

Selcuk Journal of Agriculture and Food Sciences

The effect of *in-ovo* injection of *Lactobacilla Rhamnosus* on hatching traits and growth parameters of quails

Abbas Fadhıl ABDULQADER¹, Ali AYGUN^{1, *}, Abdoulaziz Hamissou MAMAN¹, Osman OLGUN¹ ¹Selcuk University, Faculty of Agriculture, Department of Animal Science, Konya, Turkey

ARTICLE INFO

Article history: Received date: 19.06.2018 Accepted date: 23.07.2018

Keywords:

Lactobacilla Rhamnosus Quail eggs In-ovo Hatchability Growth

1. Introduction

In-ovo injection is a method to administer substances into the egg during incubation with the objective of promoting positive effects on hatchability and post-hatch growth performance (Uni and Ferket 2004). The *in-ovo* method was first used by Sharma and Burmester (1982) for the vaccination of turkey eggs against Marek's disease.

Researchers have recently used materials such as probiotic (Maiorano et al. 2012; Majidi-Mosleh et al. 2017; Triplett et al. 2018), proteins (Ohta et al. 1999; Bhanja et al. 2014; Kermanshahi et al. 2015), carbohydrates (Ipek et al. 2004; Tako et al. 2004; Zhai et al. 2011; Salmanzadeh et al. 2012), vitamins and minerals (Bello et al. 2013; Salary et al. 2014; Oliveira et al. 2015), ascorbic acid (Elibol et al. 2001; Ipek et al. 2004; Sgavioli et al. 2015), hormones (Kocamis et al. 1999), bee products (Coşkun et al. 2014; Moghaddam et al. 2014; Aygun 2016; Bozbay et al. 2016) *in-ovo* administration.

Lactobacillus rhamnosus is a member of the family Lactobacilliaceae and has received this name because it largely ferments carbohydrates to lactic acid. The digestive system of humans and animals is the natural habitat of these bacteria as well as finding a wide spreading area in the land (Mao et al. 2016). Immunity

ABSTRACT

The aim of this study was to investigate the effect of *in-ovo* injection of *Lactobacilla Rhamnosus* (0, 16×10^3 , 32×10^3 and 48×10^3 cfu) as probiotic source on hatching parameters and growth performance of quails. In this, a total of 400 quail hatching eggs, were randomly distributed among 4 experimental groups. Each experimental group contained 4 replicates of 25 eggs each. *In-ovo* injections of *Lactobacilla Rhamnosus* into quail eggs did not statistically affect incubation characteristics and some tissue weights. A high amount of *Lactobacilla Rhamnosus* injection into the breeder quail eggs negatively affected live weight and live weight gain. As a result, it can be said that low or medium levels *Lactobacilla Rhamnosus* injection into quail eggs have no effect on incubation and growth parameters of quails.

of interested hosts with Lactobacillus is increasing day by day due to its high antimicrobial effect and its ability to withstand high acidity (Naqid et al. 2015; Sugiharto et al. 2015; Xie et al. 2015). Lactobacillus rhamnosus have a high resistance to acidity and bile salts (Goldin et al. 1992; Alander et al. 1999), antimicrobial effect (Toki et al. 2009), has the properties of reducing negative effects against aflatoxin B1. (Gratz et al. 2007). Chen et al. (2017) stated that the addition of Lactobacillus rhamnosus in the broiler rations increased the growth performance and the meat quality, improvement of digestive system microflora and consequently, Lactobacillus rhamnosus may be a good probiotic for poultry. It has been reported that different probiotics in-ovo injection has a positive effect on digestive secretions and live body weight (Maiorano et al. 2012; Pruszynska-Oszmalek et al. 2015), but the feed conversion was not affected Pruszynska-Oszmalek et al. (2015) or adversely affected (Maiorano et al. 2012).

The aim of this study is to determine the effect of *in-ovo Lactobacillus rhamnosus* quail fertile eggs injection on incubation features and growth parameters.

