

## Glioblastoma stem cells: a therapeutic challenge

Erdoğan Pekcan ERKAN<sup>1,2,3</sup>, Ufuk VURGUN<sup>1,2</sup>, Reşat Serhat ERBAYRAKTAR<sup>4</sup>, Zübeyde ERBAYRAKTAR<sup>5,\*</sup>

<sup>1</sup>Izmir Biomedicine and Genome Center, Dokuz Eylül University, İzmir, Turkey

<sup>2</sup>Department of Neuroscience, Institute of Health Sciences, Dokuz Eylül University, İzmir, Turkey

<sup>3</sup>PharmaPlus Laboratories, PharmaPlus İlaç ve Sağlık Ürünleri Ltd. Şti., İzmir, Turkey

<sup>4</sup>Department of Neurosurgery, Faculty of Medicine, Dokuz Eylül University, İzmir, Turkey

<sup>5</sup>Department of Biochemistry, Faculty of Medicine, Dokuz Eylül University, İzmir, Turkey

Received: 26.08.2015 • Accepted/Published Online: 17.11.2015 • Final Version: 08.11.2016

**Abstract:** The outcome for glioblastoma patients remains extremely poor, despite the advances in surgical and medical fields. It is hypothesized that glioblastoma progression, as well as tumor recurrence, is driven by a small number of cells called cancer stem cells (CSCs), which are characterized by their ability of self-renewal and proliferation, giving rise to progeny of transformation into multiple neuroepithelial lineages. Understanding the biology of CSCs is likely to explain why existing treatment strategies fail to affect the relatively quiescent and resistant CSC compartment. Here, we review the current knowledge on CSCs in glial tumors. In addition, we discuss the importance of the CSC hypothesis in the advancement of therapies for brain tumors.

**Key words:** Glioblastoma, cancer stem cell, therapy, resistance

### 1. Introduction

Glioblastoma is considered as the most aggressive primary brain tumor and has an extremely poor prognosis. The median 5-year survival rate is less than 3%, which makes this disease a devastating condition for both patients and their caregivers. Resistance to available therapies and recurrence are common in most cases; identification and molecular characterization of cancer stem cells (CSCs) (Singh et al., 2003; Yuan et al., 2004) have shown that these cells are responsible, in part, for resistance, as well as tumor reformation.

In this review, we will try to summarize the recent advances in glioblastoma biology, with a special focus on glioblastoma CSCs (GSCs). We will also discuss the molecular features of GSCs, and how these features can be exploited as potential therapeutic strategies.

### 2. Glioblastoma

#### 2.1. Background and epidemiology

Gliomas are the most common primary brain tumors (approximately 80% of all cases). Glioblastoma in particular is the most common and aggressive form of glioma (Omuro and DeAngelis, 2013). Glioblastoma is classified as WHO grade IV astrocytoma according to the World Health Organization (WHO) classification of brain tumors. The estimated incidence of brain and nervous

system tumors is 240,000 cases per annum. According to the Central Brain Tumor Registry of the United States (CBTRUS) report in 2013, the incidence of glioblastoma is 3.19/100,000. and the median age at diagnosis is 64 years (Ostrom et al., 2013; Thakkar et al., 2014).

The tumor is generally localized in the forebrain (cerebrum). In most cases, tumor formation occurs spontaneously. However, several genetic and epidemiological risk factors have been identified, including increased age, exposure to high-dose radiation, and history of genetic disorders (e.g., Li–Fraumeni syndrome, Turcot's syndrome, retinoblastoma, and neurofibromatosis 1 and 2), allergies, ionizing radiation, and occupational exposure to chemicals (e.g., pesticides, solvents) (Schwartzbaum et al., 2006; Ostrom et al., 2014). In addition, while symptoms vary between patients depending on tumor size and localization, the common symptoms include increased intracranial pressure, visual impairment, seizures, sensory loss, mood/personality changes, and impaired cognitive function (Wen and Kesari, 2008; Ostrom et al., 2014). Magnetic resonance imaging (MRI) is performed in suspected cases, and definitive diagnosis is made after pathological examination of the biopsy specimen.

Glioblastoma patients have poor prognosis, and the 5-year survival rate after diagnosis is less than 5% (Ostrom et al., 2013; Smoll et al., 2013). Long-term survival in

\* Correspondence: zubeyde.erbayraktar@gmail.com

glioblastoma is associated with several parameters, including younger age, lower Ki-67, hypermethylation of the O-6-methylguanine-DNA methyltransferase (*MGMT*) promoter, and mutations in the *IDH1* and *IDH2* genes (Scott et al., 1999; Krex et al., 2007; Yan et al., 2009; Hartmann et al., 2010).

## 2.2. Molecular subtypes of glioblastoma

In 2010, Verhaak and colleagues integrated mRNA expression profiles from different platforms and discovered four distinct molecular subtypes (Verhaak et al., 2010). They demonstrated that the so-called classical, mesenchymal, proneural, and neural subtypes are characterized by individual genetic signatures. The classical subtype is characterized by chromosome 7 amplification (especially *EGFR* gene amplification) paired with chromosome 10 loss, lack of *TP53* mutations, focal 9p21.3 deletion (on the *CDKN2A* locus), and high expression levels of *NES*, Notch, and Sonic hedgehog (*SHH*) signaling pathway elements (Verhaak et al., 2010). The mesenchymal subtype is characterized by focal deletion of 17q11.2 (*NF1* locus), expression of previously described mesenchymal markers (*CHI3L1* and *MET*), and high levels of TNF and NF- $\kappa$ B pathway elements (Verhaak et al., 2010). The proneural subtype is characterized by mutations in the *IDH1* and *PDGFRA* genes, which are not commonly seen in other subtypes (Verhaak et al., 2010). Integrated pathway analysis of genomic alterations in glioblastoma has shown that the PI(3)K/MAPK pathway, p53 pathway, and Rb pathway are the most affected signaling pathways (Brennan et al., 2013).

