

RESEARCH

In silico investigation of potential COVID-19-associated microRNA signatures

Potansiyel COVID-19 ilişkili mikroRNA'ların bilgisayar ortamında araştırılması

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Abstract

Purpose: The global pandemic COVID-19, caused by the coronavirus SARS-CoV-2, is persistent despite the increasing vaccination rates, with new cases being reported per week. MicroRNAs, that is, non-coding RNA species that regulate gene expression at the post-transcriptional level, play a pivotal role in the SARS-CoV-2 life cycle, pathophysiology and host's anticoronaviral responses. The objective of this study was the in silico discovery of functionally associated miRNAs that likely co-regulate COVID-19-related genes

Materials and Methods: In the present study, an integrative bioinformatics approach was employed, including database searching, gene set enrichment analysis, network-based and microRNA target prediction methods, towards the discovery of epigenetic determinants of COVID-19.

Results: An intricate microRNA-target gene network was constructed, and a set of 8 highly interacting microRNAs, that potentially co-target and co-regulate key COVID-19-related genes, was detected. These miRNAs and their corresponding genes are likely involved in the host's response to SARS-CoV-2 infection.

Conclusion: The 8 functionally associated miRNAs could constitute a signature for COVID-19 diagnosis.

Keywords: bioinformatics; COVID-19 infectionassociated genes; antiviral microRNAs; protein-protein interactions; gene set enrichment analysis; microRNAgene network

Öz

Amaç: Koronavirüs SARS-CoV-2'nin neden olduğu küresel COVID-19 pandemisi, artan aşılama oranlarına rağmen varlığını sürdürmekte ve her hafta yeni vakalar rapor edilmektedir. Transkripsiyon sonrası seviyede gen ekspresyonunu düzenleyen kodlamayan RNA türleri olan mikroRNA'lar, SARS-CoV-2 yaşam döngüsünde, patofizyolojisinde ve konağın antikoronaviral yanıtlarında çok önemli bir rol oynarlar. Bu çalışmada, COVID-19 ile ilişkili genleri düzenlemesi muhtemel, işlevsel olarak ilişkili miRNA'ları in silico araştırıp bulmak amaçlanmıştır.

Gereç ve Yöntem: Bu çalışmada, COVID-19'un epigenetik belirleyicilerini bulmaya yönelik veri tabanı araştırması, gen seti zenginleştirme analizi ve internet tabanlı mikroRNA'ya yönelik hedef tahmin yöntemlerini içeren bütünleştirici bir biyoinformatik yaklaşım kullanılmıştır.

Bulgular: Karmaşık bir mikroRNA-hedef gen ağı oluşturularak, potansiyel olarak hedeflenen ve COVID-19 ile ilgili önemli genleri düzenleyen yüksek düzeyde etkileşime sahip 8 mikroRNA'dan oluşan bir dizi tespit edildi. Bu miRNA'lar ve bunlara karşılık gelen genler SARS-CoV-2 enfeksiyonuna verilen yanıtta rol oynayabilir. **Sonuç:** İşlevsel olarak ilişkili 8 miRNA, COVID-19 tanısı için önemli bulgular olabilirler.

Anahtar kelimeler: biyoinformatik, COVID-19 enfeksiyonu ile ilişkili genler, antivirüs mikroRNA'lar, protein-protein etkileşimleri, gen seti zenginleştirme analiz,; mikroRNA-gen ağı

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INTRODUCTION

The ongoing global pandemic COVID-19, caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), persists despite the vaccination efforts, with thousands of COVID-19 cases and a great number of deaths to be reported weekly¹. Of note, the recently emerged SARS-CoV-2 variants BA.2.86 ('Pirola') and EG.5 (Eris) are highly infectious and transmissible strains²⁻⁵. Moreover, the growing incidence and prevalence, as well as the high transmissibility of the JN.1 strain, an offspring of BA.2.86, led WHO to declare it as a "variant of interest"6,7. SARS-CoV-2-mediated infection is characterized by activation of the innate immune response (the first line of immune defense) and hyperinflammation⁸⁻¹⁰. COVID-19 is associated with numerous complications and comorbidities, including common cold, pneumonia, blood coagulation, lymphopenia, acute respiratory distress syndrome (ARDS), tissue injury, multiple organ dysfunction, etc.¹¹⁻¹⁴.

MicroRNAs (miRNAs) are endogenous, short (22nucleotide) single-stranded non-protein-coding RNA molecules that can regulate gene expression at the post-transcriptional level through base-pairing with the 3' untranslated region (3'UTRs) of their target mRNAs; they subsequently suppress mRNA expression by mRNA degradation or translational inhibition. A single miRNA can potentially target multiple genes, and, conversely, a gene can be targeted by many miRNAs¹⁵⁻¹⁶.

