

Menba Kastamonu Üniversitesi Su Ürünleri Fakültesi Dergisi Menba Journal of Fisheries Faculty ISSN 2147-2254 | e-ISSN: 2667-8659

Menba Kastamonu Üniversitesi Su Ürünleri Fakültesi Dergisi 2024; 10(1): 83-89

change and growth of crayfish.

Derleme Makale/Review Article

The Importance of Calcium in Crayfish Farming

Abstract

Gülşen UZUN GÖREN^{1*}, Şennan YÜCEL¹

¹Sinop University, Fisheries Faculty, Department of Aquaculture, Sinop/Türkiye

*E-mail: gulsenuzn@hotmail.com

Article Info :

Received: 09/01/2024 Accepted: 24/04/2024

Keywords:

- Crayfish
- A. Leptodactylus
- Shell change
- Growth

Kerevit Yetiştiriciliğinde Kalsiyumun Önemi

<u>Makale Bilgisi:</u>	Öz
Geliş: 09/01/2024 Kabul Ediliş: 24/04/2024	Dünya çapında ekolojik ve ekonomik olarak önemli bir tür olan kerevitler (<i>Astacus leptodactylus</i> Eschscholtz, 1823) ülkemiz içinde ihraç ürünleri arasında yer alan iç sulardan elde edilen bir türdür. Türkiye'de kerevit üretimi doğal su kaynakları, baraj gölleri ve göletlerden yapılan avcılığa dayanmaktadır. Kerevitlerin büyümesinde en önemli elementlerden biri kalsiyumdur. Bu derlemenin amacı, kerevitlerin kabuk değişimi ve büyümesinde kalsiyumun önemi incelenerek, kerevit
Anahtar Kelimeler:	yetiştiriciliğinin gelişmesine katkı sağlamaktır.
 Kerevit A. Leptodactylus Kabuk değişimi Büyüme 	Atıf bilgisi/Cite as: Uzun Gören, G. & Yücel, Ş. (2024). The Importance of calcium in crayfish farming. Menba Journal of Fisheries Faculty, 10 (1), 83-89

Crayfish (Astacus leptodactylus Eschscholtz, 1823), an ecologically and economically important

species worldwide, is a species obtained from inland waters, which is among the export products in our

country. Crayfish production in Turkey is based on fishing from natural water sources, dam lakes, and

ponds. One of the most important elements in the growth of crayfish is calcium. This review aims to contribute to the development of crayfish farming by examining the importance of calcium in shell

INTRODUCTION

Crayfish are one of the crustaceans that live in freshwater throughout their lives. Under natural conditions, most of them are cytenohaline. Some crayfish species can survive in brackish waters as well as in low salinities (Susanto & Charmantier, 2000). Crayfish living in freshwater have a constant loss of ions in their bodies and an inflow of water through osmosis. Crayfish are hyperosmotic in freshwater at low salinity and isosmotic at high salinity (Susanto & Charmantier, 2000; Khodabandeh et al., 2005).

Calcium is one of the most important elements in crayfish growth. They spend almost all of their cuticular calcium during shell change. Therefore, they need a lot of calcium after shell change (Wheatley & Ayers, 1995; Wheatley & Gannon, 1995). Alpbaz (2005) states that the shell of crayfish consists of 40% calcium carbonate and 7% calcium phosphate. Many activities and feeding are reduced before shell change. When preparing feed for crayfish, it is thought that it can be accelerated by adding calcium to the feed. During the shell change process, calcium dissolved in the lower part of the shell is first extracted to increase the flexibility of the shell (Reynolds, 2002). For the new shell to form and harden, 10-20% calcium is needed, which is supplied from calcium stored in the stomach stones (Taugbøl et al., 1996). The dissolved calcium in the stomach stones is absorbed during shell replacement and is used for recalcification of shell sections after continued feeding. Only a small fraction of the calcium needed in stomach stones is stored, whereas the calcium needed after shell exchange is met by eating the discarded shell (Reynolds, 2002).