2. Materials and Methods

This study was carried out at the Agricultural Faculty of Selcuk University, Konya, Turkey. A total of 400 quail hatching eggs were obtained from a

^{*} Corresponding author email: aaygun@selcuk.edu.tr

commercial farm (Konya, Turkey). As probiotic source *Lactobacillus rhamnosus* GG (Mansel pharmaceutical industry and trade inc.) microorganism were used. The physiological serum containing 50 ml of 0.9% NaCl was diluted to 16×10^3 , 32×10^3 and 48×10^3 cfu per egg of the probiotic source and was stirred at 500 rpm for 3 hours. A total of 400 quail hatching eggs, were randomly distributed among 4 experimental groups. Each experimental group contained 4 replicates of 25 eggs each.

The experimental groups consisted of a control group (without *in-ovo* injection) and 3 trial groups injected with $16x10^3$, $32x10^3$ and $48x10^3$ cfu of *Lactobacillus rhamnosus* GG per egg.

The incubator was set to $37.5 \, ^{\circ}$ C and 55-60% humidity for the first 14 days of the incubation and $37.2 \, ^{\circ}$ C temperature and 75% humidity for the hatching machine. The eggs were turned 12 times at 90° per day until 14 days of incubation.

Table 1	
Composition of basal d	iet

Composition o	i basai ulet		
Ingredients	%	Chemical	
		composition ²	
Maize	55.90	Metabolic energy,	2899
		kcal/kg	
Soybean meal	39.50	Crude protein, %	23.99
Vegetable oil	1.80	Calcium, %	1.150
Limestone	1.25	Total phosphorus, %	0.517
DCP	0.82	Available	0.299
		phosphorus, %	
Salt	0.30	Lysine, %	1.306
Premix ¹	0.25	Methionine, %	0.439
DL Methionine	0.18	Methionine +	0.940
		cystine, %	

¹Vitamin mineral premix ration in 1 kg, vitamin A, 8.800 IU; vitamin D3, 2.200 IU; vitamin E, 11 mg; nicotinic acid, 44 mg; Cal-D-Pan, 8.8 mg; riboflavin 4.4 mg; 2.5 mg of thiamine; vitamin B12, 6.6 mg, folic acid, 1 mg; D-Biotin, 0.11 mg; choline, 220 mg; manganese, 80 mg; copper, 5 mg; iron, 60 mg; zinc, 60 mg; cobalt, 0.20 mg; iodine, 1 mg; selenium, 0.15 mg.

² Analyzed value as feed

Table 2

Group	Fertility	Hatchability of set eggs	Hatchability fertile eggs	of	Embryonic mortality (% of fertile eggs)		
		00	00		1 to 9d	10 to 16d	17 to 18d
Control	86.54	75.96	87.52		4.56	3.42	4.50
16x10 ³ cfu	94.23	83.65	88.74		2.09	6.17	3.00
32×10^3 cfu	88.22	78.29	88.74		2.27	4.50	4.50
48x10 ³ cfu	85.58	76.92	89.44		3.57	1.19	5.80
SEM	2.48	4.50	3.40		1.69	1.40	1.91
P-value	0.117	0.694	0.988		0.726	0.244	0.816

On the 14th day of incubation, after the egg shell was disinfected with 70% ethanol, a hole was opened for injection with a micromotor (Strong 210, Korea). The prepared solutions were injected into amnion with a 26-gauge plastic disposable syringe (0.20 mL). After the injection, the hole was sealed with adhesive and transferred to the hatch basket. Eggs in the control group were kept out of the same temperature and time, so that the effect of environmental conditions was tried to be minimized. After injection, the hole was closed by dropping and placed in the hatch trays as 25 eggs in each subgroup. The incubation results were calculated as follow

Hatchability of fertile eggs (%) = number of eggs hatched out/number of fertile eggs x 100

Fertility (%) = number of fertile eggs/number of total eggs produced or set x 100.