It has been demonstrated that miRNAs play a critical role in the coronaviral life cycle, pathogenesis and host antiviral responses^{17,18}. SARS-CoV-2 infectioninduced changes in the expression patterns of the host miRNAs can lead to the down-regulation of key genes involved in the immune response and, consequently, to immunosuppression or attenuated host immune surveillance, thereby enabling the coronavirus to evade the host's immune system¹⁹⁻²¹. SARS-CoV-2 can also co-opt host miRNAs implicated in immune response suppression (e.g., hsa-miR-939 and hsa-miR-146b), in order to facilitate coronaviral replication and subvert host immune responses18,22. Moreover, miRNAs encoded by the coronaviral genome can target genes involved in host immune response/inflammation, like IFN1mediated signaling^{23,24}. SARS-CoV-2 non-coding RNAs can also deplete the host's miRNA pool, by acting as miRNA sponges^{25,26}.

In a recent study, Ahmad et al. (2023) suggest that the host miRNAs that are differentially regulated during COVID-19 infection can serve as biomarkers for prognosis in COVID-19, as well as the design of novel miRNA-based antiviral agents²⁷. According to Fayyad-Kazan and colleagues (2021), differentially expressed circulating miRNAs (e.g., miR-19a-3p, miR-19b-3p, and miR-92a-3p) are considered powerful biomarkers for the timely and accurate diagnosis of COVID-1928. In addition, deregulated plasma miRNAs were shown to have predictive and discriminatory potential for COVID-19 severity and mortality²⁹. In a study by Farr et al. (2021), it was shown that three differentially expressed miRNAs (miR-423-5p, miR-23a-3p and miR-195-5p) could accurately discriminate COVID-19 patients from healthy controls¹⁹. Furthermore, low expression of hsa-let7b-5p was detected in naso-oropharyngeal specimens from COVID-19 patients and it is implicated in the regulation of the angiotensinconverting enzyme 2 (ACE2) and dipeptidyl peptidase-4 (DPP4) receptors³⁰.

Although the co-regulatory effects of host miRNAs during SARS-CoV-2 infection have been studied fragmentarily, a comprehensive network analysis of the host COVID-19-relevant genes and miRNAs has not been conducted. Therefore, our study aims to contribute to the investigation of potential miRNA determinants of the epigenetic regulation of genes that are prominently associated with COVID-19, in order to decipher the co-regulation activities of miRNAs exerted upon those genes during coronavirus infection. This study hypothesizes that by employing a bioinformatics approach to construct a miRNA-gene network, we could provide a fundamental framework for detecting host genes and epigenetic regulators that likely respond during SARS-CoV-2 infection in a coordinated way.

MATERIALS AND METHODS

Acquisition of COVID-19-related genes

To obtain a comprehensive list of genes significantly linked to COVID-19, the GeneCards database (https://www.genecards.org/)^{31,32} (accessed March 2023) was searched by using the keywords "COVID-19" and "SARS-CoV-2"; in this way, the genes with relevance score \geq 9 were extracted. Asfa et al.

Protein-protein interaction network

The physical and functional associations among the protein products of the retrieved COVID-19-related genes were investigated and visualized through STRING (Search Tool for Retrieval of Interacting Genes/Proteins) v11.533, a knowledgebase of both experimentally supported or predicted, functional and/or physical, gene/protein association data derived from diverse resources. In order to enhance the reliability of the given interactions, a relatively high confidence interaction score (≥ 0.7) was set as cutoff; only those associations based on automated text mining, knowledge databases and experimental evidence were taken into consideration. The associations were further visualized and analyzed through the open-source platform Cytoscape (v.3.10.0)³⁴. Moreover, the Cytoscape plugin cytoHubba35, which allows network topological analysis by twelve local and global ranking algorithms (betweenness, bottleneck, clustering coefficient, closeness. degree, density of maximum neighborhood component, eccentricity, edge percolated component, maximum clique centrality, maximum neighborhood component, radiality, and stress), was utilized to select the top 26 nodes, by using the node-degree filter.

