

Managing *Helicobacter pylori* infection: transitioning from conventional to alternative treatment approaches

Serhat Öcal[✉]

Department of Internal Diseases, Keçiören VM Medical Park Hospital, Ankara, Turkey

ABSTRACT

Helicobacter pylori, an essential constituent of the gastric microbiome in those infected, is commonly associated with medical conditions such as chronic gastritis, peptic ulcer disease, and gastric cancer. In recent years, the growing resistance to antibiotics worldwide has emerged as a substantial hurdle in the effective treatment of *H. pylori* infection. Consequently, it has necessitated the exploration of innovative treatment strategies aimed at bolstering the potency of existing antibiotic-based eradication therapies. Such avant-garde strategies include the incorporation of probiotics and prebiotics as complementary measures to *H. pylori* treatment, the use of antimicrobial peptides as potential replacements for traditional antibiotics, and the application of photodynamic therapy via ingestible devices. Other advanced methodologies entail deploying drug delivery systems that utilize microparticles and nanoparticles, the invention of vaccines, the exploration of natural products, and the potential use of phage therapy. This review offers a contemporary synopsis of these burgeoning strategies designed to suppress *H. pylori*, delving into their strengths, hurdles, and aspects to consider during their development. A significant achievement would be the creation of an efficient human vaccine; however, previous attempts at developing such vaccines have met with obstacles or even cessation. Numerous natural products have displayed anti-*H. pylori* properties, predominantly in laboratory environments. Nonetheless, a requirement remains for more extensive clinical studies to fully comprehend their role in exterminating *H. pylori*. Finally, phage therapy, while demonstrating potential as a suitable alternative, grapples with considerable challenges, chiefly the isolation of highly virulent bacteriophages that specifically target *H. pylori*.

Keywords: *Helicobacter pylori*, antibiotic resistance, innovative treatment strategies

Helicobacter pylori (*H. pylori*), a Gram-negative, helical bacterium, exhibits high global prevalence, infecting over half of the worldwide population [1, 2]. It primarily colonizes the gastric mucosa, constituting the majority of the gastric microbiota in those infected [3-7]. Infections with *H. pylori*, which typically initiates in childhood and can persist lifelong if left untreated, are linked to an array of gastric and extragastric conditions. These include

chronic gastritis, peptic ulcer disease, gastric carcinoma, and gastric mucosa-associated lymphoid tissue (MALT) lymphoma [8]. The pathogenic potential of individual *H. pylori* strains significantly influences these clinical outcomes, with certain virulence factors associated with an elevated risk of disease development [9-11].

H. pylori infection also impinges upon the gastric and gut microbiota, potentially playing a contributory

Corresponding author: Serhat Öcal, MD.,
Phone: +90 312 666 08 00, E-mail: serhatocal73@hotmail.com

Received: June 28, 2023
Accepted: August 13, 2023
Published Online: August 23, 2023

How to cite this article: Öcal S. Managing *Helicobacter pylori* infection: transitioning from conventional to alternative treatment approaches. Eur Res J. 2024;10(1):136-143. doi: 10.18621/eurj.1320819

Copyright © 2024 by Prusa Medical Publishing
Available at <http://dergipark.org.tr/eurj>



This is an open access article distributed under the terms of [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/)



role in both gastric and extragastric manifestations [12]. Nevertheless, the attempt to eradicate *H. pylori* via antibiotic therapy elicits concerns regarding the emergence of antibiotic-resistant strains and disruptions in the composition of the intestinal microbiota [13-17].

Owing to the high prevalence of *H. pylori* infection and its concomitant diseases, efficient eradication therapy is vital for clinical management. Contemporary treatment guidelines take into account previous antibiotic exposure and regional resistance rates to select the apt regimen. Triple therapy involving a proton-pump inhibitor (PPI), amoxicillin, and clarithromycin for a 14-day period is recommended as the first-line therapy in regions with low clarithromycin resistance [18]. An alternative option is Bismuth-containing quadruple therapy (BQT). In cases of high clarithromycin resistance, quadruple concomitant therapy or BQT is suggested [19]. Levofloxacin-based therapy is discouraged as a first-line treatment due to the escalating fluoroquinolone resistance [20]. Other recommended options include sequential therapy and fluoroquinolone-based LOAD therapy [21, 22]. The selection of the PPI molecule is also of importance, with second-generation PPIs demonstrating higher efficacy than their first-generation counterparts [23]. BQT is considered the most effective first-line therapy, while dual therapy employing amoxicillin and vonoprazan exhibits potential in East Asia [18, 24, 25].

Antibiotic resistance poses a major obstacle in *H. pylori* treatment, with eradication rates dwindling and resistance rates surging globally [25-29]. The improvement of diagnostic tools, such as non-invasive testing methods, is instrumental in combating antibiotic resistance [18, 30]. Additionally, factors related to patient compliance, side effects, and host genetic variants necessitate the exploration of alternative therapeutic approaches for *H. pylori* treatment.

