



EFFECTS OF DRYING TEMPERATURE ON CURCUMIN AND PIPERINE DISSOLUTION AND THE RELEASE KINETICS OF SOLID DISPERSION-BASED MICROPARTICLES: A PRELIMINARY STUDY

KURUTMA SICAKLIĞININ KURKUMİN VE PİPERİN ÇÖZÜNMESİNE VE KATI DİSPERSİYON BAZLI MİKROPARTİKÜLLERİN SALINIM KİNETİĞİNE ETKİLERİ: BİR ÖN ÇALIŞMA

Monica Octaviani Tiara DEWI¹ , Dewi SETYANINGSIH^{1*} 

¹Department of Pharmacy, Faculty of Pharmacy, Sanata Dharma University, Yogyakarta, Indonesia

ABSTRACT

Objective: *One of the major challenges in developing curcumin as a pharmaceutical agent is its low bioavailability after oral administration. Co-administration of curcumin-piperine combined with employing solid dispersions (SD) approach has been shown to enhance curcumin dissolution and bioavailability. Understanding the influence of the processing temperature during spray drying is crucial in SDs preparations; the purpose of this study is to inquire the effect of inlet temperature spray-dryer on dissolution behavior and the best-fit kinetic model of dissolution itself.*

Material and Method: *The SD powder was prepared using a spray-drying method by varying the inlet temperature (105°C; 115°C; 125°C) and involved polyvinyl alcohol (PVA) as a carrier. The SD were prepared at 30% Curcuma longa and 10% Piper nigrum extracts. Yield (%) of the dried powder resulted from the spray drying process was monitored, and dissolution behavior of curcumin and piperine were analyzed using a dissolution efficiency (DE) value. Furthermore, mathematical model describing the release mechanism of curcumin and piperine from the dissolution were evaluated using a DDSolver software.*

Result and Discussion: *The variation of drying temperature on the spray dryer affects the dissolution behavior and the % yield of the PVA-based SD containing C. longa and P. nigrum extract. The most ideal mathematical model of kinetic release for curcumin and piperine were the Quadratic model, indicating that the mechanism of dissolution is diffusion through a gap between the PVA particle and the surrounding medium.*

Keywords: *Curcuma longa, DDSolver, dissolution, Piper nigrum, solid dispersion*

* Corresponding Author / Sorumlu Yazar: Dewi Setyaningsih
e-mail / e-posta: dewi@usd.ac.id, Phone / Tel.: +6282135993494

ÖZ

Amaç: Kurkuminin farmasötik bir ajan olarak geliştirilmesindeki en büyük zorluklardan biri, oral uygulamadan sonra düşük biyoyararlanımıdır. Kurkumin-piperin'in katı dispersiyon (KD) yaklaşımı kullanılarak birlikte uygulanmasının kurkumin çözünmesini ve biyoyararlanımını artırdığı gösterilmiştir. Püskürtmeli kurutma sırasında işlem sıcaklığının etkisinin anlaşılması, KD preparatlarında çok önemlidir; bu çalışmanın amacı, püskürtmeli kurutucunun giriş sıcaklığının çözünme davranışı ve çözünmenin en uygun kinetik modeli üzerindeki etkisini araştırmaktır.

Gereç ve Yöntem: KD tozu, giriş sıcaklığı değiştirilerek (105°C; 115°C; 125°C) ve taşıyıcı olarak polivinil alkol (PVA) kullanılarak püskürtmeli kurutma yöntemi ile hazırlanmıştır. KD, %30 *Curcuma longa* ve %10 *Piper nigrum* ekstraktları kullanılarak hazırlanmıştır. Püskürtmeli kurutma işleminden elde edilen kurutulmuş tozun verimi (%) izlenmiş ve kurkumin ve piperinin çözünme davranışı, çözünme etkinliği (ÇE) değeri kullanılarak analiz edilmiştir. Ayrıca, kurkumin ve piperinin çözünmeden salınım mekanizmasını tanımlayan matematiksel model bir DDSolver yazılımı kullanılarak değerlendirilmiştir.

Sonuç ve Tartışma: Püskürtmeli kurutucuda kurutma sıcaklığının değişimi, *C. longa* ve *P. nigrum* ekstresi içeren PVA bazlı KD'nin çözünme davranışını ve % verimini etkilemektedir. Kurkumin ve piperin için kinetik salınımın en ideal matematiksel modeli, çözünme mekanizmasının PVA partikülü ile çevreleyen ortam arasındaki bir boşluktan difüzyon olduğunu gösteren Kuadratik modeldir.

Anahtar Kelimeler: *Curcuma longa*, çözünme, DDSolver, katı dispersiyon, *Piper nigrum*

INTRODUCTION

Herbal secondary metabolites have proved to be valuable sources that play an essential role in Indonesian traditional medicine for maintaining health and curing various diseases. In the community, herbal preparation is often found as combining two or more plant extracts for more health benefits [1]. One of the natural polyphenols found in the rhizome of *Curcuma longa* Linn is curcuminoids, with curcumin being the most prominent component among the curcuminoids. The polyphenolic compound curcumin plays an essential role as an antioxidant with several pharmacological activities, such as anti-inflammation and anticancer properties. Although curcumin has many beneficial properties, the clinical application of curcuminoids as pharmaceutical agents is limited due to their low bioavailability after oral administration. Curcumin is classified as Biopharmaceutics Classification System (BCS) II which is less soluble in water (11 ng/ml) but has high membrane permeability. The low bioavailability of curcumin is due to its sensitivity to phase II metabolism in the gastrointestinal tract, where it is known to be a substrate for uridine 5'-diphosphate-glucuronosyltransferase (UGT) enzymes, presenting a significant barrier to its development as an active pharmaceutical ingredient [2,3].