Hatchability of set eggs (%) = number of eggs hatched out/number of eggs set in the incubator x 100.

After hatching time was ended the non-hatched eggs according to the macroscopic analyze were examined by the method described by Aygun et al. (2012).

Table 3

Effect of in-ovo probiotic injection into quail hatching eggs on some organ weights and live weight

	Organ wei	ghts,%	Live weight, g					
Group	Liver	Yolk sac	At	1. w	2. w	3. w	4. w	5. w
			hatch					
Control	2.66	8.49	8.50	19.40	36.03	76.06 ^a	116.77 ^a	151.33 ^a
16x10 ³ cfu	2.41	9.33	8.37	19.09	35.63	68.98^{ab}	112.09 ^a	141.48^{b}
32×10^3 cfu	2.51	9.16	8.53	19.25	35.59	72.31 ^a	113.64 ^a	150.08^{a}
48x10 ³ cfu	2.52	7.19	8.43	18.18	34.94	63.26 ^b	95.53 ^b	138.08^{b}
SEM	0.84	0.14	0.07	0.41	0.82	1.74	1.13	1.86
<i>P</i> -value	0.694	0.335	0.487	0.228	0.845	0.013	0.001	0.001

^{a-b} within a column, means with no common superscript letters are significantly different (P<0.05)

The growth performance of each group having 15 quail chicks were taken and placed in the quail growing cages. On the first day after incubation the house temperature was adjusted to 33 °C and decrease by 3 °C each week and was stable at 20 °C during the growth period, 23 hours of light and one hour of darkness were applied. During the growing period the quail growing feed (Table 1) and water were given ad libitum. For the determination of live weight changes, chicks were weighed weekly and weight gain was determined from these weights.

Statistical analysis

At the end of the experiment, the variance analyses were applied to all variables obtained from the trial groups (Minitab 2000), and the differences between means of the groups were determined by the Duncan test (Duncan 1955).

3. Results and Discussion

The effect of embryonic mortality on the incubation parameters of the *in-ovo* probiotic injection to breed quail eggs is shown in Table 2. No significant differences were found among the treatments groups for hatchability, fertility, and embryonic mortality. *In*- ovo Lactobacillus rhamnosus GG administration does not appear to have an adverse effect on incubation. Similar findings were reported in the previous studies. Edens et al. (1997) explained that the injection of L. reuteri in the turkey eggs did not influenced negatively the incubation results. Damron et al. (1981) stated that the injection of L. acidophilus in the turkey eggs did not influence negatively the incubation results. Triplett et al. (2018) found similar results with the injection of L. Acidophilus in the broiler laying hens eggs. The effects of treatments on the liver and yolk sac weights and live weight of quails at growth period are given in the Table 3. No significant differences were found between the experiment groups regarding the liver, yolk sac weights and weight at the hatch and 1st and 2nd week weights of growth period. Whereas the injection of probiotic in-ovo had a significant effect on the live weight of quail chicks at 3, 4 and 5 weeks of growth period .The control group at 3rd and 4th week of growth had the highest live weight value and the difference between the 48×10^3 cfu probiotic addition group was significant. The difference between the control group which was the group with the highest live weight value and the groups 16x10³ and 48x10³ cfu injected with in-ovo bacteria at 5 week was statistically significant.

Table 4

	1	• . • •	1 . 1 .	1 1
HITPOT OF IN-OVO	nrohiotic intection	1 1nto augu	hatching eggs	on live weight gain
Lincer or m=0.00		I muo quan	natening eggs	on nye weigin gam

