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REVİEW

The Genetics of Parkinson's Disease Parkinson Hastalığı Genetiği

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ABSTRACT

Parkinson's disease (PD) is one of the most common neurodegenerative diseases worldwide. Approximately 15% of PD patients have a family history of the disease in one or more first-degree relatives, and 5-10% of PD cases exhibit a classical Mendelian inheritance pattern. In 1997, the heritable transmission of PD was first documented. Recent studies have found 90 independent genome-wide signals at 78 loci th

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ÖZ

Parkinson hastalığı (PH), dünya çapında en yaygın nörodejenerdif hastalıklardan biridir. PH
hastalarının yaklaşık %15'inde bir veya daha fazla birinci derece akrabada hastalık öyküsü vardır
ve PH vakdarının %5-10'u klasik

Anahtar Kelimeler: Parkinson Hastalığı, Kalıtım, Otozomal Dominant, Otozomal Resesif

Introduction

Parkinson's disease (PD) is the second most common inheritance pattern (6). The first evidence of heritable neurodegenerative disorder after oxidative and metabolic stress, excitotoxicity, and associated with PD (8). inflammation (4). Clinical findings of PD can be divided into two categories: motor symptoms; the cardinal signs of PD, and non-motor symptoms which can even be present in the pre-clinical phase. Motor symptoms can be characterized as tremors, rigidity, bradykinesia/ akinesia, and postural instability. Non-motor symptoms may occur years before the diagnosis of the disease and include constipation, hyposmia, sleep-wake **Monogenic Forms of PD** cycle disorders, apathy and depression (5).

disease (1). Its prevalence is estimated at over six in the alpha-synuclein (SNCA) gene responsible for a million worldwide and is expected to double by 2040 monogenic form of PD was defined (7). Shortly after this (2). The pathogenesis of PD is based on the loss of discovery, many studies identified numerous genes with dopaminergic neurons in the substantia nigra pars autosomal recessive (AR) and autosomal dominant (AD) compacta (SNpc) and intraneuronal aggregation inheritances. Recently, Nalls et al. have conducted the of misfolded alpha-synuclein called Lewy bodies (3). largest study to date for the genetics of PD, analyzing 7.8 The underlying mechanisms leading to the loss of million single nucleotide polymorphisms (SNPs) in 37.688 dopaminergic neurons in the SNpc can be listed as PD cases, 18.618 United Kingdom Biobank (UKB) "proxy pathological alpha-synuclein aggregation, disruption cases (individuals who do not have Parkinson's disease of intracellular protein degradation systems such as but have a first degree relative that has)" and 1.4 the endo-lysosomal autophagy and the ubiquitin-million controls. They found 90 independent genomeproteasome pathways, mitochondrial dysfunction, wide significant signals at 78 loci that are thought to be Alzheimer's transmission of PD was reported in 1997. Rare mutations

> The acquisition of new genetic technologies is rapidly illuminating both the pathogenesis and clinical and genetic diversity of PD. Therefore, it is very important to understand the genetic factors behind PD. In this review, we summarized the genetic etiologies associated with PD.

The etiology of the disease is obscure, and most PD were detected through linkage analysis of affected cases are sporadic. Approximately 15% of patients families. On the other hand, few variants were also have a family history of PD in one or more first-degree revealed by genome-wide association studies (GWAS). relatives and 5–10% of PD show a classical Mendelian The loci associated with PD phenotypes were named Historically, the monogenic forms of PD predominantly

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the 'PARK' loci and are indicated by the number representing the chronological order of discovery. For example, the PRKN gene is also referred to as PARK2. However, multiple PARK loci may refer to the same gene, and some PARK loci do not appear to cause disease. Therefore, the current recommendation is to use gene names instead of PARK loci (9). Monogenic forms of PD are summarized in Table 1.

Autosomal dominant forms of PD

These are the genes that are clearly identified as risk factors for PD: SCNA (Synuclein, Alpha), LRRK2 (Leucine-Rich Repeat Kinase 2), and VPS35 (Vacuolar Protein Sequence 35 Retromer Complex Component). More recently, the association with LRP10 (Low Density Lipoprotein Receptor-Related Protein 10) and EIF4G1 (Eukaryotic Translation Initiation Factor 4G) are being studied. Moreover, some pathogenic variations of GBA (Beta-glucosidase) gene, which cause Gaucher's disease in biallelic state, can lead to PD with variable penetrance in monoallelic state (10).

SNCA

The SNCA (PARK1, PARK4; OMIM: 163890) gene is responsible for producing the alpha-synuclein protein. The exact function of alpha-synuclein (α-synuclein) is yet to be determined. However, its role is thought to regulate neurotransmitter release, synaptic vesicles and neuronal differentiation (11).

The SNCA gene was the first gene determined to be linked to PD. In 1997, Polymeropoulos et al. identified a missense variant that substituted the amino-acids alanine with threonine at position 53 (p.A53T) of the SNCA gene in an Italian family (7). The same variant was also found to be associated with the PD in three Greek kindreds. Afterwards, several missense SNCA variants were identified, including p.A30P, p.E46K, p.H50Q and p.G51D (12–15). Missense mutations of SNCA are reported to have a direct impact on a-synuclein conformation and function. Patients with pathogenic missense variations in this gene often tend to develop PD before 50 years of age, and the symptoms worsen rapidly; but respond well to levodopa (L-DOPA). The phenotype also differs between different missense variants of SNCA. For instance, patients with p.G51D show extremely rapid disease progression causing some patients to die within ten years of onset. Atypical findings such as pyramidal signs, cognitive decline, psychiatric disturbances, myoclonus, and seizures can be observed too (15).

