin, L., Zhang, X., Shi, H., Wang, W., Qiao, Z., Yang, W., & Du, W. (2020). Identification of a novel *N*-acyl Homoserine lactone synthase, *AhyI*, in *Aeromonas hydrophila* and structural basis for its substrate specificity. *Journal of Agricultural and Food Chemistry*, 68(8), 2516-2527.
- Kim, S.R., & Yeon, K.M. (2018). Quorum sensing as language of chemical signals. In *Comprehensive Analytical Chemistry*, 81, 57-94. Elsevier.
- Konai, M.M., Dhanda, G., & Haldar, J. (2018). Talking through chemical languages: quorum sensing and bacterial communication. In *Quorum sensing and its biotechnological applications*, 17-42. Springer, Singapore.
- Kumari, A., Pasini, P., Deo, S.K., Flomenhoft, D., Shashidhar, H., & Daunert, S. (2006). Biosensing systems for the detection of bacterial quorum signaling molecules. *Analytical Chemistry*, 78(22), 7603-7609.
- Laj, N., Elayadeth-Meethal, M., Huxley, V.A.J., Hussain, R.R., Kuruniyan, M.S., & Naseef, P.P. (2022). Quorum-sensing molecules: Sampling, identification and characterization of *N*-acyl-homoserine lactone in *Vibrio* sp. *Saudi Journal of Biological Sciences*, 29(4), 2733-2737. <https://doi.org/10.1016/j.sjbs.2021.12.062>
- LaSarre, B., & Federle, M.J. (2013). Exploiting quorum sensing to confuse bacterial pathogens. *Microbiology and Molecular Biology Reviews*, 77(1), 73-111.
- Lerat, E., & Moran, N.A. (2004). The evolutionary history of quorum-sensing systems in bacteria. *Molecular Biology and Evolution*, 21(5), 903-913.
- Lewis, H.A., Furlong, E.B., Laubert, B., Eroshkina, G.A., Batiyenko, Y., Adams, J.M., ... & Buchanan, S.G. (2001). A structural genomics approach to the study of quorum sensing: crystal structures of three LuxS orthologs. *Structure*, 9(6), 527-537.
- Liu, X., Liu, Q., Sun, S., Sun, H., Wang, Y., Shen, X., & Zhang, L. (2022). Exploring AI-2-mediated interspecies communications within rumen microbial communities. *Microbiome*, 10(1), 1-16.
- Malone, C.L., Boles, B.R., & Horswill, A.R. (2007). Biosynthesis of *Staphylococcus aureus* autoinducing peptides by using the *synechocystis* DnaB mini-intein. *Applied and Environmental Microbiology*, 73(19), 6036-6044.
- Marin, S.D.L., Xu, Y., Meijler, M.M., & Janda, K.D. (2007). Antibody catalyzed hydrolysis of a quorum sensing signal found in Gram-negative bacteria. *Bioorganic & Medicinal Chemistry Letters*, 17(6), 1549-1552.
- Mayville, P., Ji, G., Beavis, R., Yang, H., Goger, M., Novick, R.P., & Muir, T.W. (1999) Structure-activity analysis of synthetic autoinducing thiolactone peptides from *Staphylococcus aureus* responsible for virulence. *Proc Natl Acad Sci USA* 96, 1218-1223. <https://doi.org/10.1046/j.1365-2958.2001.02539.x>
- McClellan, K.H., Winson, M.K., Fish, L., Taylor, A., Chhabra, S.R., Camara, M., ... & Williams, P. (1997). Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of *N*-acylhomoserine lactones. *Microbiology*, 143(12), 3703-3711.
