Journal of Physical Chemistry and Functional Materials

Home Page of Journal: https://dergipark.org.tr/jphcfum



An Insilico Study on Structural Aspects of Caffeine by ArgusLab 4 software

DÖzlem İşcan*

Bartın University, Project and Technology Office, Bartın, Türkiye * Corresponding author: Özlem İşcan E-mail: oiscan@bartin.edu.tr

ABSTRACT

In this study, the molecular docking method investigated the quasi-experimental molecular computational studies of the caffeine compound found in many herbals and the binding properties of the 4QHO protein to the active site. For molecular docking, geometry optimization of caffeine molecule was done in the Argus Lab program with the semi-experimental PM3 method. The HOMO, LUMO, and HOMO-LUMO energy differences and potential energy surface of all optimized molecules were calculated. To understand the relationship between the caffeine molecule and the feeling of satiety, the interaction between caffeine and 4QHO protein was examined using the molecular docking method. The selected crystal structure of 4QHO protein was obtained from the protein database in *.pdb format, and optimized ligand interaction with this crystal structure was studied using ArgusLab. As a result of molecular docking studies, ligand-protein binding energies, hydrogen bond sites, and the number of interactions between ligand-protein were determined and evaluated.

1. Introduction

Caffeine is one of the methylxanthine class of stimulants for the central nervous system (CNS) [1]. As a eugeroic, encouraging wakefulness, and a mild cognitive enhancer, improving alertness and attentional performance, its main uses are for recreational purposes [2,3]. The neurotransmitter acetylcholine is released more readily when caffeine inhibits adenosine's binding to the adenosine A1 receptor [4]. Due to their structural similarities, caffeine and adenosine can bind to each other and impede adenosine receptors, which causes this effect [5]. Because it inhibits phosphodiesterase nonselectively, caffeine also raises the levels of cyclic AMP [6].

It was utilised in Dutch medicine in the first part of the 20th century as a circulatory and respiration stimulant. The first people to take advantage of coffee's medicinal properties were Islamic physicians; long before the second millennium A.D.D., the Yemeni Sufis are credited with using it as a beverage for the first time. Caffeine was banned from use because of the growing belief that it is seditious and causes diarrhoea, even though caffeine is

ARTICLE INFO

Keywords:

Caffeine ArgusLab Molecular Docking

Received: 2023-12-08 Accepted: 2023-12-16 ISSN: 2651-3080 DOI: 10.54565/jphcfum.1402117

used as a stimulant more often. Coffee was brought to Holland ten years after being introduced to England in the 1650s. The coffee plant was brought to Java by the Dutch in 1688, and because of the island's reputation for producing fine coffee, premium coffee has come to be known by the moniker "Java" [7].

Obesity is associated with many diseases, especially cardiovascular and cancer diseases [8,9]. The oxidation of N-methyl groups in nucleic acids, and specifically of N6-methyladenosine (m6A) in RNA, is catalyzed by the Fe(II) and 2-oxoglutarate (2OG)dependent oxygenase fat mass and obesity-associated protein (FTO). [10-13]. The overwhelming body of data indicates that SNPs in FTO significantly increase energy intake while decreasing satiety. [14].

Arguslab's molecule builder was used to create the caffeine molecules acquired from Pubchem [15,16]. The atoms in caffeine's molecules have a net charge of 0 and 74 valence electrons. 2D structures, ball-stick and space-

filling view in PubCHEM compound and active conformation caffeine are shown in Figure 1.



2D Structure



Space-Filling

Figure 1 Structure of Caffeine

Ball and Stick

Potential energies, molecular structures, structure optimization geometry, atom coordinate vibration frequencies, bond length, bond angle, and reaction pathways are all predicted by the quantum mechanically based electronic structure software ArgusLab [17]. Conformational analysis, a method for calculating molecular structures, conformational energies, and other molecular properties, is based on molecular mechanics. It incorporates ideas from classical mechanics. A molecule is thought to be an assembly of atoms bound together by classical forces. Bond lengths, bond angles, torsion angles,

2. Material and Method

The crystal structure of FTO was obtained from RCSB [19] (PDB ID:4QHO). We chose ArgusLab 4.0 to investigate the binding site. One (www.arguslab.com) is the study's source. ArgusLab can function on Windows 11 from the Windows operating system.

