## JOURNAL OF CONTEMPORARY MEDICINE

DOI:10.16899/jcm.1352503 J Contemp Med 2023;13(5):954-958

Original Article / Orijinal Araştırma



# Liver Protection of Hydroxytyrosol Mediated by Spexin and TRPM2

## Spexin ve TRPM2'nin Aracılık Ettiği Hidroksitirozolün Karaciğer Koruması

## Delif ONAT<sup>1</sup>, Devin KOCAMAN<sup>2</sup>

<sup>1</sup>Department of Medical Pharmacology and Clinical Pharmacology, Faculty of Medicine, Adıyaman University, Adıyaman, Turkey <sup>2</sup>Department of Histology and Embryology, Faculty of Medicine, Fırat University, Elazığ, Turkey

## Abstract

**Aim**: In the study, the role of Spexin (SPX) and Transient Receptor Potential (TRP) Melastatin-Like Subfamily Member 2 (TRPM2) in the protective effect of Hydroxytyrosol (HT) in rats given Corn Syrup was evaluated.

**Material and Method:** The rats were divided into 4 groups (6 rats in each) (Control, HT, Corn Syrup, Corn Syrup +HT). Rats were given 30% Corn Syrup with drinking water for 6 weeks. Four ml/kg/day liquid containing HT was applied by oral gavage alone and together with Corn Syrup for 6 weeks. Molecular parameters SPX and TRPM2 were examined histopathologically in liver tissue.

**Results**: The SPX levels decreased and the TRPM2 levels increased more in the Corn Syrup-given Group than in the Control Group. SPX levels increased and TRPM2 levels decreased after HT treatment. In the HT Group only, no differences were detected when compared to the control Group.

**Conclusion**: SPX and TRPM2 may mediate the protective effect of HT on the liver in rats given corn syrup.

Keywords: Corn Syrup, Hydroxytyrosol, Spexin, TRPM2

## Öz

**Amaç**: Araştırmada Mısır Şurubu verilen sıçanlarda Hidroksitirozol (HT)'ün koruyucu etkisinde Spexin (SPX) ve geçici reseptör potansiyeli (TRP) melastatin benzeri alt aile üyesi 2 (TRPM2)'nin rolü değerlendirildi.

**Gereç ve Yöntem**: Sıçanlar 4 gruba (her grupta 6 sıçan) ayrıldı (Kontrol, HT, Mısır Şurubu, Mısır Şurubu +HT). Sıçanlara 6 hafta süreyle %30'luk Mısır Şurubu içme suyuyla birlikte verildi. HT içeren 4 ml/kg/gün sıvı, tek başına ve Mısır Şurubu ile birlikte 6 hafta süreyle oral gavajla uygulandı. Karaciğer dokusunda SPX ve TRPM2 moleküler parametreleri histopatolojik olarak incelendi.

**Bulgular**: Mısır Şurubu verilen grupta SPX seviyeleri azaldı ve TRPM2 seviyeleri kontrol grubuna göre daha fazla arttı. HT tedavisinden sonra SPX seviyeleri arttı, TRPM2 seviyeleri ise azaldı. Yalnızca HT grubunda kontrol grubuyla karşılaştırıldığında herhangi bir farklılık tespit edilmedi.

**Sonuç**: SPX ve TRPM2, mısır şurubu verilen sıçanlarda HT'nin karaciğer üzerindeki koruyucu etkisine aracılık edebilir.

**Anahtar Kelimeler**: Mısır Şurubu, Hidroksitirosol, Spexin, TRPM2

Corresponding (*İletişim*): Elif ONAT, Department of Medical Pharmacology and Clinical Pharmacology, School of Medicine, Adıyaman University, Adıyaman, 02040, Turkey.
E-mail (*E-posta*): eonat@adiyaman.edu.tr
Received (*Geliş Tarihi*): 30.08.2023 Accepted (*Kabul Tarihi*): 26.09.2023



## INTRODUCTION

Glucose and fructose are closely related to simple sugars, but fructose is associated more with metabolic diseases. The main source of fructose was fruit until the 1960s, but then High Fructose Corn Syrup (HFCS) became a dominant component of the Western Diet.<sup>[11]</sup> Consumption of fructose as sugar and High Fructose Corn Syrup has increased significantly in recent years. This trend is associated with increasing metabolic diseases. The biochemical pathways of fructose metabolism were described in the early 1990s and fructose metabolism and its pathophysiological effects on the body at the organismal level have only recently been investigated.<sup>[2]</sup>

