



Screening of Primary and Secondary Metabolites Profile of Different Extracts of *Cassia fistula* Flowers and its Perspectives on the Antimicrobial and Antidiabetic Potential of Active Extract

Nivetha Arunprakash^{1*} , Christina Ruby Stella¹ , Angel Praba¹ ,
Vijayavilas Sathyan Sangeetha² 

¹Holy Cross College, Bharathidasan University, Department of Chemistry, Tiruchirappalli, India

²Dhanalakshmi Srinivasan College of Arts and Science for Women, Department of Chemistry, Perambalur, India.

Abstract: This article provides an overview of the pharmacological activity and phytochemical constituents found in *Cassia fistula* flower extract. Some phytonutrients and antioxidants of plant extract may be compromised due to the relatively high concentration of organic solvents. The main goal of this study was to find out how well an aqueous extract from *Cassia fistula* flowers fights diabetes and germs in a lab setting. The maceration extract yielded the best results among others. The antimicrobial activity of the extract was evaluated using bacterial and fungal strains, while the anti-diabetic activity was assessed at different concentrations, ranging from 10 to 50 µg/mL. The maximum inhibition of 54.07% was observed at a concentration of 50 µg/mL, compared to the standard acarbose, which showed an inhibition of approximately 62.23%. The IC₅₀ value of the flower extract was 49.3494 µg/mL, while that of the standard acarbose was 42.1726 µg/mL. The presence of phytochemicals in the extract was determined using UV and FTIR spectral studies, which are effective methods for extracting a broad range of chemical constituents. The results show that *Cassia fistula* flower extract has promising anti-diabetic and antimicrobial activity. It can be concluded that *Cassia fistula* has high alpha-amylase inhibitory activity, and these extracts have the potential to be effective antimicrobial agents against pathogenic microorganisms.

Keywords: Phytochemicals, *Cassia fistula*, Anti-microbial, Anti-diabetic, FTIR.

Submitted: February 14, 2024. **Accepted:** January 18, 2025.

Cite this: Arunprakash N, Stella CR, Praba A, Sangeetha VS. Screening of Primary and Secondary Metabolites Profile of Different Extracts of *Cassia fistula* Flowers and its Perspectives on the Antimicrobial and Antidiabetic Potential of Active Extract. JOTCSA. 2025;12(2): 77-84.

DOI: <https://doi.org/10.18596/jotcsa.1436181>

***Corresponding author's E-mail:** nivethaarun1717@gmail.com

1. INTRODUCTION

The present study aims to evaluate the antidiabetic effect of flowers of *Cassia fistula*. *Cassia fistula* is a small tree or shrub found in many western countries like Indonesia, Vietnam, Thailand, and India. Traditional folk medicine has a long history of utilizing all parts of the *Cassia fistula* plant. Studies have quantitatively evaluated the plant's antioxidant content, which has a direct impact on regulating blood sugar levels. Diabetes, a chronic and serious disease that affects people worldwide, poses a significant risk to the function of major organs such as the heart and kidneys. As a result, it is considered a dangerous illness (2,3). Currently, over 500 million people worldwide are affected by type 1 and type 2 diabetes mellitus. Medications, including insulin, are

critical for people with diabetes to survive. The high cost of treatment and the serious nature of the disease have led to a growing awareness of the need for efficient, economical, and safe substitute medications (4).

Researchers have investigated the use of herbs and isolated compounds, such as glucosides, alkaloids, and steroids, to treat different stages of diabetes. Due to its greater accessibility, the focus of research has been on plant-based alternative medicines (5). In vitro and animal tests have been conducted to evaluate the effectiveness of plants and their extracts in regulating blood sugar levels. Given the rising number of people affected by diabetes in recent years, it is essential to explore alternative remedies. Investigating the effectiveness of

medicinal plants known to be useful in many traditional systems of medicine is a promising approach to discovering new antidiabetic drugs (6,7).

To address the critical issue of growing bacteria resistance against conventional antibiotics, it is essential to find novel antimicrobial chemicals or extracts. Biodiversity in plant chemicals is a rich potential resource. The flowers were studied with polar and non-polar solvents like hexane, chloroform, methanol, and ethanol, which exhibit good antibacterial activity, especially for gram-positive bacteria. Hence, the floral part of the *Cassia fistula* can be used for good antimicrobial drug formation (8). The pod of *Cassia fistula* was evaluated for its antidiabetic activity in rats with various concentrations of ethanolic extract, which exhibited a significant lowering of the blood sugar level. It is well recognized that the plant components of *Cassia fistula* constitute a significant source of secondary metabolites, particularly phenolic compounds (9,10). The current investigation examines the various techniques for extracting *Cassia fistula* flowers and assesses the antimicrobial and antidiabetic properties of the resulting active extract.

2. EXPERIMENTAL SECTION

2.1. Materials

Ferric chloride, glacial acetic acid, ammonia, Dragendroff's reagent, ethanol, hydrochloric acid, sulfuric acid, sodium hydroxide, and double distilled water were purchased from SRL chemicals and Merck. All chemicals utilized in the research were of superior quality and high purity, with a minimum purity level of $\geq 99.0\%$. The flowers of *Cassia fistula* were collected from Trichy district, India, in May.

