Alınış / Received: 29.08.2024 Kabul / Accepted: 30.11.2024 Online Yayınlanma / Published Online: 25.04.2025 Araştırma Makalesi / Research Article DOI: 10.22312/sdusbed.1539732



Süleyman Demirel Üniversitesi Sağlık Bilimleri Dergisi Suleyman Demirel University Journal of Health Sciences



## Uncovering the Benefits of Epicatechin for Oxidative Stress in Human Health

İnsan Sağlığında Oksidatif Stres için Epikateşinin Faydalarının Ortaya Çıkarılması

## Okan SANCER<sup>1</sup>\*<sup>(b)</sup>, Muhammet Yusuf TEPEBAŞI<sup>2</sup><sup>(b)</sup>, Uğur ŞAHİN<sup>1</sup><sup>(b)</sup>, İlter İLHAN<sup>3</sup><sup>(b)</sup>

<sup>1</sup>Suleyman Demirel University, Innovative Technologies Application and Research Center, Genetic Research Unit, Isparta, Turkey

<sup>2</sup>Suleyman Demirel University, Medical Faculty, Medical Genetic Department, Isparta, Turkey <sup>3</sup>Suleyman Demirel University, Medical Faculty, Medical Biochemistry Department, Isparta, Turkey

\*Corresponding author: okansancer@hotmail.com

## ABSTRACT

Objective: Epicatechin (EC) is one of the major components of green tea (*Camellia sinensis*) catechins. This study investigated the effect of the amount of epicatechin obtained by brewing green tea under optimal conditions against oxidative stress induced by hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). Materials and Methods: In peripheral blood mononuclear cells (PBMCs), the amount of epicatechin determined by brewing green tea under optimum conditions was applied against 250 µM H<sub>2</sub>O<sub>2</sub> and cell viability was determined by 3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) test, total antioxidant status (TAS) levels and total oxidant status (TOS) levels were determined by biochemical analysis, apoptosis-related Bax, Bcl2, p53 expression analysis was determined by quantitative real time polimerase chain reaction (qRT-PCR) method. Results: The cell viability was significantly higher in the H<sub>2</sub>O<sub>2</sub>+EC group than in the  $H_2O_2$  group (p<0.001). TOS and TAS levels were changed considerably in the  $H_2O_2$ +EC group compared to the  $H_2O_2$  group (p<0.05 and p<0.001, respectively). Bax, p53 expression level decreased in  $H_2O_2$ +epicatechin treated cells compared to  $H_2O_2$  treated cells (p<0.001 and p<0.01 respectively), while Bcl2 expression level increased in H<sub>2</sub>O<sub>2</sub>+epicatechin treated cells compared to H<sub>2</sub>O<sub>2</sub> treated cells p<0.01). Conclusion: The results show that the amount of epicatechin obtained from brewing green tea under optimum conditions has a protective effect on peripheral blood mononuclear cells (PBMCs) against H<sub>2</sub>O<sub>2</sub> induced oxidative stress. Specifically, it was concluded that epicatechin increased cell viability, decreased oxidative stress markers and modulated the expression of key apoptosis-related proteins, thus promoting cell survival.