Hydroxytyrosol (HT) in olive oil is a polyphenol that has antioxidant characteristics that support the healthy characteristics of olive oil along with other components such as polyphenols and flavonoids.<sup>[3]</sup> HT includes cardioprotective, anti-inflammatory, anti-cancer, and antimicrobial effects,<sup>[3]</sup> which are the characteristics of olives as a healthy food. It was shown previously that HT prevents damage caused by Reactive Oxygen Species (ROS)<sup>[4]</sup> in endothelial cells and reduces endothelial damage and atherogenic injuries.<sup>[5]</sup> Previous studies reported that HT exerts its protective effects through the following mechanisms; (i) by preventing Low-Density Lipoprotein (LDL) oxidation, (ii) by inhibiting platelet aggregation, (iii) by mitigating mitochondrial abnormalities and preventing the metabolic syndrome (MetS) induced by a high fructose diet,<sup>[6]</sup> and (iv) by producing anti-inflammatory effects in conjunction with decreased Cyclooxygenase 1 (COX1) and COX2 enzyme activity.<sup>[7]</sup>

Spexin (SPX) is a 14 amino acid-long peptide hormone expressed extensively in central and peripheral tissues and secreted into the circulation as a response to metabolic stress. Studies show that SPX acts as a multifunctional peptide in metabolic processes. Endogenous SPX is sensitive to metabolic changes. It was reported that circulating SPX levels were reduced in chronic diseases. This suggests that SPX is a potential drug target for the development of novel pharmacological strategies.<sup>[8]</sup>

Transient Receptor Potential (TRP) Melastatin-Like Subfamily Member 2 (TRPM2) is a Ca<sup>2+</sup> permeable cation channel with extremely low Ca<sup>2+</sup> selectivity,<sup>[9,10]</sup> and is highly present in different cells.<sup>[11,12]</sup> TRPM2 is involved in different cellular and physiological processes, including cytokine production, cell death, oxidative stress, and fibrosis. Also, TRPM2 is an important factor in cell death caused by oxidative stress over the activation of caspase cleavage.<sup>[13,14]</sup> However, more studies are required to clarify its functional roles.

Although the protective effects of HT on metabolic diseases are known, data on how these effects happen are insufficient. In the present study, it was investigated whether SPX and TRPM2 had any roles in the protective effect of HT on liver damage in rats as a result of consuming Corn Syrup with drinking water (30% for six weeks).

#### **MATERIAL AND METHOD**

#### Animals and experimental design

The study was approved byAdıyaman University Animal Experiments Ethics Committee (Date: 06.10.2022, Decision No: 2022/051-2). The experiments were carried out per the "Guide for the Care and Use of Laboratory Animals". Twenty-four male, 200-250 g Sprague-Dawley rats (8-10 weeks) provided by Adıyaman University Experimental Research Center given ad libitum standard water and feed were used in the study in 4 groups (n: 6); Group I (Control), Group II (HT), Group III (Corn Syrup), and Group IV (Corn Syrup+HT). No applications were made to the Control Group. HT was supplied in liquid form from Kale Natural Herbal Products Company in Turkey. From this liquid containing HT, 4 ml/kg/day was administered orally for 6 weeks to rats in Groups II and IV. Rats in Groups III and IV were given 30% Corn Syrup with drinking water for 6 weeks.<sup>[15]</sup> At the end of 6 weeks, the rats were anesthetized with IP Ketamine (75 mg/kg)+Xylazine (10 mg/kg), and blood samples were taken from the hearts of all groups (Blood was drawn from the hearts of the rats to terminate the study). The liver tissues were fixed in a 10% formaldehyde solution for histological evaluations.

#### Immunohistochemical examination

Liver tissues of animals were passed through histological follow-up series and embedded in paraffin blocks. Immunohistochemical staining was applied with 5-µm sections as described by Kocaman and Artas.<sup>[16]</sup> Immunohistochemistry (IHC) was applied on 3-µm histological tissue microarray slides with Spexin primary antibodies (A04088-1, Booster Biological Technology, Pleasanton, CA, USA) and rabbit polyclonal anti-TRPM2 antibodies (Ab-11168), Abcem, Cambridge, UK) and were photographed with Zeiss Axio Scope A1 microscope (Carl Zeiss Microscopy GmB H 07745 Jena, Germany) and a histoscore was established for SPX and TRPM2.