Apart from nucleotide substitutions that disrupt the protein function, altered dosage of the protein can also cause protein misexpression. Several studies showed that duplication or triplication of SNCA can lead to an increase in α-synuclein expression, which increases the risk of developing PD (16,17). These studies also reported that patients with duplications or triplications of SNCA tend to experience a faster progression (16,17).

LRRK2

The LRRK2 (PARK8; OMIM: 609007) gene encodes the LRRK2 protein (18). LRKK2, also known as dardarin, is a large protein that is involved in a wide variety of cellular functions including autophagy, cytoskeletal dynamics, intracellular membrane trafficking, synaptic vesicle turnover and inflammation (19,20). Dardarin dysfunction leads to the disruption of α-synuclein degradation in cellular clearance pathways and thus causes the accumulation of misfolded α-synucleins (21).

In 2004, it was discovered that the LRRK2 gene is linked to the development of PD. A Japanese family with AD parkinsonism was found to have the c.6055G>A variant (p.G2019S). This variant is the most prevalent pathogenic mutation worldwide that causes PD (18). Its incidence is particularly high in the Ashkenazi Jewish (26%) and North African Berber (41%) populations (22– 24). Other diverse variants of the LRKK2 gene were explored, yet only eight proved to be pathogenic (N1437H, R1441 G/H/C, Y1699C, G2019S, S1761R, G2385R, R1628P and I2020T) (25). These variants result in advanced age-onset PD clinically resembling a sporadic PD (26).

VPS35

The VPS35 (PARK17; OMIM: 601501) gene enables the production of a component of the multimeric retromeric complex. The complex is one of the main conductors in endosomal sorting and trafficking (27,28).

A missense mutation (p.D620N) in the VPS35 gene was detected in Swiss and Australian families in 2011 (29). However, in a study conducted by Nuytemans et al. (30), 213 patients with PD were analyzed through whole-exome sequencing and the results showed no evidence indicating that genetic variations of VPS35 significantly impacted the development of PD. Nevertheless, considering rarity (<%1 of familial PD cases) and lack of functional evidence of VPS35 variants except p.D620N, further studies with larger samples are needed for a clearer deduction (31).

GBA

The GBA (OMIM: 606463) gene encodes for a lysosomal enzyme, beta-glucosidase. Betaglucosidase catalyzes the breakdown of the glycolipid glucosylceramide into ceramide and glucose (32). Biallelic pathogenic GBA variants can lead to Gaucher disease, a lysosomal storage disorder caused by reduced glucocerebrosidase activity (33). Whereas heterozygous carriers are in an increased risk of developing PD (10).

Various case-control studies showed that signs of PD such as tremor and bradykinesia can be exhibited in Gaucher patients; whereupon PD symptoms were included in the spectrum of the disease (34,35).

The two most prevalent variants are p.N370S and p.L444P globally; p.N370S heterozygosity raises the risk of PD by four times whereas p.L444P increases it by twelve times (36). The clinical findings of GBA-

associated PD are similar to idiopathic PD. However, studies showed earlier age of onset, higher dementia rates, and faster worsening of the symptoms in PD patients with heterozygous pathogenic GBA variants (37–39).

LRP10

The LRP10 (OMIM: 609921) gene enables formation of the LRP10 protein, which contains a class of surface receptors that play an important role in the trafficking and processing of amyloid precursor protein (40). First, a LRP10 missense mutation, c.1807G>A (p.G603R) was detected in an Italian family with hereditary PD (41). Subsequently, Kia et al. investigated the LRP10 gene in a study involving 2835 PD patients and 5343 controls (42). However, they found no significant difference in LRP10 gene variants between controls and PD patients. Currently, the pathogenic role of LRP10 mutations in PD is still unclear.

EIF4G1

The EIF4G1 (PARK18; OMIM: 600495) gene responsible for a member of eukaryotic translation initiation factors that play important roles for the ribosome/ mRNA-bridge (43). Initially, the R1205H variant in the EIF4G1 gene were identified in a French family (44). In a cohort study of 4,708 PD patients screened for R1205H, nine patients from seven families from the USA, Canada, Ireland, Italy, and Tunisia were found to carry this variant (45). Subsequently, Huttenlocher et al. (46) studied a cohort of 2,146 European PD patients to evaluate the relationship between EIF4G1 mutations and PD. They identified the R1205H mutation in only one patient. Moreover, a recent study found no association between EIF4G1 and PD (47).

Autosomal recessive forms of PD

There are specific genes that are strongly linked to the autosomal recessive (AR) types of PD, including PRKN (RBR E3 ubiquitin protein ligase), PINK1 (PTEN-derived putative kinase 1) and DJ-1 (Oncogene DJ1). These three genes interplay in a mitochondria proteolysis pathway. Patients with these variants have similar clinical manifestations to sporadic PD, although the age of onset is earlier. ATP13A2 (ATPase 13A2), PLA2G6 (Phospholipase A2, Group VI), FBXO7 (F-Box Only Protein 7), DNAJC6 (DNAJ/HSP40 Homolog, Subfamily C, Member 6), SYNJ1 (Synaptojanin 1) appear as infrequent and complex forms of autosomal recessive PD. Parkinsonism is the primary clinical feature of these patients, but they may also present with atypical manifestations such as supranuclear gaze palsy, mental retardation, or seizures.

PRKN

The PRKN (Parkin; PARK2; OMIM: 602544) gene, is one of the largest genes in humans and is responsible for producing a protein called parkin (48). The parkin protein is involved in the process of ubiquitination, a form of post-translational modification, and is responsible for the breakdown of damaged or excess proteins (49).