## 2.3. Standard of care

The current standard of care consists of gross total resection (GTR), followed by radiotherapy and adjuvant chemotherapy. GTR is the recommended approach as it reduces intracranial pressure (Ricard et al., 2012), and the extent of GTR has a significant effect on survival. Orringer and colleagues reported that 1-year survival is significantly higher in patients who undergo >90% resection (Orringer et al., 2012).

Temozolomide (TMZ) is the current standard for chemotherapy for glioblastoma. Stupp and colleagues showed that TMZ administration during and after radiotherapy significantly increases the median overall survival and 2–5-year survival compared to radiotherapy only (Stupp et al., 2009). Moreover, inactivation of *MGMT* expression through promoter methylation is correlated with higher sensitivity to TMZ (Esteller et al., 2000; Hegi et al., 2005). Hegi and colleagues identified that TMZ+radiotherapy leads to significantly higher median overall survival in patients with methylated *MGMT* promoter (21.7 months), compared to patients who received radiotherapy only (15.3 months) (Hegi et al., 2005). Taken together, these findings indicate the significant benefit

offered by TMZ treatment to glioblastoma patients with a methylated *MGMT* promoter.

External beam radiation therapy (EBRT) is the current standard in radiotherapy. Resistance to radiotherapy is quite common, and it is known that a specific EGFR variant (EGFRvIII) mediates radioresistance by inducing the genes involved in double-stranded DNA repair mechanisms (Mukherjee et al., 2009).

## 3. Glioblastoma stem cells

Different models have been proposed to explain the complicated nature of tumor development. The CSC hypothesis postulates a hierarchy in the tumor population, where CSCs are positioned at the top. Thus, CSCs give rise to different cell types through differentiation (Tang, 2012). However, it should be noted that the relationship between CSCs and differentiated tumor cells is bidirectional; in vitro and in vivo interventions (treatment modalities, silencing/overexpressing genes and/or proteins, hypoxia) may trigger dedifferentiation of tumor cells to GSCs, thus creating a dynamic equilibrium between these cell populations (Tang, 2012).

### 3.1. Molecular features of GSCs

Long-term clonal repopulation and self-renewal represent two key features of CSCs (Nguyen et al., 2012). CSCs cannot be easily distinguished from differentiated tumor cells in the case of tumors that display low levels of hierarchy, and in relatively homogeneous tumors (Nguyen et al., 2012; Kreso et al., 2014).

GSCs share several features with neural stem cells (NSCs), including expression of Nestin and CD133, and can form spheres in the presence of required growth factors (Zhu et al., 2014). Similar to NSCs, GSCs also rely on certain transcription factors, which are crucial for their maintenance. These factors include sex-determining region Y-box 2 (*SOX2*), octamer-binding transcription factor (*OCT4*), and Nanog homeobox (*NANOG*) (Schmitz et al., 2007; Ikushima et al., 2011). A list of key biological markers that are used for characterization of GSCs is provided in the Table.

CD133 has served as one of the most frequently used markers to characterize GSCs. CD133<sup>+</sup> glioblastoma cells are able to form tumors, even in low cell numbers. Singh and colleagues reported that 100 CD133<sup>+</sup> cells are able to form tumors when transplanted into the brains of severe combined immunodeficient (SCID) mice; on the other hand, injection of high numbers of CD133<sup>-</sup> cells ( $10^5$ ) does not cause tumor formation (Singh et al., 2004).

Contrary to this notion, different studies have shown the presence of CD133<sup>-</sup> GSCs (Beier et al., 2007, 2011). Comprehensive gene expression studies on molecular subtypes of glioblastoma have shown that CD133 positivity is enhanced in the mesenchymal subtype (Phillips et al., 2006).

**Table .** Important markers for characterization of GSCs.

| Marker              | Function                                                       | Reference                                      |
|---------------------|----------------------------------------------------------------|------------------------------------------------|
| CD133               | Positive association with aggressiveness                       | Brescia et al. (2013)                          |
| CD44                | Positive association with aggressiveness                       | Pietras et al. (2014)                          |
| CD15                | Enrichment marker in CD133 <sup>-</sup> tumors                 | Kahlert et al. (2012), Auffinger et al. (2014) |
| TLX                 | Self-renewal                                                   | Zou et al. (2012)                              |
| ID1                 | Self-renewal                                                   | Soroceanu et al. (2013)                        |
| Integrin $\alpha 6$ | Regulation of self-renewal, proliferation, and tumor formation | Lathia et al. (2010)                           |
| L1CAM               | Maintenance of growth and survival of CD133 <sup>+</sup> cells | Bao et al. (2008)                              |
| Nestin              | Regulation of sphere formation, tumor growth, invasion         | Matsuda et al. (2015)                          |
| SOX2                | Maintenance of self-renewal                                    | Seymour et al. (2015)                          |
| Osteopontin         | Maintenance of stemness, sphere formation, tumor growth        | Lamour et al. (2015)                           |