- Milton, D.L., Hardman, A., Camara, M., Chhabra, S.R., Bycroft, B.W., Stewart, G.S., & Williams, P. (1997). Quorum sensing in *Vibrio anguillarum*: characterization of the *vanI/vanR* locus and identification of the autoinducer *N*-(3-oxodecanoyl)-L-homoserine lactone. *J Bacteriol*, 179, 3004-3012. <https://doi.org/10.1128/jb.179.9.3004-3012.1997>
- Miller, M.B., & Bassler, B.L. (2001). Quorum sensing in bacteria. *Annual Review of Microbiology*, 55(1), 165-199.
- Miller, S.T., Xavier, K.B., Campagna, S.R., Taga, M.E., Semmelhack, M.F., Bassler, B.L., & Hughson, F.M. (2004). *Salmonella typhimurium* recognizes a chemically distinct form of the bacterial quorum-sensing signal AI-2. *Molecular Cell*, 15, 677-687. <https://doi.org/10.1016/j.molcel.2004.07.020>

- Rajput, A., Kaur, K., & Kumar, M. (2016). SigMol: repertoire of quorum sensing signaling molecules in prokaryotes. *Nucleic Acids Research*, 44(D1), D634-D639.
- Parveen, N., & Cornell, K.A. (2011). Methylthioadenosine/S-adenosylhomocysteine nucleosidase, a critical enzyme for bacterial metabolism. *Molecular Microbiology*, 79(1), 7-20.
- Pereira, H., Azevedo, F., Rego, A., Sousa, M.J., Chaves, S.R., & Côrte-Real, M. (2013) The protective role of yeast cathepsin D in acetic acid-induced apoptosis depends on ANT (Aac2p) but not on the voltage-dependent channel (Por1p). *FEBS Lett*, 587, 200–205. <https://doi.org/10.1016/j.febslet.2012.11.025>
- Pütz, E., Gazanis, A., Keltsch, N.G., Jegel, O., Pfitzner, F., Heermann, R., ... & Tremel, W. (2022). Communication Breakdown: Into the Molecular Mechanism of Biofilm Inhibition by CeO₂ Nanocrystal Enzyme Mimics and How It Can Be Exploited. *ACS Nano*, 16(10), 16091-16108.
- Qiao, Z., Li, J., & Qin, S. (2022). Bioactive Compounds for Quorum Sensing Signal-Response Systems in Marine Phycosphere. *Journal of Marine Science and Engineering*, 10(5), 699.
- Rosario, M.E., Diou-Cass, Q., Harvey, E., Whalen, K., & Rowley, D.C. (2022). Production of signaling molecules by the marine bacterium *Pseudoalteromonas galathea* and implications for antagonistic interactions with the bloom forming coccolithophore *Emiliania huxleyi*. *Chemical Investigation of Metabolites Produced by Marine Pseudoalteromonas spp.*, 42.
- Rutherford, S.T., Bassler, B.L. (2012). Bacterial quorum sensing: its role in virulence and possibilities for its control. *Cold Spring Harbor perspectives in medicine*, 2(11), a012427.
- Sahreen, S., Mukhtar, H., Imre, K., Morar, A., Herman, V., & Sharif, S. (2022). Exploring the function of quorum sensing regulated biofilms in biological wastewater treatment: A review. *International Journal of Molecular Sciences*, 23(17), 9751.
- Schaefer, A.L., Harwood, C.S., & Greenberg, E.P. (2018). “Hot stuff”: The many uses of a radiolabel assay in detecting acyl-homoserine lactone quorum-sensing signals. In *Quorum Sensing*, 35-47. Humana Press.
- Soni, K., Jesudhasan, P., Cepeda, M., Williams, B., Hume, M., Russell, W.K., ... & Pillai, S.D., (2007). Proteomic analysis to identify the role of LuxS/AI-2 mediated protein expression in *Escherichia coli* O157: H7. *Foodborne Pathogens and Disease*, 4(4), 463-471.
- Sundar, K., Prabu, R., & Jayalakshmi, G. (2022). Quorum Sensing Inhibition Based Drugs to Conquer Antimicrobial Resistance. *The Global Antimicrobial Resistance Epidemic: Innovative Approaches and Cutting-Edge Solutions*, 199.