Keywords: Bax, Bcl2, Epicatechin, Oxidative stress, p53

## ÖΖ

Amaç: Epikateşin (EC) yeşil çay (Camellia sinensis) kateşinlerinin ana bileşenlerinden biridir. Bu çalışmada, yeşil cayın optimum kosullarda demlenmesiyle elde edilen epikatesin miktarının hidrojen peroksit (H<sub>2</sub>O<sub>2</sub>) tarafından indüklenen oksidatif strese karşı etkisi araştırılmıştır. Materyal ve Metot: Periferik kan mononükleer hücrelerinde (PBMCs), yeşil çayın optimum koşullarda demlenmesi ile belirlenen epikateşin miktarı 250 µM H<sub>2</sub>O<sub>2</sub>'ye karşı uygulanmış ve hücre canlılığı 3-(4, 5-dimetiltiyazol-2-yl)-2,5-difeniltetrazolyum bromür (MTT) testi ile, total antioksidan seviyeleri (TAS) ve total oksidan seviyeleri (TOS) biyokimyasal analiz ile, apoptoz ile ilişkili Bax, Bcl2, p53 ekspresyon analizi gerçek zamanlı kantitatif polimeraz zincir reaksiyonu (qRT-PCR) yöntemi ile belirlenmiştir. Bulgular: H<sub>2</sub>O<sub>2</sub>+EC grubunda hücre canlılığı H<sub>2</sub>O<sub>2</sub> grubuna göre anlamlı derecede yüksek bulunmuştur (p<0.001). TOS ve TAS seviyeleri,  $H_2O_2$  grubuna kıyasla  $H_2O_2$ +EC grubunda önemli ölçüde değişmiştir (sırasıyla p<0.05 ve p < 0.001). Bax, p53 ifade düzeyi H<sub>2</sub>O<sub>2</sub>+epikateşin uygulanan hücrelerde H<sub>2</sub>O<sub>2</sub> uygulanan hücrelere kıyasla azalırken (sırasıyla p<0.001 ve p<0.01), Bcl2 ifade düzeyi H<sub>2</sub>O<sub>2</sub>+epikateşin uygulanan hücrelerde H<sub>2</sub>O<sub>2</sub> uygulanan hücrelere kıyasla artmıştır (p<0.01). Sonuç: Sonuçlar, yeşil çayın optimum koşullar altında demlenmesinden elde edilen epikateşin miktarının H<sub>2</sub>O<sub>2</sub> kaynaklı oksidatif strese karşı periferak kan mononükleer hücreler (PKMH) üzerinde koruyucu bir etkiye sahip olduğunu göstermektedir. Spesifik olarak, epikateşinin hücre canlılığını artırdığı, oksidatif stres belirteçlerini azalttığı ve apoptozla ilgili anahtar proteinlerin ekspresyonunu modüle ettiği, böylece hücre sağ kalımını desteklediği sonucuna ulaşılmıştır.

Anahtar Kelimeler: Bax, Bcl2, Epikateşin, Oksidatif stres, p53

#### **INTRODUCTION**

Epicatechin EC is one of the main components of green tea catechins (1). Green tea has characteristics of meta-5,7-dihydroxy groups in chain A and dihydroxy or trihydroxy groups in chain B (2). The B chain seems essential to the antioxidant reactions (3). Antioxidant activity is a molecule or ion's capacity to prevent other molecules' oxidative reactions (4). EC has effective and direct antioxidant activity. By its strong antioxidant capacity, it scavenges free radicals in cells. Compared with vitamin C and vitamin E, the antioxidant capacity of EC is 20 and 50 times greater. The biological activity of ECs is mainly a result of interactions with proteins and lipids. These interactions result in an effect on the levels of oxidants (5).

Oxidative stress, characterised by an imbalance between free radicals and antioxidants, has a significant impact on several health conditions such as infertility, cancer, diabetes, metabolic syndrome, atherosclerosis, neurodegenerative, cardiovascular, gastrointestinal and liver diseases.  $H_2O_2$  is a crucial reactive oxygen species (ROS) and plays a significant role in biological processes. Generally, when it exceeds 50  $\mu$ M, it causes oxidative damage in tissues and organs and elicits an inflammatory response. Antioxidants are essential for neutralising free radicals and thus preventing oxidative damage. They can be enzymatic or non-enzymatic and work by scavenging free radicals, chelating metal ions or upregulating other antioxidant defences (6).

Green tea is considered one of the healthiest drinks around the globe. Its acceptance in this way is due to its rich structure in polyphenols (7, 8). While many studies have shown that plant-derived flavonoids have excellent antioxidant activity,(9, 10) research on green tea in recent years has focused on the relationship between consumption and disease prevention (7).

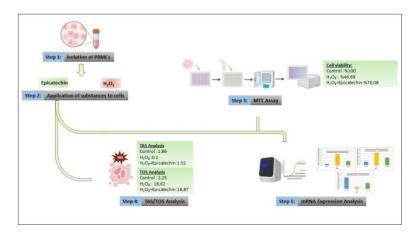
The catechins and sensory properties may differ depending on the green tea brewing conditions. Our aim with this study was to understand if the amount of epicatechin determined according to the optimum conditions during green tea brewing has a protective effect on cells against  $H_2O_2$ .