Alternative Approaches for *H. pylori* Treatment

Innovative therapies for *H. pylori* infections strive to meet WHO standards for eradication rates, patient compliance, and prevention of antimicrobial resistance. These include strategies like the use of probiotics and prebiotics, antimicrobial peptides, photodynamic therapy, natural products, vaccines, micro- and nanoparticles, and phage therapy [31-34].

Probiotics are live microorganisms that confer health benefits. They modulate the immune response, generate antimicrobial substances, and compete with *H. pylori* for adhesion sites. Lactobacillus and Bifidobacterium are the commonly used probiotic genera [31, 35, 36]. Prebiotics are non-digestible nutrients metabolized by bacteria. They stimulate the growth of specific intestinal bacteria and are utilized as supplements to bolster *H. pylori* treatment [32]. Supplementation with probiotics and prebiotics in *H. pylori* therapy has demonstrated increased eradication rates and reduced side effects in patients [33, 34]. Mechanisms of probiotic action include immunological modulation, production of antimicrobial substances, inhibition of adhesion, and enhancement of mucin secretion [35, 36].

Antimicrobial peptides (AMPs), naturally produced by organisms, play a pivotal role in innate immunity. They interact with microbial cell membranes, leading to cell lysis. Synthetic AMP analogs have exhibited anti-*H. pylori* activity [37, 38]. AMPs produced by gastric epithelial cells regulate the bacterial population in the stomach, but *H. pylori* has evolved resistance mechanisms [39-41].

Present research focuses on identifying efficacious probiotic strains and evaluating their efficiency either alone or in combination with antibiotics. Discrepant results have been reported, necessitating further studies to ascertain their impact on *H. pylori* eradication rates and gut microbiota [42-47]. In summary, while alternative approaches such as probiotics, prebiotics, and antimicrobial peptides exhibit potential in *H. pylori* treatment, additional research is required to determine their effectiveness and optimal usage [37].

Antimicrobial Peptides

Antimicrobial peptides (AMPs) are short oligopeptides possessing a positive charge, produced naturally by numerous organisms. Their role in innate immunity is essential, enabling them to defend against a variety of pathogens, including *H. pylori*. AMPs interfere with microbial cell membranes, instigating increased membrane permeability, pore formation, and leading to cell lysis. Synthetic AMP analogues, for instance, pexiganan, tilapia piscidins, and PGLa-AM1, have shown a remarkable ability to combat *H. pylori*. Bicarinalin and cathelicidins have also displayed anti-*H. pylori* activity. Despite their effectiveness, *H. pylori*

has evolved specific resistance mechanisms against host AMPs. The development of new synthetic AMP analogues and further research is actively underway [37, 38, 48-50].

Photodynamic Therapy

Photodynamic therapy (PDT) combines a photoactive molecule, termed a photosensitizer, with visible light and oxygen to generate cytotoxic reactive oxygen species (ROS). The potential of PDT as a treatment for *H. pylori* infections is currently under investigation. *H. pylori* inherently synthesizes and stores photosensitizers, making it suitable for antibacterial PDT without the requirement for external photosensitizers. In vitro research has shown significant reductions in *H. pylori* colony-forming units (CFU) following PDT using both red and blue lights. Moreover, the conjunction of PDT with antibiotics has resulted in a synergistic antibacterial effect. The use of PDT without the addition of exogenous photosensitizers has shown minimal side effects on gastric mucosa. Current research aims to refine PDT efficacy by considering tissue interactions and optical properties [51-58].

Micro- and Nanoparticles for Drug Delivery

Microparticles (MPs) and nanoparticles (NPs) have demonstrated potential in drug delivery for the treatment of microbial infections, including *H. pylori*. The small size and high surface-to-volume ratio of these particles enhance therapeutic efficacy and diminish side effects in comparison to traditional antibiotic-based treatments. Specifically, NPs can navigate physiological barriers and interact with pathogen membranes and cell walls. Drugs, such as antibiotics, can be loaded onto these particles, ensuring delivery to the infection site and protection from degradation and resistance mechanisms [57-60].

Expanding on these alternative approaches, AMPs, PDT, and micro- and nanoparticles provide potential novel treatments for *H. pylori*. Currently, research is directed towards optimizing their efficacy and suitability for clinical settings.

Microparticles and Nanoparticles for *H. pylori* Eradication

In the quest to enhance *H. pylori* eradication, microparticles and nanoparticles have come under investigation. Chitosan-based MPs and NPs have shown

promise as gastric drug delivery systems, due to their biocompatibility, antimicrobial attributes, and mucoadhesiveness. Progress has been made using chitosan-based mini-tablets, nanoparticles, or hydrogels loaded with amoxicillin. Moreover, Poly (lactic-co-glycolic acid) nanoparticles possessing pH-sensitive and acid-resistant properties have been developed for targeted drug delivery to *H. pylori* infection sites. Metal nanoparticles, including gold, silver, and zinc oxide, have demonstrated good biocompatibility and anti-*H. pylori* activity. Another breakthrough was the coating of polymeric nanoparticles with *H. pylori* outer membrane proteins, which led to reduced *H. pylori* adhesion to gastric tissues. The combination of lytic bacteriophages with nanoparticles resulted in a synergistic effect, decreasing *H. pylori* colonization. Further optimization of these methods is required [61-64].