- Swift, S., Winson, M.K., Chan, P.F., Bainton, N.J., Birdsall, M., Reeves, P.J., ... & Stewart, G.S.A.B. (1993). A novel strategy for the isolation of luxI homologues: evidence for the widespread distribution of a LuxR: LuxI superfamily in enteric bacteria. *Molecular Microbiology*, 10(3), 511-520.
- Swift, S., Karlyshev, A.V., Fish, L., Durant, E.L., Winson, M.K., Chhabra, S.R., ... & Stewart, G. (1997). Quorum sensing in *Aeromonas hydrophila* and *Aeromonas salmonicida*: identification of the LuxRI homologs *AhyRI* and *AsaRI* and their cognate N-acylhomoserine lactone signal molecules. *Journal of Bacteriology*, 179(17), 5271-5281.
- Thirunavukkarasu, R., Pandi, P., Pushparaj, S., Subramanian, K., & Arokiarajan, M. S. (2023). Quorum sensing relationship analysis of microbial symbionts. *Microbial Symbionts*, 831.
- Wang, J., Quan, C., Wang, X., Zhao, P., & Fan, S. (2011). Extraction, purification and identification of bacterial signal molecules based on N-acyl homoserine lactones. *Microbial Biotechnology*, 4(4), 479-490.
- MDowell, P., Affas, Z., Reynolds, C., Holden, M.T., Wood, S.J., Saint, S., Cockayne, A., Hill, P.J., Dodd, C.E., Bycroft, B.W., Chan, W.C., & Williams, P. (2001). Structure, activity and

- evolution of the group I thioM. M. Konai et al. lactone peptide quorum-sensing system of *Staphylococcus aureus*. *Mol Microbiol*, 41, 503–512.
- Winsberg, M. (2022). Speaking in Thumbs: A Psychiatrist Decodes Your Relationship Texts So You Don't Have to. Doubleday.
- Winzer, K., Hardie, K.R., & Williams, P. (2002). Bacterial cell-to-cell communication: sorry, can't talk now—gone to lunch!. *Current Opinion in Microbiology*, 5(2), 216-222.
- Wynendaele, E., Bronselaer, A., Nielandt, J., D'Hondt, M., Stalmans, S., Bracke, N., ... & De Spiegeleer, B. (2013). Quorumpeps database: chemical space, microbial origin and functionality of quorum sensing peptides. *Nucleic Acids Research*, 41(D1), D655-D659.
- Wynendaele, E., Gevaert, B., Stalmans, S., Verbeke, F., & De Spiegeleer, B. (2015). Exploring the chemical space of quorum sensing peptides. *Peptide Science*, 104(5), 544-551.
- von Rad, U., Klein, I., Dobrev, P.I., Kottova, J., Zazimalova, E., Fekete, A., ... & Durner, J. (2008). Response of *Arabidopsis thaliana* to *N*-hexanoyl-DL-homoserine-lactone, a bacterial quorum sensing molecule produced in the rhizosphere. *Planta*, 229(1), 73-85.
- Yang, T., Tal-Gan, Y., Paharik, A.E., Horswill, A.R., & Blackwell, H.E. (2016). Structure-function analyses of a *Staphylococcus epidermidis* autoinducing peptide reveals motifs critical for AgrC-type receptor modulation. *ACS Chem Biol*, 11, 1982–1991. <https://doi.org/10.1021/acscchembio.6b00120>
- Zavilgelsky, G.B., & Manukhov, I.V. (2001). Quorum sensing, or how bacteria “talk” to each other. *Molecular Biology*, 35(2), 224-232.
- Zhang, Z., & Pierson III, L.S. (2001). A second quorum-sensing system regulates cell surface properties but not phenazine antibiotic production in *Pseudomonas aureofaciens*. *Applied and Environmental Microbiology*, 67(9), 4305-4315.