2.2. Preparation of Flower Extracts

The fresh flowers of *Cassia fistula* were collected and dried at room temperature. To prepare the extract, the dried flower samples were ground into a fine powder. Each of the four portions of the 200g flower sample was extracted using maceration, digestion, infusion, and decoction methods with 1000 mL of double-distilled water. The methods chosen have been devised to achieve effectiveness in removing a wide range of chemical constituents. These techniques facilitate both heating and heatless extraction that enable the isolation of compounds with varying solubility. The maceration extraction method involved soaking the flower sample (200g) in 1000 mL of double distilled water for 72 hours, with periodic shaking and subsequent filtration for further use. For the digestion method, 200g of the flower sample was mixed with 1000 mL of double distilled water and heated in a water bath at 50°C for 30 minutes, followed by filtration of the extract for further studies. The infusion method involved the same solvent-to-sample ratio as the maceration process, with a shorter soaking time of up to 4 hours, followed by filtration. Finally, the decoction method entailed heating 200 g of the flower sample with 1000 mL of double distilled water continuously for 30 minutes, with the concentrated extract then filtered for further analysis. (5,11).

2.3. Phytochemical Screening of Plant Extracts

The qualitative evaluation of phytoconstituents in the four different extracts was carried out using some standard procedures. The most crucial chemical analyses are those that examine the specific physiological advantages of the phytochemicals. Four extracts were examined qualitatively for the presence of such significant phytochemicals as flavonoids, alkaloids, phenolic compounds, tannin, terpenoids, saponin, carbohydrates, and steroids; outcomes were compared (7,12).

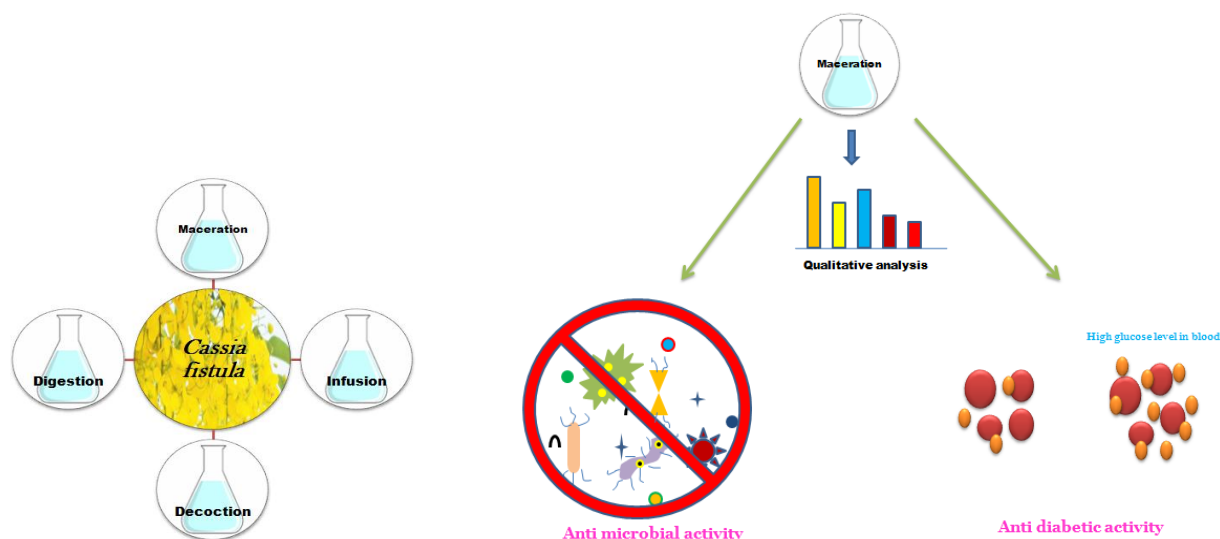


Figure 1: Schematic diagram of antimicrobial and antidiabetic activity of *Cassia fistula*.

2.4. Characterization of Flower Extract

The best outcomes of the phytochemical constituents were selected from the screening, and the extract obtained by the maceration technique was further

analyzed. The macerated extract of *Cassia fistula* was subjected to spectral analysis, like UV-Vis and Fourier transform infrared spectroscopy. The aqueous extract of *Cassia fistula* was studied by a

UV-Vis spectrometer (Perkin Elmer, Lambda 35 model) with a wavelength range of 200–800 nm (13,14). The flower extract was subjected to the FTIR spectrometer (Perkin Elmer, range 4000–400 cm^{-1}) for the functional groups (15).

2.5. Antimicrobial Activity of Flower Extract

The agar well diffusion method was used to assess the flower extract's antibacterial and antifungal properties. The microbial test employed bacterial and fungal strains of *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Candida albicans*, and *Aspergillus niger*. Gentamicin was used as a control for these antimicrobial tests. The strains were subcultured in the nutrient broth, which was prepared just before the test. The nutritional agar medium was prepared, and the petri plates were prepared with wells (16,17). Each well was treated with the flower extract at a different concentration. The inhibition rate was calculated using the diameter of the zone of inhibition formed by the flower extract (18).