Functional enrichment analysis

Gene set enrichment analysis was conducted with the online tool WebGestalt (WEB-based GEne SeT AnaLysis Toolkit)36,37 for the identification of statistically significant over-represented biological pathways in the COVID-19 genes under study. The WebGestalt parameters selected were "Organism of Interest": Homo sapiens, "Method of Interest": Over-Representation Analysis (ORA), "Functional database": pathway/Reactome for biological paths, "Select gene ID type": gene symbol, "Select Reference set": genome; the default advanced parameters were chosen, and only those pathways with a Benjamini-Hochberg-adjusted *p*-value³⁸ ≤ 0.05 were included in the analysis. Affinity propagation was applied to reduce the terms and cluster them into representative categories.

miRNA regulators of COVID-19 genes

The miRNAs potentially regulating the COVID-19 genes were investigated. To this end, both, the experimentally verified and predicted miRNAs targeting the COVID-19 genes were obtained by

applying four state-of-the-art software tools: i) (http://diana.imis.athenamicroT_CDS innovation.gr/DianaTools/index.php?r=microT_C DS/index) is based on the DIANA-microT-CDS algorithm to predict miRNA target genes³⁹; ii) TargetScan (https://www.targetscan.org/vert_80/) searches for the presence of conserved sites matching the miRNA region⁴⁰; PITA seed iv) (https://genie.weizmann.ac.il/pubs/mir07/) recognizes thermodynamically favorable miRNAmRNA interactions⁴¹; iii) miRDB (http://mirdb.org/) predicts functionally annotated miRNA gene targets by machine learning approaches^{42,43}. The miRNA-mRNA interactions retrieved from each tool were combined and the duplicates were removed. To enhance the accuracy of the prediction, only those miRNA-gene targets predicted by more than three methods were considered in this study.

Pairwise miRNA associations

The miRNA-miRNA relationships were collected from two different sources, MiRGOFS⁴⁴ and GOSemSim⁴⁵, which infer functional similarities between miRNA pairs based on the degree of functional relatedness of their corresponding target genes. The pairwise miRNA interactions from both sources were merged, and the duplicates were removed. Only those miRNA-miRNA interactions with a weight score above 0.85 (where "1.0" is the highest score) were chosen so as to enhance robustness.

Independent validation

The findings of this study were further compared against an independent transcriptomic dataset, available in the comprehensive online resource COVID19db⁴⁶; the "differential expression" module was used, which provides gene expression profiling in whole blood derived from COVID-19 patients and healthy controls.

RESULTS

A total of 149 COVID-19-relevant genes were obtained from GeneCards (Table S1). A functional network of the products of those genes was generated, and 128 nodes appear to be highly interconnected (Figure 1). By examining the topological properties of the network, we identified those key nodes that are more relevant to the overall

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function of the network and, therefore, biologically meaningful⁴⁷⁻⁴⁸. The top 26 nodes were detected in the protein-protein interaction network (Figure 1 and Table S1) based on the combined output of the twelve algorithms in cytoHubba; scores generated by all algorithms were assigned to each node in the network and, then, the top-ranked nodes were scored based on the degree method (i.e. the number of direct connections of a given node to its neighbor nodes).

The terms over-represented in the genes, the products of which are involved in the protein association network (Figure 1), are mainly associated with immune signaling pathways, e.g., cytokine, interleukin, interferon, Toll-like receptor, MYD88, NF-kF, T-cell receptor (TCR)-mediated signaling cascades (Figure 2). Notably, 109 (out of 128) and all 26 top-ranking genes are found in the over-

represented Reactome pathways, further highlighting their prominent role in COVID-19 pathophysiology.

The differential expression profiles of several COVID-19-related top-26 genes (Table S1) between SARS-CoV-2-infected patients and healthy controls were further investigated through the "differential expression" module of COVID19db⁴⁶. Among those, the cytokines *IL10*, *IL18* and *CXC10* are upregulated in the COVID-19 group (Figure 3), consistent with their pro-inflammatory effects⁴⁹⁻⁵¹.

As shown in Figure 4, 22 of the top-ranking genes are potentially targeted by more than 8 miRNAs, namely hsa-miR-1276, hsa-miR-3121-3p, hsa-miR-338-5p, hsa-miR-340-5p, hsa-miR-5692a, hsa-miR-570-3p, hsa-miR-664a-3p and hsa-miR-7-5p.



Figure 1. Interaction network of the COVID-19 proteins. The nodes denote genes/gene products, and the edges represent functional associations. The nodes corresponding to the top-ranking gene products are indicated by light red fill color.

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Figure 2. Bar plot illustrating the over-represented Reactome pathways in the COVID-19 genes. The width of bar plots is proportional to the number of genes in each pathway.

DISCUSSION

Collectively, the proteins encoded by 128 of the 149 COVID-19-associated genes are strongly connected in an interaction network, suggesting physical and functional associations among the corresponding proteins. Most of the 26 key genes in the network (Figure 1 and Table S1) encode components of the immune system (CD4, CD8A, HLA-B, IRF3, MYD88 etc.), including pro- and anti-inflammatory factors, such as chemokines and cytokines (CXCL8, CXCL10, IL1B, IL2, IL4, IL6, IL10, IL17A, IL18, IFNG) and the tumor necrosis factor (TNF). In many COVID-19 cases, exacerbated inflammatory responses ("cytokine storm") are observed, which result from the acute increase in the levels of circulating pro-inflammatory cvtokines and chemokines, and their uncontrolled release, both at local and systemic levels⁵²⁻⁵⁴. Several studies, though,

suggest that the human host's immune system is rather compromised upon SARS-CoV-2 infection and it is not capable of eliciting a sufficient immune response^{55,56}.