Parkin-associated PD includes marked degeneration of dopaminergic neurons in the main pathology of the substantia nigra pars compacta. Lewy bodies, the pathognomonic finding for idiopathic PD, may be absent in these cases (50). The characteristics of PRKN related PD share a remarkable similarity with idiopathic PD signs such as tremors, rigidity and bradykinesia (51). However, the disease usually has an earlier onset; even childhood-onset cases have been reported (51). Additionally, biallelic PRKN mutations are the most common genetic variants in juvenile PD (52). Another study revealed that PRKN mutations occur in 77% of familial cases with an age of onset <30 and in 10-20% of patients with early-onset PD (53).

Although biallelic PRKN variants are an established risk of developing PD, there is much debate on the potential influence of heterozygous PRKN variants on PD. Several small studies have claimed that heterozygous PRKN variants increase the risk of PD (54,55). However, this could not be confirmed in other studies and meta-analyses (56–58). More recently, a study involving 2809 PD patients and 3629 healthy controls has been conducted to investigate the potential link between PD and heterozygous PRKN variants, including single nucleotide variants and copy number variations (CNVs) (59). The findings have indicated that there is no connection between heterozygous PRKN variants and PD (59).

PINK1

The PINK1 (PARK6; OMIM: 608309) gene encodes a mitochondrial serine/threonine kinase (60). Twothirds of the mutations reported in PINK1 are loss-offunction mutations that affect serine/threonine kinase activity. These findings highlight the importance of mitochondrial proteolysis pathway in the pathogenesis of PD (61–63). PINK1-related PD findings clinically overlap with sporadic PD and present at early-onset. Moreover, in these patients, non-motor findings are observed more commonly (10).

DJ-1

The DJ-1 (PARK7; OMIM: 602533) gene encodes a 189 amino acid-long protein, named DJ-1, that functions in regulation of transcription, oxidative stress, and mitochondrial metabolism (64). The discovery of DJ-1 as a causative gene for PD was brought about by its occurrence in two consanguineous families of Dutch and Italian origin (65). Single nucleotide and structural variations such as Glu163Lys, Leu166Pro, and g.168-185dup have been reported (66,67). DJ-1 gene mutations occur in approximately 1-2% of early-onset PD (68). PD patients with DJ-1 mutations show earlyonset Parkinson's symptoms followed by psychiatric disturbances such as psychotic disorder, anxiety, and cognitive decline, and generally respond well to L-DOPA treatment (69–71).

DNAJC6

The DNAJC6 (PARK19; OMIM: 608375) gene encodes auxilin, a neuronal protein that regulates molecular chaperone activity by stimulating ATPase activity

(72). First, a homozygous splice mutation (c.801-2A>G) of the DNAJC6 gene is identified in two Palestinian brothers. The symptoms were rapidly progressive and unresponsive to treatment (73). The DNAJC6 gene variants that cause truncation of the protein lead to juvenile atypical parkinsonism, while non-truncating variants cause early-onset parkinsonism (74).

ATP13A2

The ATP13A2 (PARK9; OMIM: 610513) gene encodes ATP13A2 protein, acts as a critical regulator of lysosomal functions (75). A study identified upregulated ATP1A32 expression in surviving dopaminergic neurons of patients with idiopathic PD, suggesting a neuroprotective role of this protein (76). The ATP13A2 protein deficiency leads to Kufor-Rakeb syndrome (KRS; OMIM: 606693), characterized by juvenile, levodoparesponsive Parkinsonism, pyramidal manifestations, and dementia. The presence of iron accumulation in the basal ganglia in some patients with KRS suggests that it can be considered among neurodegenerative syndromes with brain iron accumulation (NBIAs) (77).

FBXO7

Mutations in the FBXO7 (PARK15; OMIM: 605648) gene, responsible for producing F-box protein 7, can cause a rare autosomal recessive Parkinsonian-pyramidal syndrome (78). The syndrome shows symptoms of early-onset parkinsonism, along with pyramidal system involvement like psychomotor retardation, eyelid apraxia, and chorea (79).

PLA2G6

The PLA2G6 (PARK14; OMIM: 603604) gene encodes a phospholipase A2 enzyme subgroup, and has a key role in inflammation, cell proliferation, apoptosis, and remodeling of membrane phospholipids (80). PLA2G6 mutations are highly heterogeneous and result in a complex group of neurodegenerative diseases. The clinical picture of PLA2G6-related neurodegeneration is classified in three overlapping phenotypes, one of which is 'PLA2G6-related dystonia-parkinsonism' (81). PLA2G6-related dystonia-parkinsonism begins in late adolescence and presents with early-onset PD, gait disturbance, and neuropsychiatric symptoms (81).

SYNJ1

The SYNJ1 (PARK20; OMIM: 604297) gene produces a protein called Synaptojanin 1, which plays a crucial role in regulating vesicle endocytosis and recycling (82,83). In several studies, biallelic mutations of SYNJ1 were associated with two distinct phenotypes: earlyonset PD and a severe neurodegenerative disorder with epilepsy and tauopathies (84–86). A recent study has revealed that homozygous missense mutations such as p.R839C and p.Y832C result in typical PD or early-onset atypical parkinsonism (87). On the other hand, it is suggested that p.Y888C homozygous missense mutations could lead to severe progressive neurodegeneration. All of these findings suggest wide clinical and genetic heterogeneity for SYNJ1 variations.

X-linked forms of PD

TAF1

TAF1 (TATA-binding protein-associated factor-1) (OMIM: 313650) is responsible for the only known X-linked PD: X-linked torsion dystonia-parkinsonism syndrome. TAF1 encodes an essential component of Transcription factor II D which is critical for RNA polymerase II–mediated gene transcription (88). SVA (short interspersed nuclear element, variable number of tandem repeats, and Alu composite) retrotransposon insertion in intron 32 of the TAF1 is a founder variant in Philippines and the only known pathogenic variation to date that causes X-linked torsion dystonia-parkinsonism syndrome (89,90).