ArgusLab software was used on a window-based computer for all conformational analysis (geometry optimization) research. Many tools for building models, minimizing, and representing molecular structures have been made possible by advances in computing [20-22]. The semi-empirical Parametric Method 3 (PM3) parameterization was utilized to complete the minimization process after ArgusLab produced the caffeine structure [23-24].

The minimum potential energy is determined using the geometry convergence function in the ArgusLab software. The surfaces were designed to show properties of the ground state and the excited conditions, such as orbitals, electron densities, spin densities, and electrostatic potentials (ESP). They also produced grid data to create and other structural features' potential energy functions are used to characterize these forces. Equation (1) can calculate the molecule's energy (E) as a sum of terms.

 $E = E_{stretching} + E_{bending} + E_{torsion} + E_{Vander Waals} + E_{electrostatic} + E_{hydrogen bond} + cross term$ (1)

The precise computation of the geometric properties of molecules depends on these terms. A force field is the collection of energy functions and the associated parameters [18].

molecular orbital surfaces, which show the molecular orbitals and map the electrostatic potential on the electron density surface. We used the geometry convergence map to calculate the minimum potential energy for caffeine. [25-28]. Using the PM3 method, the Mulliken and ZDO Atomic Charges of caffeine were determined.

After downloading the FTO crystal structure (PDB ID: 4QHO) from the PDB databank, the ArgusLab program was used to create the binding site by selecting the "Make binding site for this protein" option. Caffeine was chosen as the ligand, and hydrogen atoms were added. The ligands were then permitted to operate using the AScore scoring function and the GA algorithm. 4.0 in ArgusLab.

3. Results and Discussions

Table 1 provides the atomic input data for thecomputation above. According to ArgusLab 4.0'sRHF/PM3 method, the last minimum geometrical energyand SCF energy were determined to be -85.8936912420 au(-53899.1536 kcal/mol) in Table 2.

Atoms	Х	Y	Ζ	Atoms	Х	Y	Z
Number				Number			
1 C	-3.097000	-1.122000	-0.400000	2 N	-2.099000	-0.045000	-0.311000
3 C	-2.351000	1.305000	-0.248000	4 N	-1.242000	2.015000	-0.170000
5 C	-0.268000	1.082000	-0.161000	6 C	-0.744000	-0.191000	-0.276000
7 C	0.073000	-1.412000	-0.383000	8 O	-0.314000	-2.470000	-0.818000
9 N	1.430000	-1.150000	0.118000	10 C	1.900000	0.178000	-0.308000
11 0	2.980000	0.312000	-0.834000	12 N	1.039000	1.319000	-0.082000
13 C	1.574000	2.655000	0.216000	14 C	2.201000	-2.076000	0.924000
15 H	-4.119000	-0.681000	-0.409000	16 H	-2.934000	-1.701000	-1.336000
17 H	-2.992000	-1.797000	0.478000	18 H	-3.360000	1.743000	-0.261000
19 H	2.686000	2.615000	0.231000	20 H	1.240000	3.371000	-0.568000
21 H	1.201000	2.991000	1.210000	22 H	3.190000	-1.626000	1.163000
23 H	1.653000	-2.290000	1.869000	24 H	2.352000	-3.023000	0.358000

Table 1 Atomic coordinates of caffeine compound

 Table 2 SCF is performed by computing SCF using a single electron matrix.