One way to overcome low bioavailability is the use of bioenhancer. Piperine, the main active ingredient of *P. nigrum*, showed effective results in increased bioavailability of curcumin in combination dosage [4,5]. Co-administration of piperine with curcumin (1:100) enhanced serum curcumin concentration by 154% for 1-2 hours after the onset. At the dosages used in the study, piperine appears to increase plasma concentrations, absorption rate, and curcumin bioavailability in rats and human subjects with no adverse effects [3]. Another study discovered that when curcumin was co-administered with piperine, the absorption was increased and remained in the body tissues for significantly longer (maximum 48 hours) [6]. However, piperine is also classified as BCS II because of its poor water solubility (40 mg/L) [4]. In contemplation of overcoming the poor aqueous solubility the SD method of *C. longa* and *P. nigrum* extracts is suggested.

Spray drying is a common SD method for increasing the dissolution rate by evaporating liquid into microscopic droplets. This technique uses atomization in hot water to remove the solvent from the dispersion of the target compound in the matrix solution, resulting in a powder. The final product quality and drying yield of spray-dried product is affected by manufacturing parameters such as feed flow rate, inlet and outlet air temperatures, atomization speed or pressure, feed concentration, etc [7]. Inlet temperature is the primary variable besides the feed flow rate in spray drying that needs to be optimized [8]. Several studies have published the effect of operating parameters and drying conditions on the physical properties of spray-dried powders, such as inlet drying yield, moisture content (MC) [9-12],

and wettability [9,11,12]. Neither report studied the impacts of the spray-drying operating parameters on the dissolution rate of spray-dried powder properties.

Prior research on the bioavailability of curcumin encapsulation in different ratios with varied carriers using SD has been conducted by Hu et al. [13]. Meanwhile, Wang et al. [14] investigated the bioavailability of curcumin SD co-formed with piperine without an excipient/carrier. Meanwhile, other studies, including the investigations by Jumah et al. [15], Kumar and Muzaffar [16], and Fujita et al. [17] have observed the effects of spray-drying method operating parameters on the powder characteristics. Nonetheless, the effect of inlet spray-drying temperature on the functional properties of curcumin piperine encapsulation using PVA has not yet been investigated. Hence, in this study inlet temperatures varied from 105°C to 125°C were applied to dry the PVA-based SD containing *C. longa* and *P. nigrum* extract.

In order to estimate curcumin and piperine absorption, it is also necessary to discover the kinetics and mechanism of dissolution. To determine the mechanism of drug release, various mathematical models for assessing dissolution profiles have been proposed. A theoretical investigation of the process can produce mathematical models of a dissolution profile, but due to the variety of dosage forms and its complexity, no theoretical base exists in most situations. As a result, empirical models must be used to fit dissolution data [18,19]. For that reason, a quantitative assessment of dissolution profile character is required.

DD Solver is a valuable software that involves a non-linear regression approach to perform kinetic analysis of dissolution profile [20]. DDSolver can be used as a predominant tool for monitoring drug product reliability and stability, as well as a quick and low-cost technique for predicting *in vivo* drug absorption. To the best of our knowledge, the application of mathematical models in evaluating the curcumin piperine release from SD is limited. Therefore, in addition to investigating the temperature of the inlet spray-drying on the functional properties of curcumin piperine, this study also presents empirical mathematical models to simulate and predict *in vivo* drug absorption of SD curcumin piperine.

MATERIAL AND METHOD

Material

C. longa extract of 97.56% curcuminoid content was given by PT. Phytochemindo Reksa, Bogor, Indonesia. *P. nigrum* extract was given by Dr.rer.nat Yosi Bayu Murti, Faculty of Pharmacy, Universitas Gadjah Mada Yogyakarta, Indonesia. The piperine standard (Sigma Aldrich) and the validated reversed phase High-Performance Liquid Chromatography (RP-HPLC) method were used to extract and further define the sample (validity recovery 91.14% with >0,999 correlation coefficient) [21]. PVA was donated by PT Konimex Solo, Central Java, Indonesia. Ethanol 96%, methanol, Sodium dihydrogen phosphate, and Sodium Lauryl Sulfate (SLS) were purchased from Merck, Darmstadt, Germany. Dissolution medium consisting of sodium phosphate buffer of pH 6.0 and Milli-Q water were prepared in the laboratory.

Preparation of Spray Dried Curcumin-Piperine

The SD of *C. longa* and *P. nigrum* extract were prepared by spray drying using PVA as a carrier. The SD contained 30% w/w *C. longa* extract; 10% w/w *P. nigrum* extract; 60% w/w PVA. In brief, *C. longa*, and 0.6 mg of *P. nigrum* extracts were diluted in 600 ml ethanol, and 3.6 mg PVA was diluted in 20 ml hot water (80-90°C). The resulting solution was loaded through a two-way nozzle into a BUCHI B-290 mini spray dryer installed with a B-290 dehumidifier under the following operating conditions: feed pump rate of 8%, aspirator rate of 100%, nozzle cleaner of 2%, and inlet temperature varied 105, 115, and 125° (not exceed 180°C) [22,23]. The spray-dried powder obtained was precisely weighed for drying yield calculation (33.50%; 33.76%; 37.01%) and placed in a desiccator for further evaluation.