Multifactorial Inheritance in PD

Genetic and environmental factors play a role in the development of multifactorial diseases. Most of the adult-onset chronic diseases show multifactorial inheritance pattern, which is usually characterized by familial aggregation of disease (91). Likewise, apart from the PD cases with highly penetrant genetic variants as discussed above, majority of late-onset sporadic PD cases show no sign of Mendelian inheritance. Moreover, the variants of some genes (e.g., SNCA and GBA) show low penetrance indicating additional contribution of other genetic and environmental factors (10).

To reveal genetic factors taking part in complex diseases with multifactorial features like PD, GWAS are powerful assets. There are several GWAS pointing multiple loci with potential significance for the association with PD (92). For instance, GWAS analysis done by Nalls et al. associated seven loci containing LRRK2, GBA, CATSPER3, LAMB2, LOC442028, NFKB2, and SCARB2 genes with PD (8). Additionally, NSF gene which encodes N-ethylmaleimide sensitive fusion protein that have a role in synaptic neurotransmission, also associated with PD in another study (93). However, all variants are not associated positively with PD. An example is MAPT gene H2 haplotype, which was associated with later age at onset (93). Moreover, CRHR1 gene which encodes corticotropin releasing hormone receptor 1, has also been shown to be associated with a reduced risk of PD (94). Despite all these findings, even GWAS cannot reveal a genetic component in majority of PD patients; the broadest study to date has explained only 16%−36% of PD heritability (8).

Where some GWAS are underpowered to find an associated locus, polygenic risk scores (PRS) which utilize multiple loci in the genome including common polymorphisms, can be helpful to reveal the complex relationship between genotype and phenotype of PD cases. In PRS studies, a polygenic score, and a threshold of liability for disease are calculated by analyzing a combination of multiple common genetic variants between genomic datasets of disease and control groups. PRS studies showed to have a potential to predict the liability to some diseases with multifactorial

inheritance such as schizophrenia and bipolar disorder (95). Escott-Price et al. designed a polygenic risk score for PD and found significant correlation between higher risk score and PD liability, especially in cases with early age at onset (96). Moreover, Searles Nielsen et al. found an association between age at onset and SNPs in CYP2J2, GSTM5 and SLC11A2 genes (97). These findings are promising for a future PRS that will include a broad spectrum of ethnicities and PD subtypes.

Although both GWAS and PRS studies have shown important results for comprehension of multifactorial nature of PD, it is still preliminary to make a general deduction in this field. Therefore, more populationbased studies are necessary to fully illuminate this aspect of PD.

Genetic Testing for PD

Genetic screening is recommended for Parkinson's patients with one or more; early onset of the disease (age at onset <50 years old), positive family history suggesting autosomal dominant or autosomal recessive inheritance, and high-risk ethnicities such as Ashkenazi Jewish or North African Berber (98). The choice of genetic tests should be determined by the patient's unique circumstances. For example, the SNCA, GBA, PRKN, PINK1 and DJ1 genes may be considered in patients with age at onset <50. Disease specific genetic panels containing multiple genes rather than single gene screening are more convenient due to locus heterogeneity, lower cost, and increased efficiency. It is important to note that PD panels can vary greatly between laboratories in terms of the genes they contain.

More comprehensive genetic testing, such as Whole Exome Sequencing (WES) and Whole Genome Sequencing (WGS), can also be utilized. However, complexity of bioinformatic pipelines, issues related to variants of unknown significance and reporting of secondary findings may further complicate diagnosis and management of patients. Therefore, multidisciplinary approach is essential for the proper diagnosis, genetic counselling and management of these patients.

Conclusion

PD is a disabling neurodegenerative disorder with increasing prevalence worldwide. In the last three decades, significant strides have been made in understanding the genetics of PD. The identification of genes related to PD and their functions has uncovered novel biological pathways that play a role in the development of PD. These new pathways not only helped us better understand the disease but also shed light on potential treatment options. On the other hand, advancements in genetic information have enabled the optimization of existing treatment options specific to each patient.

Declaration of Conflicting Interests

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References

1.Kalia L V, Lang AE. Parkinson's disease. The Lancet. 2015 Aug;386(9996):896–912.

2.Rocca WA. The burden of Parkinson's disease: a worldwide perspective. Lancet Neurol. 2018 Nov;17(11):928–9.

3.Balestrino R, Schapira AHV. Parkinson disease. Eur J Neurol. 2020 Jan 27;27(1):27–42.

4.Yalçın Çakmaklı G EB. Parkinson Hastalığında Patogenez. Türkiye Klinikleri. 2021;5–10.

5.Jankovic J. Parkinson's disease: clinical features and diagnosis. J Neurol Neurosurg Psychiatry. 2008 Apr 1;79(4):368–76.

6.Lesage S, Brice A. Parkinson's disease: from monogenic forms to genetic susceptibility factors. Hum Mol Genet. 2009 Apr 15;18(R1):R48–59.

7.Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, et al. Mutation in the α-Synuclein Gene Identified in Families with Parkinson's Disease. Science (1979). 1997 Jun 27;276(5321):2045–7.

8. Nalls MA, Blauwendraat C, Valleraa CL, Heilbron K, Bandres-Ciaa S, Chang D, et al. Identification of novel risk loci, causal insights, and heritable risk for Parkinson's disease: a meta-analysis of genome-wide association studies. Lancet Neurol. 2019 Dec;18(12):1091–102.

9.Marras C, Lang A, van de Warrenburg BP, Sue CM, Tabrizi SJ, Bertram L, et al. Nomenclature of genetic movement disorders: Recommendations of the international Parkinson and movement disorder society task force. Movement Disorders. 2016 Apr;31(4):436– 57.

