# The Effect of Mineral Admixtures on Alkali-silica Reaction in Concrete

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#### Abstract

In this research, antagonistic activities of 30 *Bacillus* species isolated from various fish samples were studied. Isolated *Bacillus* species were analyzed using the agar diffusion method in terms of their general inhibition effects against some food pathogen/contaminant bacteria and lactic acid bacteria isolated from the fish intestinal tracts. Some of these strains exhibited antimicrobial activity against *Bacillus subtilis*, *Pseudomonas fluorescens*, *Lactobacillus coryneformis*, *Lactobacillus plantarum* and *Lactobacillus xylosus*. Furthermore, the inhibitory effects of the lactic acid bacteria on the *Bacillus* isolates were analyzed by using the same method. It was determined that the lactic acid bacteria inhibited *Bacillus* spp. strains at different levels of inhibition zones.

Keywords: Bacillus, Lactobacillus, Fishes, Isolation, Antimicrobial.

# INTRODUCTION

The antibiotics are widely distributed in the nature, where they play an important role in regulating the microbial population of soil, water, sewage, and compost. The Gram-positive, aerobic, rod-shaped endospore-forming bacteria of the genus Bacillus are most impressively produced antibiotics as secondary metabolites [1-3]. Kümmerer [4] is stated that, within the last decade, an increasing number of studies covering antibiotic input, occurrence, fate and effects have been published, but there is still a lack of understanding and knowledge about antibiotics in the aquatic environment despite the numerous studies performed. The increased practice of aquaculture has led to a high number of disease outbreaks with an increasing range of pathogens. Consequently, the extensive use of broadspectrum antibiotics in aquaculture has led, as in other fields, to drug resistance problems [5]. In recent years, many studies have been determined the antimicrobial properties of strains of Bacillus [6-10]. Shelar et al. [11] determined that a bacteriocin produced by the isolate of B. atrophaeus JS-2 was showed antimicrobial properties against some Grampositive and Gram-negative bacteria. In a study, 50 out of the 118 Bacillus strains isolated from soil samples exhibited antagonistic activities against at least two or more strain from a panel of pathogenic and non-pathogenic microorganisms [12]. Probiotics may provide an alternative way to reduce the use of antibiotics in aquaculture and simultaneously may avoid the development of antibioticresistant bacteria. Most probiotics proposed as biological control agents in aquaculture belong to the lactic acid bacteria family [13]. Most studies have reported that the different strains of *Bacillus* spp. and the lactic acid bacteria found in fish and fish products [14-17].

The aim of this study was to evaluate the antagonistic activity of *Bacillus* spp. strains isolated from the fishes and to determine the antimicrobial activity of fish intestinal lactic acid bacteria on the *Bacillus* isolates.

# MATERIALS AND METHODS

### **Isolation, Identification and Growth Conditions**

Fresh fishes (anchovy, bluefish, gray mullet, horse mackerel, red sea bream, and whiting) were procured from fish retail markets in Ankara, TURKEY and packed in iceboxes and transferred to the laboratory within 1 h. Each 1 g of the fish samples (the intestinal digestive tract) was suspended in 9 mL sterile distilled water and shaken vigorously for 2 min. The samples were heated at 60 °C for 60 min in a water bath. Then, the liquid was serially diluted in sterile distilled water, and the dilution from  $10^{-1}$  to  $10^{-6}$  was plated on nutrient agar medium (Oxoid). The plates were incubated at  $37\pm1$  °C for 24-48 h [18].

Based on Bergey's Manual [19], the strains were classified using the following criteria: Gram reaction, spore occurrence, growth temperature range (5, 30, 40, 50, 55, 65 °C), pH values of the growth (5.7, 6.8), salt resistance (2, 5, 7, 10 g NaCl/100 mL), catalase activity,

lecitinase activity, production of gas from glucose, sugar fermentation (D-glucose, D-mannitol, lactose, sucrose, melibiose, D-xylose, L-arabinose, maltose and salicin, 1 g/100 mL), reduction of nitrate, and gelatin and starch hydrolysis.