Dissolution Test

The dissolution test was performed in 900 ml of 0.5 w/v % SLS (0.5 gram of SLS is used to make up a total volume of 100 ml) in a 20 mM phosphate buffer solution with a pH of 6 (USP Apparatus II). The study was conducted for 120 minutes under a stirring speed of 75 rpm (37 ± 0.5°C). To maintain

the sink condition, at predetermined time intervals, the sample (5.0 ml) was removed and replaced with a fresh dissolution medium at the same temperature.

Curcumin and piperine concentrations in dissolution samples were determined using validated spectrophotometry and Vierordt's method for simultaneous determination of curcumin and piperine (equations 1 and 2) [24]. The compound absorptivity for each wavelength was determined by plotting the absorbance obtained at the respective wavelength into the calibration equation of $y = 0.1606x + 0.0045$ (curcumin) and $y = 0.09x - 0.0088$ (piperine) (piperine).

Simultaneous Equation (Vierordt's Method):

$$C_c = \frac{(A2.ap1) - (A1.ap2)}{(ac2.ap1) - (ac1.ap2)} \quad (1)$$

$$C_p = \frac{(A1.ac2) - (A2.ac1)}{(ac2.ap1) - (ac1.ap2)} \quad (2)$$

Cc: Concentration of curcumin

Cp: Concentration of Piperine

A1: Absorbance measured at wavelength 1

A2: Absorbance measured at wavelength 2

Ac1: Curcumin absorptivity at wavelength 1 in absorbance/(g/100ml)

Ac2: Curcumin absorptivity at wavelength 2 in absorbance/(g/100ml)

Ap1: Piperine absorptivity at wavelength 1 in absorbance/(g/100ml)

Ap2: Piperine absorptivity at wavelength 2 in absorbance/(g/100ml)

The curcumin-piperine concentrations of the samples were measured using a verified method UV spectrophotometer at 430,5 and 344,4 nm. The dissolution data were expressed as a percentage (%) dissolved and dissolution efficiency at 120 min (DE120).

$$DE_t = \int_0^t \left(\frac{Y dt}{Y_{100t}} \right) \times 100\%$$

Statistical Analysis

All experiments were performed in triplicate, and statistical tests were carried out using one-way ANOVA Test for more than two data (obtained to describe the closeness of dissolution profiles). Statistically significant data was accepted if the p-value was less than 0.05. *In-vitro* dissolution data were fitted to the mathematical kinetics model in DD Solver with (1) statistical parameters-based evaluation of the release kinetics model: $R^2_{adjusted}$, Akaike Information Criterion (AIC), and Model Selection Criterion (MSC).

RESULT AND DISCUSSION

Drying Yield

Drying yield is the most important criterion when evaluating the viability of the spray-drying process in SD methodology. Table 1 shows the drying yields for spray-dried curcumin-piperine at different temperature of the inlet. The drying yield was increased along with the increasing temperature, and the highest yield was attained when the inlet temperature reached 125°C. In a study, the authors observed that increasing inlet temperature from 130°C–180°C led to increased process yield of micro-sized maltodextrin (MDX) (6.75 – 40.25%) [25]. Our findings were similar to this result. Another study reported an increase in the drying yield of waxy rice starch, from 74.83% to 88.66%, when the temperature on inlet spray drier was elevated from 40°C to 80°C. However, because of the sticking problem, the drying yield of waxy rice starch decreased at 100°C, indicating that when the drying temperature exceeds the gelatinization onset temperature, the droplets become drier and stick to the cyclone wall [26,27]. The inlet temperature positively affects the drying yield, with higher inlet air

temperatures increasing the effectiveness of mass and heat transfer processes while reducing the risk of inadequate drying particles hitting and forming crust on the drying chamber wall [28].

Table 1. Drying yield of spray-dried curcumin-piperine at different temperatures

| | Formula 1 (105°C) | Formula 2 (115°C) | Formula 3 (125°C) |
|-------------------|-------------------|-------------------|-------------------|
| Curcumin (30%) | 2.3995 g | 1.8002 g | 1.8001 g |
| Piperine (10%) | 0.7998 g | 0.6001 g | 0.6000 g |
| PVA (60%) | 4.8003 g | 3.6001 g | 3.6001 g |
| Total weight | 7.9996 g | 6.0004 g | 6.0002 g |
| Yield calculation | 33.50% | 33.76% | 37.01% |

In-Vitro Dissolution Study

An *in-vitro* dissolution is an essential approach in the pharmaceutical sector for drug development and estimating a drug product's *in vivo* performance as a quality control test. A dissolution study can be used in place of determining bioequivalence (biowaiver). An *in-vitro* dissolution study also can be used to identify pharmaceutical products' long-term stability and shelf life [29].