Cycle	Energy (au)	Difference	Cycle	Energy (au)	Difference
1	-43.879246		2	-60.473763689	-16.5945
3	-54.788619925	5.68514	4	-71.721995461	-16.9334
5	-76.790156961	-5.06816	6	-80.425637025	-3.63548
7	-83.516250964	-3.09061	8	-85.318098861	-1.80185
9	-85.866961190	-0.548862	10	-85.889610524	-0.0226493
11	-85.892988983	-0.00337846	12	-85.893496721	-0.000507737
13	-85.893629512	-0.000132792	14	-85.893669687	-4.01751e-005
15	-85.893683293	-1.36062e-005	16	-85.893688229	-4.93609e-006
17	-85.893690076	-1.84619e-006	18	-85.893690783	-7.06986e-007
19	-85.893691059	-2.75879e-007	20	-85.893691168	-1.09324e-007
21	-85.893691210	-4.20114e-008	22	-85.893691229	-1.91109e-008
23	-85.893691236	-7.53471e-009	24	-85.893691240	-3.2112e-009
25	-85.893691241	-1.29899e-009	26	-85.893691242	-5.52518e-010
27	-85.893691242	-2.38742e-010	28	-85.893691242	-9.93623e-011
29	-85.893691242	-4.18368e-011	30	-85.893691242	-1.86446e-011
31	-85.893691242	-7.7307e-012	32	-85.893691242	-1.81899e-012
33	-85.893691242	-2.72848e-012	34	-85.893691242	9.09495e-013
35	-85.893691242	-1.13687e-012	36	-85.893691242	-3.41061e-013
37	-85.893691242	5.68434e-013	38	-85.893691242	-1.13687e-013

The ZDO and Mulliken atomic charges of caffeine are listed in Table 3.

Table 3 List of Mulliken and ZDO Atomic Charges of Caffeine by using ArgusLab software

Atoms	ZDO	Mulliken	Atoms	ZDO	Mulliken	Atoms	ZDO	Mulliken
Number	Atomic	Atomic	Number	Atomic	Atomic	Number	Atomic	Atomic
	Charges	Charges		Charges	Charges		Charges	Charges
1 C	-0.1183	-0.3342	9 N	-0.0328	-0.0834	17 H	0.0832	0.1623
2 N	0.3608	0.3283	10 C	0.2317	0.2739	18 H	0.1666	0.2666

3 C	-0.1940	-0.2706	11 0	-0.3766	-0.3855	19 H	0.0662	0.1411
4 N	-0.1358	-0.1458	12 N	0.1181	0.0802	20 H	0.0748	0.1519
5 C	-0.0531	-0.0298	13 C	-0.0771	-0.2855	21 H	0.0455	0.1163
6 C	-0.4072	-0.4384	14 C	-0.0620	-0.2670	22 H	0.0690	0.1443
7 C	0.3708	0.4203	15 H	0.0626	0.1346	23 H	0.0446	0.1153
8 O	-0.3739	-0.3829	16 H	0.0716	0.1480	24 H	0.0654	0.1399

The heat released from individual atoms during forming the element's stable form under standard conditions is known as the atomic heat of formation. It is important to remember that thermodynamic adjustments not be included in the formation energy since the parametrization implicitly includes them. ArgusLab software is used to determine the most energetically favorable conformation of caffeine, which has a heat of formation of -52.0215 kcal/mol.

Known as Frontier Orbitals, the HOMO and LUMO orbitals are very helpful in explaining chemical reactivity. HOMO (Highest Occupied Molecular Orbital I) and HOMO -1 (MO 36). LUMO (Lowest Unoccupied Molecular Orbital I) and e MO 37. Fig. 5 shows caffeine's LUMO +1 (MO 39) and MO 38. This was done theoretically using PM3.

Examining the E_{HOMO} and E_{LUMO} is crucial to understanding the complex's electronic characteristics. Using PM3, this was accomplished theoretically. The two colors signify the positive and negative areas of the orbital; the blue regions correspond to an increase in electron density, and the red areas to a decrease in electron density. Nevertheless, these calculations were also investigated in a vacuum and the ground state. By comparing them with other compounds that are similar to them, one can use them to obtain information.



Figure 2. Visualise the LUMO+1, LUMO, HOMO, HOMO-1 of Caffeine, blue shows positive and red shows negative.
Molecular orbital representations of the caffeine in each of its four energy states (GAP1, GAP2, GAP3, and GAP4), along with their corresponding GAP values (energy variation between frontier orbitals HOMO and LUMO).

The entire surface of caffeine is depicted with a color map in Figure 3. To display a cutaway of the same surface and the underlying molecular structure, this figure makes use of a clipping plane. The ESP energy (in hartrees) for each color is displayed on the color m The electrostatic potential's polarity and magnitude are reflected in the color of the surface. The regions of greatest stability for a positive test charge are shown by the red end of the spectrum, while the areas of least stability are indicated by the magenta/blue end.