As mentioned above, calcium can also be obtained from feed (Aiken & Waddy, 1987). The availability and importance of exogenous calcium varies between species. After shell change, ionized calcium in water also helps to harden the shell (Malley, 1980; Taugbøl et al., 1996).

The most important function of the gills in crayfish is to regulate ion exchange between water and hemolymph (Wheatly &Gannon, 1995; Wheatly, 1999; Barradas et al. 1999). They have mechanisms to take up, assimilate and store calcium from the aquatic environment through their gills. Assimilation is reduced at low pH, in the absence of bicarbonate and at low temperature. Depending on nutrient requirements and after shell replacement, nutrient deficiency leads to reduced calcium assimilation (Greenaway, 1985). Calcium intake from different sources is important and a significant amount of calcium is consumed during shell formation (Malley, 1980). In this review, the importance of calcium in shell change and growth of crayfish was examined (Figure 1).



Figure 1. Crayfish (Astacus leptodactylus Eschscholtz, 1823)

The Importance Of Calcium In Shell Change And Growth Of Crayfish

The growth process of crayfish is characterized by the shedding of each old shell and the formation of a new one. In other words, crayfish growth is based on a series of shell changes. The molting event is the secretion of a new and larger shell that occurs before hardening. The soft stage, when the new skeleton develops, is a sensitive period for crayfish and they have a soft shell. If there is insufficient shelter to cope with the population density, there are losses due to cannibalism. The calcium needed during the hardening of the skeleton is supplied from the body or water. The molting activity is normal or slow depending on the calcium status. Calcium deficiency causes molting to proceed slowly. They also need calcium for the new shell to harden. As can be understood, the new shell uses calcium gradually as it forms. During the molting phase calcium is used for calcification and feeding is continued during this phase. Calcium stored in the gastrolith is used in small amounts as needed. During the hardening of the new shell, the body rapidly increases its mass by taking in water and air into its tissues. The crayfish then builds new tissues to fill its new expanding shell. Very rapid growth is achieved during this period. The main tissue increase in crayfish occurs during the intermolt stage (Lowery, 1988; Huner & Barr, 1991).

Calcium is very important in the formation of the shell. In this respect, the water for crayfish farming should be rich in calcium content (Alpbaz, 1993; Groves, 1985; Hunner, 1994).

Calcium ions in the water help hardening of the new shell. However, optimum calcium values for *A. leptodactylus* in crayfish populations are 50-100 mg/L, with 5-130 mg/L being the limit values (Köksal, 1988). Decreasing calcium concentration has a limiting effect on growth and reproduction, with the growth and reproduction of Astacus juveniles slowing down when the calcium content in the water is lower than 1 mg/L (Hessen et al., 1991; Rukke, 2002).

Freshwater crayfish are known to eat old shells that they discard from their bodies. On the other hand, when the calcium saturation level is 5 mg/L or less, freshwater crayfish cannot successfully form new shells because shell calcification is incomplete (James & Huner, 1985; Rukke, 2002). The literature states that the calcium level in water should be 2 mg/L for crayfish to survive. Some research results have shown that the amount of calcium in the water should be more than 10 mg/L for crayfish to survive. It has been reported that increasing the calcium concentration in the habitat increases the survival rate of crayfish and partially slows down shell change (Parkyn, 2002). Although calcium deficiency in the environment affects shell change, metabolic activity, and growth of crayfish, high calcium concentration in water can be toxic to crayfish (Winkler, 1986). Studies have shown that the calcium requirements of cold-water crayfish species are higher than those of temperate-water species (Berber & Mazlum, 2009; Mazlum, 2007).

Environmental factors such as water temperature, water quality (pH, calcium, and magnesium), light transmittance, photoperiod, and stocking density play an important role in crayfish growth (McClain et al., 1992; Nyström, 1994; Jover et al., 1999; Paglianti and Gherardi, 2004; Ramalho et al., 2008). The physicochemical properties of water are also important for crayfish. Calcium has a wide range of biological functions in crayfish life, such as growth and shell change, and has an important structural role in crayfish. They must obtain the calcium needed for the hardening of the new shell from water or feed (Hammond et al., 2006). Studies have shown that different calcium concentrations have effects on crayfish survival, growth and shell change (Holdich, 2002).