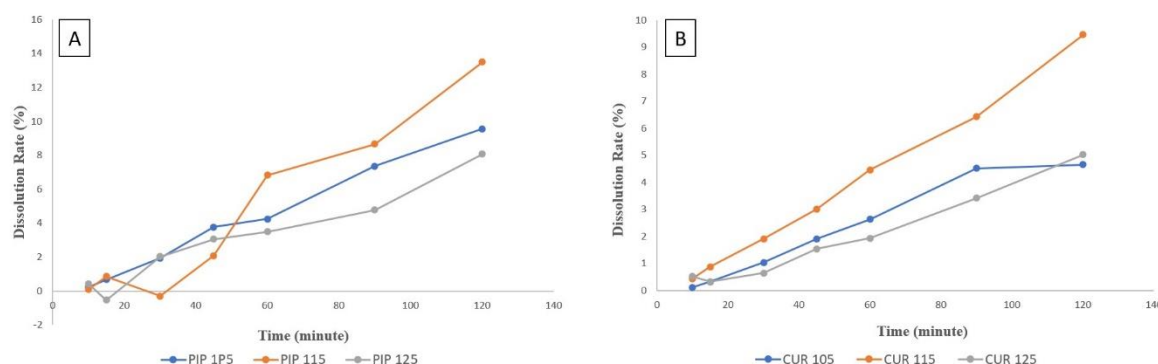


Figure 1. The dissolution rate of Curcumin (A); and Piperine (B) in sodium phosphate buffer (pH 6.0) at temperature $37\pm 0.5^\circ\text{C}$

The dissolution profiles represented as percentage dissolution rate versus time dilution of SD at various inlet temperatures are illustrated in Figure 1. The percent dissolution value was also calculated as the Area Under the Curve (AUC) and Dissolution Efficiency (DE) values at 120 minutes. Figure 2 depicts the DE₁₂₀ values of curcumin and piperine which were calculated to compare the dissolution profile. The data from the three formulas were found to be normally distributed by the normality test results, hence the ANOVA test was carried out. The ANOVA test revealed that piperine and curcumin had significantly different dissolution rates amongst the SD formulations obtained ($p > 0.05$). The dissolution rate of curcumin and piperine increased from 105°C to 115°C, but then slightly decreased when the temperature of the inlet was raised to 125°C. The slight decreased dissolution rate obtained at 125°C could be explained by the crust formation which prevents water penetration during the dissolution study. The crust was formed due to rapid moisture evaporation occurring at 125°C drying temperature [16,30]. Thereby, the crust formation during drying in a spray dryer can be prevented by conducting it at lower temperatures. However, if the temperature of the inlet is defectively lower, the particle will retain moisture for a longer period of time and shrinks resulting in a smaller particle size which can affect the dissolution rate [17]. Shi et al. [10] reported similar dissolution behaviors in the

dried watermelon SD which was processed at varied inlet temperatures from 120°C to 150°C. The particle size decreased from 21.64 nm (120°C drying temperature) to 13.44 nm (140°C drying temperature). Then, the particle size increased to 21.21 nm when the inlet temperature was raised to 150°C, resulting in a lower dissolution rate. Another study by Santhalakshmy et al. [11] obtained similar results, analyzing the production of spray-dried jamun fruit juice powder at 140°C to 160°C. Furthermore, Figure 1 was shown that the highest dissolution rate only reached $13.49\% \pm 4.07\%$ at 115°C. This is most likely due to the PVA carrier. The dissolution results in the three formulas where the remaining capsules are not dissolved at 120 minutes show that PVA can form a gel layer, making it difficult to dissolve. The formation of the gel layer causes the diffusion layer to thicken, affected in delaying dissolution [31]. Water molecules are prevented from penetrating through the particles due to the surface layer. By decreasing the particle's wettability, the dissolution powder and transfer rate provided an enormous force for evaporation, allowing powders with lower water content to be formed [32,33]. In addition, due to the formation of the gel layer, the PVA carrier also increased the viscosity of a preparation which affected the course of dissolution. Viscosity is inversely proportional to dissolution rate, so as viscosity increases, the dissolution profile in a medium will presumably decrease [34,35].

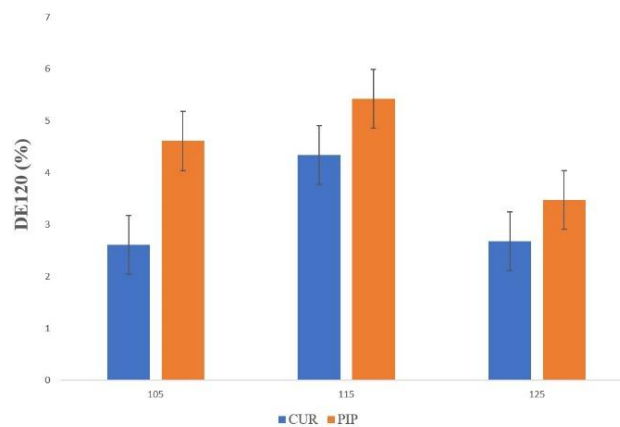


Figure 2. Dissolution Efficiency (DE120) of curcumin and piperine at 120 minutes

Dissolution Study of Curcumin-Piperine Kinetic Release using DDSolver

Mathematical models of kinetics drug release are useful for predicting the exact transport mechanism that affects a drug's *in vivo* dissolution profile [34]. Dissolution profiles of curcumin-piperine in various inlet temperatures were analyzed using drug release kinetic models. The obtained dissolution profiles of curcumin-piperine were fitted onto several mathematical models conducted by a non-linear regression approach using the DDSolver. The DDSolver was used to plot the dissolution profile data, which included the dissolution time (minutes) and the percentage of drug dissolved.