SOX2 is a transcription factor that has a critical role in maintenance of self-renewal of stem cells, and especially neural stem cells (Ellis et al., 2004; Thiel, 2013). SOX2 overexpression at mRNA and protein levels has been identified in tumor tissues (Alonso et al., 2011; Annovazzi et al., 2011). Gene amplification (Brennan et al., 2013), as well as hypomethylation of the SOX2 promoter (Alonso et al., 2011), can explain SOX2 overexpression in glioblastoma. GSCs express SOX2, which maintains stemness through the TGF- $\beta$  signaling pathway (Ikushima et al., 2009). Gangemi and colleagues showed that loss of SOX2 expression impairs cell proliferation and tumorigenicity of glioblastoma cells in vivo (Gangemi et al., 2009). Given the limited expression of SOX2 in the adult brain (Baer et al., 2007; Seymour et al., 2015), targeting SOX2 can be a potential strategy for treatment of glioblastoma.

Previously, Chen and colleagues showed that ablation of Nestin-expressing glioblastoma cells leads to increased survival of tumor-bearing mice (Chen et al., 2012). This finding highlights the importance of Nestin with respect to tumor propagation.

TLX is a nuclear receptor that is specifically expressed in adult NSCs, and its presence is required for neurogenesis in the subventricular zone (SVZ) (Liu et al., 2008). In addition, mouse models of glioblastoma have shown that the combination of forced TLX overexpression and loss of tumor suppressor genes (TP53 and INK4A/ARF) is sufficient to cause tumor formation (Liu et al., 2010; Park et al., 2010; Zou et al., 2012). Liu and colleagues identified that TLX overexpression leads to migration of progenitor and/or stem cells from their natural niche, and combination of p53 mutations and TLX overexpression lead to glioblastoma initiation in vivo (Liu et al., 2010).

While Tlx has been shown to be druggable (Benod et al., 2014), identification and characterization of potent Tlx inhibitors warrant further studies. However, given the close link between TLX and histone deacetylases (HDACs), HDAC inhibitors can be used to target TLX<sup>+</sup> GSCs (Xie et al., 2014).

L1CAM is a neural adhesion molecule that regulates different cellular processes, including migration, invasion, adhesion, survival, and growth (Maness and Schachner, 2007). Bao and colleagues showed that L1CAM is differentially overexpressed in CD133<sup>+</sup> glioblastoma cells (Bao et al., 2008). Cheng and colleagues provided supporting evidence for this phenomenon, showing that L1CAM is differentially overexpressed in the invasive fronts of glioblastoma (Cheng et al., 2011). In another study, Held-Feindt and colleagues showed that TGF- $\beta 1$  signaling regulates L1CAM expression in glioblastoma, and L1CAM confers resistance to TMZ (Held-Feindt et al., 2012). L1CAM also participates in regulation of DNA damage checkpoint response. Cheng and colleagues showed that the intracellular domain of L1CAM is cleaved from the membrane-bound form through ADAM10- (A Disintegrin and Metalloprotease 10) and Presenilin-mediated cleavage. This, in turn, leads to its translocation to the nucleus, where it induces NBS1 expression through c-Myc (Cheng et al., 2011). As a result, L1CAM enhances DNA damage checkpoint activation and confers radioresistance to GSCs.

Osteopontin is a secreted phosphoprotein that is critical for osteoblast function (Jan et al., 2010). In addition to its role in bone formation, osteopontin is an important angiogenic molecule for glioblastoma, as it is found in the tumor microvasculature (Takano et al., 2000). Osteopontin also functions as a driver of

invasion and tumor growth. Jan and colleagues showed that osteopontin enhances invasion of glioblastoma cells by inducing MMP-2 secretion and vimentin expression (Jan et al., 2010). They also demonstrated that 5-aza-2'-deoxycytidine, an anticancer agent, reduces cell invasion and inhibits glioblastoma tumor growth by suppressing osteopontin expression (Jan et al., 2010).

Inhibitor of DNA binding/differentiation (Id) proteins are negative regulators of the basic HLH family of transcription factors (Perk et al., 2005). Id proteins are well known for their functions related to differentiation, as well as self-renewal of stem cells (O'Brien et al., 2012; Romero-Lanman et al., 2012; Lasorella et al., 2014). Id1 overexpression is a common feature of different cancers (Perk et al., 2006; Lasorella et al., 2014) and is also related to metastasis of breast cancer to the lungs (Gupta et al., 2007). Id1 overexpression has been documented in glioblastoma, which is also positively correlated with tumor grade and proliferation index (Vandeputte et al., 2002). Given the link between TGF- $\beta$  and Id1 expression, inhibition of TGF- $\beta$  signaling can be a potential strategy for glioblastoma treatment. Indeed, Anido and colleagues found that inhibition of the TGF- $\beta$  signaling pathway decreases the number of CD44<sup>high</sup>/Id1<sup>high</sup> GSCs through suppression of Id1 and Id3 expression (Anido et al., 2010). They concluded that this strategy can be employed to overcome tumor recurrence, which is driven through GSCs.

### 3.2. Deregulated signaling pathways in GSCs

Deregulation of cellular signaling pathways is one of the major features that distinguishes CSCs from NSCs. Previous studies have shown that several key signaling pathways are deregulated in glioblastoma. These include Notch signaling, Wnt/beta-catenin signaling, receptor tyrosine kinase (RTK) signaling, and Sonic hedgehog (SHH) signaling.