In the microscopic evaluation of the staining density; the negative staining areas were given "0", areas with < 25 % staining were given "0.1", areas with 26-50% staining were given "0.4", areas with 51-75% staining were given "0.6", and areas with near-homogeneous staining (76-100%) were given "0.9". The final histoscore was calculated with the following formula. Histoscore = Distribution × Intensity.<sup>[16]</sup>

#### **Statistical analysis**

The SPSS 22 (IBM Corporation, USA) was used for the analysis. The One-Way ANOVA Test was used and post-hoc multiple comparisons were made with Tukey HSD. The data are given as Mean  $\pm$  SD. P<0.05 was taken as statistically significant.

## RESULTS

#### Immunohistochemical Findings

With the immunohistochemical staining of SPX and TRPM2 immunoreactivity in liver tissue under the light microscope, the following results were achieved.

SPX immunoreactivity was lower in the Corn Syrup Group when compared to the Control and HT Groups (p<0.001). SPX immunoreactivity was elevated in the Corn Syrup+HT Group than in the Corn Syrup Group (p<0.001) (**Table 1**). SPX immunoreactivity histoscores for the four groups are given in **Figure 1**.

Table 1: Immunohistochemical findings for SPX in the liver tissues							
Groups	Control	HT	Corn Syrup	Corn Syrup+HT			
SPX	1.05±0.16	1.1±0.15	0.08±0.03 <sup>ab</sup>	0.55±0.08 abc			
Error bars show SD; a. p<0.05 compared to control; b. p<0.05 compared to HT; c. p<0.05 compared to Corn Syrup.							

TRPM2 immunoreactivity was elevated in the Corn Syrup Group when compared to the Control and HT Groups (p<0.001). TRPM2 immunoreactivity was lower in the Corn Syrup+HT Group than in the Corn Syrup Group (p<0.001) (**Table 2**). TRPM2 immunoreactivity histoscores for the four groups are given in **Figure 2**.

Table 2: Immunohistochemical findings for TRPM2 in the liver tissues						
Groups	Control	HT	Corn Syrup	Corn Syrup+HT		
TRPM2	0.06±0.02	0.08±0.03	1.2±0,33 <sup>ab</sup>	0.5±0.08 <sup>abc</sup>		
Error bars show SD; a. p<0.05 compared to control; b. p<0.05 compared to HT; c. p<0.05 compared to Corn Syrup.						

## DISCUSSION

The role of SPX and TRPM2 molecules in the protective effect of HT on the pathological changes in the liver because of Corn Syrup consumption in rats was evaluated histopathologically in the present study. It was shown in the study for the first time that SPX and TRPM2 may mediate the protective effects of HT, whose metabolic protective effect is known, against liver damage because of Corn Syrup.

SPX is commonly found in endocrine and epithelial tissue<sup>[17]</sup> and is considered to be involved in metabolic disorders. SPX was lower in patients with MetS in a clinical study. Also, an inverse relationship was detected between SPX and glucose, lipid, and blood pressure in MetS.<sup>[18]</sup> SPX treatment decreased fatty acid uptake into hepatocytes.<sup>[19]</sup> However, Subcutaneous (SC) injection of SPX was shown to reduce appetite and reduce caloric intake by approximately 32% in rats.<sup>[20]</sup> Also, Behrooz et al.<sup>[21]</sup> reported an inverse relationship between SPX levels and dietary fat intake in obese children. SPX has potential regulatory roles in metabolic status. SPX treatment reduced hepatic lipid storage, Aspartate Aminotransferase (AST), and Alanine Aminotransferase (ALT) in Diet-Induced Obese (DIO) mice. Also, the uptake of Long-Chain Fatty Acids (LCFAs) in hepatocytes was reduced by SPX.<sup>[19]</sup> Similarly, another study conducted with DIO mice showed that SPX reduced lipid accumulation and glycogen levels.<sup>[18]</sup> Also, hepatic glucose production was reduced due to SPX in DIO rats, and CRISPR/Cas9-mediated silencing of SPX in Human Liver Cancer Cells (HepG2) triggered gluconeogenesis. <sup>[22]</sup> Similarly, plasma SPX levels were found to be lower in Non-Alcoholic Fatty Liver Disease (NAFLD) patients when compared to controls,<sup>[23]</sup> which indicates the potential therapeutic value of SPX in the treatment of hepatic steatosis/ NAFLD.<sup>[8]</sup> It was found in the present study that the SPX levels decreased in the liver tissue in the Corn Syrup Group when compared to the Control Group, and the SPX level increased after HT treatment, which suggests that SPX may also contribute to this characteristic of HT, which is known to have protective effects on the liver. Because HT consumption, which is one of the main components of olive oil and has significant antioxidant, anti-inflammatory, and antimicrobial characteristics, is associated with the improvement of MetS and related disorders,<sup>[24]</sup> it has been recently found to