2.6. Antidiabetic Activity of Flower Extract

200 μL of alpha-amylase solution was mixed with various concentrations of aqueous flower extract (10, 20, 30, 40, and 50 μL) and incubated for 10 mins (19). 200 mL dinitrosalicylic acid reagent was prepared using 12 g of sodium potassium tartrate

tetrahydrate in 8.0 mL of 2 M NaOH and 20 mL of 96 mM of 3,5-dinitrosalicylic acid solution, and the mixture was boiled, cooled and diluted for the absorbance of 540 nm. Acarbose was used as the standard reference. The inhibition rate and IC_{50} values were calculated (20).

3. RESULTS AND DISCUSSION

The occurrence of phytochemicals in the four different extracts was studied. The maceration method of extraction was found to be the most effective, as it is a low-tech technique that requires minimal equipment. This method does not involve heat, which may contribute to the preservation of bioactive compounds. In contrast, the decoction method, which involves continuous heating, is commonly used for extracting compounds from tough plant materials. This method is particularly useful for extracting less soluble compounds. The majority of the phytochemicals were present in all of the extracts since polar solvents were used. In comparison to other extracts, the maceration extraction process produced significantly better results. Some phytochemicals were absent or showed mild occurrence in the digestion and infusion processes, which may be due to the heating process of the plant extract.

Table 1: Phytochemical screening of aqueous flower extract of *Cassia fistula*.

S.No.	Phytochemicals	Extraction method			
		Maceration	Infusion	Digestion	Decoction
1	Flavonoid	+++	++	++	++
2	Alkaloid	+++	+	+	++
3	Phenols	++	-	+	+
4	Tannin	++	-	+	+
5	Terpenoid	++	+	+	+
6	Saponin	-	+	-	+
7	Carbohydrates	+++	++	++	++
8	Steroid	++	++	+	+

(+ : slightly present, ++ : moderately present, +++ : strongly present, - : absent)



Figure 2: Flower of *Cassia fistula*.

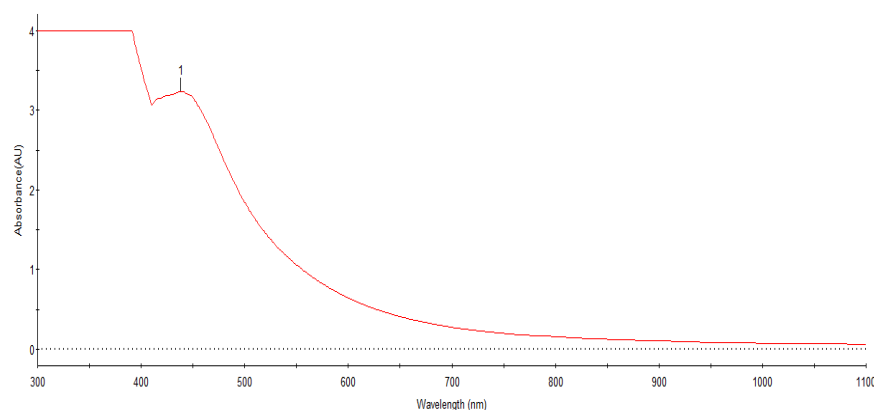


Figure 3: UV-Vis spectrum of flower extract of *Cassia fistula*.

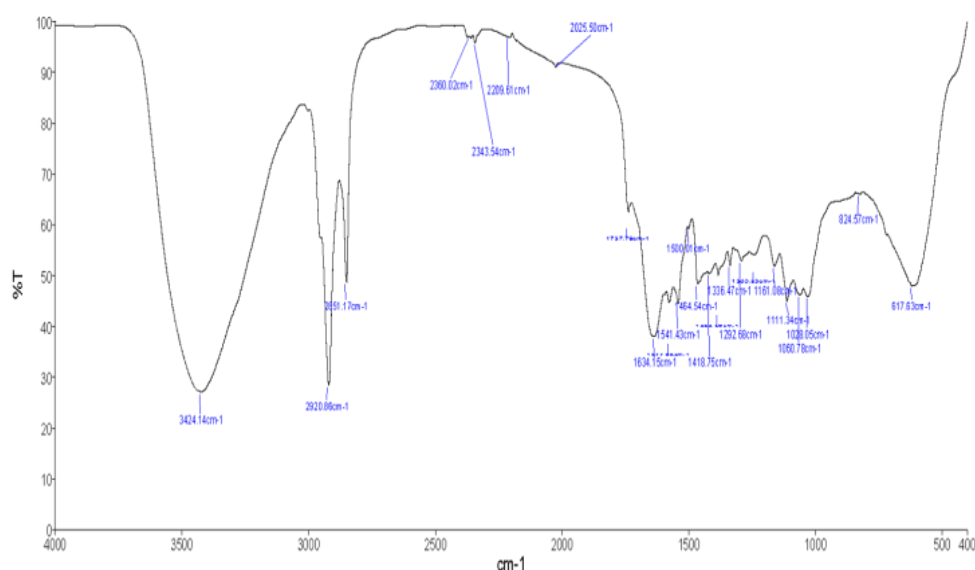


Figure 4: FTIR spectrum of flower extract of *Cassia fistula*.