Vaccines against *H. pylori*

The development of a highly effective vaccine against *H. pylori* is perceived as a cost-effective method to prevent infection and associated diseases. Although prophylactic and therapeutic vaccines have been tested, no large-scale vaccine has been successfully produced yet. Clinical trials of different vaccine formulations have exhibited some immune responsiveness but have not consistently reduced bacterial load or stimulated protective immunity. Current research is centered on multivalent epitope-based vaccines, recombinant or fusion protein-based vaccines, and mucosal adjuvants. Oral vaccines face obstacles, but nanoparticles have shown promise in safeguarding antigens and eliciting immune responses. For instance, chitosan nanoparticles have displayed stability, enhanced immune response, and prolonged release of DNA vaccines. Immunoinformatics has also contributed to the screening of antigen targets and designing epitope-based vaccines. Vaccine development necessitates further research [65-67].

Natural Products against *H. pylori*

Natural products like plants, fruits, and spices have been explored for their anti-*H. pylori* properties. These products exhibit inhibitory effects on bacterial enzymes, possess anti-adhesive and anti-inflammatory properties, and provide bactericidal or bacteriostatic effects. Investigations have been carried out on Citrus bergamia derivatives, blueberry, grape seed extract,

mastic gum, cinnamon, ginger, curcumin, chestnut, oak honey, and propolis. However, further studies are needed to identify the specific compounds and mechanisms responsible for their activity [68-71].

Phage Therapy for *H. pylori*

The concept of phage therapy, which employs bacteriophages to target and eradicate bacterial infections, has sparked interest in treating *H. pylori*. Phages are known for their ability to selectively lyse specific bacterial strains while leaving the microbiota unharmed. Deemed safe for clinical use, phages replicate inside bacterial host cells, enabling them to target new bacterial cells. Phage therapy has proven effective in treating infections caused by pathogenic and antibiotic-resistant bacteria. Despite its potential, further research is required to translate phage therapy into a clinical application for *H. pylori* [72-76].

Interestingly, phage therapy for *H. pylori* infections has shown potential. In a study, a therapy was developed combining phage Hp ϕ with lactoferrin adsorbed on hydroxyapatite nanoparticles (NPs). This novel treatment demonstrated enhanced antimicrobial effects against *H. pylori* in human gastric cancer cells [77]. Yet, comprehensive data on phages and phage-*H. pylori* interactions in the gastric environment are lacking. The absence of sequenced phage genomes restricts our understanding of key factors like toxins, antimicrobial resistance genes, and virulence in *H. pylori* phages. Moreover, reports of endolysins present in *H. pylori* phages are nonexistent [78].

Endolysins are phage proteins responsible for breaking down bacterial cell walls and present an alternative approach in phage therapy. Endolysins show specificity to the bacterial host, and to date, bacterial resistance to endolysins hasn't been reported [79]. However, treating Gram-negative bacteria like *H. pylori* is challenging due to the presence of an outer membrane barrier. Strategies involving engineering approaches or combining lysins with weak acids have shown success in permeabilizing the outer membrane [80, 81].

CONCLUSION

The landscape for *H. pylori* eradication continues to

evolve, with promising alternative approaches emerging from recent research. Antimicrobial peptides, phage therapy, nanoparticle-enhanced delivery, vaccines, and natural products represent novel strategies that are redefining the treatment modalities available for *H. pylori* infection.

Antimicrobial peptides (AMPs), both natural and synthetic, offer a new way to combat *H. pylori*. The ongoing development of synthetic AMP analogues and the further understanding of their interaction with microbial cell membranes are expected to enhance their potential as an effective therapeutic approach.

Phage therapy has also shown significant potential for *H. pylori* treatment. The ability of bacteriophages to selectively lyse specific bacterial strains could lead to more targeted therapies that do not disrupt the microbiota. The combined use of endolysins and engineered approaches presents a compelling opportunity to overcome the challenges presented by the outer membrane barrier of Gram-negative bacteria such as *H. pylori*.

Nanotechnology has emerged as a potent tool for improved drug delivery. The properties of microparticles and nanoparticles, including their small size, high surface-to-volume ratio, and ability to overcome physiological barriers, hold significant promise for enhanced therapeutic effectiveness.

Moreover, the development of a vaccine against *H. pylori* has been considered a cost-effective method to prevent infection and associated diseases. Despite various challenges, ongoing research in multivalent epitope-based vaccines and recombinant or fusion protein-based vaccines, coupled with the use of mucosal adjuvants and nanoparticles, suggests that a successful vaccine may be within reach.

Finally, natural products also offer an appealing direction for research, with their demonstrated inhibitory effects on *H. pylori* and the potential for fewer side effects compared to synthetic drugs.