DDSolver offers a few statistical metrics to assess the dissolution model, including adjusted coefficient of determination, correlation coefficient, and coefficient determination. The most well-known and extensively used of these criteria for identifying *in vitro* drug release data modeling are R^2_{adjusted} , AIC, and MSC [36]. The R^2_{adjusted} was considered the most acceptable parameter to compare the dissolution models [19]. As shown in Table 2, the result revealed that the prepared curcumin and piperin SD at all temperatures exhibited Quadratic model tendencies, with R^2_{adjusted} values of 0.98427, 0.98787, and 0.95503 for curcumin and 0.97992, 0.96202, and 0.87643 for piperin. However, the results showed a high similarity between models at different temperatures. As a result, other statistical criteria, AIC and MSC were applied using DDSolver.

Table 2. Results of statistical parameters to describe the release of SD curcumin and piperine by each model

| SD | Model | Dissolution Model Parameters (mean) | | | | | |
|-----|------------------|-------------------------------------|----------|----------|----------|----------|----------|
| | | R^2_{adjusted} | | MSC | | AIC | |
| | | Curcumin | Piperine | Curcumin | Piperine | Curcumin | Piperine |
| 105 | Zero-order | 0.95864 | 0.95839 | 2.79162 | 2.91618 | 2.55799 | 11.57415 |
| | First-order | 0.95912 | 0.95643 | 2.79969 | 2.81064 | 2.49343 | 12.41852 |
| | Higuchi | 0.79417 | 0.77688 | 1.18165 | 1.11619 | 15.43776 | 25.97409 |
| | Hixson-Crowell | 0.95899 | 0.95716 | 2.79756 | 2.84498 | 2.51046 | 12.14375 |
| | Hopfenberg | 0.95235 | 0.95275 | 2.55256 | 2.68150 | 4.47048 | 13.45165 |
| | Weibull | 0.86903 | 0.92924 | 1.50097 | 3.01295 | 12.88314 | 10.80005 |
| | Quadratic | 0.98427 | 0.97992 | 2.64386 | 4.03150 | 3.74001 | 2.65158 |
| | Korsmeyer-Peppas | 0.84935 | 0.91540 | 1.53594 | 2.24456 | 12.60343 | 16.94712 |
| | Gompertz | 0.96055 | 0.96712 | 2.79172 | 3.32432 | 2.55720 | 8.30906 |
| 115 | Zero-order | 0.95973 | 0.91568 | 2.92895 | 2.07868 | 3.45045 | 13.89547 |
| | First-order | 0.95625 | 0.91392 | 2.83752 | 2.05761 | 4.18193 | 14.06399 |
| | Higuchi | 0.73873 | 0.77688 | 0.96011 | 1.11619 | 19.20115 | 25.97409 |
| | Hixson-Crowell | 0.95743 | 0.91454 | 2.86762 | 2.06493 | 3.94110 | 14.00548 |
| | Hopfenberg | 0.95302 | 0.90180 | 2.67895 | 1.83021 | 5.45045 | 15.88320 |
| | Weibull | 0.92859 | 0.88359 | 2.26581 | 1.60994 | 8.75560 | 17.64536 |
| | Quadratic | 0.98787 | 0.96202 | 4.05118 | 2.95320 | 5.52737 | 14.89926 |
| | Korsmeyer-Peppas | 0.96224 | 0.91540 | 2.96048 | 2.24456 | 3.19825 | 16.94712 |
| | Gompertz | 0.92975 | 0.90005 | 2.22296 | 1.81909 | 9.09839 | 15.97214 |
| 125 | Zero-order | 0.88825 | 0.84292 | 1.88969 | 1.54995 | 18.23976 | 27.37747 |
| | First-order | 0.88563 | 0.83605 | 1.85928 | 1.49783 | 18.48306 | 27.79443 |
| | Higuchi | 0.70210 | 0.63066 | 0.82793 | 0.64051 | 26.73386 | 34.65304 |
| | Hixson-Crowell | 0.88657 | 0.83848 | 1.86973 | 1.51559 | 18.39942 | 27.65239 |
| | Hopfenberg | 0.86974 | 0.81676 | 1.64046 | 1.30003 | 20.23357 | 29.37687 |
| | Weibull | 0.78403 | 0.73191 | 1.06917 | 0.86727 | 24.80395 | 32.83897 |
| | Quadratic | 0.95503 | 0.87643 | 2.50284 | 2.39663 | 13.33454 | 20.60408 |
| | Korsmeyer-Peppas | 0.61860 | 0.80884 | 0.60494 | 1.35567 | 28.51776 | 28.93170 |
| | Gompertz | 0.87629 | 0.82748 | 2.43229 | 1.43095 | 13.89894 | 28.32953 |

The ideal model is the one with the lowest AIC value, while the most accurate model has the highest MSC value. MSC is a modified opposite form of the AIC that has been validated to be independent of the underlying point scaling. The appropriate MSC value is greater than 2 or 3 [19,36]. Thus, the MSC and AIC values for all the kinetic dissolution models of varied temperatures were evaluated. From the kinetic model parameters as depicted in Table 2, the quadratic model is the most appropriate model to explain the phenomenon of curcumin piperine SD dissolution reverse to the result of AIC, MSC, and R^2_{adjusted} value. Curve fitting results also indicated that the quadratic model is the best model for explaining the behavior of curcumin and piperine dissolution profiles at all inlet temperatures (Figure 3).