Although crayfish consumption in Turkey is not sufficient, they have a high economic value in foreign markets. Natural fishing in different geographical regions of Turkey is very rare. On the other hand, diseases in natural production cannot be intervened. In this sense, crayfish farming should be cultivated. Different fish feeds are used in crayfish farming for research purposes and trials are completed by adding different additives to these feeds for experimental purposes.

Taugbol et al. (1996) reported that the survival rate of crayfish was proportional to the increase in calcium concentration in the water, but partially decreased due to mortality caused by shell change. Hammond et al. (2006), in a study on *Paranephrops zealandicus*, reported that crayfish survival was proportional to the increase in the amount of calcium in the water (up to 10 mg/L), but above certain values of calcium in the water (e.g. 80 mg/L) the growth rate did not change. However, some studies have shown that neither high nor low calcium concentrations have a positive effect on growth.

In a related study, it was reported that although calcium deficiency in water affects shell change, metabolic activity and growth of crayfish, high calcium concentrations in water may have toxic effects on crayfish (Winkler, 1986). In similar studies on *Penaeus/Litopenaeus vannamei*, it was found that when Ca/Mg was 1:3, survival and growth rates increased with increasing calcium and magnesium ions, but when calcium concentration reached 30 mg/L, growth and survival rates decreased with increasing calcium and magnesium ions (Chen Ji Wang and Chen, 2004).

Calcium accumulates in the outer shell of many crustaceans, such as freshwater crayfish, which require sufficient calcium for each shell change (Rukke, 2002). Apparently, the calcium content of aquatic environments is important for the growth of freshwater crayfish (Hessen et al., 1989). When there is a calcium deficiency, the molting activity proceeds more slowly. Therefore, the soft-shell period lasts longer and crayfish are vulnerable to intense predator attacks. Cannibalism also increases during this period. Rapid growth and frequent shell changes increase cannibalism and predation. Hammond et al. (2006) showed that cannibalism in crayfish decreases with increasing calcium concentration in water.

Zahmetkesh et al. (2007), examined the growth and survival rates of *Astacus leptodactylus* juveniles by feeding them diets with different levels of calcium (0, 1, 2, 3, and 4%). As a result of the study, the best weight and length increases were observed in the group fed with 3-4% calcium, but there was no significant difference between these experimental groups. The highest growth (11.65 g) was observed in the group fed with 4% calcium. The lowest survival rate (30%) was observed in the group fed with 2% calcium. In this study, it was found that there was a significant increase in growth due to different concentrations of calcium compounds.

The lack of statistical difference between the experimental groups suggests that it is not correct to attribute the results only to calcium compounds. Because water temperature is the primary factor in shell change, survival, feeding, and growth of crayfish (Lowery, 1988; Whitledge & Rabeni, 2003). In general, low temperatures are recommended for the survival of juvenile crayfish and high temperatures are recommended for their rapid growth. Different researchers working with crayfish belonging to Astacid and other families have reported that although temperatures of 20°C or higher give the best results for crayfish growth, the survival rate of crayfish kept at 15°C or lower has increased (Westman et al., 1993; Gydemo & Westin, 1989; Mazlum 2007).

Taugbol et al. (1996) *Astacus leptodactylus* reported that 10-20% calcium is needed for the hardening of the new shell and storage of gastroliths. Another result obtained from this study was that gastroliths were found to be larger and heavier in soft-shelled crayfish that had recently changed their shells. One of the most important reasons for this is that new-shelled individuals cannot use gastroliths. Because during shell change, calcium carbonate in the stomach is brought back to dissolved form and used in the formation of the mouth and outer shell. 1/3 of the amount of calcium carbonate required for new shell development is supplied from the stomach, hepatopancreas, and blood (Taugbol et al., 1996; Holdich, 2002).