However, despite these promising alternatives, further in vivo studies and well-designed clinical trials are necessary to fully understand and realize their potential. As our understanding of *H. pylori* and its resistance mechanisms expands, these targeted therapies could provide personalized treatment options that selectively deplete *H. pylori* without disrupting the normal gastric microbiome.

In this context, these alternatives represent not only a departure from the traditional antibiotic-based treatments, but also a promising pathway towards more effective and sustainable solutions for the management and eradication of *H. pylori* infections. Such advancements could significantly improve patient outcomes, reduce the societal and economic burden of these infections, and usher in a new era of personalized and effective treatments for *H. pylori*.

In summary, the research landscape for *H. pylori* eradication is expanding, with alternative approaches like phage therapy, antimicrobial peptides, nanoparticles, vaccines, and more showing great potential. However, comprehensive in vivo studies and well-structured clinical trials are crucial to fully grasp their efficacy. These targeted therapies can provide personalized treatment options that selectively deplete *H. pylori* without disrupting the normal gastric microbiome, making them exciting alternatives to broad-spectrum antibiotics and non-selective treatments for *H. pylori* infections.

Authors' Contribution

Study Conception: SÖ; Study Design: SÖ; Supervision: SÖ; Funding: N/A; Materials: N/A; Data Collection and/or Processing: SÖ; Statistical Analysis and/or Data Interpretation: SÖ; Literature Review: SÖ; Manuscript Preparation: SÖ and Critical Review: SÖ.

Conflict of interest

The author disclosed no conflict of interest during the preparation or publication of this manuscript.

Financing

The author disclosed that they did not receive any grant during conduction or writing of this study.

REFERENCES

- Azevedo NF, Pinto AR, Reis NM, Vieira MJ, Keevil CW. Shear stress, temperature, and inoculation concentration influence the adhesion of water-stressed *Helicobacter pylori* to stainless steel 304 and polypropylene. *Appl Environ Microbiol*. 2006;72(4):2936-2941. doi: 10.1128/AEM.72.4.2936-2941.2006.
- Hooi JKY, Lai WY, Ng WK, et al. Global Prevalence of *Helicobacter pylori* Infection: Systematic Review and Meta-Analysis. *Gastroenterology*. 2017;153(2):420-429. doi: 10.1053/j.gastro.2017.04.022.
- Bik EM, Eckburg PB, Gill SR, et al. Molecular analysis of the bacterial microbiota in the human stomach. *Proc Natl Acad Sci U S A*. 2006;103(3):732-737. doi: 10.1073/pnas.0506655103.
- Klymiuk I, Bilgiler C, Stadlmann A, Thannesberger J, et al. The Human Gastric Microbiome Is Predicated upon Infection with *Helicobacter pylori*. *Front Microbiol*. 2017;8:2508. doi: 10.3389/fmicb.2017.02508.
- Parsons BN, Ijaz UZ, D'Amore R, et al. Comparison of the human gastric microbiota in hypochlorhydric states arising as a result of *Helicobacter pylori*-induced atrophic gastritis, autoimmune atrophic gastritis and proton pump inhibitor use. *PLoS Pathog*. 2017;13(11):e1006653. doi: 10.1371/journal.ppat.1006653.
- Wang Z, Bafadhel M, Haldar K, et al. Lung microbiome dynamics in COPD exacerbations. *Eur Respir J*. 2016;47(4):1082-1092. doi: 10.1183/13993003.01406-2015.
- Ferreira RM, Pereira-Marques J, Pinto-Ribeiro I, et al. Gastric microbial community profiling reveals a dysbiotic cancer-associated microbiota. *Gut*. 2018;67(2):226-236. doi: 10.1136/gutjnl-2017-314205.
- Cover TL, Blaser MJ. *Helicobacter pylori* in health and disease. *Gastroenterology*. 2009;136(6):1863-1873. doi: 10.1053/j.gastro.2009.01.073.
- Gerhard M, Rad R, Prinz C, Naumann M. Pathogenesis of *Helicobacter pylori* infection. *Helicobacter*. 2002;7 Suppl 1:17-23. doi: 10.1046/j.1523-5378.7.s1.3.x.
- Blaser MJ. *Helicobacter pylori* and the pathogenesis of gastroduodenal inflammation. *J Infect Dis*. 1990;161(4):626-633. doi: 10.1093/infdis/161.4.626.
- Ferreira RM, Machado JC, Figueiredo C. Clinical relevance of *Helicobacter pylori vacA* and *cagA* genotypes in gastric carcinoma. *Best Pract Res Clin Gastroenterol*. 2014;28(6):1003-1015. doi: 10.1016/j.bpg.2014.09.004.
- Franceschi F, Annalisa T, Teresa DR, et al. Role of *Helicobacter pylori* infection on nutrition and metabolism. *World J Gastroenterol*. 2014;20(36):12809-12817. doi: 10.3748/wjg.v20.i36.12809.
- Liou JM, Chen CC, Chen MJ. Sequential versus triple therapy for the first-line treatment of *Helicobacter pylori*: a multicentre, open-label, randomised trial. *Lancet*. 2013;381(9862):205-213. doi: 10.1016/S0140-6736(12)61579-7.
- Tai WC, Liang CM, Kuo CM, et al. A 14 day esomeprazole and amoxicillin-containing high-dose dual therapy regimen achieves a high eradication rate as first-line anti-*Helicobacter pylori* treatment in Taiwan: a prospective randomized trial. *J Antimicrob Chemother*. 2019;74(6):1718-1724. doi: 10.1093/jac/dkz046.
- Hsu PI, Wu DC, Chen WC, et al. Randomized controlled trial comparing 7-day triple, 10-day sequential, and 7-day concomitant therapies for *Helicobacter pylori* infection. *Antimicrob Agents Chemother*. 2014;58(10):5936-5942. doi: 10.1128/AAC.02922-14.
- Hanada K, Graham DY. *Helicobacter pylori* and the molecular pathogenesis of intestinal-type gastric carcinoma. *Expert Rev Anticancer Ther*. 2014;14(8):947-954. doi: 10.1586/14737140.2014.911092.
- Fallone CA, Chiba N, van Zanten SV, et al. The Toronto Consensus for the Treatment of *Helicobacter pylori* Infection in Adults. *Gastroenterology*. 2016;151(1):51-69. doi: 10.1053/j.gastro.2016.04.006.
- Malfertheiner P, Megraud F, O'Morain CA, et al. Manage-