The NF-kB subunits, NFKB1 and RELA (Table S1), are tightly connected to several pro-inflammatory agents. NF-kB can modulate the host's immediate innate immune response to SARS-CoV-2 infection. In particular, SARS-CoV-2-mediated activation of NF-kB was found to induce the expression of the genes *IL1*, *IL2*, *IL6*, *IL8* and *TNF*⁵⁷. Human miRNAs could attenuate the effects of the COVID-19-induced inflammatory cascade (e.g., organ injuries), by targeting and suppressing the gene coding for NFKB1¹⁷.

The effector gene OAS1 affects susceptibility to SARS-CoV-2 and has a protective effect against COVID-19 severity⁵⁸⁻⁶⁰. SARS-CoV-2 is suggested to bind and activate TLR4 (Toll-like receptor 4), a

sensor for innate immunity, leading to increased expression of ACE2, the major receptor for SARS-CoV-2 cell entry⁶¹, thereby facilitating coronavirus entry and amplifying the inflammatory response^{62,63}. CSF2 (colony-stimulating factor 2) was also found to be over-expressed in SARS-CoV-2-infected human lung epithelial cells⁶⁴.

Over-expression of the cytokine IL6 has been reported to be highly associated with the mortality risk of COVID-19; IL6 also acts as a perpetrator of the SARS-CoV-2-induced cytokine storm65-67. However, IL6 appears to be down-regulated in the blood of COVID-19 patients (Figure 3), consistent with the findings of a previous bioinformatics study by Özbek and colleagues (2023), wherein IL6 was not found to be significantly dysregulated in diverse SARS-CoV-2-infected tissues⁵⁵. This is probably due to the dual role of IL6 as a pro- and antiinflammatory cytokine68-69, as well as the diverse immune response patterns observed in COVID-19 patients^{55,70}. The pleiotropic anti-inflammatory cytokine IL471 is also down-regulated in COVID-19 blood samples (Figure 3), in agreement with previous studies suggesting cytokine storm suppression during SARS-CoV-2 infection55,56.

Increased expression of TLR4 greatly affects heart failure in COVID-19 patients63,72. TLR4 is known to trigger the activation of pro-inflammatory cytokines73, including TNF, which is also upregulated in COVID-19 (Figure 3); increased expression of TNF was found to be a prognostic factor for mortality among COVID-19 patients with comorbidities and disease progression74. MYD88, which plays a central role in TLR/IL1R-mediated signalling in innate and adaptive immunity¹¹, is overexpressed in the COVID-19 group (Figure 3); MYD88 polymorphisms were shown to be tightly associated with COVID-19 severity75. Finally, the expression level of the interferon-inducible gene OAS1 is markedly higher in the SARS-CoV-2 infected patients, as compared to the healthy samples; this is in agreement with the findings of a recent study, wherein SARS-CoV-2 infection significantly increased the expression of OAS176.

The causal genes of diseases/disorders usually share common regulatory mechanisms in order to ensure their coordinated regulation. Similarly, disease-related miRNAs act in a cooperative manner so as to exert their regulatory effect upon their corresponding target genes⁷⁷⁻⁷⁸. In our study, all eight miRNAs that target the top-22 genes are associated in a pairwise

fashion (Figure 4), suggesting participants in an intricate regulatory network, wherein these miRNAs act in a synergistic fashion to co-target and co-regulate the expression of COVID-19 genes.

There is evidence that several of those eight miRNAs are associated with SARS-CoV-2 and COVID-19. Vastrad and colleagues (2020) analyzed gene expression data from whole blood from human patients with COVID-19 and they found hsa-miR-5692a among those miRNAs that target genes that are up-regulated in COVID-19 samples as compared to healthy controls79. Moreover, hsa-miR-340-5p was down-regulated in the peripheral blood collected from COVID-19 patients⁸⁰. According to Katopodis et al. (2022), hsa-miR-3121-3p and hsa-miR-570-3p could regulate the expression of mediators of the SARS-CoV-2 cell entry81. Also, hsa-miR-1276 was shown to have the capacity to bind viral RNA and to be modulated significantly in pneumocytes, suggesting a role in SARS-CoV-2 infection⁸². Bartoszewsk et al. (2020) suggested potential SARS-CoV-2-mediated modulation of host miRNAs, including hsa-miR-664a-3p, in order to perpetuate coronaviral persistence/replication and evade host immune defences²⁵.

