

# Eurasian Journal of Molecular and Biochemical Sciences

https://dergipark.org.tr/en/pub/ejmbs

## The Apoptotic Effects of a Proteasome Inhibitor on Caco-2 Colon Carcinoma Cells

### Selina AKSAK KARAMEŞE 1\* 💿

<sup>\*</sup> Kafkas University, Medical Faculty, Department of Histology and Embryology, 36100, Kars, Turkey

Cite: Aksak Karameşe S. The Apoptotic Effects of a Proteasome Inhibitor on Caco-2 Colon Carcinoma Cells. Eurasian Mol Biochem Sci 2023;2(3): 21-26.

Received: 03 March 2023, Accepted: 02 April 2023

#### Abstract

The first and reversible inhibitor of the 26S proteasome (Velcade, formerly PS-341) has a crucial role with its antitumor activity against a number of malignancies and acts as a prominent inducer of apoptosis. In the present work, it was demonstrated the enhanced anticancer activity of a proteasome inhibitor by evaluating the levels of Bcl-2 protein family members on Caco-2 cell line. For this reason, Caco-2 colon carcinoma cells were treated with Velcade at four different doses (0, 5, 10, and 20nM). Western blotting was performed to detect the expression levels of apoptotic protein. Additionally, apoptotic cell death was demonstrated with in situ TUNEL staining by immunohistochemistry (IHC). An increase in the expression levels of both pro-apoptotic proteins was detected in a dose-dependent manner; however, the effective dose was detected as 10 nM. According to the IHC results, it can be said that a serious increase in the amount of apoptotic cells was determined. Taking everything into account, this proteasome inhibitor has a potential to be effective in inducing apoptotic mechanisms on colon carcinoma cells.

Keywords: Pro-apoptotic proteins, Bax, Bcl-2, Velcade, Caco-2, Colon carcinoma

#### Introduction

Colon carcinoma is one of the most common and aggressive cancer types in both men and women all around the world (1, 2). In recent years, approximately two million new cases and one million deaths were reported; however, this high mortality rate has decreased year by year, because of earlier diagnosis through screening and better treatment possibilities. At that point, standard therapies for colon carcinoma con-

\*Correspondence: Selina AKSAK KARAMEŞE

Kafkas University, Medical Faculty, Department of Histology and Embryology, 36100, Kars, Turkey. E-mail: selin.atauni@hotmail.com tain firstly chemotherapy and then biological therapy followed by tumor resection (3-8). It is known that standard chemotherapy in this disease is 5-fluorouracil plus either oxaliplatin or irinotecan (3, 5, 6, 9). Although large clinical experiments are still trying to find the best way to combine these therapeutics, unfortunately advanced unresectable colorectal cancer remains incurable and new targets and new approaches to treatment are needed (10).

Bortezomib (Velcade, PS-341), a dipeptide boronic acid derivative and the first 26S proteasome inhibitor, shows antitumor and anticarcinogenic activity in most human cancers (11-15). It is mediated through



reversible binding to the N-terminus threonine residue in the  $\beta$ -1 subunit of the catalytic core complex of the 26S proteasome, leading to reversible inhibition of the proteolytic activity of the proteasome. Bortezomib is also a primary subcellular component of the protein degradation pathway that regulates the turnover of proteins involved in cell cycle arrest and apoptosis, such as the p21 cyclin-dependent kinase inhibitor, cyclins, and IKB, a regulator of nuclear factor-KB transcriptional activity (1, 10).

For the apoptotic cellular death, two important groups of proteins have a key role: the Bcl-2 family and a class of cysteine proteases known as caspases (16). The Bcl-2 family proteins have key functions especially in the regulation of a cellular mechanism, apoptosis, by targeting the mitochondria to show their pro- or antiapoptotic activities. Bcl-2, an anti-apoptotic protein, is known to regulate apoptotic pathways and promote cell survival. Bax, a pro-apoptotic molecule of that family, is expressed abundantly and selectively during apoptosis and essential to mitochondrial release of apoptogenic factors (17). Increasing the ratio of Bcl-2 to Bax has commonly been used to determine the induction of apoptosis in several tissues. On the other hand, lack of Bax markedly decreases the cytotoxicity of proteasome inhibitors (18). It is still unclear the molecular mechanisms of Bax activation by proteasome inhibition; however, most scientific studies are focused on this issue (19).