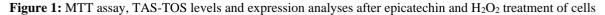
## **MATERIAL and METHOD**

#### **Isolation of Human PBMC**

Peripheral venous blood was collected from heparin tubes from healthy volunteer who had not been exposed to radiation or any drugs or smoked for six months. The Declaration of Helsinki was followed in the study. This study was approved by the Süleyman Demirel University Medical Faculty Ethics Committee (decision dated 05.12.2023 and numbered 15/270). PBMCS were isolated by Histopaque 1077 (Sigma-Aldrich, Switzerland).

Cell viability was determined to be 98% using trypan blue stain. The medium was changed once every 24 hours (11). The workflow of the study is given in Figure 1.





## MTT Assay

PBMCs were seeded in 96-well flat-bottomed microplates (Sarstedt AG, Germany) at  $1 \times 10^4$  cells/well density. A 5% CO<sub>2</sub> incubator at 37 °C was incubated for 24 h before any treatments.

For the best result of taste and sensory properties, brewing at 85 °C for 3 minutes represents the optimal condition. Under these conditions, an epicatechin maximum of 6.75 mg/100 mL was determined (12). Epicatechin (Sigma Chemical Co., USA) of 67.5 ppm was cultured with cells in an incubator for 24 h (37 °C, 5% CO<sub>2</sub>). During the last hour, these cells (except the control group) were incubated with 250  $\mu$ M H<sub>2</sub>O<sub>2</sub>(13).

The medium consisted of the following components: RPMI-1640 (Biological Industries, Israel) medium, 10% FBS (Sigma - Aldrich, USA) and 100 IU/mL penicillin, 100  $\mu$ g/mL streptomycin (Sigma - Aldrich, USA). MTT (Sigma, USA) final concentration was adjusted to 0.5 mg/mL. The resulting formazan crystals were dissolved in DMSO. A multiscan plate reader (Synergy HTX BioTek, USA) was used to measure cell viability at 570 nm (14). Cell viability was evaluated in percentage relative to the control group, denoted as 100%. Three individual wells were measured per treatment point.

## **Biochemical Analysis**

After the culture step, plates were centrifuged at 1800g for 6 minutes, and the pellet was washed with PBS. An ultrasonic homogenizer was used to homogenize the cells. After the second centrifugation, supernatants were transferred to Eppendorf tubes for analysis (15).

TAS and TOS levels were measured by spectrophotometric method (Beckman Coulter AU 5800, USA) in triplicate with commercial kits (Rel Assay Diagnostics, Türkiye) according to the kit protocol (16). The results were expressed as millimolar Trolox equivalents per liter in TAS and micromolar hydrogen peroxide equivalent per liter ( $\mu$ mol H<sub>2</sub>O<sub>2</sub> Eqv/L) in TOS (17).

## **RT-qPCR** Analysis on mRNA

Total RNA extraction, purity and concentration measurement, cDNA extraction were performed similar to our previous study and according to the manufacturer's protocol (18). Primers were designed to detect specific mRNA sequences. The NCBI website was used to test possible primer sequences. Bax (F:5'-CAGGGGGCCCTTTTGCTTCA-3' R:5'-GGAAAAAGACCTCTCGGGGGG-3'), Bcl-2 (F:5'-AAAAATACAACATCACAGAGGAAGT-3' R:5'-TCCCGGTTATCGTACCCTGT-3'), p53 (F:5'-ACCTATGGAAACTACTTCCTGAAA-3' R:5'-GCTGCCCTGGTAGGTTTTCT -3') primers were designed to amplify. ACTB (F: 5'-GCTCGCCTTGCCGAT-3' R:5'- AGGTAGTCAGTCAGGTCCCG-3') expression was used for normalization. The manufacturer's instructions were followed for real-time RT-PCR conditions. The  $2^{-\Delta\Delta Ct}$  comparative method was used for relative quantification of gene expression. To determine amplification specificity, qPCR products were evaluated using melting curves. Each sample was run in triplicate.