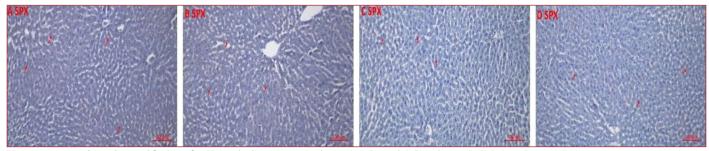


Figure 1: Immunohistochemical findings for SPX in the liver tissues (red arrow). A.Control, B.Corn Syrup, C.HT, D.Corn Syrup+HT

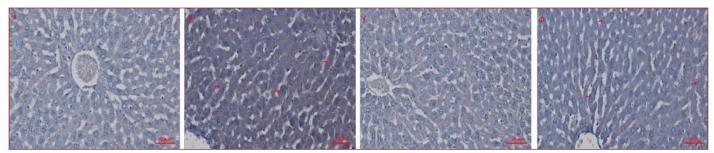


Figure 2: Immunohistochemical findings for TRPM2 in the liver tissues (red arrow). A.Control, B.Corn Syrup, C.HT, D.Corn Syrup+HT

improve Insulin Resistance and obesity by modulating the gut microbiota.[25] HT-rich olive leaf extracts were shown to have hypolipidemic and hepatoprotective effects on high-fat dietinduced lipid metabolism disorder and liver injury in rats after improving the antioxidative defense system and blocking protein expression in inflammation and liver damage and against metabolic disorders induced by high-fructose diet. <sup>[28]</sup> In another study, dietary supplementation with 5 mg of HT attenuated the deleterious metabolic effects that were produced by a high fructose diet in mice. HT's protective effects in the liver are considered to be associated with (i) the restoration of the activity of A-5 and A-6 desaturase enzymes by preventing depletion of n-3 LCPUFAs, (ii) reduced oxidative stress, (iii) the down-regulation of lipogenic factor SREBP-1c, and (iv) preservation of n-3 LCPUFA levels in extrahepatic tissues.<sup>[29]</sup> SPX is likely to play roles in these mechanisms of action, which belong to the protective characteristics of HT. However, more studies are needed to understand this

TRPM2 activity is indispensable for many physiological processes, including insulin secretion in pancreatic β-cells, monocyte chemokine production, and heat sensation of hypothalamic neurons.<sup>[30]</sup> Because of its Ca<sup>2+</sup> permeability, TRPM2 is also involved in many pathophysiological processes that cause cell death because of the production of Reactive Oxygen Species (ROS).<sup>[31]</sup> For this reason, TRPM2 has become an attractive pharmacological target. In a previous study, evaluation of acetaminophen-induced liver injury due to blood liver enzyme concentration and liver histology exhibited less severe liver injury in TRPM2 knockout mice compared to WT mice.<sup>[32]</sup> TRPM2 channels are an integral part of the acetaminophen-induced hepatocellular death mechanism. For this reason, TRPM2-mediated cell death is an important mechanism in NAFLD-induced liver injury.<sup>[33]</sup> In our study, it was found that the TRPM2 levels increased in the Corn Syrup given Group when compared to the Control Group, and the TRPM2 level decreased after HT treatment, which suggests that TRPM2 may be involved in the mechanism of action of HT's protective characteristics, which has antiinflammatory and antioxidant characteristics. TRPM2, which is proinflammatory, provides the basis for discoveries regarding this pathology in terms of liver damage.