The bacterial strains were cultivated in Nutrient Broth (NB) (Oxoid), which contained (per L) 1 g lab-lemco powder, 2 g yeast extract, 5 g peptone and 5 g NaCl. The pH was adjusted to 6.8 with 0.01 M HCl and 0.01 M NaOH. The incubation temperature was maintained at  $37\pm1$  °C, and the agitation was maintained at 100 rpm. The cultures were inoculated into broth medium with 2% (v/v) inocula.

#### **Test Microorganisms**

Some pathogen/contaminant bacteria and lactic acid bacteria isolated from the intestinal tracts of rainbow trout and mirror carp used in this study were obtained from Culture Collections of the Biotechnology Laboratory at the Department of Biology in Gazi University. The names and codes of the test bacteria are presented in Table 1. The pathogen and contaminant bacteria were activated through incubation in a NB (Oxoid) for 24 h. The lactic acid bacteria were cultured in MRS broth (Oxoid) for 24 h. *Micrococcus flavus* TIM and lactic acid bacteria were incubated at 30±1 °C while the other test bacteria were incubated at 37±1 °C.

**Table 1.** Names and codes of the test microorganisms.

TEST MICROORGANISMS										
Food Pathogen and Contaminant Bacteria	Lactic Acid Bacteria									
Bacillus subtilis ATCC 6633	Lactobacillus casei HS1									
Escherichia coli ATCC 11230	Lactobacillus coryneformis HS18									
Micrococcus flavus TIM	Lactobacillus jensenii HS30									
Pseudomonas aeroginosa ATCC 29212	Lactobacillus plantarum HC13									
Pseudomonas fluorescens RSKK 240	Lactobacillus xylosus HC9									
Staphylococcus aureus 4-43	-									
Yersinia enterolitica ATCC 1501	-									

### Inhibitory Effect by Agar-Well Diffusion Method

The inhibitory effects of isolates on test bacteria were determined by agar-well diffusion method [20]. Similarly, the inhibitory effects of the lactic acid bacteria on the Bacillus isolates were analyzed by using the same method. All the bacteria were incubated at the appropriate temperature and medium for 24 h. Nutrient agar and MRS agar media (20 mL) were poured into each sterile petri dish (100 mm diameter). 100 µL suspensions of target strain cultured for 24 h were spread on the plates, and wells of 6 mm diameter were punched in the agar with a sterile steel borer. The Bacillus and lactobacilli cultures were centrifuged at 6000 g for 15 min to remove cell debris. After centrifugation, supernatant samples (100 µL) were filled into the wells of agar plates directly. The inoculated plates were incubated for 24 h at their optimum growth temperatures, and the diameter of the inhibition zone was measured with calipers as mm. The measurements were done from the edge of the zone to the edge of the wall.

# RESULTS AND DISCUSSION

Thirty *Bacillus* species were isolated from various fish samples. In the identification tests, 30 *Bacillus* strains were identified as 4 *B. pasteurii*, 3 *B. badius*, 3 *B. circulans*, 3 *B. licheniformis*, 3 *B. megaterium*, 3 *B. thuringiensis*, 2 *B. brevis*, 2 *B. cereus*, 2 *B. sphaericus*, 2 *B. subtilis*, 1 *B. coagulans*, 1 *B. lentus*, and 1 *B. pumilus*. Names of isolated species and their codes are given in Table 2.

Table 2. Isolated species and their codes.

Code	Species	
P1, P4, P7	Bacillus megaterium	
P2, P3, P6, P12	Bacillus pasteurii	
P5, P16, P17	Bacillus circulans	
P8, P9	Bacillus subtilis	
P10	Bacillus coagulans	
P11, P13	Bacillus cereus	
P14, P15	Bacillus sphaericus	
P18	Bacillus lentus	
P19, P21, P22	Bacillus badius	
P20, P25, P26	Bacillus licheniformis	
P23, P29	Bacillus brevis	
P24	Bacillus pumilus	
P27, P28, P30	Bacillus thuringiensis	

This study examined the antagonistic activities of the *Bacillus* spp. strains against the test bacteria. *B. subtilis* P9, *B. pasteurii* P12 and *B. licheniformis* P25 showed an inhibition zone diameter of 5.80 mm, 4.95 mm, and 5.00 mm against *P. fluorescens* RSKK 240, respectively. It was also determined that *B. cereus* P11 (5.05 mm) and *B. thuringiensis* P30 (5.70 mm) had inhibitory effects on *B. subtilis* ATCC 6633 (Table 3).