Cilbiz (2010) study in crayfish fry, 1.5%, 2.0%, 2.5%, 2.5%, and 3.0% calcium was added to the feed, and high levels of feeds containing calcium (2.5%-3.0%) were found to have better specific growth rate and feed conversion ratio.

Türel & Berber (2016) in the experiment, calcium carbonate was added to the feed at the rates of 1%, 3%, and 5%. Control calcium carbonate was not added to the group. 1% after 90 days of research, crayfish fry fed with feed containing 3% and 5% calcium carbonate (CaCO3) Although not significant, weight gain and specific growth rates were found to be higher. The best survival rate was obtained in feeds containing 5% calcium carbonate. Survival rates in the groups were found to be significantly higher than the control group (p < 0.05).

Recently, studies have shown that to maximize the growth of crayfish, they should also be fed with other feed sources high in protein (Uzun, 2007). In addition, it is important to add vitamins, minerals, antioxidants, and various other additives to the feed in fry production under artificial conditions (Öz, 2005). One of these additives is calcium. As it is known, the growth of crayfish involves many shell change processes. It follows the shell change. During this shell change, crayfish are exposed to calcium. They need calcium. Calcium deficiency in water can cause crayfish to change their shells, affecting metabolic activity and growth. However, high calcium can also have a toxic effect (Winkler, 1986). Therefore, the calcium content needs of crayfish of different sizes should be determined.

CONCLUSION

Calcium is one of the most important elements in the growth and development of crayfish. Research has shown that; it has been determined that dietary calcium is utilized in at least two ways in crayfish (Hessen et al., 1989).

These are expressed as the absorption of calcium through the digestive system wall and the intake of calcium into the body through the gills. Researchers have stated that calcium intake in crayfish increases with increasing calcium content in the feed.

Crayfish get some of the calcium they need from water. Research has shown that the amount of calcium they get from water is not always sufficient. In this case, the amount needed should be met by adding calcium to the feed.

Studies were carried out with 1%, 2%, 3%, and 4% calcium concentrations and it was found that the study with 4% concentration was effective on growth. However, the levels of effect on growth between the experimental groups were not statistically significant.

Determining the concentration ratio of statistically significant results by adding different concentrations of calcium to the feeds will be important for the culturing of crayfish.

In the trials to be conducted, crayfish trials of different sizes and species should be established and the ideal calcium ratios for each size should be determined. Cultivation should be encouraged by determining feed rations suitable for different geographical conditions of the country.