The extract made using the maceration and decoction methods included significant amounts of flavonoids and phenolic substances. These phytochemicals have antioxidant properties, supporting several therapeutic qualities (7,39). Flavonoids are one of the major antioxidant agents present in most plant species. It is significantly present in maceration and decoction extracts. This plays a vital role in antimicrobial and antidiabetic activity. This secondary metabolite is widely studied for anticancer and anti-inflammatory activity (21,22). Many flavonoids were successfully isolated, and their structures were identified. The polyphenolic compounds and functional groups are responsible for the chelation of metals by scavenging free radicals and acting as the best antioxidant agents (12,36).

All different kinds of extracts contain substantial amounts of tannin. Typically, tannin is found in the tree's bark, protecting it from microbes. As a result, tannins have built-in antibacterial properties. These specific tannins, which were extracted from plant samples, have antiviral and antibacterial properties (7,21,35). Previous studies back up the claim that it slows tumor growth, which has a positive impact on numerous cancer types, like breast and lung cancer. Steroid content in *Cassia fistula* flower extracts is noticeable. These phytosterols are very beneficial for

lowering cholesterol and blood sugar. Important phytosterols like stigmasterol and sitosterol have reduced the absorption of cholesterol (23,27,34). The maceration technique-adapted extract had the highest concentration of phytoconstituents of all the extracts. Hence, this extract was used for further antimicrobial and antidiabetic studies. The process of digestion necessitates high temperatures, which may cause the degradation of heat-sensitive compounds, resulting in the loss of biological activity. Furthermore, this extraction method exhibits reduced efficiency in extracting non-polar compounds. The infusion method displays limited extraction efficiency and possesses a brief shelf-life with aqueous extracts, allowing for microbial growth and oxidation. The decoction technique may cause the evaporation of volatile compounds in the extract, which are responsible for numerous medicinal properties. Each of these methods has its unique advantages and disadvantages, and the study assists in selecting the most appropriate extraction procedure for further research based on the specific requirements of biological studies.

The *Cassia fistula* flower extract was subjected to UV-Vis spectroscopy. The spectrum gives the details of σ and n bonds, chromophores, and other lone pairs of electrons (13,28). Hence, this information can help

identify the phytochemicals present in the plant extract. The aqueous extract shows absorption at a wavelength range of 438 nm, which indicates the presence of unsaturated groups (35,40). The peaks formed may be due to the presence of tannin, flavonoids, and carotenoids. This absorption range indicates the $\pi - \pi^*$ transition in the aromatic ring (24,29).

The FTIR spectrum of the flower extract of *Cassia fistula* is shown in figure 4. The well-defined absorbance peaks indicate the presence of many alkyl and alkane groups, as well as some hydroxyl groups. The absorption peaks at 3424 cm^{-1} and 2920 cm^{-1} indicated stretching of the hydroxyl group and symmetric stretching of saturated compounds in the extract. The peak that appears at 2209 cm^{-1} indicates the alkyne group. The peaks at 1541 and 1292 cm^{-1} may be due to the $\text{C}=\text{C}$ stretching and $\text{C}=\text{O}$ stretching (15,30). Hence, this FTIR spectrum proved the presence of phenolic compounds, aromatic compounds, amines, and alkanes in the aqueous extract of *Cassia fistula* flowers.

The aqueous extract of *Cassia fistula* flower was tested at various concentrations (25, 50, 75, and 100 μL) in gram-positive, gram-negative, and fungal strains. The microorganisms chosen for this study were selected based on their clinical relevance, specifically *Staphylococcus aureus* and *Escherichia coli*, which are the most common causes of skin infections and pneumonia, respectively, and are known to exhibit heavy antibiotic-resistant strains. Furthermore, these microorganisms represent a

diverse range of organisms, including both gram-negative and gram-positive bacteria, as well as fungal strains, which allows for a broader understanding of the antimicrobial activity of *Cassia fistula* flowers. These microorganisms are ideal for studying antimicrobial resistance mechanisms and can be easily cultivated with well-defined growth requirements. Collectively, the pathogens responsible for a wide range of human diseases are these microorganisms and studying them can provide valuable insights into treatment strategies for various types of infections. The growth of *Staphylococcus aureus* bacteria and *Aspergillus niger* fungi was inhibited better than that of other microorganisms. On increasing the concentration of the flower extract, the zone of inhibition increased. The *Bacillus subtilis* and *Pseudomonas aeruginosa* have less inhibition at 100 μL of the extract when compared to other microorganisms. Urinary tract infections caused by *Escherichia coli* can be treated with the *Cassia fistula* flower extract (25,31,33). Hence, the extract can be used as an alternative antimicrobial medicine and for further drug discovery studies. This kind of plant-based alternative medicine has been well appreciated in recent years for its less toxic and more effective antimicrobial behavior. Lethal diseases like pneumonia were caused by *Bacillus subtilis*, which can be treated by the *Cassia fistula* flower extract (17,32,41). Natural antioxidant agents like flavonoids and phenolic compounds may be responsible for the good antimicrobial activity of the *Cassia fistula* extract, which could be a suitable alternative to synthetic medicine.

Table 2: Antimicrobial activity of aqueous extract of *Cassia fistula* flower.