RTK signaling pathways have been extensively studied in glioblastoma. Of note, comprehensive genetic analyses have shown that EGFR gene amplification and activating mutations, as well as PDGFR amplification, are common events in glioblastoma (Verhaak et al., 2010). Activation of RTK signaling pathways leads to constitutive activation of the downstream PI3K/Akt signaling, which is responsible for maintenance of cell growth and proliferation.

Notch signaling is critical for stem cells, as it functions in regulation of self-renewal and differentiation. Tchorz and colleagues showed that constitutive activation of Notch signaling leads to tumor formation and astroglial lineage entry (Tchorz et al., 2012).

SHH signaling is a key pathway for maintenance of self-renewal and regulates proliferation of GSCs (Clement et al., 2007; Xu et al., 2008; Takezaki et al., 2011). In addition, hyperactivation of SHH signaling and PTEN coexpression are associated with reduced survival (Xu et

al., 2008). Bar and colleagues showed that cyclopamine-mediated inhibition of SHH signaling depletes GSCs (Bar et al., 2007). Their findings suggest that SHH inhibition can be a potential strategy to specifically target GSCs.

CSCs also rely on Wnt/beta-catenin signaling to regulate stemness and differentiation. In addition, Kim and colleagues reported that activation of Wnt/beta-catenin signaling can contribute to radioresistance in GSCs (Kim et al., 2012). Thus, targeting Wnt/beta-catenin signaling may serve as an alternative therapeutic strategy. Recently, De Robertis and colleagues identified and characterized a small molecule inhibitor (SEN461) of the canonical Wnt/beta-catenin signaling pathway. They found that in vivo administration of SEN461 reduces tumor growth (De Robertis et al., 2013).

### 4. Glioblastoma stem cells: a therapeutic challenge

It is hypothesized that treatment failure results from insufficient drug delivery and the targeting of differentiated tumor cells rather than CSCs (Beier et al., 2011). CSCs use different mechanisms to escape chemotherapy-induced cell death, including activation of DNA damage response (Bao et al., 2006) and functions of specific proteins including MGMT and multidrug resistance proteins (e.g., ABCB1) (Beier et al., 2011).

Another factor affecting chemoresistance is tumor evolution. It has been suggested that tumors adapt a chemoresistant phenotype through selection of preexisting clones or formation of de novo subclones (Prados et al., 2015). Supporting evidence for this notion has come from a recent study, where Johnson and colleagues analyzed the origin and evolution of recurrent glioma (Johnson et al., 2014). Through exome sequencing, they identified that TMZ treatment causes a significant portion of the recurrent tumors to follow an alternative path to high-grade glioma. Moreover, they found that recurrent tumors have a TMZ-induced mutagenesis signature (in RB and Akt-mTOR genes) (Johnson et al., 2014).

In another study, Auffinger and colleagues demonstrated for the first time that glioblastoma cells are capable of interconverting between non-CSCs and CSCs upon chemotherapy (Auffinger et al., 2014). They showed that TMZ treatment increases the proportion of GSCs in vitro and in vivo. In addition, lineage-tracing analysis showed that this increase is not a result of enhanced cell proliferation, but rather a result of a phenotypic shift to the CSC state (as demonstrated by stem cell markers, including CD133, SOX2, Oct4, and Nestin). Overall, their results suggest a potential mechanism for escape from chemotherapy.

Targeting self-renewal capacity of GSCs has been used as a promising strategy for treatment of glioblastoma. Recently, Hale and colleagues showed that GSCs are

enriched in CD36 (a scavenger receptor), which can be used to distinguish self-renewing cells. In addition, they found that reduction of CD36 expression leads to loss of self-renewal, tumor initiation capacity, and loss of integrin alpha 6 expression. Overall, they concluded that glioblastoma CSCs selectively use CD36 for their maintenance (Hale et al., 2014).

Deregulated miRNA expression (i.e. downregulation of tumor-suppressor miRNAs) can also contribute to tumor formation and/or progression in glioblastoma. Gal and colleagues compared miRNA expression profiles of CD133<sup>+</sup> and CD133<sup>-</sup> glioblastoma CDCs and found that several miRNAs (including miR-451, miR-486, and miR-425) are overexpressed in CD133<sup>-</sup> glioblastoma CDCs compared to CD133<sup>+</sup> cells. They also showed that exogenous overexpression of miR-451 disperses neurosphere formation and inhibits cell proliferation (Gal et al., 2008). Their results indicate that restoring expression of tumor-suppressive miRNAs can be used as an alternative strategy for treatment of glioblastoma.

Hitomi and colleagues reported that connexins, which are structural elements of gap junctions, show differential expression between GSCs and differentiated tumor cells (Hitomi et al., 2015). Their results show that differentiated tumor cells predominantly express connexin 43 (Cx43), whereas GSCs express Cx46. In addition, they found that

Cx46 expression decreases and Cx43 expression increases during differentiation of GSCs. Reduced expression of Cx46 impaired the tumor-forming capacity and self-renewal of glioblastoma CSCs.

Gamma-secretase inhibitors and RNA interference have been previously used to inhibit Notch signaling, which leads to reduced radioresistance and impaired formation of tumor spheres (Fan et al., 2010; Wang et al., 2010).