Figure 3 shows a potential electrostatic map of the stemonal molecule produced by applying the Mulliken charges with scale.

The results concerning the molecular docking of caffeine over 4QHO are shown in Fig. 4. One issue involved the different parameters between different docking programs. The grid box size for ArgusLab (151 x 140 x 151 Å) was set. The best molecular docking position of caffeine is -6.09979 kcal/mol. 2D images were created with the help of biovia discovery studio (Discovery Studio Client version 19. 1.0. 18287). Analysis on the PLIP server [29] revealed that two hydrogen bonds were involved in binding each compound with the protein. The amino acids involved in hydrogen bond formation for caffeine are shown in Table 4.



Figure 4 a. Demonstrate the interaction of Caffeine at the binding site of 4QHO b. 3D structure of 4QHO-Caffeine complex c.2D structure of 4QHO-Caffeine complex

II. J D J.										
Hydrogen Bonds										
Index	Residue	AA	Distance	Distance	Donor	Protein	Side	Donor	Acceptor	
			H-A	D-A	Angle	donor?	chain	Atom	Atom	
1	84A	ARG	1.60	2.54	158.85	\checkmark	\checkmark	452 [Ng+]	3416 [O ₂]	
2	87A	GLY	2.17	2.84	124.12	\checkmark	\checkmark	470 [Nam]	3419 [O ₂]	

Table 4 Hydrogen bond interactions between 4qho protein and caffeine

4. Discussion

It has been observed that the ArgusLab program has a very easy-to-understand interface and completes the calculations in a very short time. In this research study, we included some calculation studies that can be done with ArgusLab and observed that the results are stable. Using the ArgusLab software, caffeine's lowest energetically favorable conformation is found to have a minumun energy conformation of -52.0215 kcal/mol. The studies used ArgusLab software to determine the minimum potential energy at which the minimum energy conformation of caffeine is found to be at -85.8936912420 au (-53899.1536 kcal/mol). It can now be demonstrated that caffeine has an active effect on FTO protein.

The lowest energy conformations were employed in molecular modeling calculations after the geometric variables related to caffeine were finally fully optimized for the compound. The calculated thermodynamic parameter, dipole moment, Mulliken and ZDO Atomic Charge, and optimized geometry were all well within the computational results' accuracy range. The data derived from the calculated parameter was analyzed. Studies have demonstrated a modest impact of caffeine compounds on obesity.

It has been shown that caffeine molecules have a moderate effect against obesity in an insilico environment with the help of molecular docking studies. According to the results of this research, it is thought that consuming a cup of Turkish coffee containing plenty of caffeine can reduce the feeling of eating by suppressing hunger.

Because of this, modeling, computation, and docking studies have given us new insight into the caffeine compound and knowledge about challenging problems to observe through experimentation.

Acknowledgement:

The author would like to thank Bartin University for all its support.

Competing interests

The authors declare that they have no competing interests.