Microorganisms	Concentration of extract and zone of inhibition (mm/mL)				
	25 μL	50 μL	75 μL	100 μL	Control
<i>Bacillus subtilis</i>	14	16	19	22	20
<i>Staphylococcus aureus</i>	16	18	21	25	20
<i>E. coli</i>	15	18	20	23	20
<i>Pseudomonas aeruginosa</i>	15	18	20	22	25
<i>Candida albicans</i>	15	18	21	24	23
<i>A. Niger</i>	16	19	22	26	23

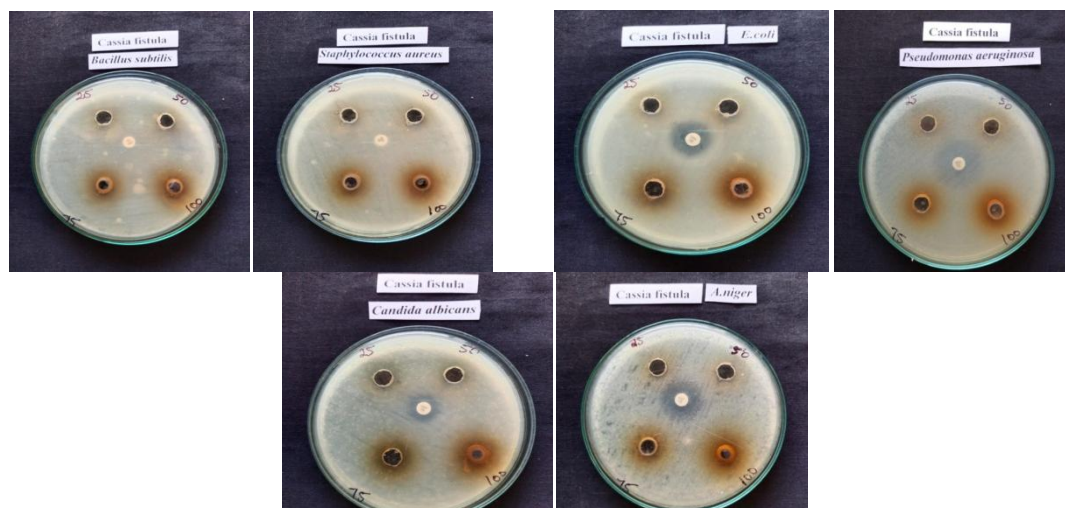
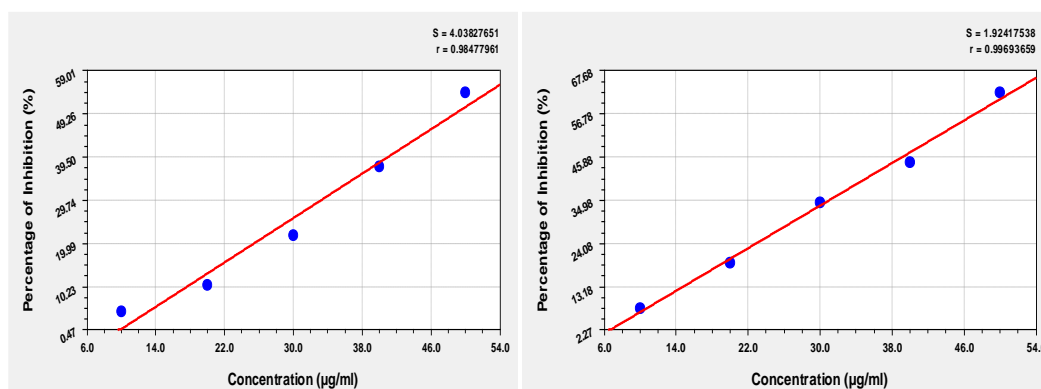


Figure 5: Antimicrobial activity of flower extract of *Cassia fistula*.

Table 3: Antidiabetic activity of aqueous extract of *Cassia fistula* flower.

Volume of the extract (µg/mL)	Absorbance at 540 nm		Percentage inhibition (%)	
	Standard	Extract	Standard	Extract
10	0.215	0.222	7.7253	4.7210
20	0.188	0.208	19.3133	10.7296
30	0.153	0.182	34.3347	21.8884
40	0.129	0.146	44.6351	37.3390
50	0.088	0.107	62.2317	54.0772