In the present work, it was demonstrated the enhanced anticancer activity of a proteasome inhibitor by evaluating the levels of Bcl-2 protein family members (Bcl-2 and Bax) on Caco-2 cell line.

#### **Materials and Methods**

**Culture of Caco-2 Colon Carcinoma Cells:** A kind of human colon cancer cell line (Caco-2) was purchased from ATCC by the special code of HTB-37. The Caco-2 cells were incubated with a mixture containing 10% heat-inactivated fetal bovine serum, 100 units/ml penicillin-streptomycin-gentamicin

antibiotic solution. Caco-2 cells were cultured in T75 culture flasks (Corning) in a special CO<sub>2</sub> incubator at 37 °C. Subconfluent cultures were detached by using trypsin and counted with firstly trypan blue and then automated cell counting machine, and cells were then seeded in 6-well plates in DMEM (Dulbecco's Modified Eagle Medium). Then, bortezomib (Velcade, PS-341; Millennium Pharmaceuticals) application was performed with the following doses; 5 nM, 10 nM and 20 nM. After co-incubation, the media were collected; then the cells were washed with PBS and finally, MTT cell proliferation assay were performed.

**MTT Cell Viability Assay:** The viability of incubated cells was measured by using MTT. Firstly, incubated cells were inoculated into the 96-well plate at a density of 106 cells in each well. Then, colon cells were cultured with 3-different concentrations of Velcade (5nM, 10 nM and 20 nM), and then incubated with 1 ml of MTT solution for 4 h at 37oC. Then, the culture medium was removed and 1 ml of 0.1 mol/l HCl in absolute isopropanol was added into the plate. The absorbance was measured at 490 nm with an automatic microplate reader (Thermo, Multiskan GO, USA). The final calculation for cell viability was as follows: cell viability (fold) <sup>1</sup>/<sub>4</sub> the absorbance of treated group/the average absorbance of untreated group

TUNEL Assay: For detection of DNA breakage, cells were air dried and fixed in 4% paraformaldehyde for 1 h at room temperature. In Situ Cell Death Kit from Roche (California, USA) was used for TUNEL assav according to the manufacturer's protocol. Briefly, fixed cells were rinsed in PBS, blocked, and permeabilized. TUNEL reaction mixture was then added to the cells and incubated at 37 C for 1 h. Apoptotic signals were converted with peroxidase and stained with 3,3'diaminobenzidine tetrahydrochloride. Apoptotic cells were detected and scored under fluorescence microscopy with 4',6-diamidino-2-phenylindole counter staining. The average of the TUNEL-positive nuclei ratio in at least 10 representative microscopic fields was calculated to compare the apoptosis ratio **F** within the different groups.

#### Results

Western Blotting and analysis: After treatment, cells were collected using a cell scraper at 4°C and lysed in a whole-cell lysis buffer. Approximately 25 µg total cellular protein from each sample was subjected to SDS-PAGE, proteins were transferred to nitrocellulose membranes, and the membranes were blocked with 5% nonfat milk in a TBS solution containing 0.1 % Tween 20 for 1 h. The blots were then probed overnight with anti-Bax and anti-Bcl-2 antibodies, washed, and probed with specific secondary antibodies coupled to HRP. Immuno-reactive material was detected by enhanced chemiluminescence (Fusion Fx, Fisher Biotek, Australia).

The anti-proliferative effect of Velcade/bortezomib (in 5, 10, and 20 nM doses) was evaluated by cell viability MTT method in Caco-2 cells in both 24 and 48 h. It was seen that bortezomib caused both time- and dose-dependent strong decreases in the viability of Caco-2 cells. According to the MTT findings, a decrease was detected in the viability of Caco-2 cells in 24-h evaluation. The higher doses of Velcade/bortezomib (10 and 20 nM) showed expected results in both 24 and 48-hour as seen in Figure 1. According to Figure 1, a time- and dose-dependent inhibition in cell viability of colon cancer Caco-2 cells were detected.



Figure 1. MTT analysis of Caco-2 colon cancer cells treated with bortezomib for 24 and 48 h., inhibitory concentration 50 (IC50) values were calculated by obtained 2 results of MTT.