REFERENCES

- Aiken, D.E., & Waddy, S.L., (1987). Moulting and Gowth in Crayfish: A Review. Canadian
- Technical Report of Fisheries and Aquatic Sciences. No:1587. Canada.
- Alpbaz, A., (1993). Crustacean and Arthropod Breeding, Ege University, Faculty of Fisheries Publications, Izmir.
- Alpbaz, A., (2005). Aquaculture, Alp Publications, 549s, Izmir.
- Barradas, C., Dunel-Erb, Lignon, J., & Pequeux, A., (1999). Superimposed morphofunctional
- study of ion regulation and respiration in single gill filaments of the crayfish Astacus leptodactylus. Journal of Crustacean Biology, 19, 14-25.
- Berber, S., & Mazlum, Y., (2009). Reproductive efficiency of the narrow-clawed crayfish,
- Astacus leptodactylus, in several populations in Turkey. Crustaceana, 82 (5), 531-542.
- Chen, C.S., Ji, D.H., Wang, X.B., & Chen, Z.Q., (2004). Effects of Ca2+ and Mg2+ on survival
- and growth of Penaeus vannamei. Journal of Fisheries of China, 28, 413-418.
- Cilbiz, M., (2010). Feeding freshwater lobster with feeds containing different calcium content
- (Astacus leptodactylus) on growth, survival rate and shell change, Master's Thesis, Institute of Science, Isparta.
- Greenaway, P., (1985). Calcium balance and moulting in Crustacea. Biological Reviews, 60
- (3), 425-454.
- Groves, R.E., (1985). The Crayfish: Its nature and nurture. Fishing New Boks Ltd., England. 72p.
- Gydemo, R., & Westin, L., (1989). Growth and survival of juvenile Astacus astacus L. at
- optimized water temperature, in: De Pauw, N. et al. Aquaculture: a biotechnology in progress, 383-391.
- Hammond, K.S., Hollows, J.W., Townsend, C.R., & Lokman, P.M., (2006). Effects of
- temperature and water calcium concentration on growth, survival and molting of freshwater crayfish, *Paranephrops zealandicus*. Aquaculture, 251, 271-279.
- Hessen, D.O., Agerberg, A., Kjellberg, G., Odelstrom, T., & Westman, K., 1989. Food,
- Nutrition Growth, Reproduction and Genetics. In: Crayfish Culture in Europe. Report from the Workshop on Crayfish culture, 39-48.
- Hessen, D.O., Kristiansen, G., & Lid, I., 1991. Calcium uptake from food and water in the
- crayfish Astacus astacus (L. 1758), measured by radioactive 45CA (Decapoda, Astacidea)., Crustaceana, 60 (1), 76-83.
- Holdich, D.M. (2002). General biology-background and functional morphology. in: biology of
- freshwater crayfish (edited by: Holdich, D.M.). pp: 3-30.
- Huner, J.V., & Barr, J.E., (1991). Red swamp crawfish: biology and exploitation. The
- Louisiana Sea Grant College Program, Center for Wetland Resources. Louisiana State University, Baton Rouge.
- Hunner, J.V., (1994). Freshwater Crayfish Aquaculture in North America, Europe and Australia: Families Astacidae, Cambaridae and Parastacidae. 312 p.
- James, W.A.Jr., & Huner, J.V., (1985). Freshwater prawns, In: Huner, J.V. and Brown, E.E.
- (Eds), Crustasean and Mollusk Aquaculture in the United States, Avi publish Company, Inc., Wesport, Connecticut, 1-54.
- Jover, M., Fernandez-Carmona, J., Del Rio, M.C. & Soler, M., (1999). Effect of feeding
- cooked-extruded diets, containing different levels of protein, lipid and carbohydrate on growth of red swamp crayfish (*Procambarus clarkii*). Aquaculture, 178, 127-137.
- Khodabandeh, S., Charmantier, G., Blasco, C., Grousset, E., & Charmantier, D.M., (2005).
- Ontogeny of the antennal glands in the crayfish *Astacus leptodactylus* (Crustacea, Decapoda): anatomical and cell differentiation. Cell and Tissue Research, 319, 153-165.
- Köksal, G., (1988). A. leptodactylus in Europe. In: Freshwater Crayfish: Biology, Management

and Exploitation (eds D.M. Holdich and R.S. Lowery), Croom Helm Press, pp. 365-400.