**Figure 6:** Graph of concentration of flower extract/standard and percentage of inhibition.

Various concentrations of *Cassia fistula* flower extract were evaluated for their antidiabetic activity using the α -amylase enzyme inhibition method. An *in vitro* α -amylase assay was carried out using 10, 20, 30, 40, and 50 µg/mL of *Cassia fistula* flowers and acarbose as a standard drug. The aqueous extract of *Cassia fistula* flower exhibits an inhibitory activity of α -amylase as IC_{50} 49.3494 µg/mL, whereas acarbose exhibits an IC_{50} 42.1726 µg/mL. Maximum inhibition of 54.07% was found at a concentration of 50 µg/mL of extract, which is compared with standard acarbose, which shows 62.23%. The IC_{50} value serves as a crucial parameter for assessing the efficacy of substances in biological functions. The IC_{50} value obtained for *Cassia fistula* extract is considered decent, indicating a high potency to inhibit 50% achievement at a lower concentration. While IC_{50} is a valuable metric, it is only one aspect of an extract's profile. It should be evaluated in conjunction with other factors to assess its therapeutic potential for antidiabetic treatment fully. Therefore, the obtained IC_{50} value for *Cassia fistula* flower is essential for understanding the extract's potency, effectiveness, and potential for development into therapeutic agents. Diabetes can be treated by reducing the production and absorption of glucose in the body. This can be done through the inhibition of enzymes like α -amylase, which helps digest carbohydrates. The results show that a significant amount of flower extract inhibits the α -amylase enzymes; hence, it can be used for the therapeutic approach to diabetes mellitus (37,38). The flowers of *Cassia fistula* could be used as an alternative natural antidiabetic agent as the disease is genetic, and a high number of cases have been recorded in recent years. Future research could focus on isolating and characterizing bioactive compounds from *Cassia fistula* extract and elucidating their mechanism of action. Additionally, optimizing the

extraction method and formulating *Cassia fistula* for pharmacological uses could be explored.

4. CONCLUSION

Cassia fistula flower contains a significant source of bioactive compounds. The traditional medicinal system is becoming increasingly apparent as a global concern. Because of its low toxicity and widespread usage of its therapeutic properties, *Cassia fistula* could be one of them. UV and IR characterization of the aqueous extract of *Cassia fistula* flowers was used to predict the absorbance pattern and functional present groups. The antimicrobial activity of the *Cassia fistula* flower extract shows a maximum zone of inhibition against tested microbes, especially against fungi. The findings show that the antidiabetic activity of the aqueous extract reveals high α -amylase inhibitory activity. It can be used as a green medicine for diabetes mellitus. It can be concluded that the aqueous extract of *Cassia fistula* flower can be used as a better alternative for treating diabetes mellitus and is also effective for pathogenic disorders.

5. CONFLICT OF INTEREST

The authors declare that they have no conflict of interest in this work.

6. ACKNOWLEDGMENTS

The authors are thankful to the Holy Cross College management for providing facilities to conduct this research work.

7. REFERENCES

1. Abubakar A, Haque M. Preparation of medicinal plants: Basic extraction and fractionation procedures

for experimental purposes. J Pharm Bioallied Sci [Internet]. 2020 Jan 1;12(1):1–10. Available from: [<URL>](#).

2. Ambadiang MMM, Atontsa BCK, Tankeo SB, Nayim P, Wamba BEN, Bitchagno GTM, et al. Bark extract of *Cassia sieberiana* DC. (Caesalpiniaceae) displayed good antibacterial activity against MDR gram-negative phenotypes in the presence of phenylalanine-arginine β -naphthylamide. BMC Complement Med Ther [Internet]. 2020 Dec 12;20(1):342. Available from: [<URL>](#).

3. Capetti F, Cagliero C, Marengo A, Bicchi C, Rubiolo P, Sgorbini B. Bio-Guided Fractionation Driven by In Vitro α -Amylase Inhibition Assays of Essential Oils Bearing Specialized Metabolites with Potential Hypoglycemic Activity. Plants [Internet]. 2020 Sep 21;9(9):1242. Available from: [<URL>](#).

4. Chinnathambi A, Alharbi SA, Joshi D, V S, Jhanani GK, On-uma R, et al. Synthesis of AgNPs from leaf extract of *Naringi crenulata* and evaluation of its antibacterial activity against multidrug resistant bacteria. Environ Res [Internet]. 2023 Jan 1;216:114455. Available from: [<URL>](#).

5. Daisy P, Saipriya. Biochemical analysis of *Cassia fistula* aqueous extract and phytochemically synthesized gold nanoparticles as hypoglycemic treatment for diabetes mellitus. Int J Nanomedicine [Internet]. 2012 Mar 7;7:1189–202. Available from: [<URL>](#).

6. Dedvisitsakul P, Watla-iad K. Antioxidant activity and antidiabetic activities of Northern Thai indigenous edible plant extracts and their phytochemical constituents. Heliyon [Internet]. 2022 Sep 1;8(9):e10740. Available from: [<URL>](#).

7. Duraipandian V, Ignacimuthu S. Antibacterial and antifungal activity of *Cassia fistula* L.: An ethnomedicinal plant. J Ethnopharmacol [Internet]. 2007 Jul 25;112(3):590–4. Available from: [<URL>](#).

8. Grover M, Behl T, Sehgal A, Singh S, Sharma N, Virmani T, et al. In vitro phytochemical screening, cytotoxicity studies of *Curcuma longa* extracts with isolation and characterisation of their isolated compounds. Molecules [Internet]. 2021 Dec 11;26(24):7509. Available from: [<URL>](#).

9. Gutiérrez-Grijalva E, Picos-Salas M, Leyva-López N, Criollo-Mendoza M, Vazquez-Olivo G, Heredia J. Flavonoids and phenolic acids from oregano: Occurrence, biological activity and health benefits. Plants [Internet]. 2017 Dec 26;7(1):2. Available from: [<URL>](#).

10. Hameed B, Ali Q, Hafeez MM, Malik A. Antibacterial and antifungal activity of fruit, seed and root extracts of *Citrullus colocynthis* plant. Biol Clin Sci Res J [Internet]. 2020;2020(1):33. Available from: [<URL>](#).