10.Cook Shukla L SJFJ et al. Parkinson Disease Overview.2004 May 25 [Updated 2019 Jul 25]. In: In: Adam MP MGPR et al. , editors. G [Internet]., editor. GeneReviews® [Internet] Seattle (WA): University of Washington, Seattle; 1993-2023 [Internet]. [cited 2023 Jul 14]. Available from: https://www.ncbi.nlm.nih.gov/books/NBK1223/

11.Bonini NM, Giasson BI. Snaring the Function of α-Synuclein. Cell. 2005 Nov;123(3):359–61.

12.Lesage S, Anheim M, Letournel F, Bousset L, Honoré A, Rozas N, et al. G51D α-synuclein mutation causes a novel Parkinsonian-pyramidal syndrome. Ann Neurol. 2013 Apr;73(4):459–71.

13.Zarranz JJ, Alegre J, Gómez-Esteban JC, Lezcano E, Ros R, Ampuero I, et al. The new mutation, E46K, of α-synuclein causes parkinson and Lewy body dementia. Ann Neurol. 2004 Feb;55(2):164–73.

14.Krüger R, Kuhn W, Müller T, Woitalla D, Graeber M, Kösel S, et al. AlaSOPro mutation in the gene encoding α-synuclein in Parkinson's disease. Nat Genet. 1998 Feb;18(2):106–8.

15.Kiely AP, Asi YT, Kara E, Limousin P, Ling H, Lewis P, et al. α-Synucleinopathy associated with G51D SNCA mutation: a link between Parkinson's disease and multiple system atrophy? Acta Neuropathol. 2013 May 12;125(5):753–69.

16.Ross OA, Braithwaite AT, Skipper LM, Kachergus J, Hulihan MM, Middleton FA, et al. Genomic investigation of α-synuclein multiplication and parkinsonism. Ann Neurol. 2008 Jun;63(6):743–50.

17.Singleton AB, Farrer M, Johnson J, Singleton A, Hague S, Kachergus J, et al. α-Synuclein Locus Triplication Causes Parkinson's Disease. Science (1979). 2003 Oct 31;302(5646):841–841.

18.Funayama M, Hasegawa K, Kowa H, Saito M, Tsuji S, Obata F. A new locus for Parkinson's disease (PARK8) maps to chromosome 12p11.2-q13.1. Ann Neurol. 2002 Mar;51(3):296–301.

19.Wallings R, Manzoni C, Bandopadhyay R. Cellular processes associated with LRRK2 function and dysfunction. FEBS Journal. 2015 Aug;282(15):2806–26.

20.Cookson MR. LRRK2 Pathways Leading to Neurodegeneration. Curr Neurol Neurosci Rep. 2015 Jul 26;15(7):42.

21.Lee HJ, Khoshaghideh F, Patel S, Lee SJ. Clearance of α-Synuclein Oligomeric Intermediates via the Lysosomal Degradation Pathway. The Journal of Neuroscience. 2004 Feb 25;24(8):1888–96.

22.Lee AJ, Wang Y, Alcalay RN, Mejia-Santana H, Saunders-Pullman R, Bressman S, et al. Penetrance estimate of LRRK2 p.G2019S mutation in individuals of non-Ashkenazi Jewish ancestry. Movement Disorders. 2017 Oct;32(10):1432–8.

23.Marder K, Wang Y, Alcalay RN, Mejia-Santana H, Tang MX, Lee A, et al. Age-specific penetrance of LRRK2 G2019S in the Michael J. Fox Ashkenazi Jewish LRRK2 Consortium. Neurology. 2015 Jul 7;85(1):89–95.

24.Healy DG, Falchi M, O'Sullivan SS, Bonifati V, Durr A, Bressman S, et al. Phenotype, genotype, and worldwide genetic penetrance of LRRK2-associated Parkinson's disease: a case-control study. Lancet Neurol. 2008 Jul;7(7):583–90.

25.Simpson C, Vinikoor-Imler L, Nassan FL, Shirvan J, Lally C, Dam T, et al. Prevalence of ten LRRK2 variants in Parkinson's disease: A comprehensive review. Parkinsonism Relat Disord. 2022 May;98:103– 13.

26.Trinh J, Zeldenrust FMJ, Huang J, Kasten M, Schaake S, Petkovic S, et al. Genotype-phenotype relations for the Parkinson's disease genes SNCA, LRRK2, VPS35: MDSGene systematic review. Movement Disorders. 2018 Dec;33(12):1857–70.

27.Hanss Z, Larsen SB, Antony P, Mencke P, Massart F, Jarazo J, et al. Mitochondrial and Clearance Impairment in p. <scp>D620N VPS35</ scp> Patient□Derived Neurons. Movement Disorders. 2021 Mar 3;36(3):704–15.

28.Bono K, Hara-Miyauchi C, Sumi S, Oka H, Iguchi Y, Okano HJ. Endosomal dysfunction in iPSC-derived neural cells from Parkinson's disease patients with VPS35 D620N. Mol Brain. 2020 Dec 8;13(1):137.

29.Vilariño-Güell C, Wider C, Ross OA, Dachsel JC, Kachergus JM, Lincoln SJ, et al. VPS35 Mutations in Parkinson Disease. The American Journal of Human Genetics. 2011 Jul;89(1):162–7.

30.Nuytemans K, Bademci G, Inchausti V, Dressen A, Kinnamon DD, Mehta A, et al. Whole exome sequencing of rare variants in EIF4G1 and VPS35 in Parkinson disease. Neurology. 2013 Mar 12;80(11):982–9.