The most important limitation of the study was that methods such as PCR and Western Blot Analysis could not be used because of financial reasons. Further studies involving larger numbers of animals to explain the association of HT with SPX and TRPM2 will support the molecular mechanisms of the study. Also, the protective effects of HT on the liver must be supported by clinical findings.

### CONCLUSION

It is considered that some novel molecules such as SPX and TRPM2 may contribute to the protective effects of HT against the harmful effects of Corn Syrup on the liver.

#### **ETHICAL DECLARATIONS**

**Ethics Committee Approval**: The study was approved by Adiyaman University Animal Experiments Ethics Committee (Date: 06.10.2022, Decision No: 2022/051-2).

**Informed Consent:** Because the study was designed retrospectively, no written informed consent form was obtained from patients.

Referee Evaluation Process: Externally peer-reviewed.

**Conflict of Interest Statement**: The authors have no conflicts of interest to declare.

**Financial Disclosure**: The authors declared that this study has received no financial support.

**Author Contributions**: All of the authors declare that they have all participated in the design, execution, and analysis of the paper, and that they have approved the final version.

#### REFERENCES

- 1. Febbraio MA, Karin M. "Sweet death": Fructose as a metabolic toxin that targets the gut-liver axis. Cell Metab 2021;33(12):2316–28.
- 2. Jung S, Bae H, Song WS, Jang C. Dietary Fructose and Fructose-Induced Pathologies. Annu Rev Nutr 2022;42:45–66.
- Cicerale S, Lucas LJ, Keast RS. Antimicrobial, antioxidant and antiinflammatory phenolic activities in extra virgin olive oil. Curr Opin Biotechnol 2012;23:129–35.
- Vilaplana-Pérez C, Auñón D, García-Flores LA, Gil-Izquierdo A. Hydroxytyrosol and potential uses in cardiovascular diseases, cancer, and AIDS. Front Nutr 2014;27:1–18.
- Catalán U, López de Las Hazas MC, Rubió L et al. Protective effect of hydroxytyrosol and its predominant plasmatic human metabolites against endothelial dysfunction in human aortic endothelial cells. Mol Nutr Food Res 2015;59:2523–36.
- Cao K, Xu J, Zou X et al. Hydroxytyrosol prevents diet-induced metabolic syndrome and attenuates mitochondrial abnormalities in obese mice. Free Radic Biol Med 2014;67:396–407.
- 7. Scoditti E, Nestola A, Massaro M et al. Hydroxytyrosol suppresses MMP-9 and COX-2 activity and expression in activated human monocytes via PKC $\alpha$  and PKC $\beta$ 1 inhibition. Atherosclerosis 2014;232:17–24.
- Türkel İ, Memi G, Yazgan B. Impact of spexin on metabolic diseases and inflammation: An updated minireview. Experimental Biology and Medicine 2022;247:567–73.
- 9. Zhang Z, Tóth B, Szollosi A, Chen J, Csanády L. Structure of a TRPM2 channel in complex with Ca2+ explains unique gating regulation. eLife 2018;7:e36409.
- 10. Hasan R, Zhang X. Ca2+ regulation of TRP ion channels. Int J Mol Sci 2018;19(4):1256.
- 11. Sumoza-Toledo A, Penner R. TRPM2: A multifunctional ion channel for calcium signaling. J Physiol 2011;589:1515–25.
- 12. Nagamine K, Kudoh J, Minoshima S et al. Molecular cloning of a novel putative Ca2+ channel protein (TRPC7) highly expressed in brain. Genomics 1998;5:124–31.
- 13. Hara Y, Wakamori M, Ishii M et al. LTRPC2 Ca2+-permeable channel activated by changes in redox status confers susceptibility to cell death. Mol Cell 2002;9:163–73.
- 14.Zhang W, Hirschler-Laszkiewicz I, Tong Q et al. TRPM2 is an ion channel that modulates hematopoietic cell death through activation of caspases and PARP cleavage. Am J Physiol Cell Physiol 2006;290:C1146–C1159.
- 15.15.Gün A, Özer MK, Bilgiç S, Kocaman N, Ozan G. Effect of Caffeic Acid Phenethyl Ester on Vascular Damage Caused by Consumption of High Fructose Corn Syrup in Rat. Oxid Med Cell Longev 2016; 2016:3419479.

mechanism.