## 5. Concluding remarks

The advances in stem cell biology in the past decade have helped us to better understand the development and pathogenesis of brain tumors. Despite the identification and characterization of GSCs, the existence of a specific cell population and their specific roles in the context of tumor development are still debated. By utilizing the key molecular features of CSCs, new therapeutic strategies can be developed to achieve more durable clinical responses. Alternatively, given the inherent tumor tropism of NSCs, endogenous and/or engineered NSCs can be used as therapeutic delivery vehicles for treatment of glioblastoma. Taken together, translating the accumulated knowledge on the biology of NSCs and CSCs with well-designed studies may bring new possibilities for effective therapies to glioblastoma patients.

## References

- Alonso MM, Diez-Valle R, Manterola L, Rubio A, Liu D, Cortes-Santiago N, Urquiza L, Jauregi P, Lopez de Munain A, Sampron N et al. (2011). Genetic and epigenetic modifications of Sox2 contribute to the invasive phenotype of malignant gliomas. *PLoS One* 6: e26740.
- Anido J, Saez-Borderias A, Gonzalez-Junca A, Rodon L, Folch G, Carmona MA, Prieto-Sanchez RM, Barba I, Martinez-Saez E, Prudkin L et al. (2010). TGF- $\beta$  receptor inhibitors target the CD44<sup>high</sup>/Id1<sup>high</sup> glioma-initiating cell population in human glioblastoma. *Cancer Cell* 18: 655-668.
- Annovazzi L, Mellai M, Caldera V, Valente G, Schiffer D (2011). SOX2 expression and amplification in gliomas and glioma cell lines. *Cancer Genomics Proteomics* 8: 139-147.
- Auffinger B, Tobias AL, Han Y, Lee G, Guo D, Dey M, Lesniak MS, Ahmed AU (2014). Conversion of differentiated cancer cells into cancer stem-like cells in a glioblastoma model after primary chemotherapy. *Cell Death Diff* 21: 1119-1131.
- Baer K, Eriksson PS, Faull RL, Rees MI, Curtis MA (2007). Sox-2 is expressed by glial and progenitor cells and Pax-6 is expressed by neuroblasts in the human subventricular zone. *Exp Neurol* 204: 828-831.
- Bao S, Wu Q, Li Z, Sathornsumetee S, Wang H, McLendon RE, Hjelmeland AB, Rich JN (2008). Targeting cancer stem cells through L1CAM suppresses glioma growth. *Cancer Res* 68: 6043-6048.
- Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, Dewhirst MW, Bigner DD, Rich JN (2006). Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 444: 756-760.
- Bar EE, Chaudhry A, Lin A, Fan X, Schreck K, Matsui W, Piccirillo S, Vescovi AL, DiMeco F, Olivi A et al. (2007). Cyclopamine-mediated hedgehog pathway inhibition depletes stem-like cancer cells in glioblastoma. *Stem Cells* 25: 2524-2533.
- Beier CP, Beier D (2011). CD133 negative cancer stem cells in glioblastoma. *Front Biosci* 3: 701-710.
- Beier D, Hau P, Proescholdt M, Lohmeier A, Wischhusen J, Oefner PJ, Aigner L, Brawanski A, Bogdahn U, Beier CP (2007). CD133(+) and CD133(-) glioblastoma-derived cancer stem cells show differential growth characteristics and molecular profiles. *Cancer Res* 67: 4010-4015.
- Beier D, Schulz JB, Beier CP (2011). Chemoresistance of glioblastoma cancer stem cells--much more complex than expected. *Mol Cancer* 10: 128.
- Benod C, Villagomez R, Filgueira CS, Hwang PK, Leonard PG, Poncet-Montange G, Rajagopalan S, Fletterick RJ, Gustafsson JA, Webb P (2014). The human orphan nuclear receptor tailless (TLX, NR2E1) is druggable. *PLoS One* 9: e99440.