**Table 3.** Antimicrobial activity of *Bacillus* strains on some food pathogen/contaminant bacteria.

Ant	ibacterial Activity					
Strains Tested Bacteria						
	B. subtilis ATCC 6633	P. fluorescens RSKK 240				
B. subtilis P9	-	+++				
B. cereus P11	+++	-				
B. pasteurii P12	-	++				
B. licheniformis P25	-	+++				
B. thuringensis P30	+++	-				

Antimicrobial activity: zone of inhibition.

According to Pinchuck et al. [21], probiotic strain *B. subtilis* 3, whose safety was previously demonstrated, is known to have antagonistic properties against species of the family *Enterobacteriaceae* as well as inhibit *Helicobacter pylorii*. Binnet [22] reported that *Bacillus* strains isolated from milk displayed antimicrobial activity against *E. coli*, *Y. enterolitica* and *P. aeroginosa*. In the study by Perez et al. [23, 24], *B. subtilis* MIR 15 strain did not show antimicrobial activity against *E. coli*, *M. luteus* and *P. aeroginosa*. Aslim et al. [25] found that *B. megaterium*, *B. subtilis* and *B. thuringiensis* strains were active against *E. coli* a

<sup>-:</sup> No effect.

<sup>+:</sup> Zone width 1-3 mm; ++: Zone width 3-5 mm; +++: Zone width 5-7 mm

coli and Y. enterolitica. In a study on the antimicrobial activity of 29 Bacillus strains isolated from the soil against some tested bacteria, Yilmaz et al. [26] determined only 5 isolates with antimicrobial activity. They concluded that not all the Bacillus isolates showed inhibitory effects on E. coli ATCC 11230, M. flavus TIM, P. aeroginosa ATCC 29212, S. aureus 4-43 and Y. enterolitica ATCC 1501. This study deduced that the isolates used have no inhibitory effects regarding E. coli ATCC 11230, M. flavus TIM, P. aeroginosa ATCC 29212, S. aureus 4-43, Y. enterolitica ATCC 1501. Oscariz et al. [27] reported that B. cereus strain isolated from soil was active against most Gram-positive but not Gram-negative bacteria.

The inhibitory effects of *Bacillus* isolates against lactic acid bacteria are given in Tablo 4. *B. cereus* P13 (3.90 mm), *B. thuringiensis* P27 (4.50 mm) and P28 (4.55 mm) were active against *L. coryneformis* HS18. *B. licheniformis* P25 (2.90 mm) and P26 (3.25 mm) had inhibitory effects on *L. plantarum* HC13. In addition, *B. megaterium* P4 (3.50 mm) and P7 (2.75 mm) showed antimicrobial activity against *L. xylosus* HC9. The other *Bacillus* strains did not show antimicrobial activity against the lactic acid bacteria tested. Kalayli [28] determined that *Bacillus* strains isolated from milk and fermented products did not show any antimicrobial activities against *L. casei*, but the strains inhibited the growth of *L. plantarum* 

Table 4. Antimicrobial activity of *Bacillus* strains on lactic acid bacteria.

bacteria.			
	Antibacteria	Activity	
Strains			
	Tes	sted Bacteria	
	L. coryneformis	L. plantarum	L. xylosus
	HS18	HC13	HC9
B. megaterium P4	-	-	++
B. megaterium P7	-	-	+
B. cereus P13	++	-	-
B. licheniformis P25	-	+	-
B. licheniformis P26	-	++	-
B. thuringensis P27	++	-	-
B. thuringensis P28	++	-	-

Antimicrobial activity: zone of inhibition.