31.Dulski J, Uitti RJ, Ross OA, Wszolek ZK. Genetic architecture of Parkinson's disease subtypes – Review of the literature. Front Aging Neurosci. 2022 Oct 20;14.

32.National Center for Biotechnology Information (NCBI) United States National Library of Medicine (NLM). GBA1 glucosylceramidase beta 1 [Homo sapiens (human) [Internet]. 2023 [cited 2023 May 23]. Available from: https://www.ncbi.nlm.nih.gov/gene/2629

33.Brady RO, Kanfer JN, Bradley RM, Shapiro D. Demonstration of a deficiency of glucocerebroside-cleaving enzyme in Gaucher's disease. Journal of Clinical Investigation. 1966 Jul 1;45(7):1112–5.

34.Goker-Alpan O. Parkinsonism among Gaucher disease carriers. J Med Genet. 2004 Dec 1;41(12):937–40.

35.Neudorfer O, Giladi N, Elstein D, Abrahamov A, Turezkite T, Aghai E, et al. Occurrence of Parkinson's syndrome in type 1 Gaucher disease. QJM. 1996 Sep 1;89(9):691–4.

36.Gan-Or Z, Amshalom I, Kilarski LL, Bar-Shira A, Gana-Weisz M, Mirelman A, et al. Differential effects of severe vs mild GBA mutations on Parkinson disease. Neurology. 2015 Mar 3;84(9):880–7.

37.Mata IF, Leverenz JB, Weintraub D, Trojanowski JQ, Chen-Plotkin A, Van Deerlin VM, et al. GBA Variants are associated with a distinct pattern of cognitive deficits in Parkinson's disease. Movement Disorders. 2016 Jan;31(1):95–102.

38.Lopez G, Kim J, Wiggs E, Cintron D, Groden C, Tayebi N, et al. Clinical course and prognosis in patients with Gaucher disease and parkinsonism. Neurol Genet. 2016 Apr 4;2(2):e57.

39.Winder-Rhodes SE, Evans JR, Ban M, Mason SL, Williams-Gray CH,

Foltynie T, et al. Glucocerebrosidase mutations influence the natural history of Parkinson's disease in a community-based incident cohort. Brain. 2013 Feb;136(2):392–9.

40.Brodeur J, Thériault C, Lessard-Beaudoin M, Marcil A, Dahan S, Lavoie C. LDLR-related protein 10 (LRP10) regulates amyloid precursor protein (APP) trafficking and processing: evidence for a role in Alzheimer's disease. Mol Neurodegener. 2012 Dec 26;7(1):31.

41.Quadri M, Mandemakers W, Grochowska MM, Masius R, Geut H, Fabrizio E, et al. LRP10 genetic variants in familial Parkinson's disease and dementia with Lewy bodies: a genome-wide linkage and sequencing study. Lancet Neurol. 2018 Jul;17(7):597–608.

42.Pihlstrøm L, Schottlaender L, Chelban V, Houlden H, Al-Sarraj S, Arzberger T, et al. LRP10 in α-synucleinopathies. Lancet Neurol. 2018 Dec;17(12):1033–4.

43.National Center for Biotechnology Information (NCBI) United States National Library of Medicine (NLM). Eif4g1 eukaryotic translation initiation factor 4, gamma 1 [Mus musculus (house mouse)].

44.Chartier-Harlin MC, Dachsel JC, Vilariño-Güell C, Lincoln SJ, Leprêtre F, Hulihan MM, et al. Translation Initiator EIF4G1 Mutations in Familial Parkinson Disease. The American Journal of Human Genetics. 2011 Sep;89(3):398–406.

45.Chartier-Harlin MC, Dachsel J, Hulihan M, Kachergus J, Lepretre F, Le Rhun E, et al. P2.206 EIF4G1 mutations in familial parkinsonism. Parkinsonism Relat Disord. 2009 Dec;15:S145–6.

46.Huttenlocher J, Krüger R, Capetian P, Lohmann K, Brockmann K, Csoti I, et al. EIF4G1 is neither a strong nor a common risk factor for Parkinson's disease: evidence from large European cohorts: Table1. J Med Genet. 2015 Jan;52(1):37–41.

47.Saini P, Rudakou U, Yu E, Ruskey JA, Asayesh F, Laurent SB, et al. Association study of DNAJC13, UCHL1, HTRA2, GIGYF2, and EIF4G1 with Parkinson's disease. Neurobiol Aging. 2021 Apr;100:119.e7-119. e13.

48.Yoshii SR, Kishi C, Ishihara N, Mizushima N. Parkin Mediates Proteasome-dependent Protein Degradation and Rupture of the Outer Mitochondrial Membrane. Journal of Biological Chemistry. 2011 Jun;286(22):19630–40.

49.Seirafi M, Kozlov G, Gehring K. Parkin structure and function. FEBS J. 2015 Jun 16;282(11):2076–88.

50.Ishikawa A, Takahashi H. Clinical and neuropathological aspects of autosomal recessive juvenile parkinsonism. J Neurol. 1998 Sep 10;245(S3):P4–9.

51.Lücking CB, Dürr A, Bonifati V, Vaughan J, De Michele G, Gasser T, et al. Association between Early-Onset Parkinson's Disease and Mutations in the Parkin Gene. New England Journal of Medicine. 2000 May 25;342(21):1560–7.

52.Klein C, Lohmann-Hedrich K. Impact of recent genetic findings in Parkinson's disease. Curr Opin Neurol. 2007 Aug;20(4):453–64.

53.Cazeneuve C, Sân C, Ibrahim SA, Mukhtar MM, Kheir MM, LeGuern E, et al. A new complex homozygous large rearrangement of the PINK1 gene in a Sudanese family with early onset Parkinson's disease. Neurogenetics. 2009 Jul 12;10(3):265–70.