- Brennan CW, Verhaak RG, McKenna A, Campos B, Noushmehr H, Salama SR, Zheng S, Chakravarty D, Sanborn JZ, Berman SH et al. (2013). The somatic genomic landscape of glioblastoma. *Cell* 155: 462-477.
- Brescia P, Ortensi B, Fornasari L, Levi D, Broggi G, Pelicci G (2013). CD133 is essential for glioblastoma stem cell maintenance. *Stem Cells* 31: 857-869.
- Chen J, Li Y, Yu TS, McKay RM, Burns DK, Kernie SG, Parada LF (2012). A restricted cell population propagates glioblastoma growth after chemotherapy. *Nature* 488: 522-526.
- Cheng L, Wu Q, Guryanova OA, Huang Z, Huang Q, Rich JN, Bao S (2011). Elevated invasive potential of glioblastoma stem cells. *Biochem Bioph Res Co* 406: 643-648.
- Cheng L, Wu Q, Huang Z, Guryanova OA, Huang Q, Shou W, Rich JN, Bao S (2011). L1CAM regulates DNA damage checkpoint response of glioblastoma stem cells through NBS1. *EMBO J* 30: 800-813.
- Clement V, Sanchez P, de Tribolet N, Radovanovic I, Ruiz i Altaba A (2007). HEDGEHOG-GLI1 signaling regulates human glioma growth, cancer stem cell self-renewal, and tumorigenicity. *Curr Biol* 17: 165-172.
- De Robertis A, Valensin S, Rossi M, Tunci P, Verani M, De Rosa A, Giordano C, Varrone M, Nencini A, Pratelli C et al. (2013). Identification and characterization of a small-molecule inhibitor of Wnt signaling in glioblastoma cells. *Mol Cancer Ther* 12: 1180-1189.
- Ellis P, Fagan BM, Magness ST, Hutton S, Taranova O, Hayashi S, McMahon A, Rao M, Pevny L (2004). SOX2, a persistent marker for multipotential neural stem cells derived from embryonic stem cells, the embryo or the adult. *Dev Neurosci* 26: 148-165.
- Fan X, Khaki L, Zhu TS, Soules ME, Talsma CE, Gul N, Koh C, Zhang J, Li YM, Maciaczyk J et al. (2010). NOTCH pathway blockade depletes CD133-positive glioblastoma cells and inhibits growth of tumor neurospheres and xenografts. *Stem Cells* 28: 5-16.
- Gal H, Pandi G, Kanner AA, Ram Z, Lithwick-Yanai G, Amariglio N, Rechavi G, Givol D (2008). MIR-451 and imatinib mesylate inhibit tumor growth of glioblastoma stem cells. *Biochem Bioph Res Co* 376: 86-90.
- Gangemi RM, Griffiro F, Marubbi D, Perera M, Capra MC, Malatesta P, Ravetti GL, Zona GL, Daga A, Corte G (2009). SOX2 silencing in glioblastoma tumor-initiating cells causes stop of proliferation and loss of tumorigenicity. *Stem Cells* 27: 40-48.
- Gupta GP, Perk J, Acharyya S, de Candia P, Mittal V, Todorova-Manova K, Gerald WL, Brogi E, Benezra R, Massague J (2007). ID genes mediate tumor reinitiation during breast cancer lung metastasis. *P Natl Acad Sci USA* 104: 19506-19511.
- Hale JS, Otvos B, Sinyuk M, Alvarado AG, Hitomi M, Stoltz K, Wu Q, Flavahan W, Levison B, Johansen ML et al. (2014). Cancer stem cell-specific scavenger receptor 36 drives glioblastoma progression. *Stem Cells* 32: 1746-1758.
- Hartmann C, Hentschel B, Wick W, Capper D, Felsberg J, Simon M, Westphal M, Schackert G, Meyermann R, Pietsch T et al. (2010). Patients with IDH1 wild type anaplastic astrocytomas exhibit worse prognosis than IDH1-mutated glioblastomas, and IDH1 mutation status accounts for the unfavorable prognostic effect of higher age: implications for classification of gliomas. *Acta Neuropathol* 120: 707-718.
- Held-Feindt J, Schmelz S, Hattermann K, Mentlein R, Mehdorn HM, Sebens S (2012). The neural adhesion molecule L1CAM confers chemoresistance in human glioblastomas. *Neurochem Int* 61: 1183-1191.
- Hitomi M, Deleyrolle LP, Mulkearns-Hubert EE, Jarrar A, Li M, Sinyuk M, Otvos B, Brunet S, Flavahan WA, Hubert CG et al. (2015). Differential connexin function enhances self-renewal in glioblastoma. *Cell Reports* 11: 1031-1042.
- Ikushima H, Todo T, Ino Y, Takahashi M, Miyazawa K, Miyazono K (2009). Autocrine TGF-beta signaling maintains tumorigenicity of glioma-initiating cells through Sry-related HMGB-box factors. *Cell Stem Cell* 5: 504-514.
- Ikushima H, Todo T, Ino Y, Takahashi M, Saito N, Miyazawa K, Miyazono K (2011). Glioma-initiating cells retain their tumorigenicity through integration of the Sox axis and Oct4 protein. *J Biol Chem* 286: 41434-41441.
- Jan HJ, Lee CC, Shih YL, Hueng DY, Ma HI, Lai JH, Wei HW, Lee HM (2010). Osteopontin regulates human glioma cell invasiveness and tumor growth in mice. *Neuro-Oncology* 12: 58-70.
- Kahlert UD, Bender NO, Maciaczyk D, Bogiel T, Bar EE, Eberhart CG, Nikkhah G, Maciaczyk J (2012). CD133/CD15 defines distinct cell subpopulations with differential in vitro clonogenic activity and stem cell-related gene expression profile in in vitro propagated glioblastoma multiforme-derived cell line with a PNET-like component. *Folia Neuropathol* 50: 357-368.
- Kim Y, Kim KH, Lee J, Lee YA, Kim M, Lee SJ, Park K, Yang H, Jin J, Joo KM et al. (2012). Wnt activation is implicated in glioblastoma radioresistance. *Lab Invest* 92: 466-473.
- Kreso A, Dick JE (2014). Evolution of the cancer stem cell model. *Cell Stem Cell* 14: 275-291.
- Krex D, Klink B, Hartmann C, von Deimling A, Pietsch T, Simon M, Sabel M, Steinbach JP, Heese O, Reifenberger G et al. (2007). Long-term survival with glioblastoma multiforme. *Brain* 130: 2596-2606.
- Lamour V, Henry A, Kroonen J, Nokin MJ, von Marschall Z, Fisher LW, Chau TL, Chariot A, Sanson M, Delattre JY et al. (2015). Targeting osteopontin suppresses glioblastoma stem-like cell character and tumorigenicity in vivo. *Int J Cancer* 137: 1047-1057.
- Lasorella A, Benezra R, Iavarone A (2014). The ID proteins: master regulators of cancer stem cells and tumour aggressiveness. *Nat Rev Cancer* 14: 77-91.
- Lathia JD, Gallagher J, Heddleston JM, Wang J, Eyler CE, Macswords J, Wu Q, Vasani A, McLendon RE, Hjelmeland AB et al. (2010). Integrin alpha 6 regulates glioblastoma stem cells. *Cell Stem Cell* 6: 421-432.