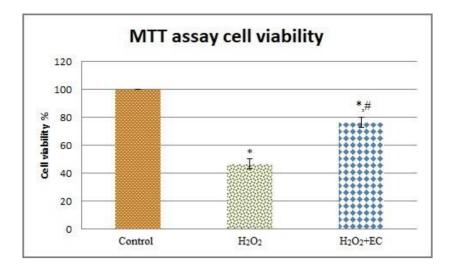
## **Statistical Analysis**

The results of the expression study were evaluated using SPSS 18.0 statistical analysis software (SPSS Inc., Chicago, IL). One-way ANOVA was used to analyze the results of the expression, MTT, TAS, and TOS levels. LSD and TUKEY tests were used as post-hoc tests. p<0.05 was considered to be significant.

# **RESULTS**

# MTT Assay Results

Cell viability was significantly lower in the  $H_2O_2$  and  $H_2O_2$ +epicatechin groups compared with the control group (p<0.001). In the comparison of cell viability in the  $H_2O_2$  group and the  $H_2O_2$ +epicatechin group, the  $H_2O_2$ +epicatechin group was significantly higher (p<0.001) (Figure 2).



**Figure 2:** Cell viabilty. Data expressed as mean±SD. Comparision between groups and result were assessed by oneway ANOVA test. \*p<0.001 ascompared to the control group, <sup>#</sup>p<0.001 compared to the H<sub>2</sub>O<sub>2</sub>-treated group.

#### **Biochemical Results**

TOS level was significantly higher in the  $H_2O_2$  and  $H_2O_2$ +epicatechin groups than in the control group (p<0.001). TOS level was considerably higher in the  $H_2O_2$  group than in the  $H_2O_2$ +epicatechin group (p<0.05). TAS level was significantly lower in the  $H_2O_2$  group than in the control group (p<0.001) and considerably higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001). TAS level was significantly higher in the  $H_2O_2$ +epicatechin group (p<0.001).

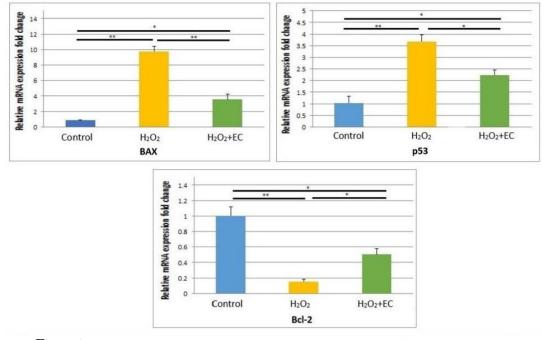
Groups	Negative Control	$H_2O_2$	H <sub>2</sub> O <sub>2+</sub> EC
TOS (μmol H <sub>2</sub> O <sub>2</sub> Eq./L)	2.25±0.74	18.62±0.82*	16.87±0.53*,#
TAS (mmol TroloxEq./L)	1.86±0.08	$0.20\pm0.04^*$	1.52±0.04*,##
OSI (µmol H2O2 equiv./lt)/(mmol Trolox equiv./lt x 10)	0.12±0.04	9.36±1.49	1.11±0.02

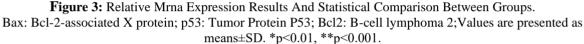
Table 1: TOS and TAS levels of PBMCs

Data are expressed as mean $\pm$ SD. One-way ANOVA test was used to assess comparisons between groups and results of oxidative stress markers. LSD tests were used as post-hoc tests. \*p<0.001 compared to the control group; #p<0.05, ##p<0.001 compared to the H<sub>2</sub>O<sub>2</sub> treated group.

#### **Expression Analysis of Bax, Bcl, p53**

The relative mRNA level of Bax in the  $H_2O_2$ ,  $H_2O_2$ +epicatechin groups increased significantly compared to the control group (respectively; p<0.001 and p<0.01) and lower significantly in  $H_2O_2$ +epicatechin group compared to the  $H_2O_2$  group (p<0.001). The relative mRNA level of p53 in the  $H_2O_2$ ,  $H_2O_2$ +epicatechin groups increased significantly compared to the control group (respectively; p<0.001 and p<0.01) and lower significantly in  $H_2O_2$ +epicatechin group compared to the H<sub>2</sub>O<sub>2</sub> group ( p<0.001 and p<0.01) and lower significantly in  $H_2O_2$ +epicatechin group compared to the H<sub>2</sub>O<sub>2</sub> group ( p<0.01). The relative mRNA level of Bcl-2 in the H<sub>2</sub>O<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>+epicatechin groups decreased significantly compared to the control group (respectively; p<0.001 and p<0.01) and lower significantly compared to the control group (respectively; p<0.001 and p<0.01) and lower significantly compared to the control group (respectively; p<0.001 and p<0.01).