The limitations of this study are that the differential expression status of the predicted miRNAs, and their corresponding pairwise interactions, have not been validated experimentally in the context of COVID-19.

In conclusion, in this study, by applying an integrative bioinformatics pipeline and stringent criteria, we discovered eight functionally related miRNAs that potentially co-target and co-regulate pivotal COVID-19-associated genes. This panel of miRNAs could represent potential signature components for COVID-19, which can be taken into consideration in the clinical setting for updating and complementing the current biomarkers towards improving the accuracy of diagnosing SARS-CoV-2-infected patients.

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Ethical Approval: Ethics Committee Approval is not required for this article, since we have used publicly accessible data and resources for our analyses. In these cases, our institution does not require any Ethics Committee Approval.

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Figure 3. Differential gene expression analysis between COVID-19 (blue) and healthy control (golden brown) samples.



Figure 4. MiRNA-target gene network in COVID-19. The miRNAs are represented by polygons, and the miRNA target genes are denoted by circles.

REFERENCES

- 1. WHO Coronavirus (COVID-19) Dashboard. https://covid19.who.int/.
- Mahase E. Covid-19: New "Pirola" variant BA.2.86 continues to spread in UK and US. BMJ. 2023;382:2097.
- Satapathy P, Kumar P, Gupta JK, Rabaan AA, Al Kaabi NA, Mohanty D et al. The emergence and implications of SARS-CoV-2 omicron subvariant BA.2.86 on global health. Int J Surg. 2024.
- Abdolreza E, Fereshteh E, Armin JM, Amir S. EG.5 (Eris) and BA.2.86 (Pirola) two new subvariants of SARS-CoV-2: a new face of old COVID-19. Infection. 2024.
- Zhang L, Kempf A, Nehlmeier I, Cossmann A, Richter A, Bdeir N et al. SARS-CoV-2 BA.2.86 enters lung cells and evades neutralizing antibodies with high efficiency. Cell. 2024.
- Wang X, Lu L, Jiang S. SARS-CoV-2 evolution from the BA.2.86 to JN.1 variants: unexpected consequences. Trends Immunol. 2024;45:81-4.
- Khan SA, Bhuiyan MA, Dewan SMR. JN.1: The present public health concern pertains to the emergence of a novel variant of COVID-19. Environ Health Insights. 2024;18:11786302241228958.
- Diamond MS, Kanneganti TD. Innate immunity: the first line of defense against SARS-CoV-2. Nat Immunol. 2022;23:165-76.

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- Gustine JN, Jones D. Immunopathology of Hyperinflammation in COVID-19. Am J Pathol. 2021;191:4-17.
- Tufan A, Avanoglu Guler A, Matucci-Cerinic M. COVID-19, immune system response, hyperinflammation and repurposing antirheumatic drugs. Turk J Med Sci. 2020;50:620-32.
- Chen YM, Zheng Y, Yu Y, Wang Y, Huang Q, Qian F et al. Blood molecular markers associated with COVID-19 immunopathology and multi-organ damage. EMBO J. 2020;39:e105896.
- Krynytska I, Marushchak M, Birchenko I, Dovgalyuk A, Tokarskyy O. COVID-19-associated acute respiratory distress syndrome versus classical acute respiratory distress syndrome (a narrative review). Iran J Microbiol. 2021;13:737-47.
- Nalbandian A, Sehgal K, Gupta A, Madhavan MV, McGroder C, Stevens JS et al. Post-acute COVID-19 syndrome. Nat Med. 2021;27:601-15.
- Sanyaolu A, Okorie C, Marinkovic A, Patidar R, Younis K, Desai P et al. Comorbidity and its impact on patients with COVID-19. SN Compr Clin Med. 2020;2:1069-76.
- 15. Ambros V. The functions of animal microRNAs. Nature. 2004;431:350-5.
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell. 2004;116:281-97.
- Abedi F, Rezaee R, Hayes AW, Nasiripour S, Karimi G. MicroRNAs and SARS-CoV-2 life cycle, pathogenesis, and mutations: biomarkers or therapeutic agents? Cell Cycle. 2021;20:143-53.
- Arghiani N, Nissan T, Matin MM. Role of microRNAs in COVID-19 with implications for therapeutics. Biomed Pharmacother. 2021;144:112247.
- Farr RJ, Rootes CL, Rowntree LC, Nguyen THO, Hensen L, Kedzierski L et al. Altered microRNA expression in COVID-19 patients enables identification of SARS-CoV-2 infection. PLoS Pathog. 2021;17:e1009759.
- Yang CY, Chen YH, Liu PJ, Hu WC, Lu KC, Tsai KW. The emerging role of miRNAs in the pathogenesis of COVID-19: Protective effects of nutraceutical polyphenolic compounds against SARS-CoV-2 infection. Int J Med Sci. 2022;19:1340-56.
- Liang Y, Fang D, Gao X, Deng X, Chen N, Wu J et al. Circulating microRNAs as emerging regulators of COVID-19. Theranostics. 2023;13:125-47.
- Panda M, Kalita E, Singh S, Kumar K, Rao A, Prajapati VK. MiRNA-SARS-CoV-2 dialogue and prospective anti-COVID-19 therapies. Life Sci. 2022;305:120761.
- Khan MA, Sany MRU, Islam MS, Islam A. Epigenetic regulator mirna pattern differences among SARS-CoV, SARS-CoV-2, and SARS-CoV-2 world-wide isolates delineated the mystery behind the epic pathogenicity and distinct clinical characteristics of pandemic COVID-19. Front Genet. 2020;11:765.