Additionally in our study, the lactic acid bacteria were assayed for their ability to produce inhibitory substances against the growth of *Bacillus* strains. The results concerning the determination of the antimicrobial effects are presented in Table 5. *L. xylosus* HC9 did not show antimicrobial activity against *B. megaterium* P1, *B. pasteurii* P6 and *B. cereus* P13. *L. casei* HS1 and *L. jensenii* HS30 did not have inhibitory effects on *B. subtilis* P9. The other lactic acid bacteria inhibited the growth of the *Bacillus* strains, at different levels of inhibition zones. The highest antimicrobial activity of lactic acid bacteria (*L. xylosus* HC9) was against *B. megaterium* P4 (5.60 mm) whereas the lowest antimicrobial activity of bacteria (*L. casei* HS1) was against *B thuringiensis* P30 (1.00 mm).

**Table 5.** Antimicrobial activity of lactic acid bacteria on the *Bacillus* strains.

		Antibacterial Activity													
Lactic Acid		Bacillus strains													s
Bacteria	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10	P11	P12	P13	P14	P15
L. casei HS1	++	++	++	+	++	+	++	++	-	+	+	++	++	+	++
L.coryneformis HS18	++	++	++	++	+	++	++	++	++	+	++	++	+	+	++
L. jensenii HS30	++	+	+	++	++	+	++	++	-	+	++	++	++	++	++
L.plantarum HC13	++	++	++	++	++	++	++	+	‡ +	+	++	++	++	+	++
L. xylosus HC9	-	+	++	+ +	+	-	++	+	+	++	+	++	- 1	+	+

Antimicrobial activity: zone of inhibition.

**Table 5** (Continued). Antimicrobial activity of lactic acid bacteria on the *Bacillus* strains.

		Antibacterial Activity													
Lactic Acid		Bacillus strains												š	
Bacteria	P16	P17	P18	P19	P20	P21	P22	P23	P24	P25	P26	P27	P28	P29	P30
L. casei HS1	++	++	+	+	+	+	+	+	+	+	+	+	+	+	+
L. coryneformis HS18	++	+	+	++	+	++	+	++	++	++	++	++	++	++	++
L. jensenii HS30	++	++	+	+	+	+	+	+	+	+	+	+	+	+	+
L. plantarum HC13	++	++	++	+	++	++	+	+	+	+	+	+	+	+	+
L. xylosus HC9	++	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Antimicrobial activity: zone of inhibition.

Among the bacteria used as probiotics, lactic acid bacteria have an important role because they are beneficial to human and animal health [29, 30]. Probiotic strains should have desirable antibiotic resistance and sensitivity patterns, be antagonistic toward potentially pathogenic microorganisms, and have metabolic activities beneficial to the well-being of the host [31]. In our study, L. xylosus HC9 did not show antimicrobial activity against B. megaterium P1, B. pasteurii P6 and B. cereus P13. L. casei HS1, and L. jensenii HS30 did not have inhibitory effects on B. subtilis P9. The other lactic acid bacteria inhibited the growth of the Bacillus strains at different levels of inhibition zones. Katircioğlu [32] reported that the lactic acid bacteria inhibited Bacillus spp. at different levels of inhibition zones. Jiranvanichpaisal et al. [33] emphasized the use of Lactobacillus sp. as the probiotic bacteria in the giant tiger shrimp (Penaeus monodon Fabricus). Inhibiting activity of two Lactobacillus sp. against Vibrio sp., E. coli, Staphylococcus sp. and B. subtilis was also determined.

In conclusion, *Bacillus* spp. strains isolated from the intestinal tracts of various fishes are antimicrobial activity against some food pathogen/contaminant bacteria. Also, it has been found that lactic acid bacteria isolated from the intestinal tracts of fish have inhibitory effects against *Bacillus* spp. Strains

<sup>-:</sup> No effect.

<sup>+:</sup> Zone width 1-3 mm; ++: Zone width 3-5 mm; +++: Zone width 5-7 mm.

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<sup>-:</sup> No effec

<sup>+:</sup> Zone width 1-3 mm; ++: Zone width 3-5 mm; +++: Zone width 5-7 mm.