54.Lesage S, Lohmann E, Tison F, Durif F, Durr A, Brice A. Rare heterozygous parkin variants in French early-onset Parkinson disease patients and controls. J Med Genet. 2007 Oct 26;45(1):43–6.

55.Oliveira SA, Scott WK, Martin ER, Nance MA, Watts RL, Hubble JP, et al. Parkin mutations and susceptibility alleles in late-onset Parkinson's disease. Ann Neurol. 2003 May;53(5):624–9.

56.MacArthur J, Bowler E, Cerezo M, Gil L, Hall P, Hastings E, et al. The new NHGRI-EBI Catalog of published genome-wide association studies (GWAS Catalog). Nucleic Acids Res. 2017 Jan 4;45(D1):D896–901.

57.Lücking CB, Dürr A, Bonifati V, Vaughan J, De Michele G, Gasser T, et al. Association between Early-Onset Parkinson's Disease and Mutations in the Parkin Gene. New England Journal of Medicine. 2000 May 25;342(21):1560–7.

58.Kay DM, Moran D, Moses L, Poorkaj P, Zabetian CP, Nutt J, et al. Heterozygous parkin point mutations are as common in control subjects as in Parkinson's patients. Ann Neurol. 2007 Jan;61(1):47–54.

59.Yu E, Rudakou U, Krohn L, Mufti K, Ruskey JA, Asayesh F, et al. Analysis of Heterozygous PRKN Variants and Copy□Number Variations in Parkinson's Disease. Movement Disorders. 2021 Jan 24;36(1):178–87.

60.Poole AC, Thomas RE, Andrews LA, McBride HM, Whitworth AJ, Pallanck LJ. The PINK1/Parkin pathway regulates mitochondrial morphology. Proceedings of the National Academy of Sciences. 2008 Feb 5;105(5):1638–43.

61.Cazeneuve C, Sân C, Ibrahim SA, Mukhtar MM, Kheir MM, LeGuern E, et al. A new complex homozygous large rearrangement of the PINK1 gene in a Sudanese family with early onset Parkinson's disease. Neurogenetics. 2009 Jul 12;10(3):265–70.

62.Camargos ST, Dornas LO, Momeni P, Lees A, Hardy J, Singleton A, et al. Familial Parkinsonism and early onset Parkinson's disease in a Brazilian movement disorders clinic: Phenotypic characterization and frequency of SNCA , PRKN , PINK1 , and LRRK2 mutations. Movement Disorders. 2009 Apr 15;24(5):662–6.

63.Klein C, Westenberger A. Genetics of Parkinson's Disease. Cold Spring Harb Perspect Med. 2012 Jan 1;2(1):a008888–a008888.

64.Junn E, Taniguchi H, Jeong BS, Zhao X, Ichijo H, Mouradian MM. Interaction of DJ-1 with Daxx inhibits apoptosis signal-regulating kinase 1 activity and cell death. Proceedings of the National Academy of Sciences. 2005 Jul 5;102(27):9691–6.

65.Bonifati V, Rizzu P, van Baren MJ, Schaap O, Breedveld GJ, Krieger E, et al. Mutations in the DJ-1 Gene Associated with Autosomal Recessive Early-Onset Parkinsonism. Science (1979). 2003 Jan 10;299(5604):256–9.

66.Bonifati V, Rizzu P, van Baren MJ, Schaap O, Breedveld GJ, Krieger E, et al. Mutations in the DJ-1 Gene Associated with Autosomal Recessive Early-Onset Parkinsonism. Science (1979). 2003 Jan 10;299(5604):256–9.

67.Annesi G, Savettieri G, Pugliese P, D'Amelio M, Tarantino P, Ragonese P, et al. DJ-1 mutations and parkinsonism-dementia-amyotrophic lateral sclerosis complex. Ann Neurol. 2005 Nov;58(5):803–7.

68.Pankratz N, Pauciulo MW, Elsaesser VE, Marek DK, Halter CA, Wojcieszek J, et al. Mutations in DJ-1 are rare in familial Parkinson disease. Neurosci Lett. 2006 Nov;408(3):209–13.

69.Annesi G, Savettieri G, Pugliese P, D'Amelio M, Tarantino P, Ragonese P, et al. DJ-1 mutations and parkinsonism-dementia-amyotrophic lateral sclerosis complex. Ann Neurol. 2005 Nov;58(5):803–7.

70.Abou-Sleiman PM, Healy DG, Quinn N, Lees AJ, Wood NW. The role of pathogenic DJ-1 mutations in Parkinson's disease. Ann Neurol. 2003 Sep;54(3):283–6.

71.Bonifati V, Rizzu P, van Baren MJ, Schaap O, Breedveld GJ, Krieger E, et al. Mutations in the DJ-1 Gene Associated with Autosomal Recessive Early-Onset Parkinsonism. Science (1979). 2003 Jan 10;299(5604):256–9.

72.Yim YI, Sun T, Wu LG, Raimondi A, De Camilli P, Eisenberg E, et al. Endocytosis and clathrin-uncoating defects at synapses of auxilin knockout mice. Proceedings of the National Academy of Sciences. 2010 Mar 2;107(9):4412–7.

73.Elsayed LEO, Drouet V, Usenko T, Mohammed IN, Hamed AAA, Elseed MA, et al. A Novel Nonsense Mutation in DNAJC6 Expands the Phenotype of Autosomal-Recessive Juvenile-Onset Parkinson's Disease. Ann Neurol. 2016 Feb;79(2):335–7.

74.Olgiati S, Quadri M, Fang M, Rood JPMA, Saute JA, Chien HF, et al. DNAJC6 Mutations Associated With Early-Onset Parkinson's Disease. Ann Neurol. 2016 Feb;79(2):244–56.