Lifect of <i>in-ov</i>	o probiblic inje	ction into quan	natening eggs on	nve weight gam		
Group	0-1 w	1-2 w	2-3 w	3-4 w	4-5 w	0-5 w
Control	10.90	16.63	40.03 ^a	40.71 ^a	34.56 ^{ab}	142.83 ^a
16x10 ³ cfu	10.72	16.54	33.36 ^{ab}	43.11 ^a	29.39^{b}	133.11 ^{bc}
32×10^3 cfu	10.72	16.34	36.72 ^a	41.33 ^a	36.44 ^{ab}	141.55^{ab}
48×10^3 cfu	9.74	16.76	28.32^{b}	32.27 ^b	42.55 ^a	129.64 ^c
SEM	0.43	0.72	1.93	1.34	1.87	1.89
<i>P</i> -value	0.290	0.985	0.023	0.011	0.007	0.002

^{a-c} within a column, means with no common superscript letters are significantly different (P<0.05)

The effect of in-ovo probiotic injection on growth period live weight gain at 1st and 2nd week periods was not significant, while in other periods this trial effect became statistically significant. In the experiment 2-3, 3-4 and 0-5 the lowest live weight gain was observed in the chicks that hatched from the eggs with the highest probiotic injections. However, the live weight gain in the last week of the growth period was again obtained in the group with the highest probiotic injection. (Maiorano et al. 2012) reported that the coadministration of Lactococcus acidophilus (500 cfu) and Strepcoccus faecium (500 cfu) at the 12th day of incubation to the breeding broiler chicken eggs increased the growth period live weight but negatively affected the feed conversion. Similarly Pruszynska-Oszmalek et al. (2015) carried a research in which the prebiotics (inulin and Bi2tos) and synbiotics (inulin +Lactococcus lactissub sp. lactisand Bi2tos

+Lactococcus lactissubsp.cre-moris) were injected in-

ovo into the air cell on the 12th d embryonic development, they demonstrated the beneficial effects of prebiotics and synbiotics inoculated *in-ovo*. Bi2tos and inulin given with *Lactococcus lactis ssp.* when injected in-ovo on the 12th d chicken embryo development elevated body weight at the end of the rearing period. Simultaneously, the investigated compounds significantly increased the total activity of pancreatic enzymes; amylase, lipase, and trypsin.

4. Conclusion

It was concluded that injection of *Lactobacillus rhamnosus* in the breeding quail eggs at 14 day of incubation had no negative effect on the incubation parameter. However injection at the high level of *Lactobacillus rhamnosus* had a negative effect on the growth parameters.

5. References

- Alander M, Satokari R, Korpela R, Saxelin M, Vilpponen-Salmela T, Mattila-Sandholm T, von Wright A (1999) Persistence of colonization of human colonic mucosa by a probiotic strain, Lactobacillus rhamnosusGG, after oral consumption. *Applied and environmental microbiology* 65, 351-4.
- Aygun A (2016) The effects of in-ovo injection of propolis on egg hatchability and starter live performance of japanese quails. *Brazilian Journal of Poultry Science* **18**, 83-9.
- Aygun A, Sert D, Copur G (2012) Effects of propolis on eggshell microbial activity, hatchability, and chick performance in Japanese quail (Coturnix coturnix japonica) eggs. *Poultry Science* **91**, 1018-25.
- Bello A, Zhai W, Gerard P, Peebles E (2013) Effects of the commercial in ovo injection of 25hydroxycholecalciferol on the hatchability and hatching chick quality of broilers. *Poultry Science* 92, 2551-9.
- Bhanja S, Sudhagar M, Goel A, Pandey N, Mehra M, Agarwal S, Mandal A (2014) Differential expression of growth and immunity related genes influenced by in ovo supplementation of amino acids in broiler chickens. *Czech J. Anim. Sci* **59**, 399-408.
- Bozbay CK, Konanc K, Nuh O, Öztürk E (2016) Yumurta içi (İn Ovo) propolis enjeksiyonunun ve enjeksiyon yerinin kuluçka randımanı, civciv çıkış ağırlığı ve yaşama gücüne etkileri. *Türkiye Tarımsal Araştırmalar Dergisi* **3**, 48-54.
- Chen F, Gao S, Zhu L, Qin S, Qiu H (2017) Effects of dietary Lactobacillus rhamnosus CF supplementation on growth, meat quality, and microenvironment in specific pathogen-free chickens. *Poultry Science* 97, 118-23.
- Coşkun I, Çayan H, Yilmaz Ö, Taskin A, Tahtabiçen E, Samli HE (2014) Effects of in-ovo pollen extract injection to fertile broiler eggs on hatchability and subsequent chick weight. *Türk Tarım ve Doğa Bilimleri Dergisi* **1**, 485-9.
- Damron B, Wilson H, Voitle R, Harms R (1981) A mixed Lactobacillus culture in the diet of Broad Breasted Large White turkey hens. *Poultry Science* **60**, 1350-1.
- Duncan DB (1955) Multiple range and multiple F tests. *Biometrics* **11**, 1-42.
- Edens F, Parkhurst C, Casas I, Dobrogosz W (1997) Principles of ex ovo competitive exclusion and in ovo administration of Lactobacillus reuteri. *Poultry Science* **76**, 179-96.
- Elibol O, Türkoğlu M, Akan M, Erol H (2001) Effects of ascorbic acid injection during incubation on the