# REFERENCES

- [1]. Drablos F, Nicholson D, Ronning M. 1999. EXAFS study of zinc coordination in bacitracin A. Biochemical and Biophysical Research Communications. 1431: 433-442.
- [2]. Bhavani SM, Ballow CH. 2000. New agents for Gram-positive bacteria. Current Opinion Microbiology. 3: 528-534.
- [3]. Muhammad SA, Ahmad S, Hameed A. 2009. Antibiotic production by thermophilic *Bacillus* specie SAT-4. Pakistan Journal of Pharmaceutical Sciences. 22: 339-345.
- [4]. Kümmerer K. 2009. Antibiotics in the aquatic environment-A review-part II. Chemosphere. 75: 435-441.
- [5]. Villamil L, Tafalla C, Figueras A, Novoa B. 2002. Evaluation of immunomodulatory effects of some lactic acid bacteria in turbot (*Scophthalmus maximus*). Clinical and Diagnostic Laboratory Immunology. 9: 131-132.
- [6]. Violeta O, Oana S, Matilda C, Maria CD, Catalina V, Gheorghe C, Petruta CC. 2011. Production of biosurfactants and antifungal compounds by new strains of *Bacillus* spp. isolated from different sources. Romanian Biotechnological Letters. 16: 84-91.
- [7]. Mathur A, Rawat A, Bhatt G, Baweja S, Ahmad F, Grover A, Madhav K, Dhand M, Mathur D, Verma SK, Singh K, Dua VK. 2011. Isolation of *Bacillus* producing chitinase from soil: production and purification of chitooligosaccharides from chitin extracted from fresh water crustaceans and antimicrobial activity of chitinase. Recent Research in Science and Technology. 3: 01-06.
- [8]. Ghribi D, Abdelkefi–Mesrati L, Mnif I, Kammoun R, Ayadi I, Saadaoui I, Maktouf S, Ellouze SC. 2012. Investigation of antimicrobial activity and statistical optimization of *Bacillus subtilis* SPB1 biosurfactant production in solid-state fermentation. Journal of Biomedicine and Biotechnology. 2012: 1-12.
- [9]. Issazadeh K, Rad SK, Zarrabi S, Rahimibashar MR. 2012. Antagonism of *Bacillus* species against *Xanthomonas campestris* pv. *campestris* and *Pectobacterium carotovorum* subsp. *carotovurum*. African Journal of Microbiology Research. 6: 1615-1620.
- [10]. Kumar SN, Siji JV, Ramya R, Nambisan B, Mohandas C. 2012. Improvement of antimicrobial activity of compounds produced by *Bacillus* sp. associated with a *Rhabditid* sp. (entomopathogenic nematode) by chancing carbon and nitrogen sources in fermentation media. Journal of Microbiology, Biotechnology and Food Sciences. 1: 1424-1438.
- [11]. Shelar SS, Warang SS, Mane SP, Sutar RL, Ghosh JS. Characterization of bacteriocin produced by *Bacillus atrophaeus* strain JS-2. International Journal of Biological Chemistry. 6: 10-16.
- [12]. Al-Ajlani MM, Hasnain S. 2010. Bacteria exhibiting antimicrobial activities; screening for antibiotics and the associated genetic studies. The Open Conference Proceedings Journal. 1: 230-238.
- [13]. Salminen S, Roberfroid M, Ramos P, Fonden R. 1998. Prebiotic substrates and lactic acid bacteria. In: Lactic Acid Bacteria: Microbiology and Functional Aspects (ed. Salminen S, von Wright A), 2nd edition, pp. 343-350. Marcel Dekker Inc., New York.
- [14]. Kim DP, Chang DS, Kim SJ. 1985. Bacterial quality of fish meat paste products and isolation of thermoduric bacteria. Korean Journal of Applied Microbiology Bioengineering. 13: 409-415.
- [15]. Choorit W, Prasertsan P. 1992. Characterization of proteases produced by newly isolated and identified