The effect of bortezomib on apoptotic pathways in Caco-2 cell line: In the analysis of protein densities, bortezomib was increased pro-apoptotic Bax level in the 24 h period of colon cancer cells (Caco-2) and higher concentration of bortezomib markedly increased Bax protein level. Besides treatment of bortezomib, a particually higher dose of bortezomid decreased the Bcl-2 level in the both 24 and 48 h periods in colon cancer cells. Bax and Bcl-2 protein densities were demonstrated in Fig 2.

According to the analyses of TUNEL results, it was seen that there were increased apoptotic cells in 24 and the 48 h periods of bortezomib treated Caco-2 colon carcinoma cells. Also, 48 h period of bortezomib increased apoptotic cells more than the 24 h period of bortezomib treatment in colon cancer cells. The results of TUNEL positive cell densities for all groups were presented in Fig 3.



**Figure 2.** Western blot analysis of Caco-2 cells lysates, treated with 5, 10, 20 nM bortezomib for 24 and 48 h periods, A: Western blot reveals expression levels of Bcl-2 and Bax in Caco-2 cells, B: western blot analysis of Bcl-2 protein levels, and C: Bax protein levels using ECL Western blotting detection system (described in material and method).

#### Discussion

Bortezomib, a first and specific inhibitor of a proteasome (26S), (20) is generally used for the treatment of some human cancer types especially multiple myeloma and known as acting as a prominent inducer of apoptosis (21). In current literature, most scientific studies reported and suggested that Bortezomib (Velcade) might also be applied for the treatment of a variety of additional cancer types (22). Colon cancer therapy is based on the surgical removal of solid tumor masses, usually combined with a series of chemical treatments. Most drugs, especially bortezomib are thought to ultimately induce apoptosis of tumor cells through mitochondrial or death-receptor pathways. Recently, other cell death mechanisms are being considered for therapeutic application, such as autophagy and necrosis (1, 23, 24).

Several conclusions can be come out upon consideration of the data presented here. First, our data supported that a treatment with bortezomib may induce apoptosis. Second, the Bcl-Bax pathway plays an important role in the apoptosis through activating



**Figure 3.** Tunnel staining of Caco-2 colon cancer cells for all groups, brown color shows tunnel positive cells.

the mitochondria-dependent pathway. Finally, using 26S proteasome inhibitor in colon carcinoma in vitro model shows an apoptotic effect by inducing some proapoptotic proteins including Bcl-2 and Bax.

In the present study, we chose the human colon carcinoma cell line Caco-2 as a model to explore the ability of bortezomib to modulate the main proteolytic especially cellular pathways: apoptosis. The importance of the mitochondrial component of the apoptotic pathway is perhaps best approached by the overexpression of the anti-apoptotic proteins Bcl-2 to block the mitochondrial contribution to the intrinsic pathway. Our results apoptotic showed that bortezomib was suppressing the Bcl-2 expression level whereas it was increasing the Bax level. Some scientific studies in current literature showed that treatment with bortezomib has been reported to cause alterations in the levels of apoptosis-regulating proteins including Bcl-xL (25), Bcl-2, and other Bcl-2-related proteins (26, 27). Kretowski et al. (28) pointed out that bortezomib has a potential to induce apoptosis via anti-apoptotic Bcl-2 family members in DLD-1 colon cancer cell line. In another study, (1) it was claimed that apoptosis was induced by Bcl-2/Bcl-xL phosphorylation in response to treatment with bortezomib. Pitts et al. also reported that bortezomib had anti-proliferative and proapoptotic effects against colon cancer cell lines by effecting Bax, Bcl-2, caspase 3, and caspase 7 levels (10).

As a conclusion, the proteasome is involved in the proteolysis of cell cycle regulators and damaged proteins, therefore the identification of proteasome modulators is a promising approach in a number of pathologies, such as cancer, neurodegenerative, cardiovascular, and metabolic diseases (29). On the other side, autophagy can be involved in cell defense mechanisms or in type II programmed cell death, in response to cellular stress conditions (30, 31).

Proteasome inhibitor bortezomib promotes cell death in colon cancer cell line Caco-2 through apoptosis. In conclusion, we suggest that bortezomib may be candidates for further evaluation as chemotherapeutic agents for human colon cancer. On the other hand, future studies will be needed to investigate other cellular mechanisms including autophagy and related parameters.