- Liu HK, Belz T, Bock D, Takacs A, Wu H, Lichter P, Chai M, Schutz G (2008). The nuclear receptor tailless is required for neurogenesis in the adult subventricular zone. *Gene Dev* 22: 2473-2478.
- Liu HK, Wang Y, Belz T, Bock D, Takacs A, Radlwimmer B, Barbus S, Reifenberger G, Lichter P, Schutz G (2010). The nuclear receptor tailless induces long-term neural stem cell expansion and brain tumor initiation. *Gene Dev* 24: 683-695.
- Maness PF, Schachner M (2007). Neural recognition molecules of the immunoglobulin superfamily: signaling transducers of axon guidance and neuronal migration. *Nat Neurosci* 10: 19-26.
- Matsuda Y, Ishiwata T, Yoshimura H, Hagio M, Arai T (2015). Inhibition of nestin suppresses stem cell phenotype of glioblastomas through the alteration of post-translational modification of heat shock protein HSPA8/HSC71. *Cancer Lett* 357: 602-611.
- Nguyen LV, Vanner R, Dirks P, Eaves CJ (2012). Cancer stem cells: an evolving concept. *Nat Rev Cancer* 12: 133-143.
- O'Brien CA, Kreso A, Ryan P, Hermans KG, Gibson L, Wang Y, Tsatsanis A, Gallinger S, Dick JE (2012). ID1 and ID3 regulate the self-renewal capacity of human colon cancer-initiating cells through p21. *Cancer Cell* 21: 777-792.
- Omuro A, DeAngelis LM (2013). Glioblastoma and other malignant gliomas: a clinical review. *JAMA-J Am Med Assoc* 310: 1842-1850.
- Orringer D, Lau D, Khatri S, Zamora-Berridi GJ, Zhang K, Wu C, Chaudhary N, Sagher O (2012). Extent of resection in patients with glioblastoma: limiting factors, perception of resectability, and effect on survival. *J Neurosurg* 117: 851-859.
- Ostrom QT, Bauchet L, Davis FG, Deltoro I, Fisher JL, Langer CE, Pekmezci M, Schwartzbaum JA, Turner MC, Walsh KM et al. (2014). The epidemiology of glioma in adults: a "state of the science" review. *Neuro-Oncology* 16: 896-913.
- Ostrom QT, Gittleman H, Farah P, Ondracek A, Chen Y, Wolinsky Y, Stroup NE, Kruchko C, Barnholtz-Sloan JS (2013). CBTRUS statistical report: primary brain and central nervous system tumors diagnosed in the United States in 2006-2010. *Neuro-Oncology* 15 (Suppl. 2): ii1-56.
- Park HJ, Kim JK, Jeon HM, Oh SY, Kim SH, Nam DH, Kim H (2010). The neural stem cell fate determinant TLX promotes tumorigenesis and genesis of cells resembling glioma stem cells. *Mol Cells* 30: 403-408.
- Perk J, Gil-Bazo I, Chin Y, de Candia P, Chen JJ, Zhao Y, Chao S, Cheong W, Ke Y, Al-Ahmadie H et al. (2006). Reassessment of id1 protein expression in human mammary, prostate, and bladder cancers using a monospecific rabbit monoclonal anti-id1 antibody. *Cancer Res* 66: 10870-10877.
- Perk J, Iavarone A, Benezra R (2005). Id family of helix-loop-helix proteins in cancer. *Nat Rev Cancer* 5: 603-614.
- Phillips HS, Kharbanda S, Chen R, Forrester WF, Soriano RH, Wu TD, Misra A, Nigro JM, Colman H, Soroceanu L et al. (2006). Molecular subclasses of high-grade glioma predict prognosis, delineate a pattern of disease progression, and resemble stages in neurogenesis. *Cancer Cell* 9: 157-173.
- Pietras A, Katz AM, Ekstrom EJ, Wee B, Halliday JJ, Pitter KL, Werbeck JL, Amankulor NM, Huse JT, Holland EC (2014). Osteopontin-CD44 signaling in the glioma perivascular niche enhances cancer stem cell phenotypes and promotes aggressive tumor growth. *Cell Stem Cell* 14: 357-369.
- Ricard D, Idbaih A, Ducray F, Lahutte M, Hoang-Xuan K, Delattre JY (2012). Primary brain tumours in adults. *Lancet* 379: 1984-1996.
- Romero-Lanman EE, Pavlovic S, Amlani B, Chin Y, Benezra R (2012). Id1 maintains embryonic stem cell self-renewal by up-regulation of Nanog and repression of Brachyury expression. *Stem Cells Dev* 21: 384-393.
- Schmitz M, Temme A, Senner V, Ebner R, Schwind S, Stevanovic S, Wehner R, Schackert G, Schackert HK, Fussell M et al. (2007). Identification of SOX2 as a novel glioma-associated antigen and potential target for T cell-based immunotherapy. *Brit J Cancer* 96: 1293-1301.
- Schwartzbaum JA, Fisher JL, Aldape KD, Wrensch M (2006). Epidemiology and molecular pathology of glioma. *Nat Clin Pract Neurol* 2: 494-503.
- Scott JN, Rewcastle NB, Brasher PM, Fulton D, MacKinnon JA, Hamilton M, Cairncross JG, Forsyth P (1999). Which glioblastoma multiforme patient will become a long-term survivor? A population-based study. *Ann Neurol* 46: 183-188.
- Seymour T, Nowak A, Kakulas F (2015). Targeting aggressive cancer stem cells in glioblastoma. *Front Oncol* 5: 159.
- Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, Dirks PB (2003). Identification of a cancer stem cell in human brain tumors. *Cancer Res* 63: 5821-5828.
- Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, Henkelman RM, Cusimano MD, Dirks PB (2004). Identification of human brain tumour initiating cells. *Nature* 432: 396-401.
- Smoll NR, Schaller K, Gautschi OP (2013). Long-term survival of patients with glioblastoma multiforme (GBM). *J Clin Neurosci* 20: 670-675.
- Soroceanu L, Murase R, Limbad C, Singer E, Allison J, Adrados I, Kawamura R, Pakdel A, Fukuyo Y, Nguyen D et al. (2013). Id-1 is a key transcriptional regulator of glioblastoma aggressiveness and a novel therapeutic target. *Cancer Res* 73: 1559-1569.
- Takano S, Tsuboi K, Tomono Y, Mitsui Y, Nose T (2000). Tissue factor, osteopontin,  $\alpha_v\beta_3$  integrin expression in microvasculature of gliomas associated with vascular endothelial growth factor expression. *Br J Cancer* 82: 1967-1973.
- Takezaki T, Hide T, Takanaga H, Nakamura H, Kuratsu J, Kondo T (2011). Essential role of the Hedgehog signaling pathway in human glioma-initiating cells. *Cancer Sci* 102: 1306-1312.
- Tang DG (2012). Understanding cancer stem cell heterogeneity and plasticity. *Cell Res* 22: 457-472.
- Tchorz JS, Tome M, Cloetta D, Sivasankaran B, Grzmil M, Huber RM, Rutz-Schatzmann F, Kirchhoff F, Schaeren-Wiemers N, Gassmann M et al. (2012). Constitutive Notch2 signaling in neural stem cells promotes tumorigenic features and astroglial lineage entry. *Cell Death Dis* 3: e325.