75.Dehay B, Martinez-Vicente M, Ramirez A, Perier C, Klein C, Vila M, et al. Lysosomal dysfunction in Parkinson disease. Autophagy. 2012 Sep 14;8(9):1389–91.

76.Ramirez A, Heimbach A, Gründemann J, Stiller B, Hampshire D, Cid LP, et al. Hereditary parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5 P-type ATPase. Nat Genet. 2006 Oct 10;38(10):1184–91.

77.Brüggemann N, Hagenah J, Reetz K, Schmidt A, Kasten M, Buchmann I, et al. Recessively Inherited Parkinsonism. Arch Neurol. 2010 Nov 1;67(11).

78.Fonzo AD, Dekker MCJ, Montagna P, Baruzzi A, Yonova EH, Guedes LC, et al. FBXO7 mutations cause autosomal recessive, early-onset parkinsonian-pyramidal syndrome. Neurology. 2009 Jan 20;72(3):240– 5.

79.Davison C. Pallido-Pyramidal Disease. J Neuropathol Exp Neurol. 1954 Jan;13(1):50–9.

80.National Center for Biotechnology Information (NCBI) United States National Library of Medicine (NLM). PLA2G6 phospholipase A2 group VI [Homo sapiens (human)] [Internet]. [cited 2023
May 27]. Available from: https://www.ncbi.nlm.nih.gov/ May 27]. Available from: https://www.ncbi.nlm.nih.gov/ gene?Db=gene&Cmd=ShowDetailView&TermToSearch=8398

81.Gregory A KMME et al. PLA2G6-Associated Neurodegeneration. . In: GeneReviews®. 2008.

82.McPherson PS, Czernik AJ, Chilcote TJ, Onofri F, Benfenati F, Greengard P, et al. Interaction of Grb2 via its Src homology 3 domains with synaptic proteins including synapsin I. Proceedings of the National Academy of Sciences. 1994 Jul 5;91(14):6486–90.

83.Cremona O, Nimmakayalu M, Haffner C, Bray-Ward P, Ward DC, De Camilli P. Assignment of SYNJ1 to human chromosome 21q22.2 and Synj12; to the murine homologous region on chromosome 16C3–4 by in situ hybridization. Cytogenet Genome Res. 2000;88(1–2):89–90.

84.Al Zaabi N, Al Menhali N, Al-Jasmi F. SYNJ1 gene associated with neonatal onset of neurodegenerative disorder and intractable seizure. Mol Genet Genomic Med. 2018 Jan;6(1):109–13.

85.Dyment DA, Smith AC, Humphreys P, Schwartzentruber J, Beaulieu CL, Bulman DE, et al. Homozygous nonsense mutation in SYNJ1 associated with intractable epilepsy and tau pathology. Neurobiol Aging. 2015 Feb;36(2):1222.e1-1222.e5.

86.Hardies K, Cai Y, Jardel C, Jansen AC, Cao M, May P, et al. Loss of SYNJ1 dual phosphatase activity leads to early onset refractory seizures and progressive neurological decline. Brain. 2016 Sep;139(9):2420–30.

87.Lesage S, Mangone G, Tesson C, Bertrand H, Benmahdjoub M, Kesraoui S, et al. Clinical Variability of SYNJ1-Associated Early-Onset Parkinsonism. Front Neurol. 2021 Mar 25;12.

88.Makino S, Kaji R, Ando S, Tomizawa M, Yasuno K, Goto S, et al. Reduced Neuron-Specific Expression of the TAF1 Gene Is Associated with X-Linked Dystonia-Parkinsonism. The American Journal of Human Genetics. 2007 Mar;80(3):393–406.

89.Lee L, Pascasio FM, Fuentes FD, Viterbo G. Torsion dystonia in Panay, Philippines. Adv Neurol. 1976;14:137–51.

90.Johnston AW,, McKusick VA. Sex-linked recessive inheritance in spastic paraplegia and parkinsonism. In: Gedda L, editor. Proceedings of the Second International Congress on Human Genetics,. 1961. p. 6–12.

91.Lobo Ingrid. Environmental Influences on Gene Expression. In: Hoopes Laura, editor. GENE EXPRESSION AND REGULATION. 2008.

92.Day JO, Mullin S. The Genetics of Parkinson's Disease and Implications for Clinical Practice. Genes (Basel). 2021 Jun 30;12(7):1006.

93.Davis AA, Andruska KM, Benitez BA, Racette BA, Perlmutter JS, Cruchaga C. Variants in GBA , SNCA , and MAPT influence Parkinson disease risk, age at onset, and progression. Neurobiol Aging. 2016 Jan;37:209.e1-209.e7.

94.Nalls MA, Pankratz N, Lill CM, Do CB, Hernandez DG, Saad M, et al. Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. Nat Genet. 2014 Sep 1;46(9):989–93.

95.International Schizophrenia Consortium; Shaun M Purcell NRWJLSPMVMCOPFSPS. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. Nature. 2009 Aug 1;460(7256):748–52.

96.Escott□Price V, Nalls MA, Morris HR, Lubbe S, Brice A, Gasser T, et

al. Polygenic risk of Parkinson disease is correlated with disease age at onset. Ann Neurol. 2015 Apr 13;77(4):582–91.

97.Susan Searles Nielsen 1 TKBLGGFMFWLJGMFPDSHC. Genotype and age at Parkinson disease diagnosis. Int J Mol Epidemiol Genet . 2013;4(1):61–9.

98.Cook L, Schulze J, Kopil C, Hastings T, Naito A, Wojcieszek J, et al. Genetic Testing for Parkinson Disease. Neurol Clin Pract. 2021 Feb;11(1):69–77.