**Declaration of Interest:** The author declares that there is no conflict of interest regarding the publication of this paper.

#### **Author Contribution**

SAK contributed all parts of the study.

#### **ORCID:**

Selina AKSAK KARAMEŞE 💿 0000-0002-4820-2096

#### References

- 1. Kim SY, Song X, Zhang L, Bartlett DL, Lee YJ. Role of BclxL/Beclin-1 in interplay between apoptosis and autophagy in oxaliplatin and bortezomib-induced cell death. Biochemical pharmacology. 2014;88(2):178-88.
- 2. Aldahhan R, Almohazey D, Khan FA. Emerging trends in the application of gold nanoformulations in colon cancer diagnosis and treatment. Semin Cancer Biol. 2022;86(Pt 2):1056-65.
- Hong YS, Hong SW, Kim SM, Jin DH, Shin JS, Yoon DH, et al. Bortezomib induces G2-M arrest in human colon cancer cells through ROS-inducible phosphorylation of ATM-CHK1. International journal of oncology. 2012;41(1):76-82.
- 4. Cunningham D, Humblet Y, Siena S, Khayat D, Bleiberg H, Santoro A, et al. Cetuximab monotherapy and cetuximab plus irinotecan in irinotecan-refractory metastatic colorectal cancer. The New England journal of medicine. 2004;351(4):337-45.
- Douillard JY, Cunningham D, Roth AD, Navarro M, James RD, Karasek P, et al. Irinotecan combined with fluorouracil compared with fluorouracil alone as first-line treatment for metastatic colorectal cancer: a multicentre randomised trial. Lancet. 2000;355(9209):1041-7.
  Hochster HS, Hart LL, Ramanathan RK, Childs BH, Hainsworth
- 6. Hochster HS, Hart LL, Ramanathan RK, Childs BH, Hainsworth JD, Cohn AL, et al. Safety and efficacy of oxaliplatin and fluoropyrimidine regimens with or without bevacizumab as first-line treatment of metastatic colorectal cancer: results of the TREE Study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2008;26(21):3523-9.
- Bokemeyer C, Bondarenko I, Makhson A, Hartmann JT, Aparicio J, de Braud F, et al. Fluorouracil, leucovorin, and oxaliplatin with and without cetuximab in the first-line treatment of metastatic colorectal cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2009;27(5):663-71.
- 8. Van Cutsem E, Kohne CH, Hitre E, Zaluski J, Chang Chien CR, Makhson A, et al. Cetuximab and chemotherapy as initial treatment for metastatic colorectal cancer. The New England journal of medicine. 2009;360(14):1408-17.
- 9. de Gramont A, Figer A, Seymour M, Homerin M, Hmissi A, Cassidy J, et al. Leucovorin and fluorouracil with or without oxaliplatin as first-line treatment in advanced colorectal cancer. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2000;18(16):2938-47.
- Pitts TM, Morrow M, Kaufman SA, Tentler JJ, Eckhardt SG. Vorinostat and bortezomib exert synergistic antiproliferative and proapoptotic effects in colon cancer cell models. Molecular cancer therapeutics. 2009;8(2):342-9.
- 11. Fennell DA, Chacko A, Mutti L. BCL-2 family regulation by the 20S proteasome inhibitor bortezomib. Oncogene.

2008;27(9):1189-97.