hatchability of large broiler eggs. *Turkish Journal* of Veterinary and Animal Sciences **25**, 245-8.

- Goldin BR, Gorbach SL, Saxelin M, Barakat S, Gualtieri L, Salminen S (1992) Survival ofLactobacillus species (strain GG) in human gastrointestinal tract. *Digestive diseases and sciences* **37**, 121-8.
- Gratz S, Wu Q, El-Nezami H, Juvonen R, Mykkänen H, Turner P (2007) Lactobacillus rhamnosus strain GG reduces aflatoxin B1 transport, metabolism, and toxicity in Caco-2 cells. *Applied and environmental microbiology* **73**, 3958-64.
- Ipek A, Sahan U, Yilmaz B (2004) The effect of in ovo ascorbic acid and glucose injection in broiler breeder eggs on hatchability and chick weight. *Archiv Fur Geflugelkunde* 68, 132-5.
- Kermanshahi H, Daneshmand A, Emami NK, Tabari DG, Doosti M, Javadmanesh A, Ibrahim SA (2015) Effect of in ovo injection of threonine on Mucin2 gene expression and digestive enzyme activity in Japanese quail (Coturnix japonica). *Research in Veterinary Science* **100**, 257-62.
- Kocamis H, Yeni Y, Kirkpatrick-Keller D, Killefer J (1999) Postnatal growth of broilers in response to in ovo administration of chicken growth hormone. *Poultry Science* 78, 1219-26.
- Maiorano G, Sobolewska A, Cianciullo D, Walasik K, Elminowska-Wenda G, Sławińska A, Tavaniello S, Żylińska J, Bardowski J, Bednarczyk M (2012) Influence of in ovo prebiotic and synbiotic administration on meat quality of broiler chickens. *Poultry Science* **91**, 2963-9.
- Majidi-Mosleh A, Sadeghi A, Mousavi S, Chamani M, Zarei A (2017) Effects of in Ovo Infusion of Probiotic Strains on Performance Parameters, Jejunal Bacterial Population and Mucin Gene Expression in Broiler Chicken. *Revista Brasileira de Ciência Avícola* 19, 97-102.
- Mao X, Gu C, Hu H, Tang J, Chen D, Yu B, He J, Yu J, Luo J, Tian G (2016) Dietary Lactobacillus rhamnosus GG supplementation improves the mucosal barrier function in the intestine of weaned piglets challenged by porcine rotavirus. *PLOS ONE* **11**, e0146312.
- Minitab I (2000) MINITAB statistical software. *Minitab Release* **13**.
- Moghaddam A, Borji M, Komazani D (2014) Hatchability rate and embryonic growth of broiler chicks following in ovo injection royal jelly. *British poultry science* **55**, 391-7.
- Naqid IA, Owen JP, Maddison BC, Gardner DS, Foster N, Tchórzewska MA, La Ragione RM, Gough KC (2015) Prebiotic and probiotic agents enhance antibody-based immune responses to Salmonella Typhimurium infection in pigs. *Animal Feed Science and Technology* 201, 57-65.