- ment of *Helicobacter pylori* infection-the Maastricht V/Florence Consensus Report. *Gut*. 2017;66(1):6-30. doi: 10.1136/gutjnl-2016-312288.
19. Molina-Infante J, Romano M, Fernandez-Bermejo M, et al. Optimized nonbismuth quadruple therapies cure most patients with *Helicobacter pylori* infection in populations with high rates of antibiotic resistance. *Gastroenterology*. 2013;145(1):121-128. doi: 10.1053/j.gastro.2013.03.050.
20. Li M, Oshima T, Horikawa T, Tozawa K, Tomita T, Fukui H, Watari J, Miwa H. Systematic review with meta-analysis: Vonoprazan, a potent acid blocker, is superior to proton-pump inhibitors for eradication of clarithromycin-resistant strains of *Helicobacter pylori*. *Helicobacter*. 2018;23(4):e12495. doi: 10.1111/hel.12495.
21. Guevara B, Cogdill AG. *Helicobacter pylori*: A Review of Current Diagnostic and Management Strategies. *Dig Dis Sci*. 2020;65(7):1917-1931. doi: 10.1007/s10620-020-06193-7.
22. Gisbert JP, Romano M, Gravina AG, et al. *Helicobacter pylori* second-line rescue therapy with levofloxacin- and bismuth-containing quadruple therapy, after failure of standard triple or non-bismuth quadruple treatments. *Aliment Pharmacol Ther*. 2015;41(8):768-775. doi: 10.1111/apt.13128.
23. Murakami K, Sakurai Y, Shiino M, Funao N, Nishimura A, Asaka M. Vonoprazan, a novel potassium-competitive acid blocker, as a component of first-line and second-line triple therapy for *Helicobacter pylori* eradication: a phase III, randomised, double-blind study. *Gut*. 2016;65(9):1439-1446. doi: 10.1136/gutjnl-2015-311304.
24. Suzuki S, Gotoda T, Kusano C, Iwatsuka K, Moriyama M. The Efficacy and Tolerability of a Triple Therapy Containing a Potassium-Competitive Acid Blocker Compared With a 7-Day PPI-Based Low-Dose Clarithromycin Triple Therapy. *Am J Gastroenterol*. 2016;111(7):949-956. doi: 10.1038/ajg.2016.182.
25. Gatta L, Vakil N, Vaira D, Scarpignato C. Global eradication rates for *Helicobacter pylori* infection: systematic review and meta-analysis of sequential therapy. *BMJ*. 2013;347:f4587. doi: 10.1136/bmj.f4587.
26. O'Morain NR, Dore MP, O'Connor AJP, Gisbert JP, O'Morain CA. Treatment of *Helicobacter pylori* infection in 2018. *Helicobacter*. 2018;23 Suppl 1:e12519. doi: 10.1111/hel.12519.
27. Savoldi A, Carrara E, Graham DY, Conti M, Tacconelli E. Prevalence of Antibiotic Resistance in *Helicobacter pylori*: A Systematic Review and Meta-analysis in World Health Organization Regions. *Gastroenterology*. 2018;155(5):1372-1382. doi: 10.1053/j.gastro.2018.07.007.
28. Megraud F, Coenen S, Versporten A, et al. *Helicobacter pylori* resistance to antibiotics in Europe and its relationship to antibiotic consumption. *Gut*. 2013;62(1):34-42. doi: 10.1136/gutjnl-2012-302254.
29. Savoldi A, Carrara E, Graham DY, Conti M, Tacconelli E. Prevalence of Antibiotic Resistance in *Helicobacter pylori*: A Systematic Review and Meta-analysis in World Health Organization Regions. *Gastroenterology*. 2018;155(5):1372-1382. doi: 10.1053/j.gastro.2018.07.007.
30. Malfertheiner P, Megraud F, O'Morain C, et al. Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. *Gut*. 2007;56(6):772-781. doi: 10.1136/gut.2006.101634.
31. Hill C, Guarner F, Reid G, et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat Rev Gastroenterol Hepatol*. 2014;11(8):506-514. doi: 10.1038/nrgastro.2014.66.
32. Davani-Davari D, Negahdaripour M, Karimzadeh I, et al. Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods*. 2019;8(3):92. doi: 10.3390/foods8030092.
33. Ahmad K, Fatemeh F, Mehri N, Maryam S. Probiotics for the treatment of pediatric *Helicobacter pylori* infection: a randomized double blind clinical trial. *Iran J Pediatr*. 2013;23(1):79-84.
34. Lionetti E, Miniello VL, Castellaneta SP, et al. *Lactobacillus reuteri* therapy to reduce side-effects during anti-*Helicobacter pylori* treatment in children: a randomized placebo controlled trial. *Aliment Pharmacol Ther*. 2006;24(10):1461-1468. doi: 10.1111/j.1365-2036.2006.03145.x.
35. Sgouras DN, Panayotopoulou EG, Martinez-Gonzalez B, Petraki K, Michopoulos S, Mentis A. *Lactobacillus johnsonii* La1 attenuates *Helicobacter pylori*-associated gastritis and reduces levels of proinflammatory chemokines in C57BL/6 mice. *Clin Diagn Lab Immunol*. 2005;12(12):1378-1386. doi: 10.1128/CDLI.12.12.1378-1386.2005.
36. Kulkarni T, Majarikar S, Deshmukh M, et al. Probiotic sepsis in preterm neonates-a systematic review. *Eur J Pediatr*. 2022;181(6):2249-2262. doi: 10.1007/s00431-022-04452-5.
37. Joshi A, Agrawal A, Bhattacharya S. Formulation and clinical advancement of nanourchins: a novel multibranching nanoparticulate drug-delivery system. *Nanomedicine (Lond)*. 2022;17(20):1477-1499. doi: 10.2217/nmm-2022-0096.
38. Luong AD, Buzid A, Luong JHT. Important Roles and Potential Uses of Natural and Synthetic Antimicrobial Peptides (AMPs) in Oral Diseases: Cavity, Periodontal Disease, and Thrush. *J Funct Biomater*. 2022;13(4):175. doi: 10.3390/jfb13040175.
39. Gao JH, Guo LJ, Huang ZY, Rao JN, Tang CW. Roles of cellular polyamines in mucosal healing in the gastrointestinal tract. *J Physiol Pharmacol*. 2013;64(6):681-693.
40. Ruan Q, Guan P, Qi W, et al. *Porphyromonas gingivalis* regulates atherosclerosis through an immune pathway. *Front Immunol*. 2023;14:1103592. doi: 10.3389/fimmu.2023.1103592.
41. Park SC, Park Y, Hahm KS. The role of antimicrobial peptides in preventing multidrug-resistant bacterial infections and biofilm formation. *Int J Mol Sci*. 2011;12(9):5971-5992. doi: 10.3390/ijms12095971.
42. Jungersen M, Wind A, Johansen E, Christensen JE, Stuer-Lauridsen B, Eskesen D. The Science behind the Probiotic Strain *Bifidobacterium animalis* subsp. *lactis* BB-12(®). *Microorganisms*. 2014;2(2):92-110. doi: 10.3390/microorganisms2020092.
43. Michalak A, Kasztelan-Szczerbińska B, Cichoż-Lach H. Impact of Obesity on the Course of Management of Inflammatory Bowel Disease-A Review. *Nutrients*. 2022;14(19):3983. doi: 10.3390/nu14193983.
44. Zhang MM, Qian W, Qin YY, He J, Zhou YH. Probiotics in *Helicobacter pylori* eradication therapy: a systematic review and meta-analysis. *World J Gastroenterol*. 2015;21(14):4345-4357. doi: 10.3748/wjg.v21.i14.4345.
45. McFarland LV, Huang Y, Wang L, Malfertheiner P. System-