The Quadratic model is based on this equation $F = 100 (k_1.t^2 + k_2.t)$ [36]. A study by Delfour and Garon explained how a quadratic model could be applied in the case of this study. It was revealed that the quadratic model can be applied not only in time-dependent or nonlinear diffusion but also through a circumstance of the polymer-medium interface [37]. From those findings, the dissolution kinetic of curcumin-piperine from the PVA based microparticle containing *C. longa* and *P. nigrum* extracts in this study can be assumed that the PVA polymer might form tiny holes or cracks at which the drug curcumin-piperine could diffuse through the gap between the PVA particle and the medium.

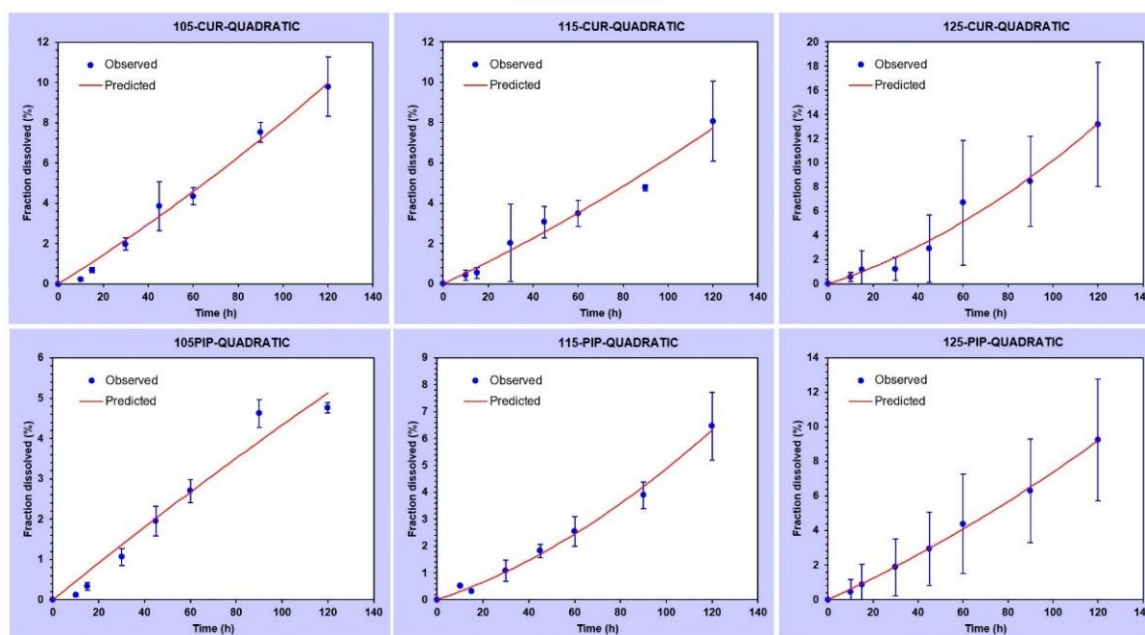


Figure 3. The most precise model prediction of the kinetic release of curcumin (CUR) and piperine (PIP)

Understanding the mechanism of oral drug absorption is important for efficacy and safety. Drug disintegration and dissolution, degradation, stomach emptying, intestinal transit, intestinal permeation and transport, intestinal metabolism, and hepatic metabolism are all possible phases in oral drug absorption. Dosage form, physicochemical and biological properties of the active pharmaceutical ingredient, and gastrointestinal (GI) tract physiology are all factors that may influence the rate and extent of drug absorption [37]. The kinetic modeling dissolution is known to be essential for estimating the absorption process. The absorption process itself is also a major challenge to control and maintain in oral administration. However, dissolution kinetic modeling cannot always illustrate the complex relationship between formulation attributes and oral absorption *in vivo*. The absorption modeling method is also considered necessary to investigate the effect of formulation attributes on oral absorption [19]. A study by Stillhart et al. [38] observed the beneficial effect of combining absorption modeling and *in vitro* dissolution tests of Basmisanil rather than only dissolution test to identify the rate-limiting processes in oral drug absorption. Therefore, predictive absorption modeling is required for further evaluation in future studies to demonstrate the compatibility between dissolution testing and the absorption process for finding the best strategy of formulation development.

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AUTHOR CONTRIBUTIONS

Concept: D.S.; Design: D.S.; Control: D.S.; Sources: D.S.; Materials: D.S.; Data Collection and/or Processing: M.O.T.D.; Analysis and/or Interpretation: M.O.T.D.; Literature Review: M.O.T.D.; Manuscript Writing: M.O.T.D.; Critical Review: D.S.; Other: -

CONFLICT OF INTEREST

The authors declare that there is no real, potential, or perceived conflict of interest for this article.

ETHICS COMMITTEE APPROVAL

The authors declare that the ethics committee approval is not required for this study.