178

- Ohta Y, Tsushima N, Koide K, Kidd M, Ishibashi T (1999) Effect of amino acid injection in broiler breeder eggs on embryonic growth and hatchability of chicks. *Poultry Science* **78**, 1493-8.
- Oliveira T, Bertechini A, Bricka R, Kim E, Gerard P, Peebles E (2015) Effects of in ovo injection of organic zinc, manganese, and copper on the hatchability and bone parameters of broiler hatchlings. *Poultry Science* **94**, 2488-94.
- Pruszynska-Oszmalek E, Kolodziejski P, Stadnicka K, Sassek M, Chalupka D, Kuston B, Nogowski L, Mackowiak P, Maiorano G, Jankowski J (2015) In ovo injection of prebiotics and synbiotics affects the digestive potency of the pancreas in growing chickens. *Poultry Science* 94, 1909-16.
- Salary J, Sahebi-Ala F, Kalantar M, Matin HRH (2014) In ovo injection of vitamin E on post-hatch immunological parameters and broiler chicken performance. Asian Pacific Journal of Tropical Biomedicine 4, S616-S9.
- Salmanzadeh M, Ebrahimnezhad Y, Shahryar HA, Beheshti R (2012) The effects of in ovo injection of glucose and magnesium in broiler breeder eggs on hatching traits, performance, carcass characteristics and blood parameters of broiler chickens. *Arch. Geflugelkunde* **76**, 277-84.
- Sgavioli S, Matos Júnior J, Borges L, Praes M, Morita V, Zanirato G, Garcia R, Boleli I (2015) Effects of ascorbic acid injection in incubated eggs submitted to heat stress on incubation parameters and chick quality. *Revista Brasileira de Ciência Avícola* 17, 181-9.
- Sharma J, Burmester B (1982) Resistance of Marek's disease at hatching in chickens vaccinated as embryos with the turkey herpesvirus. *Avian Diseases*, 134-49.
- Sugiharto S, Lauridsen C, Jensen BB (2015) Gastrointestinal ecosystem and immunological responses in E. coli challenged pigs after weaning fed liquid diets containing whey permeate fermented with different lactic acid bacteria. *Animal Feed Science and Technology* **207**, 278-82.
- Tako E, Ferket P, Uni Z (2004) Effects of in ovo feeding of carbohydrates and beta-hydroxy-betamethylbutyrate on the development of chicken intestine. *Poultry Science* **83**, 2023-8.
- Toki S, Kagaya S, Shinohara M, Wakiguchi H, Matsumoto T, Takahata Y, Morimatsu F, Saito H, Matsumoto K (2009) Lactobacillus rhamnosus GG and Lactobacillus casei suppress Escherichia coliinduced chemokine expression in intestinal epithelial cells. *International archives of allergy* and immunology 148, 45-58.
- Triplett M, Zhai W, Peebles E, McDaniel C, Kiess A (2018) Investigating commercial in ovo technology as a strategy for introducing probiotic bacteria to broiler embryos. *Poultry Science* 97, 658-66.

- Uni Z, Ferket R (2004) Methods for early nutrition and their potential. *World's Poultry Science Journal* **60**, 101-11.
- Xie J, Yu Q, Nie S, Fan S, Xiong T, Xie M (2015) Effects of Lactobacillus plantarum NCU116 on intestine mucosal immunity in immunosuppressed mice. *Journal of Agricultural and Food Chemistry* 63, 10914-20.
- Zhai W, Gerard P, Pulikanti R, Peebles E (2011) Effects of in ovo injection of carbohydrates on embryonic metabolism, hatchability, and subsequent somatic characteristics of broiler hatchlings. *Poultry Science* **90**, 2134-43.