- atic review and meta-analysis: Multi-strain probiotics as adjunct therapy for *Helicobacter pylori* eradication and prevention of adverse events. *United European Gastroenterol J.* 2016;4(4):546-561. doi: 10.1177/2050640615617358.
46. Milner E, Stevens B, An M, et al. Utilizing Probiotics for the Prevention and Treatment of Gastrointestinal Diseases. *Front Microbiol.* 2021;12:689958. doi: 10.3389/fmicb.2021.689958.
47. Ji J, Yang H. Using Probiotics as Supplementation for *Helicobacter pylori* Antibiotic Therapy. *Int J Mol Sci.* 2020;21(3):1136. doi: 10.3390/ijms21031136.
48. Igarashi M, Kitada Y, Yoshiyama H, Takagi A, Miwa T, Koga Y. Ammonia as an accelerator of tumor necrosis factor alpha-induced apoptosis of gastric epithelial cells in *Helicobacter pylori* infection. *Infect Immun.* 2001;69(2):816-821. doi: 10.1128/IAI.69.2.816-821.2001.
49. Guzmán-Rodríguez JJ, López-Gómez R, Suárez-Rodríguez LM, et al. Antibacterial activity of defensin PaDef from avocado fruit (*Persea americana* var. *drymifolia*) expressed in endothelial cells against *Escherichia coli* and *Staphylococcus aureus*. *Biomed Res Int.* 2013;2013:986273. doi: 10.1155/2013/986273.
50. Foligne B, Nutten S, Grangette C, Dennin V, et al. Correlation between in vitro and in vivo immunomodulatory properties of lactic acid bacteria. *World J Gastroenterol.* 2007;13(2):236-243. doi: 10.3748/wjg.v13.i2.236.
51. Hamblin MR, Hasan T. Photodynamic therapy: a new antimicrobial approach to infectious disease? *Photochem Photobiol Sci.* 2004;3(5):436-450. doi: 10.1039/b311900a.
52. Sokic-Milutinovic A, Todorovic V, Milosavljevic T, Micev M, Drmdarevic N, Mitrovic O. Gastrin and antral G cells in course of *Helicobacter pylori* eradication: six months follow up study. *World J Gastroenterol.* 2005;11(27):4140-4147. doi: 10.3748/wjg.v11.i27.4140.
53. Kellesarian SV, Malignaggi VR, Al-Kheraif AA, Al-Askar M, Yunker M, Javed F. Effect of antimicrobial photodynamic therapy and laser alone as adjunct to mechanical debridement in the management of halitosis: A systematic review. *Quintessence Int.* 2017;48(7):575-583. doi: 10.3290/j.qi.a38264.
54. Calvino-Fernández M, García-Fresnadillo D, Benito-Martínez S, et al. *Helicobacter pylori* inactivation and virulence gene damage using a supported sensitiser for photodynamic therapy. *Eur J Med Chem.* 2013;68:284-290. doi: 10.1016/j.ejmech.2013.07.023.
55. Denis TGS, Hamblin MR. An introduction to photoantimicrobials: photodynamic therapy as a novel method of microbial pathogen eradication. In: Méndez-Vilas A, ed., *Science against microbial pathogens: communicating current research and technological advances.* Microbiology Series. No:3., Badajoz, Spain: Formatex, 2011: pp. 675-683.
56. Wilder-Smith CH, Wilder-Smith P, Grosjean P, et al. Photo-eradication of *Helicobacter pylori* using 5-aminolevulinic acid: preliminary human studies. *Lasers Surg Med.* 2002;31(1):18-22. doi: 10.1002/lsm.10066.
57. Aguilera-Correa JJ, Esteban J, Vallet-Regí M. Inorganic and Polymeric Nanoparticles for Human Viral and Bacterial Infections Prevention and Treatment. *Nanomaterials (Basel).* 2021;11(1):137. doi: 10.3390/nano11010137.