- Singh M, Chazal M, Quarato P, Bourdon L, Malabat C, Vallet T et al. A virus-derived microRNA targets immune response genes during SARS-CoV-2 infection. EMBO Rep. 2022;23:e54341.
- Bartoszewski R, Dabrowski M, Jakiela B, Matalon S, Harrod KS, Sanak M et al. SARS-CoV-2 may regulate cellular responses through depletion of specific host miRNAs. Am J Physiol Lung Cell Mol Physiol. 2020;319:L444-L55.
- Li C, Wang R, Wu A, Yuan T, Song K, Bai Y et al. SARS-COV-2 as potential microRNA sponge in COVID-19 patients. BMC Med Genomics. 2022;15:94.
- Ahmad W, Gull B, Baby J, Panicker NG, Khader TA, Akhlaq S et al. Differentially-regulated miRNAs in COVID-19: A systematic review. Rev Med Virol. 2023;33:e2449.
- Fayyad-Kazan M, Makki R, Skafi N, El Homsi M, Hamade A, El Majzoub R et al. Circulating miRNAs: Potential diagnostic role for coronavirus disease 2019 (COVID-19). Infect Genet Evol. 2021;94:105020.
- Fernandez-Pato A, Virseda-Berdices A, Resino S, Ryan P, Martinez-Gonzalez O, Perez-Garcia F et al. Plasma miRNA profile at COVID-19 onset predicts severity status and mortality. Emerg Microbes Infect. 2022;11:676-88.
- 30. Latini A, Vancheri C, Amati F, Morini E, Grelli S, Matteucci C et al. Expression analysis of miRNA hsalet7b-5p in naso-oropharyngeal swabs of COVID-19 patients supports its role in regulating ACE2 and DPP4 receptors. J Cell Mol Med. 2022;26:4940-48.
- Rebhan M, Chalifa-Caspi V, Prilusky J, Lancet D. GeneCards: a novel functional genomics compendium with automated data mining and query reformulation support. Bioinformatics. 1998;14:656-64.
- 32. Stelzer G, Rosen N, Plaschkes I, Zimmerman S, Twik M, Fishilevich S et al. The genecards suite: from gene data mining to disease genome sequence analyses. Curr Protoc Bioinformatics. 2016;54:1.30.1-1.30.33.
- 33. Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S et al. The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. Nucleic Acids Res. 2021;49:D605-12.
- Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003;13:2498-504.
- Chin CH, Chen SH, Wu HH, Ho CW, Ko MT, Lin CY. cytoHubba: identifying hub objects and subnetworks from complex interactome. BMC Syst Biol. 2014;8:11.
- 36. Kirov S, Ji R, Wang J, Zhang B. Functional annotation of differentially regulated gene set using WebGestalt: a gene set predictive of response to

ipilimumab in tumor biopsies. Methods Mol Biol. 2014;1101:31-42.