- Thakkar JP, Dolecek TA, Horbinski C, Ostrom QT, Lightner DD, Barnholtz-Sloan JS, Villano JL (2014). Epidemiologic and molecular prognostic review of glioblastoma. *Cancer Epidemiol Biomarkers Prev* 23: 1985-1996.
- Thiel G (2013). How Sox2 maintains neural stem cell identity. *Biochem J* 450: e1-2.
- Vandeputte DA, Troost D, Leenstra S, Ijlst-Keizers H, Ramkema M, Bosch DA, Baas F, Das NK, Aronica E (2002). Expression and distribution of id helix-loop-helix proteins in human astrocytic tumors. *Glia* 38: 329-338.
- Verhaak RG, Hoadley KA, Purdom E, Wang V, Qi Y, Wilkerson MD, Miller CR, Ding L, Golub T, Mesirov JP et al. (2010). Integrated genomic analysis identifies clinically relevant subtypes of glioblastoma characterized by abnormalities in PDGFRA, IDH1, EGFR, and NF1. *Cancer Cell* 17: 98-110.
- Wang J, Wakeman TP, Lathia JD, Hjelmeland AB, Wang XF, White RR, Rich JN, Sullenger BA (2010). Notch promotes radioresistance of glioma stem cells. *Stem Cells* 28: 17-28.
- Wen PY, Kesari S (2008). Malignant gliomas in adults. *New Engl J Med* 359: 492-507.
- Xie Q, Flavahan WA, Bao S, Rich J (2014). The tailless root of glioma: cancer stem cells. *Cell Stem Cell* 15: 114-116.
- Xu Q, Yuan X, Liu G, Black KL, Yu JS (2008). Hedgehog signaling regulates brain tumor-initiating cell proliferation and portends shorter survival for patients with PTEN-coexpressing glioblastomas. *Stem Cells* 26: 3018-3026.
- Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, Yuan W, Kos I, Batinic-Haberle I, Jones S, Riggins GJ et al. (2009). IDH1 and IDH2 mutations in gliomas. *New Engl J Med* 360: 765-773.
- Yuan X, Curtin J, Xiong Y, Liu G, Waschmann-Hogiu S, Farkas DL, Black KL, Yu JS (2004). Isolation of cancer stem cells from adult glioblastoma multiforme. *Oncogene* 23: 9392-9400.
- Zhu Z, Khan MA, Weiler M, Blaes J, Jestaedt L, Geibert M, Zou P, Gronych J, Bernhardt O, Korshunov A et al. (2014). Targeting self-renewal in high-grade brain tumors leads to loss of brain tumor stem cells and prolonged survival. *Cell Stem Cell* 15: 185-198.
- Zou Y, Niu W, Qin S, Downes M, Burns DK, Zhang CL (2012). The nuclear receptor TLX is required for gliomagenesis within the adult neurogenic niche. *Mol Cell Biol* 32: 4811-4820.