#### **DISCUSSION and CONCLUSION**

PBMCs, consisting mainly of lymphocytes and monocytes, are a readily available blood cell fraction with high research potential for testing the effects of dietary intake. At the gene expression level, they reflect the impact of environmental changes. Many studies demonstrated the utility of PBMCs in showing the effects of nutrition, training, and vigorous exercise on mitochondrial oxidative balance, biosynthesis, dynamics, and antioxidant capabilities (19,21). In this context, PBMCs were considered more appropriate for our research.

Oxidative stress is the result of an imbalance between free radicals and antioxidants. It causes changes in the structure of cell membranes, lipids, proteins, lipoproteins, and DNA. Mitochondria are the most important endogenous source of ROS generation, as they play a role in forming ATP through oxidative phosphorylation, which reduces molecular  $O_2$  to  $H_2O$  via the electron transport chain (22). A study showed that heavy exercise enhances the oxidative stress-induced apoptosis (23). Another study demonstrated that exopolysaccharide-selenium nanoparticles promoted cell survival by maintaining over 90% cell viability under oxidative stress caused by 0.4 mM  $H_2O_2$  in HepG2 cells (24).  $H_2O_2$  is a source of reactive oxygen species, and functions act to induce oxidative stress (25).  $H_2O_2$  is commonly considered a cytotoxic substance that must be reduced by antioxidant defense enzymes (26).

An ex-vivo study has suggested that green tea consumption prevents LDH oxidation in humans, while Epicatechin also plays a role in the prevention of neurodegenerative diseases such as Alzheimer's and Parkinson's (27). In addition, in an in vivo study, tea catechins were observed to reduce the formation of atherosclerosis in mice with apolipoprotein E deficiency (28), while another study found that the antioxidant capacity of epicatechin in human plasma increased by 40% compared to non-users (29).

Our results show that the treatment with  $H_2O_2$  led to a significant decrease in the viability of the PBMCs. However, epicatechin had a protective effect on cell viability when used against  $H_2O_2$ , which causes oxidative stress. In addition, our results showed that the  $H_2O_2$ -induced increase in TOS levels in PBMCs decreased with epicatechin treatment, while TAS levels increased.

In a study, it was found that after treatment of mouse granulosa cells and human granulosa cells with  $H_2O_2$ , the viability rate of the cells decreased with concentration, while the expression levels of p53 and Bax increased. These findings suggest that  $H_2O_2$ -oxidative stress may be a factor in the onset of apoptosis. Because when DNA damage is limited and reversible, cells stop proliferating. Some cells exposed to DNA damage enter the cell cycle. However, when DNA damage is irreparable, cells undergo immediate apoptosis, thereby causing induction of p53 and p21 expression (30). In another study, (-)-epicatechin was found to upregulate death receptors (DR4/DR5) and modulate pro-apoptotic proteins in MDA-MB-231 cells. In contrast, it did not activate the death receptor in MCF-7 cells (31).

Similar to the studies conducted with cell culture in the literature, in our research, Bax, p53 expression increased while Bcl2 expression decreased in cells treated with  $H_2O_2$  only compared to control cells. In  $H_2O_2$ +EC treated cells, Bax, p53 expression decreased while Bcl2 expression level decreased compared to the  $H_2O_2$  group. Bax, p53, and Bcl2 genes networks and functions are present in Figure 4 (https://genemania.org/).

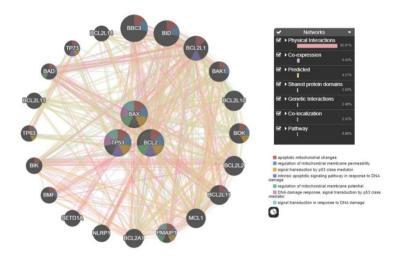


Figure 4: Bax, p53, Bcl2 genes networks and functions

The mechanisms of action of natural antioxidants are unclear. Further research is needed to identify the active target sites. In this direction, in our study, the mechanism by which the amount of epicatechin determined by brewing green tea under optimal conditions plays a protective role against  $H_2O_2$ -induced oxidative stress was revealed by genetic and biochemical tests. However, in addition to cellular studies, it is thought that the amount of epicatechin obtained by brewing and consuming green tea under optimal conditions will be insufficient to reduce the effects of oxidative stress in people who exercise excessively. We believe that this study may provide a basis for in vivo studies.