- Lowery, R.S., (1988). Growth, moulting and reproduction. In: Holdich, D. M., Lowery, R.S.
- (Eds.). Freshwater Crayfish: Biology, Management and Exploitation. Chapman and Hall, London, 83-113.
- Malley, D.F., (1980). Decreased survival and calcium uptake by the crayfish O. virilis in low
- pH. Canadian Journal of Fisheries and Aquatic Sciences, 37, 364-372.
- Mazlum, Y., (2007). Effects of temperature on the survival and growth of two cambarid crayfish
- juveniles. Crustaceana, 80(8), 947-954.
- McClain, W.R., Neil W.H. & Gatlin III D.M. (1992). Partitioning the contributions of forage-
- based production system components to weight gain of juvenile crayfish (Procambarus clarkii). Aquaculture, 101, 267-281.
- Nyström, P. (1994) Survival of juvenile signal crayfish (Pacifastacus leniusculus) in relation to
- light intensity and density. Nordic Journal of Freshwater Research, 69, 162-166.
- Öz, B., (2005). Keban Baraj Gölü'nde Yaşayan Tatlısu Istakozu (Astacus leptodactylus
- Esch.1823) Rasyonuna Farklı Oranlarda İlave Edilen Vitamin E'nin Etkileri, Doktora Tezi, Fırat Üniversitesi, Fen Bilimleri Enstitüsü, Elazığ,
- Paglianti, A., & Gherardi, F. (2004) Combined effects of temperature and diet on growth and
- survival of young of year crayfish: a comparison between indigenous and invasive species. Journal of Crustacean Biology, 24, 140-148.
- Parkyn, S., (2002). Farmed koura-a new niche for an old favourite. In: http://www.crayfishworld.com/nz.htm, 03/06/2002.
- Ramalho, R.O., Correia, A.M., & Anastacio P.M., (2008). Effects of density on growth and
- survival of juvenile red swamp crayfish, *Procambarus clarkii* (Girard), reared under laboratory conditions. Aquaculture Research, 39, 577-586.
- Reynolds, J.D., (2002). Gowth and reproduction. In: Holdich, D.M. (Ed.), Biology of
- Freshwater Crayfish. Blackwell Science, 152-163, United Kingdom.
- Rukke, N.A., (2002). Effects of low calcium consentration an two common freshwater
- crusteceans, Gammarus lacustris and Astacus astacus. Functional Ecology, Vol. 16, 161-169.
- Susanto, G.N., & Charmantier, G., (2000). Ontogeny of osmoregulation in the crayfish Astacus
- leptodactylus. Physiological and Biochemical Zoology, 73 (2), 169-176.
- Taugbol, T., Waevagen, S.B., Linlokken, A.N., & Skurdal, J., (1996). Post-molt exoskeleton
- mineralization in adult noble crayfish, *Astacus astacus*, in three lakes with different calcium levels. Freshw. Crayfish 11, 219-226.
- Türel, S., Berber, S. (2016). The effects of calcium supplemented diets on growth performance
- of crayfish (Eschscholtz, 1823) Adıyaman University Journal of Science ADYUSCI 6 (1):96-109.
- Uzun, C., (2007). Farklı stok yoğunluğunun juvenil tatlısu kerevitlerinin (Astacus
- *leptodactylus*, Eschscholtz, 1823) büyümeleri, yem değerlendirme oranları ve hayatta kalma oranları üzerine etkileri, Yüksek Lisans Tezi, Mustafa Kemal Üniversitesi, Fen Bilimleri Enstitüsü, Hatay,
- Westman, K., Savolainen, R., & Pursiainen, M., (1993). A comparative study on the growth
- and moulting of the noble crayfish, *Astacus astacus* (L) and the signal crayfish *Pacifastacus leniusculus* (Dana), in a small forest lake in southern Finland. Freshwater Crayfish, 9, 451-465
- Wheatly, M.G., & Gannon A.T., (1995). Ion regulation in crayfish: freshwater adaptations and
- the problem of moulting. American Zoologist, 35, 49-59.
- Wheatly, M.G., (1999). Calcium homeostasis in crustacea: the evolving role of branchial, renal,
- digestive and hypodermal epithelia. Journal of Experimental Zoology. 283, 620-640.

Wheatly, M.G., & Ayers, J., (1995). Scaling of calcium, inorganik contents and organic

contents to body mass during the moulting cycle of the fresh-water crayfish *Procambarus clarkii* (Girard). Journal of Crustacean Biology. 15, 409-417.

Whitledge, G.W., & Rabeni, C.F., (2003). Maximum daily consumption and respiration rates

at four temperatures for five species of crayfish from Missouri U.S.A (Decapoda,

Orconectes spp.). Crustaseana, 75(9), 1119-1132.

- Winkler, A., (1986). Effects of inorganic seawater constituents on branchial Na K ATPase
- activity in the shore crab Carcinus maenas. Marine Biology 92, 537-544.
- Zahmetkesh, A., Poorreza, J., Abedian, A., Shariatmadari, F., Valipoor, A., & Karimzadeh, K.,
- (2007). Effects of different levels of calcium on growth criteria and survival of freshwater crayfish, *Astacus leptodactylus*. Journal of Science and Technology of Agriculture and Natural Resources, 11 (40), 385-397.