References

- Nehlig, A., Daval, J. L., & Debry, G. (1992). Caffeine and the central nervous system: mechanisms of action, biochemical, metabolic and psychostimulant effects. *Brain research reviews*, 17(2), 139-170.
- [2] Camfield, D. A., Stough, C., Farrimond, J., & Scholey, A. B. (2014). Acute effects of tea constituents L-theanine, caffeine, and epigallocatechin gallate on cognitive function and mood: a systematic review and meta-analysis. Nutrition reviews, 72(8), 507-522.
- [3] Wood, S., Sage, J. R., Shuman, T., & Anagnostaras, S. G. (2014). Psychostimulants and cognition: a continuum of behavioral and cognitive activation. *Pharmacological reviews*, 66(1), 193-221.
- [4] Ribeiro, J. A., & Sebastiao, A. M. (2010). Caffeine and adenosine. Journal of Alzheimer's Disease, 20(s1), S3-S15.
- [5] Hillis DM, Sadava D, Hill RW, Price MV (2015). Principles of Life (2 ed.). Macmillan Learning. pp. 102–103.
- [6] Faudone, G., Arifi, S., & Merk, D. (2021). The medicinal chemistry of caffeine. Journal of Medicinal Chemistry, 64(11), 7156-7178.
- [7] Bhailume Meenal V. and Shinde Shubhangi R (2020) Biodegradation of Caffeine, International Journal of Science and Research, Vol 9, Issue 10.
- [8] Tzenios, N. (2023). Obesity as a risk factor for cancer. EPRA
- International Journal of Research and Development (IJRD), 8(2),
- 101-104.
- [9] Perone, F., Pingitore, A., Conte, E., Halasz, G., Ambrosetti, M., Peruzzi, M., & Cavarretta, E. (2023, March). Obesity and Cardiovascular Risk: Systematic Intervention Is the Key for Prevention. In Healthcare (Vol. 11, No. 6, p. 902). MDPI.
- [10] Jia, G., Fu, Y. E., Zhao, X. U., Dai, Q., Zheng, G., Yang, Y., ... & He, C. (2011). N 6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. Nature chemical biology, 7(12), 885-887.
- [11] Mauer, J., Sindelar, M., Despic, V., Guez, T., Hawley, B. R., Vasseur, J. J., ... & Jaffrey, S. R. (2019). FTO controls reversible m6Am RNA methylation during snRNA biogenesis. Nature chemical biology, 15(4), 340-347.
- [12] Gerken, T., Girard, C. A., Tung, Y. C. L., Webby, C. J., Saudek, V., Hewitson, K. S., ... & Schofield, C. J. (2007). The obesity-associated FTO gene encodes a 2-oxoglutaratedependent nucleic acid demethylase. Science, 318(5855), 1469-1472.
- [13] Shishodia, S., Demetriades, M., Zhang, D., Tam, N. Y., Maheswaran, P., Clunie-O'Connor, C., ... & Schofield, C. J. (2021). Structure-based design of selective fat mass and obesity associated protein (FTO) inhibitors. Journal of Medicinal Chemistry, 64(22), 16609-16625.
- [14] Gulati, P., Cheung, M. K., Antrobus, R., Church, C. D., Harding, H. P., Tung, Y. C. L., ... & Yeo, G. S. (2013). Role for the obesity-related FTO gene in the cellular sensing of amino acids. Proceedings of the National Academy of Sciences, 110(7), 2557-2562.
- [15] http://pubchem.ncbi.nlm.nih.gov/
- [16] Thompson MA. (2004). Molecular docking using ArgusLab, an efficient shape-based search algorithm and the AScoring function. ACS meeting Philadelphia 172, CINF 42, PA.

- [17] Peng C, Ayali PY, Schlegel HB and Frisch MJ, (1995) J. Comp Chem., 16: 49-51.
- [18] Cramer CJ and Truhlar DG(1992), Computer-Aided Mol. Design, 6: 629-666.
- [19] https://www.rcsb.org/
- [20] Martin YC (1998). Perspective in drug discovery and design. Springer Publisher, USA, 12th Volume, pp.3-23.
- [21] Cruciani G, Clementi S and Pastor M (1998). GOLPEguided region selection. Perspectives in
- Drug Discovery and Design, 12-14(16): 71-86.
- [22] Dunn III and Hopfinger AJ (1998). Drug Discovery, Kluwer Academic Publishers. Chapter 12, pp.167-182
- [23] Dewar MJS, Zoobisch EG, Healy EF and Stewart JJP (1985). AM1: A new general purpose quantum mechanical molecular model. J. Am. Chem. Soc. 107: 3902-3910.
- [24] James J. P. Stewart, J. (1989) Comp. Chem., 10, 209-220 and 221-264, (1989)
- [25] Thompson, M. A., Zerner M. C. J. Am. Chem. Soc., 113,
- [26] Mark A. Thompson, Eric D. Glendening, and David Feller (1994). J. Phys. Chem. 98, 10465-10476
- [27] Mark A. Thompson, and Gregory K. Schenter J. (1995). Phys. Chem. 99, 6374-6386,
- [28] Mark A. Thompson, (1996). J. Phys. Chem. 100, 14492-14507,
- [29] https://plip-tool.biotec.tu-dresden.de/plipweb/plip/result /47049a63-a21b-4f90-9705 2218ba8be383