58. Xiong MH, Bao Y, Yang XZ, Zhu YH, Wang J. Delivery of antibiotics with polymeric particles. *Adv Drug Deliv Rev.* 2014;78:63-76. doi: 10.1016/j.addr.2014.02.002.
59. Matricardi P, Meo CD, Coviello T, Alhaique F. Recent advances and perspectives on coated alginate microspheres for modified drug delivery. *Expert Opin Drug Deliv.* 2008;5(4):417-425. doi: 10.1517/17425247.5.4.417.
60. Malaekheh-Nikouei B, Bazzaz BSF, Mirhadi E, Tajani AS, Khameneh B. The role of nanotechnology in combating biofilm-based antibiotic resistance. *J Drug Deliv Sci Technol* 2020;60:101880. doi:10.1016/j.jddst.2020.101880
61. Mehta DK, Rai SR. Microencapsulation of *Lactobacillus acidophilus* NCDC 291 using emulsion technique and sensory and physico-chemical analysis of the incorporated microcapsules in dairy and non-dairy food product. In: Singhee D, Bhattacharyya K, Tuteja S, Sarkar A. eds., *Reflections.* JD Birla Institute: Kolkota (West Bengal), India, 2017: pp. 51-57.
62. Liang J, Yan H, Puligundla P, Gao X, Zhou Y, Wan X. Applications of chitosan nanoparticles to enhance absorption and bioavailability of tea polyphenols: a review. *Food Hydrocoll* 2017;69:286-292. doi:10.1016/j.foodhyd.2017.01.041
63. Gupta R, Prasad Y. Efficacy of polyvalent bacteriophage P-27/HP to control multidrug resistant *Staphylococcus aureus* associated with human infections. *Curr Microbiol.* 2011;62(1):255-260. doi: 10.1007/s00284-010-9699-x.
64. de Bortoli N, Leonardi G, Ciancia E, et al. *Helicobacter pylori* eradication: a randomized prospective study of triple therapy versus triple therapy plus lactoferrin and probiotics. *Am J Gastroenterol.* 2007;102(5):951-956. doi: 10.1111/j.1572-0241.2007.01085.x.
65. Malfertheiner P, Selgrad M, Bornschein J. *Helicobacter pylori*: clinical management. *Curr Opin Gastroenterol.* 2012;28(6):608-14. doi: 10.1097/MOG.0b013e32835918a7.
66. Dar HA, Zaheer T, Shehroz M, et al. Immunoinformatics-Aided Design and Evaluation of a Potential Multi-Epitope Vaccine against *Klebsiella Pneumoniae*. *Vaccines (Basel).* 2019;7(3):88. doi: 10.3390/vaccines7030088.
67. Chen Y, Hu H, Huang F, et al. Cocktail of isobavachalcone and curcumin enhance eradication of *Staphylococcus aureus* biofilm from orthopedic implants by gentamicin and alleviate inflammatory osteolysis. *Front Microbiol.* 2022;13:958132. doi: 10.3389/fmicb.2022.958132.
68. Abd Eldaim MA, Tousson E, Soliman MM, El Sayed IET, Abdel Aleem AAH, Elsharkawy HN. Grape seed extract ameliorated Ehrlich solid tumor-induced hepatic tissue and DNA damage with reduction of PCNA and P53 protein expression in mice. *Environ Sci Pollut Res Int.* 2021;28(32):44226-44238. doi: 10.1007/s11356-021-13904-8.
69. Muhammad SNH, Yaacob NS, Safuwani NAM, Fauzi AN. Antiglicolytic Activities of *Strobilanthes crispus* Active Fraction and its Bioactive Components on Triple-Negative Breast Cancer Cells In Vitro. *Anticancer Agents Med Chem.* 2022;22(7):1363-1369. doi: 10.2174/1871520621666210427104804.
70. Nzeako BC, Al-Namaani F. The antibacterial activity of honey on *Helicobacter pylori*. *Sultan Qaboos Univ Med J.* 2006 Dec;6(2):71-76.
71. Viertel TM, Ritter K, Horz HP. Viruses versus bacteria—novel approaches to phage therapy as a tool against multidrug-resistant pathogens. *J Antimicrob Chemother.* 2014;69(9):2326-2336. doi: 10.1093/jac/dku173.