- 12. Garrido C, Brunet M, Didelot C, Zermati Y, Schmitt E, Kroemer G. Heat shock proteins 27 and 70: anti-apoptotic proteins with tumorigenic properties. Cell cycle. 2006;5(22):2592-601.
- Song X, Kim SY, Zhang L, Tang D, Bartlett DL, Kwon YT, et al. Role of AMP-activated protein kinase in cross-talk between apoptosis and autophagy in human colon cancer. Cell death & disease. 2014;5:e1504.
- 14. Zhu K, Dunner K, Jr., McConkey DJ. Proteasome inhibitors activate autophagy as a cytoprotective response in human prostate cancer cells. Oncogene. 2010;29(3):451-62.
- Ebert B, Kisiela M, Wsol V, Maser E. Proteasome inhibitors MG-132 and bortezomib induce AKR1C1, AKR1C3, AKR1B1, and AKR1B10 in human colon cancer cell lines SW-480 and HT-29. Chem Biol Interact. 2011;191(1-3):239-49.
- 16. Chang HK, Shin MS, Yang HY, Lee JW, Kim YS, Lee MH, et al. Amygdalin induces apoptosis through regulation of Bax and Bcl-2 expressions in human DU145 and LNCaP prostate cancer cells. Biol Pharm Bull. 2006;29(8):1597-602.
- 17. Danial NN, Korsmeyer SJ. Cell death: critical control points. Cell. 2004;116(2):205-19.
- 18. Yu J, Tiwari S, Steiner P, Zhang L. Differential apoptotic response to the proteasome inhibitor Bortezomib [VELCADE, PS-341] in Bax-deficient and p21-deficient colon cancer cells. Cancer biology & therapy. 2003;2(6):694-9.
- Ding WX, Ni HM, Chen X, Yu J, Zhang L, Yin XM. A coordinated action of Bax, PUMA, and p53 promotes MG132-induced mitochondria activation and apoptosis in colon cancer cells. Molecular cancer therapeutics. 2007;6(3):1062-9.
- Karin M, Yamamoto Y, Wang QM. The IKK NF-kappa B system: a treasure trove for drug development. Nature reviews Drug discovery. 2004;3(1):17-26.
- Hideshima T, Chauhan D, Richardson P, Mitsiades C, Mitsiades N, Hayashi T, et al. NF-kappa B as a therapeutic target in multiple myeloma. The Journal of biological chemistry. 2002;277(19):16639-47.
- 22. Green DR, Kroemer G. Pharmacological manipulation of cell death: clinical applications in sight? The Journal of clinical

investigation. 2005;115(10):2610-7.

- 23. Amaravadi RK, Lippincott-Schwartz J, Yin XM, Weiss WA, Takebe N, Timmer W, et al. Principles and current strategies for targeting autophagy for cancer treatment. Clinical cancer research : an official journal of the American Association for Cancer Research. 2011;17(4):654-66.
- 24. Waters JP, Pober JS, Bradley JR. Tumour necrosis factor and cancer. The Journal of pathology. 2013;230(3):241-8.
- 25. Bai J, Demirjian A, Sui J, Marasco W, Callery MP. Histone deacetylase inhibitor trichostatin A and proteasome inhibitor PS-341 synergistically induce apoptosis in pancreatic cancer cells. Biochemical and biophysical research communications. 2006;348(4):1245-53.
  26. Jiang X, Tsang YH, Yu Q. c-Myc overexpression sensitizes Bim-
- 26. Jiang X, Tsang YH, Yu Q. c-Myc overexpression sensitizes Bimmediated Bax activation for apoptosis induced by histone deacetylase inhibitor suberoylanilide hydroxamic acid (SAHA) through regulating Bcl-2/Bcl-xL expression. The international journal of biochemistry & cell biology. 2007;39(5):1016-25.
- 27. Pei XY, Dai Y, Grant S. Synergistic induction of oxidative injury and apoptosis in human multiple myeloma cells by the proteasome inhibitor bortezomib and histone deacetylase inhibitors. Clinical cancer research : an official journal of the American Association for Cancer Research. 2004;10(11):3839-52.
- 28. Kretowski R, Stypulkowska A, Cechowska-Pasko M. Efficient apoptosis and necrosis induction by proteasome inhibitor: bortezomib in the DLD-1 human colon cancer cell line. Molecular and cellular biochemistry. 2015;398(1-2):165-73.
- 29. Ciechanover A, Orian A, Schwartz AL. The ubiquitin-mediated proteolytic pathway: mode of action and clinical implications. Journal of cellular biochemistry Supplement. 2000;34:40-51.
- 30. Bonfili L, Cuccioloni M, Cecarini V, Mozzicafreddo M, Palermo FA, Cocci P, et al. Ghrelin induces apoptosis in colon adenocarcinoma cells via proteasome inhibition and autophagy induction. Apoptosis : an international journal on programmed cell death. 2013;18(10):1188-200.
- 31. McCray BA, Taylor JP. The role of autophagy in age-related neurodegeneration. Neuro-Signals. 2008;16(1):75-84.