- Kocaman N, Artas G. Can novel adipokines, asprosin and meteorinlike, be biomarkers for malignant mesothelioma? Biotech Histochem 2020;95:171-5.
- 17. Mirabeau O, Perlas E, Severini C et al. Identification of novel peptide hormones in the human proteome by hidden Markov model screening. Genome Res 2007;17:320–7.
- Kolodziejski PA, Leciejewska N, Chmurzynska A et al. 30-Day spexin treatment of mice with diet-induced obesity (DIO) and type 2 diabetes (T2DM) increases insulin sensitivity, improves liver functions and metabolic status. Mol Cell Endocrinol 2021;536:111420.
- 19. Ge JF, Walewski JL, Anglade D, Berk PD. Regulation of hepatocellular fatty acid uptake in mouse models of fatty liver disease with and without functional leptin signaling: roles of NfKB and SREBP-1C and the effects of spexin. Semin Liver Dis 2016;36:360–72.
- 20. Walewski JL, Ge F, Lobdell H et al. Spexin is a novel human peptide that reduces adipocyte uptake of long chain fatty acids and causes weight loss in rodents with diet-induced obesity. Obesity 2014;22:1643–52.
- 21. Behrooz M, Vaghef-Mehrabany E, Ostadrahimi A. Different spexin level in obese vs normal weight children and its relationship with obesity related risk factors. Nutr Metabol Cardiovasc Dis 2020;30:674–82.
- 22. Gu L, Ding X, Wang Y et al. Spexin alleviates insulin resistance and inhibits hepatic gluconeogenesis via the FoxO1/PGC-1alpha pathway in high-fatdietinduced rats and insulin resistant cells. Int J Biol Sci 2019;15:2815–29.
- 23. Zhang L, Li G, She Y, Zhang Z. Low levels of spexin and adiponectin may predict insulin resistance in patients with non-alcoholic fatty liver. Pract Lab Med 2021;24:e00207.
- 24. V Goulas, V Exarchou, A.N Troganis et al. Phytochemicals in olive-leaf extracts and their antiproliferative activity against cancer and endothelial cells. Mol Nutr Food Res 2009;53(5):600-8.
- 25. Brown WV, Fujioka K, Wilson PWF, Woodworth KA. Obesity: why be concerned? Am J Med 2009;122:S4-11.
- 26. Rietjens SJ, Bast A, Haenen GRMM. New insights into controversies on the antioxidant potential of the olive oil antioxidant hydroxytyrosol. J Agric Food Chem 2007;55(18):7609-14.
- Bouallagui Z, Han J, Isoda H, Sayadi S. Hydroxytyrosol rich extract from olive leaves modulates cell cycle progression in MCF-7 human breast cancer cells. Food Chem Toxicol 2011;49(1):179-84.
- 28. Fki İ, Sayadi S, Mahmoudi A, Daoued İ, Marrekchi R, Ghorbe H. Comparative Study on Beneficial Effects of Hydroxytyrosol- and Oleuropein-Rich Olive Leaf Extracts on High-Fat Diet-Induced Lipid Metabolism Disturbance and Liver Injury in Rats. BioMed Research International 2020; 15:1315202.
- 29. Valenzuela R, Echeverria F, Ortiz M et al. Hydroxytyrosol prevents reduction in liver activity of  $\Delta$ -5 and  $\Delta$ -6 desaturases, oxidative stress, and depletion in long chain polyunsaturated fatty acid content in different tissues of high-fat diet fed mice. Lipids in Health and Disease 2017;16:64.
- 30. Song K, Wang H, Kamm G.B et al. The TRPM2 channel is a hypothalamic heat sensor that limits fever and can drive hypothermia. Science 2016;353:1393–98.
- 31. Nilius B, Owsianik G, Voets T, Peters J.A. Transient Receptor Potential Cation Channels in Disease. Physiol Rev 2007;87:165–217.
- 32. Kheradpezhouh E, Ma L, Morphett A, Barritt GJ, Rychkov GY. TRPM2 channels mediate acetaminophen-induced liver damage. Proc Natl Acad Sci USA 2014;111:3176–81.
- 33. Malko P,Jiang LH. TRPM2 channel-mediated cell death: an important mechanism linking oxidative stress-inducing pathological factors to associated pathological conditions. Redox Biol 2020;37: 101755.