72. Casey E, van Sinderen D, Mahony J. In Vitro Characteristics of Phages to Guide 'Real Life' Phage Therapy Suitability. *Viruses*. 2018;10(4):163. doi: 10.3390/v10040163.
73. Uyttebroek S, Chen B, Onsea J. Safety and efficacy of phage therapy in difficult-to-treat infections: a systematic review. *Lancet Infect Dis*. 2022;22(8):e208-e220. doi: 10.1016/S1473-3099(21)00612-5.
74. Sousa C, Ferreira R, Azevedo NF, et al. *Helicobacter pylori* infection: from standard to alternative treatment strategies. *Crit Rev Microbiol*. 2022;48(3):376-396. doi: 10.1080/1040841X.2021.1975643.
75. Galtier M, De Sordi L, Maura D, et al. Bacteriophages to reduce gut carriage of antibiotic resistant uropathogens with low impact on microbiota composition. *Environ Microbiol*. 2016;18(7):2237-2245. doi: 10.1111/1462-2920.13284.
76. Elbehiry A, Marzouk E, Aldubaib M, et al. *Helicobacter pylori* Infection: Current Status and Future Prospects on Diagnostic, Therapeutic and Control Challenges. *Antibiotics (Basel)*. 2023;12(2):191. doi: 10.3390/antibiotics12020191.
77. Oliveira H, Thiagarajan V, Walmagh M, et al. A thermostable *Salmonella* phage endolysin, Lys68, with broad bactericidal properties against gram-negative pathogens in presence of weak acids. *PLoS One*. 2014;9(10):e108376. doi: 10.1371/journal.pone.0108376.
78. Oliveira H, Melo LD, Santos SB, et al. Molecular aspects and comparative genomics of bacteriophage endolysins. *J Virol*. 2013;87(8):4558-4570. doi: 10.1128/JVI.03277-12.
79. Oliveira H, São-José C, Azeredo J. Phage-Derived Peptidoglycan Degrading Enzymes: Challenges and Future Prospects for In Vivo Therapy. *Viruses*. 2018;10(6):292. doi: 10.3390/v10060292.
80. Lukacik P, Barnard TJ, Keller PW, et al. Structural engineering of a phage lysin that targets gram-negative pathogens. *Proc Natl Acad Sci U S A*. 2012;109(25):9857-9862. doi: 10.1073/pnas.1203472109.
81. Lood R, Winer BY, Pelzek AJ, et al. Novel phage lysin capable of killing the multidrug-resistant gram-negative bacterium *Acinetobacter baumannii* in a mouse bacteremia model. *Antimicrob Agents Chemother*. 2015;59(4):1983-1991. doi: 10.1128/AAC.04641-14.