- Liao Y, Wang J, Jaehnig EJ, Shi Z, Zhang B. WebGestalt 2019: gene set analysis toolkit with revamped UIs and APIs. Nucleic Acids Res. 2019;47:W199-W205.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. Journal of the Royal Statistical Society. Series B (Methodological). 1995;57:289-300
- Paraskevopoulou MD, Georgakilas G, Kostoulas N, Vlachos IS, Vergoulis T, Reczko M et al. DIANAmicroT web server v5.0: service integration into miRNA functional analysis workflows. Nucleic Acids Res. 2013;41:W169-73.
- Agarwal V, Bell GW, Nam JW, Bartel DP. Predicting effective microRNA target sites in mammalian mRNAs. Elife. 2015;4:e05005.
- Kertesz M, Iovino N, Unnerstall U, Gaul U, Segal E. The role of site accessibility in microRNA target recognition. Nat Genet. 2007;39:1278-84.
- Chen Y, Wang X. miRDB: an online database for prediction of functional microRNA targets. Nucleic Acids Res. 2020;48:D127-31.
- Liu W, Wang X. Prediction of functional microRNA targets by integrative modeling of microRNA binding and target expression data. Genome Biol. 2019;20:18.
- 44. Yang Y, Fu X, Qu W, Xiao Y, Shen HB. MiRGOFS: a GO-based functional similarity measurement for miRNAs, with applications to the prediction of miRNA subcellular localization and miRNA-disease association. Bioinformatics. 2018;34:3547-56.
- 45. Yu G, Li F, Qin Y, Bo X, Wu Y, Wang S. GOSemSim: an R package for measuring semantic similarity among GO terms and gene products. Bioinformatics. 2010;26:976-8.
- 46. Zhang W, Zhang Y, Min Z, Mo J, Ju Z, Guan W et al. COVID19db: a comprehensive database platform to discover potential drugs and targets of COVID-19 at whole transcriptomic scale. Nucleic Acids Res. 2022;50:D747-D57.
- 47. Barabasi AL, Gulbahce N, Loscalzo J. Network medicine: a network-based approach to human disease. Nat Rev Genet. 2011;12:56-68.
- Kontou PI, Pavlopoulou A, Dimou NL, Pavlopoulos GA, Bagos PG. Network analysis of genes and their association with diseases. Gene. 2016;590:68-78.
- Lauw FN, Pajkrt D, Hack CE, Kurimoto M, van Deventer SJ, van der Poll T. Proinflammatory effects of IL-10 during human endotoxemia. J Immunol. 2000;165:2783-9.
- Dinarello CA. Interleukin-18, a proinflammatory cytokine. Eur Cytokine Netw. 2000;11:483-6.
- Callahan V, Hawks S, Crawford MA, Lehman CW, Morrison HA, Ivester HM et al. The proinflammatory chemokines CXCL9, CXCL10 and CXCL11 are upregulated following SARS-CoV-2

Infection in an AKT-Dependent Manner. Viruses. 2021;13:1062.

- Coperchini F, Chiovato L, Croce L, Magri F, Rotondi M. The cytokine storm in COVID-19: An overview of the involvement of the chemokine/chemokinereceptor system. Cytokine Growth Factor Rev. 2020;53:25-32.
- Hu B, Huang S, Yin L. The cytokine storm and COVID-19. J Med Virol. 2021;93:250-56.
- Montazersaheb S, Hosseiniyan Khatibi SM, Hejazi MS, Tarhriz V, Farjami A, Ghasemian Sorbeni F et al. COVID-19 infection: an overview on cytokine storm and related interventions. Virol J. 2022;19:92.
- 55. Ozbek M, Toy HI, Takan I, Asfa S, Arshinchi Bonab R, Karakulah G et al. a counterintuitive neutrophilmediated pattern in COVID-19 patients revealed through transcriptomics analysis. Viruses. 2022;15:104.
- 56. Remy KE, Mazer M, Striker DA, Ellebedy AH, Walton AH, Unsinger J et al. Severe immunosuppression and not a cytokine storm characterizes COVID-19 infections. JCI Insight. 2020;5:e140329.
- Hariharan A, Hakeem AR, Radhakrishnan S, Reddy MS, Rela M. The role and therapeutic potential of nfkappa-b pathway in severe COVID-19 Patients. Inflammopharmacology. 2021;29:91-100.
- Asgari S, Pousaz LA. Human genetic variants identified that affect COVID susceptibility and severity. Nature. 2021;600:390-91.
- Huffman JE, Butler-Laporte G, Khan A, Pairo-Castineira E, Drivas TG, Peloso GM et al. Multiancestry fine mapping implicates OAS1 splicing in risk of severe COVID-19. Nat Genet. 2022;54:125-27.
- Zhou S, Butler-Laporte G, Nakanishi T, Morrison DR, Afilalo J, Afilalo M et al. A Neanderthal OAS1 isoform protects individuals of European ancestry against COVID-19 susceptibility and severity. Nat Med. 2021;27:659-67.
- Scialo F, Daniele A, Amato F, Pastore L, Matera MG, Cazzola M et al. ACE2: the major cell entry receptor for SARS-CoV-2. Lung. 2020;198:867-77.
- Aboudounya MM, Heads RJ. COVID-19 and Toll-Like Receptor 4 (TLR4): SARS-CoV-2 may bind and activate TLR4 to Increase ACE2 expression, facilitating entry and causing hyperinflammation. Mediators Inflamm. 2021;2021:8874339.
- 63. Mukherjee S. Toll-like receptor 4 in COVID-19: friend or foe? Future Virol. 2022;17:415–17.
- Chandrashekar DS, Athar M, Manne U, Varambally S. Comparative transcriptome analyses reveal genes associated with SARS-CoV-2 infection of human lung epithelial cells. Sci Rep. 2021;11:16212.
- 65. Santa Cruz A, Mendes-Frias A, Oliveira AI, Dias L, Matos AR, Carvalho A et al. Interleukin-6 Is a biomarker for the development of fatal severe acute respiratory syndrome coronavirus 2 pneumonia. Front Immunol. 2021;12:613422.