- proteolytic microorganisms from fermented fish (Budu). World Journal of Microbiology & Biotechnology. 8: 284-286.
- [16]. Fuselli SR, Casales Fritz MR, Yeannes MI. 1994. Microbiology of the marination process used in anchovy (*E. anchoita*) production. Lebensmittel Wissenchaft & Technologie. 27: 214-218.
- [17]. Ringo E, Strom E, Tabacheck JA. 1995. Intestinal microflora of Salmonids: a review. Aquaculture Research. 26: 773-789.
- [18]. Chilcott CN, Wigley PJ. 1993. Isolation and toxicity of *Bacillus thuringiensis* from soil and insect habitats in New Zealand. Journal of Invertebrate Pathology. 61: 244-247.
- [19]. Sneath PHA. 1986. Endospore-forming Grampositive rods and cocci. In: Bergey's Manual of Systematic Bacteriology (ed. Sneath PHA, Mair NS, Sharpe ME, Holt JG), vol. 2, pp 1104-1139. Williams and Wilkins, Baltimore, MD.
- [20]. Reinheimer JA, Demkow MR, Condioti MC. 1990. Inhibition of coliform bacteria by lactic acid cultures. Australian Journal of Dairy Technology. 5: 5-9.
- [21]. Pinchuk IV, Bressollier P, Verneuil B, Fenet B, Serokulova I, Mégraud F, Urdaci C. 2001. In vitro anti-Helicobacter pylorii activity of the probiotic strain Bacillus subtilis 3 is due to secretion of antibiotics. Antimicrobial Agents and Chemotherapy. 45: 3156-3161.
- [22]. Binnet DH. 1998. Studies on resistance against antibiotics being betalactamase inhibitory, betalactamase activity of *Bacillus* spp. and inhibitory effects in salt-glucose consantrations on *Listeria monocytogenes* and other bacteria of *Bacillus* species isolated from raw milk (MSc Thesis). University of Gazi, Institute of Sciences and Technology, Ankara-TURKEY.
- [23]. Perez C, Suarez C, Castro GR. 1992. Production of antimicrobials by *Bacillus subtilis* MIR 15. Journal of Biotechnology. 26: 331-336.
- [24]. Perez C, Suarez C, Castro GR. 1993. Antimicrobial activity determined in strains of *Bacillus circulans* cluster. Folia Microbiologica. 38: 25-28.
- [25]. Aslim B, Sağlam N, Beyatli Y. 2002. Determination of some properties of *Bacillus* isolated from soil. Turkish Journal of Biology. 26: 41-48.
- [26]. Yilmaz M, Soran H, Beyatli Y. 2006. Antimicrobial activities of some *Bacillus* spp. strains isolated from the soil. Microbiological Research. 161: 127-131.
- [27]. Oscariz JC, Lasa I, Pisabarro AG. 1999. Detection and characterization of cerein 7, a new bacteriocin produced by *Bacillus cereus* with a broad spectrum of activity. FEMS Microbiology Letters. 178: 337-341.
- [28]. Kalayli E. 2001. Determination of plasmid DNA, PHB production and antimicrobial activity of *Bacillus* bacteria isolated from milk and fermented products (MSc Thesis). University of Gazi, Institute of Sciences and Technology Ankara-TURKEY.
- [29]. Rönkä E, Malinen E, Saarela M, Rinta-Koski M, Aarnikunnas J, Palva A. 2003. Probiotic and milk technological properties of *Lactobacillus brevis*. International Journal of Food Microbiology. 83: 63-74.
- [30]. Gilliland SE, Nelson CR, Maxwell C. 1985. Assimilation of cholesterol by *Lactobacillus acidophilus*. Applied Environmental Microbiology. 49: 377-381.
- [31]. Austin B, Baudet E, Stobie M. 1992. Inhibition of bacteria fish pathogens by *Tetraselmis suecica*. Journal of Fish Diseases. 15: 53-61.
- [32]. Katircioğlu H. 2001. Studies on metabolic and antimicrobial activity of lactic acid bacteria isolated from

rainbow trout (*Onchorynchus mykiss* Richardson, 1846) and mirror carp (*Cyprinus carpio* Linnaeus, 1758) (PhD Thesis). University of Gazi, Institute of Sciences and Technology Ankara-TURKEY.

[33]. Jiranvanichpaisal P, Chuaychuwong P, Menasveta P. 1997. The use of *Lactobacillus* sp. as the probiotic bacteria in the giant tiger shrimp (*Penaeus monodon* Fabricus). Poster Session of the 2nd Asia-Pasific Marine Biotechnology Conference and 3rd Asia-Pacific Conference on Algal Biotechnology, Phuket-Thailand.