**Declaration of Ethical Code:** In this study, we undertake that all the rules required to be followed within the scope of the "Higher Education Institutions Scientific Research and Publication Ethics Directive" are complied with, and that none of the actions stated under the heading "Actions Against Scientific Research and Publication Ethics" are not carried out.

This study was approved by the Süleyman Demirel University Medical Faculty Ethics Committee (decision dated 05.12.2023 and numbered 15/270).

#### REFERENCES

1. Salem YN, Sheribah ZA, Sharaf El-Din MK, Fathy MES. A novel optimized eco-friendly simple spectrofluorimetric method for the determination of total catechins in green tea extract: Application to commercial tablet. Luminescence 2024;39(3):e4727.

- 2. Balentine DA, Wiseman SA, Bouwens LC. The chemistry of tea flavonoids. Crit Rev Food Sci Nutr 1997;37(8):693-704.
- 3. Roh E, Kim J-E, Kwon JY, Park JS, Bode AM, Dong Z, et al. Molecular mechanisms of green tea polyphenols with protective effects against skin photoaging. Crit Rev Food Sci Nutr 2017;57(8):1631-7.
- 4. El-Lateef HMA, El-Dabea T, Khalaf MM, Abu-Dief AM. Recent overview of potent antioxidant activity of coordination compounds. Antioxidants 2023;12(2):213.
- 5. Zhang R, Wang J, Xia R, Li D, Wang F. Antioxidant processes involving epicatechin decreased symptoms of pine wilt disease. Front Plant Sci 2022;13:1015970.
- 6. Kumar H, Dhalaria R, Guleria S, Cimler R, Sharma R, Siddiqui SA, et al. Anti-oxidant potential of plants and probiotic spp. in alleviating oxidative stress induced by H<sub>2</sub>O<sub>2</sub>. Biomed Pharmacother 2023;165:115022.
- Sarma A, Bania R, Das MK. Green tea: Current trends and prospects in nutraceutical and pharmaceutical aspects. J Herb Med 2023:100694.
- Tural B, Ertaş E, Batıbay H, Tural S. Comparative Study on Silver Nanoparticle Synthesis Using Male and Female Pistacia Khinjuk Leaf Extracts: Enhanced Efficacy of Female Leaf Extracts. ChemistrySelect 2024;9(30):e202402117.
- 9. Shen N, Wang T, Gan Q, Liu S, Wang L, Jin B. Plant flavonoids: Classification, distribution, biosynthesis, and antioxidant activity. Food Chem. 2022;383:132531.
- 10. Wang Z, Zhou X, Chang H, Shu Z, Gou H, Zheng Y, et al. Antioxidant analysis of flavonoids extracted from Artemisia argyi leaf and their antibacterial activities against food-borne pathogens Escherichia coli and Staphylococcus aureus. Biologia 2024:1-9.
- 11. Millones-Gómez PA, De la Garza-Ramos MA, Urrutia-Baca VH, Hernandez-Martinez HC, Marín DAH, Medina CAM. Cytotoxicity of Peruvian propolis and Psidium guajava on human gingival fibroblasts, PBMCs and HeLa cells. F1000Research 2022;11.
- 12. Saklar S, Ertas E, Ozdemir IS, Karadeniz B. Effects of different brewing conditions on catechin content and sensory acceptance in Turkish green tea infusions. J Food Technol 2015;52(10):6639-46.
- 13. Jeong M-J, Lim D-S, Kim SO, Park C, Leem S-H, Lee H, et al. Protection of oxidative stress-induced DNA damage and apoptosis by rosmarinic acid in murine myoblast C2C12 cells. Biotechnol Bioprocess Eng 2022;27(2):171-82.
- 14. Kang KA, Lee KH, Zhang R, Piao M, Chae S, Kim KN, et al. Caffeic acid protects hydrogen peroxide induced cell damage in WI-38 human lung fibroblast cells. Biol Pharm Bull 2006;29(9):1820-4.