REFERENCES

1. Ashraf, K., Mujeeb, M., Ahmad, A., Ahmad, N., Amir, M. (2015). Determination of curcuminoids in *Curcuma longa* Linn. by UPLC/Q-TOF-MS: An application in turmeric cultivation. *Journal of Chromatographic Science*, 53(8), 1346-1352. [\[CrossRef\]](#)
2. Wan, S., Sun, Y., Qi, X., Tan, F. (2012). Improved bioavailability of poorly water-soluble drug curcumin in cellulose acetate solid dispersion. *AAPS PharmSciTech*, 13(1), 159-166. [\[CrossRef\]](#)
3. Shoba, G., Joy, D., Joseph, T., Majeed, M., Rajendran, R., Srinivas, P.S.S.R. (1998). Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Medica*, 64(4), 353-356. [\[CrossRef\]](#)
4. Gorgani, L., Mohammadi, M., Najafpour, G.D., Nikzad, M. (2017). Piperine-The bioactive compound of black pepper: From isolation to medicinal formulations. *Comprehensive Reviews in Food Science and Food Safety*, 16(1), 124-140. [\[CrossRef\]](#)
5. Chopra, B., Dhingra, A.K., Kapoor, R.P., Prasad, D.N. (2017). Piperine and its various physicochemical and biological aspects: A review. *Open Chemistry Journal*, 3(1), 75-96. [\[CrossRef\]](#)
6. Sachin, S., Nirmala, G., Subir, B., Suresh, J., Dushyant, G. (2012). Development and validation of UV spectroscopic method for the quick estimation of gingerol from *Zingiber officinale* rhizome extract. *International Research Journal of Pharmacy*, 3(5), 234-237.
7. Patil, V., Chauhan, A.K., Singh, R.P. (2014). Optimization of the spray-drying process for developing guava powder using response surface methodology. *Powder Technology*, 253, 230-236. [\[CrossRef\]](#)
8. Cano-Chauca, M., Stringheta, P.C., Ramos, A.M., Cal-Vidal, J. (2005). Effect of the carriers on the microstructure of mango powder obtained by spray drying and its functional characterization. *Innovative Food Science and Emerging Technologies*, 6(4), 420-428. [\[CrossRef\]](#)
9. Lee, K.C., Yoon, Y.S., Li, F.Z., Eun, J.B. (2017). Effects of inlet air temperature and concentration of carrier agents on physicochemical properties, sensory evaluation of spray-dried mandarin (*Citrus unshiu*) beverage powder. *Applied Biological Chemistry*, 60(1), 33-40. [\[CrossRef\]](#)
10. Shi, Y., Wang, J., Wang, Y., Zhang, H., Ma, Y., Zhao, X., Zhang, C. (2018). Inlet temperature affects spray drying quality of watermelon powder. *Czech Journal of Food Sciences*, 36(4), 316-323. [\[CrossRef\]](#)
11. Santhalakshmy, S., Don Bosco, S.J., Francis, S., Sabeena, M. (2015). Effect of inlet temperature on physicochemical properties of spray-dried jamun fruit juice powder. *Powder Technology*, 274, 37-43. [\[CrossRef\]](#)
12. Teera-Arunsiri, A., Suphantharika, M., Ketunuti, U. (2009). Preparation of spray-dried wettable powder formulations of *Bacillus thuringiensis*-based biopesticides. *Journal of Economic Entomology*, 96(2), 292-299. [\[CrossRef\]](#)
13. Hu, L., Shi, Y., Li, J.H., Gao, N., Ji, J., Niu, F., Chen, Q., Yang, X., Wang, S. (2015). Enhancement of oral bioavailability of curcumin by a novel solid dispersion system. *AAPS PharmSciTech*, 16(6), 1327-1334. [\[CrossRef\]](#)
14. Wang, R., Han, J., Jiang, A., Huang, R., Fu, T., Wang, L., Zheng, Q., Li, W., Li, J. (2019). Involvement of metabolism-permeability in enhancing the oral bioavailability of curcumin in excipient-free solid dispersions co-formed with piperine. *International Journal of Pharmaceutics*, 561, 9-18. [\[CrossRef\]](#)
15. Jumah, R.Y., Tashtoush, B., Shaker, R.R., Zraiy, A.F. (2000). Manufacturing parameters and quality characteristics of spray dried jameed. *Drying Technology*, 18(4-5), 967-984. [\[CrossRef\]](#)
16. Muzaffar, K., Kumar, P. (2017). Spray drying of tamarind pulp: Effect of process parameters using protein as carrier agent. *Journal of Food Processing and Preservation*, 41(2), e12781. [\[Crossref\]](#)