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- Chen LYC, Hoiland RL, Stukas S, Wellington CL, Sekhon MS. Confronting the controversy: interleukin-6 and the COVID-19 cytokine storm syndrome. Eur Respir J. 2020;56:2003006.
- Geronikolou SA, Takan I, Pavlopoulou A, Mantzourani M, Chrousos GP. Thrombocytopenia in COVID-19 and vaccine-induced thrombotic thrombocytopenia. Int J Mol Med. 2022;49:35.
- Borsini A, Di Benedetto MG, Giacobbe J, Pariante CM. Pro- and anti-inflammatory properties of interleukin (IL6) in vitro: relevance for major depression and for human hippocampal neurogenesis. Int J Neuropsychopharmacol. 2020;23:738-50.
- Scheller J, Chalaris A, Schmidt-Arras D, Rose-John S. The pro- and anti-inflammatory properties of the cytokine interleukin-6. Biochim Biophys Acta. 2011;1813:878-88.
- Galbraith MD, Kinning KT, Sullivan KD, Baxter R, Araya P, Jordan KR et al. Seroconversion stages COVID19 into distinct pathophysiological states. Elife. 2021;10:e65508.
- Chatterjee P, Chiasson VL, Bounds KR, Mitchell BM. Regulation of the anti-inflammatory cytokines interleukin-4 and interleukin-10 during pregnancy. Front Immunol. 2014;5:253.
- Choudhury A, Mukherjee S. Taming the storm in the heart: exploring different therapeutic choices against myocardial inflammation in COVID-19. Recent Adv Antiinfect Drug Discov. 2021;16:89-93.
- Swanson L, Katkar GD, Tam J, Pranadinata RF, Chareddy Y, Coates J et al. TLR4 signaling and macrophage inflammatory responses are dampened by GIV/Girdin. Proc Natl Acad Sci U S A. 2020;117:26895-906.
- Mohd Zawawi Z, Kalyanasundram J, Mohd Zain R, Thayan R, Basri DF, Yap WB. prospective roles of tumor necrosis factor-alpha (tnf-alpha) in COVID-19: prognosis, therapeutic and management. Int J Mol Sci. 2023;24:6142.

- Martinez-Gomez LE, Martinez-Armenta C, Medina-Luna D, Ordonez-Sanchez ML, Tusie-Luna T, Ortega-Pena S et al. Implication of myddosome complex genetic variants in outcome severity of COVID-19 patients. J Microbiol Immunol Infect. 2023;56:939-50.
- Assou S, Ahmed E, Morichon L, Nasri A, Foisset F, Bourdais C et al. The transcriptome landscape of the in vitro human airway epithelium response to SARS-CoV-2. Int J Mol Sci. 2023;24:12017.
- Zinani OQH, Keseroglu K, Ozbudak EM. Regulatory mechanisms ensuring coordinated expression of functionally related genes. Trends Genet. 2022;38:73-81.
- Arshinchi Bonab R, Asfa S, Kontou P, Karakulah G, Pavlopoulou A. Identification of neoplasm-specific signatures of miRNA interactions by employing a systems biology approach. PeerJ. 2022;10:e14149.
- Vastrad B, Vastrad C, Tengli A. Identification of potential mRNA panels for severe acute respiratory syndrome coronavirus 2 (COVID-19) diagnosis and treatment using microarray dataset and bioinformatics methods. 3 Biotech. 2020;10:422.
- Li C, Hu X, Li L, Li JH. Differential microRNA expression in the peripheral blood from human patients with COVID-19. J Clin Lab Anal. 2020;34:e23590.
- Katopodis P, Randeva HS, Spandidos DA, Saravi S, Kyrou I, Karteris E. Host cell entry mediators implicated in the cellular tropism of SARS-CoV-2, the pathophysiology of COVID-19 and the identification of microRNAs that can modulate the expression of these mediators (Review). Int J Mol Med. 2022;49:20.
- Milenkovic D, Ruskovska T, Rodriguez-Mateos A, Heiss C. Polyphenols could prevent SARS-CoV-2 infection by modulating the expression of mirnas in the host cells. Aging Dis. 2021;12:1169-82.