11. Jain PK, Soni A, Jain P, Bhawsar J. Phytochemical analysis of *Mentha spicata* plant extract using UV-VIS, FTIR and GC/MS technique. Available Journal Chem Pharm Res [Internet]. 2016;8(2):1–6.

Available from: [<URL>](#).

12. Jangir RN, Jain GC. Evaluation of antidiabetic activity of hydroalcoholic extract of *Cassia fistula* Linn. pod in streptozotocin-induced diabetic Rats. Pharmacogn J [Internet]. 2017 Jul 15;9(5):599–606. Available from: [<URL>](#).

13. Jani DK, Goswami S. Antidiabetic activity of *Cassia angustifolia* Vahl. and *Raphanus sativus* Linn. leaf extracts. J Tradit Complement Med [Internet]. 2020 Mar 1;10(2):124–31. Available from: [<URL>](#).

14. Karpagasundari C, Kulothungan Sjj. Analysis of bioactive compounds in *Physalis minima* leaves using GC MS, HPLC, UV-VIS and FTIR techniques. J Pharmacogn Phytochem. 2014;3(4):196–201.

15. Kifle ZD, Abdelwuhab M, Melak AD, Genet G, Meseret T, Aduagna M. Pharmacological evaluation of medicinal plants with antidiabetic activities in Ethiopia: A review. Metab Open [Internet]. 2022 Mar 1;13:100174. Available from: [<URL>](#).

16. Mechchate H, Es-safi I, Louba A, Alqahtani AS, Nasr FA, Noman OM, et al. In vitro α -amylase and α -glucosidase inhibitory activity and in vivo antidiabetic activity of *Withania frutescens* L. foliar extract. Molecules [Internet]. 2021 Jan 8;26(2):293. Available from: [<URL>](#).

17. Moteriya P, Padalia H, Chanda S. Characterization, synergistic antibacterial and free radical scavenging efficacy of silver nanoparticles synthesized using *Cassia roxburghii* leaf extract. J Genet Eng Biotechnol [Internet]. 2017 Dec 1;15(2):505–13. Available from: [<URL>](#).

18. Nandhini S, Geethalakshmi S, Selvam S, Radha R, Muthusamy P. Preliminary phytochemical and anti-diabetic activity of *Cassia sophora* Linn. J Pharmacogn Phytochem. 2016;5(1):87–91.

19. Oirere EK, Anusooriya P, Raj CA, Gopalakrishnan VK. Phytochemical analysis of N-hexane leaf extract of *Alpinia purpurata* (Vieill.) K. Schum using Uv-Vis, FTIR and GC-MS. Int J Pharm Pharm Sci [Internet]. 2015;7(8):387–9. Available from: [<URL>](#).

20. Oliveira ESC, Acho LDR, Morales-Gamba RD, do Rosário AS, Barcellos JFM, Lima ES, et al. Hypoglycemic effect of the dry leaf extract of *Myrcia multiflora* in streptozotocin-induced diabetic mice. J Ethnopharmacol [Internet]. 2023 May 10;307:116241. Available from: [<URL>](#).

21. Pham DQ, Pham HT, Han JW, Nguyen TH, Nguyen HT, Nguyen TD, et al. Extracts and metabolites derived from the leaves of *Cassia alata* L. exhibit *in vitro* and *in vivo* antimicrobial activities against fungal and bacterial plant pathogens. Ind Crops Prod [Internet]. 2021 Aug 1;166:113465. Available from: [<URL>](#).

22. Rehana D, Mahendiran D, Kumar RS, Rahiman AK. In vitro antioxidant and antidiabetic activities of zinc oxide nanoparticles synthesized using different plant extracts. Bioprocess Biosyst Eng [Internet]. 2017 Jun 30;40(6):943–57. Available from: [<URL>](#).