17. Fujita, A., Souza, V.B., Daza, L.D., Fávoro-Trindade, C.S., Granato, D., Genovese, M.I. (2017). Effects of spray-drying parameters on *in vitro* functional properties of camu-camu (*Myrciaria dubia* Mc. Vaugh): A typical Amazonian fruit. *Journal of Food Science*, 82(5), 1083-1091. [[CrossRef](#)]
18. Zuo, J., Gao, Y., Bou-Chacra, N., Löbenberg, R. (2014). Evaluation of the DDSolver software applications. *BioMed Research International*, 2014, 204925. [[CrossRef](#)]
19. Abdul Rasool, B.K., Mohammed, A.A., Salem, Y.Y. (2021). The optimization of a dimenhydrinate transdermal patch formulation based on the quantitative analysis of *in vitro* release data by DDSolver through skin penetration studies. *Scientia Pharmaceutica*, 89(3), 33. [[CrossRef](#)]
20. Mendyk, A., Jachowicz, R., Fijorek, K., Doroczyński, P., Kulinowski, P., Polak, S. (2012). KinetDS: An open source software for dissolution test data analysis. *Dissolution Technologies*, 19(1), 6-11. [[CrossRef](#)]
21. Setyaningsih, D., Santoso, Y.A., Hartini, Y.S., Murti, Y.B., Hinrichs, W.L.J., Patramurti, C. (2021). Isocratic high-performance liquid chromatography (HPLC) for simultaneous quantification of curcumin and piperine in a microparticle formulation containing *Curcuma longa* and *Piper nigrum*. *Heliyon*, 7(3), E06541. [[CrossRef](#)]
22. Shingate, P.N., Dongre, P.P., Kannur, D.M. (2013). New method development for extraction and isolation of piperine from black pepper. *International Journal of Pharmaceutical Sciences and Research*, 4(8), 3165. [[CrossRef](#)]
23. Yu, Y., Wei, R., Jia, X., Zhang, X., Liu, H., Xu, B., Xu, B. (2022). Preparation of *Piper nigrum* Microcapsules by Spray Drying and Optimization with Response Surface Methodology. *Journal of Oleo Science*, 71(12), 1789-1797. [[CrossRef](#)]
24. Murti, Y.B., Hartini, Y.S., Hinrichs, W.L.J., Frijlink, H.W., Setyaningsih, D. (2019). UV-Vis spectroscopy to enable determination of the dissolution behavior of solid dispersions containing curcumin and piperine. *Journal of Young Pharmacists*, 11(1), 26-30. [[CrossRef](#)]
25. Magri, G., Franzé, S., Musazzi, U.M., Selmin, F., Cilurzo, F. (2019). Data on spray-drying processing to optimize the yield of materials sensitive to heat and moisture content. *Data in Brief*, 23, 103792. [[CrossRef](#)]
26. Tay, J.B.J., Chua, X., Ang, C., Subramanian, G.S., Tan, S.Y., Lin, E.M.J., Wu, W.Y., Goh, K.K.T., Lim, K. (2021). Effects of spray-drying inlet temperature on the production of high-quality native rice starch. *Processes*, 9(9), 1557. [[CrossRef](#)]
27. Gul, O., Dervisoglu, M. (2020). Optimization of spray drying conditions for microencapsulation of *Lactobacillus casei* Shirota using response surface methodology. *European Food Science and Engineering*, 1(1), 1-8.
28. Fazaeli, M., Emam-Djomeh, Z., Kalbasi Ashtari, A., Omid, M. (2012). Effect of spray drying conditions and feed composition on the physical properties of black mulberry juice powder. *Food and Bioproducts Processing*, 90(4), 667-675. [[CrossRef](#)]
29. Brown, C.K., Friedel, H.D., Barker, A.R., Buhse, L.F., Keitel, S., Cecil, T.L., Kraemer, J., Morris, J.M., Reppas, C., Stickelmeyer, M.P., Yomota, C., Shah, V.P. (2011). FIP/AAPS joint workshop report: dissolution/*in vitro* release testing of novel/special dosage forms. *AAPS PharmSciTech*, 12(2), 782-794. [[CrossRef](#)]
30. Tonon, R.V., Brabet, C., Hubinger, M.D. (2008). Influence of process conditions on the physicochemical properties of açai (*Euterpe oleracea* Mart.) powder produced by spray drying. *Journal of Food Engineering*, 88(3), 411-418. [[CrossRef](#)]
31. Silva, F.E.F., Di-Medeiros, M.C.B., Batista, K.A., Fernandes, K.F. (2013). PVA/polysaccharides blended films: Mechanical properties. *Journal of Materials*, 2013, 1-6. [[CrossRef](#)]
32. Tran, T.A.T., Nguyen, H.V.H. (2018). Effects of spray-drying temperatures and carriers on physical and antioxidant properties of lemongrass leaf extract powder. *Beverages*, 4(4), 84. [[CrossRef](#)]
33. Dak, M., Verma, R.C., Jaaffrey, S.N.A. (2007). Effect of temperature and concentration on Rheological properties of “Kesar” mango juice. *Journal of Food Engineering*, 80(4), 1011-1015. [[CrossRef](#)]
34. van Duong, T., van den Mooter, G. (2016). The role of the carrier in the formulation of pharmaceutical solid dispersions. Part I: crystalline and semi-crystalline carriers. *Expert Opinion on Drug Delivery*, 13(11), 1583-1594. [[CrossRef](#)]
35. Taheri, A., Atyabi, F., Dinarvand, R. (2011). Temperature-responsive and biodegradable PVA:PVP k30:Poloxamer 407 hydrogel for controlled delivery of human growth hormone (hGH). *Journal of Pediatric Endocrinology and Metabolism*, 24(3-4), 175-179. [[CrossRef](#)]
36. Zhang, Y., Huo, M., Zhou, J., Zou, A., Li, W., Yao, C., Xie, S. (2010). DDSolver: An add-in program for modeling and comparison of drug dissolution profiles. *AAPS Journal*, 12(3), 263-271. [[CrossRef](#)]
37. Huang, W., Lee, S.L., Yu, L.X. (2009). Mechanistic approaches to predicting oral drug absorption. *The AAPS Journal*, 11, 217-224. [[CrossRef](#)]

38. Stillhart, C., Parrott, N.J., Lindenberg, M., Chalus, P., Bentley, D., Szepes, A. (2017). Characterising drug release from immediate-release formulations of a poorly soluble compound, basmisanil, through absorption modelling and dissolution testing. *The AAPS Journal*, 19(3), 827-836. [\[CrossRef\]](#)