23. Paulraj MS, Rajkumar RJ, Thanapaul S. Phytochemicals as a potential source for anti-microbial, anti-oxidant and wound healing - a review. *MOJ Bioorganic Org Chem* [Internet]. 2018;2(2):61–70. Available from: [<URL>](#).
24. Tanveer S, Latif A, Ashiq K, Qayyum M, Bajwa MA. A comprehensive review on pharmacological and phytochemical potential of *Cassia fistula* Linn: A Magical Herb. *IJBPA* [Internet]. 2019;8(6):1134–57. Available from: [<URL>](#).
25. Solikhah TI, Setiawan B, Ismukada R. Antidiabetic Activity of Papaya Leaf Extract (*Carica Papaya* L.) Isolated with Maceration Method in Alloxan-Induces Diabetic Mice. *Syst Rev Pharm* [Internet]. 2020;11(9):774–8. Available from: [<URL>](#).
26. Stevens-Barrón JC, de la Rosa LA, Wall-Medrano A, Álvarez-Parrilla E, Rodríguez-Ramírez R, Robles-Zepeda RE, et al. Chemical composition and *in vitro* bioaccessibility of antioxidant phytochemicals from selected edible nuts. *Nutrients* [Internet]. 2019 Sep 27;11(10):2303. Available from: [<URL>](#).
27. Mwalimu RJ, Packirisamy ASB. Phytochemical assessment and evaluation of the anti-oxidant, anti-microbial, anti-cholesterol, and anti-diabetic activities of *Triumfetta Pentandra* methanol based green leaf extract. *J Inorg Organomet Polym Mater* [Internet]. 2023 Dec 30;33(12):3779–93. Available from: [<URL>](#).
28. Jangid R, Jain S, Sharma MK, Chatterjee S. In vitro antioxidant and antidiabetic activity of ethanolic extract of *Prosopis* species growing in Rajasthan, India. *Vegetos* [Internet]. 2022 Sep 28;36(1):62–9. Available from: [<URL>](#).
29. Deepthi K, Renjith PK, Shameem K, Habeeb Rahman K, Chandramohanakumar N. Phytochemical screening of leaves and flower extracts of *Sesbania grandiflora* (L.) Pers. and its antimicrobial activity against fish pathogens. *Vegetos* [Internet]. 2022 Aug 4;36(2):626–33. Available from: [<URL>](#).
30. Shekwa W, Maliehe TS, Masoko P. Antimicrobial, antioxidant and cytotoxic activities of the leaf and stem extracts of *Carissa bispinosa* used for dental health care. *BMC Complement Med Ther* [Internet]. 2023 Dec 15;23(1):462. Available from: [<URL>](#).
31. Oloya B, Namukobe J, Ssengooba W, Afayoa M, Byamukama R. Phytochemical screening, antimycobacterial activity and acute toxicity of crude extracts of selected medicinal plant species used locally in the treatment of tuberculosis in Uganda. *Trop Med Health* [Internet]. 2022 Dec 17;50(1):16. Available from: [<URL>](#).
32. KC BM, Gauchan DP, Khanal SN, Lamichhane J. Phytochemical screening, antioxidant and antibacterial activity of bamboo leaf collected from agroecosystem of the Central Siwalik region, Nepal. *Vegetos* [Internet]. 2023 Nov 28;37(6):2600–6. Available from: [<URL>](#).
33. Sousa BCM de, Gomes D do A, Viana AF da S, Silva BA da, Barata LES, Sartoratto A, et al. Phytochemical analysis and antioxidant activity of ethanolic extracts from different parts of *Dipteryx punctata* (S. F. Blake) amshoff. *Appl Sci* [Internet]. 2023 Aug 24;13(17):9600. Available from: [<URL>](#).
34. Tavanappanavar AN, Mulla SI, Shekhar Seth C, Bagewadi ZK, Rahamathulla M, Muqtader Ahmed M, et al. Phytochemical analysis, GC–MS profile and determination of antibacterial, antifungal, anti-inflammatory, antioxidant activities of peel and seeds extracts (chloroform and ethyl acetate) of *Tamarindus indica* L. *Saudi J Biol Sci* [Internet]. 2024 Jan 1;31(1):103878. Available from: [<URL>](#).
35. Kattil A, Hamid, Dash KK, Shams R, Sharma S. Nutritional composition, phytochemical extraction, and pharmacological potential of mulberry: A comprehensive review. *Futur Foods* [Internet]. 2024 Jun 1;9:100295. Available from: [<URL>](#).
36. Azmat F, Safdar M, Ahmad H, Khan MRJ, Abid J, Naseer MS, et al. Phytochemical profile, nutritional composition of pomegranate peel and peel extract as a potential source of nutraceutical: A comprehensive review. *Food Sci Nutr* [Internet]. 2024 Feb 8;12(2):661–74. Available from: [<URL>](#).
37. Kasali FM, Kadima JN, Peter EL, Mtewa AG, Ajayi CO, Tusiimire J, et al. Antidiabetic medicinal plants used in Democratic Republic of Congo: A critical review of ethnopharmacology and bioactivity data. *Front Pharmacol* [Internet]. 2021 Oct 27;12:757090. Available from: [<URL>](#).
38. Shahzad N, Alzahrani AR, Aziz Ibrahim IA, Shahid I, Alanazi IM, Falemban AH, et al. Therapeutic strategy of biological macromolecules based natural bioactive compounds of diabetes mellitus and future perspectives: A systematic review. *Heliyon* [Internet]. 2024 Jan 30;10(2):e24207. Available from: [<URL>](#).
39. Iglesias-Guevara D, Sánchez-Torres P. Characterization of antifungal properties of avocado leaves and majagua flowers extracts and their potential application to control *Alternaria alternata*. *Int J Food Microbiol* [Internet]. 2024 Mar 2;413:110579. Available from: [<URL>](#).
40. Chau TP, Devanesan S, Ayub R, Perumal K. Identification and characterization of major bioactive compounds from *Andrographis paniculata* (Burm. f.) extracts showed multi-biomedical applications. *Environ Res* [Internet]. 2024 Feb 1;242:117763. Available from: [<URL>](#).
41. Oke-Altuntas F, Ipekcioglu S, Sahin Yaglioglu A, Behcet L, Demirtas I. Phytochemical analysis, antiproliferative and antioxidant activities of *Chrozophora tinctoria*: A natural dye plant. *Pharm Biol* [Internet]. 2017 Jan 5;55(1):966–73. Available from: [<URL>](#).