- 15. Oğuz EK, Arihan O, Oğuz AR. Oxidative and genotoxic effects of bisphenol A on primary gill cell culture of Lake Van Fish (Alburnus tarichi Güldenstädt, 1814). Chem Ecol 2018;34(10):914-24.
- 16. Altindag O, Erel O, Soran N, Celik H, Selek S. Total oxidative/anti-oxidative status and relation to bone mineral density in osteoporosis. Rheumatol Int 2008;28:317-21.
- 17. Erel O. A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. Clin Biochem 2004;37(4):277-85.
- 18. Sancer O, Şahin U, Çetin ES, Tepebaşi MY, Cezaroğlu Y, Bilir G, et al. Effect of Laurus nobilis on bacteria and human transforming growth factor-β1. AMB Rev Assoc Med Bras 2024;70(3):e20230683.
- Sureda A, Martorell M, Bibiloni MdM, Bouzas C, Gallardo-Alfaro L, Mateos D, et al. Effect of free fatty acids on inflammatory gene expression and hydrogen peroxide production by ex vivo blood mononuclear cells. Nutrients 2020;12(1):146.
- 20. Da Rosa PC, Bertomeu JB, Royes LFF, Osiecki R. The physical exercise-induced oxidative/inflammatory response in peripheral blood mononuclear cells: signaling cellular energetic stress situations. Life Sci 2023;321:121440.
- 21. Minutolo A, Gismondi A, Chirico R, Di Marco G, Petrone V, Fanelli M, et al. Antioxidant Phytocomplexes Extracted from Pomegranate (Punica granatum L.) Using Hydrodynamic Cavitation Show Potential Anticancer Activity In Vitro. Antioxidants 2023;12(8):1560.
- 22. Hajam YA, Rani R, Ganie SY, Sheikh TA, Javaid D, Qadri SS, et al. Oxidative stress in human pathology and aging: Molecular mechanisms and perspectives. Cells 2022;11(3):552.
- 23. Wang J-S, Huang Y-H. Effects of exercise intensity on lymphocyte apoptosis induced by oxidative stress in men. Eur J Appl Physiol 2005;95:290-7.
- 24. Xiao Y, Zhang X, Huang Q. Protective effects of Cordyceps sinensis exopolysaccharide-selenium nanoparticles on H2O2-induced oxidative stress in HepG2 cells. Int J Biol Macromol 2022;213:339-51.
- 25. Jeong Y-M, Choi Y-G, Kim D-S, Park S-H, Yoon J-A, Kwon S-B, et al. Cytoprotective effect of green tea extract and quercetin against hydrogen peroxide-induced oxidative stress. Arch Pharm Res 2005;28:1251-6.
- 26. Halliwell B, Clement MV, Long LH. Hydrogen peroxide in the human body. FEBS Lett 2000;486(1):10-3.
- 27. Miura Y, Chiba T, Miura S, Tomita I, Umegaki K, Ikeda M, et al. Green tea polyphenols (flavan 3-ols) prevent oxidative modification of low density lipoproteins: an ex vivo study in humans. J Nutr Biochem 2000;11(4):216-22.
- 28. Miura Y, Chiba T, Tomita I, Koizumi H, Miura S, Umegaki K, et al. Tea catechins prevent the development of atherosclerosis in apoprotein E–deficient mice. J Nutr 2001;131(1):27-32.
- 29. Rein D, Lotito S, Holt RR, Keen CL, Schmitz HH, Fraga CG. Epicatechin in human plasma: in vivo determination and effect of chocolate consumption on plasma oxidation status. J Nutr 2000;130(8):2109S-14S.
- 30. Wang Y, Li Q, Ma Z, Xu H, Peng F, Chen B, et al. β-Nicotinamide Mononucleotide Alleviates Hydrogen Peroxide-Induced Cell Cycle Arrest and Death in Ovarian Granulosa Cells. Int J Mol Sci 2023;24(21):15666.
- 31. Pereyra-Vergara F, Olivares-Corichi IM, Perez-Ruiz AG, Luna-Arias JP, García-Sánchez JR. Apoptosis induced by (-)-epicatechin in human breast cancer cells is mediated by reactive oxygen species. Molecules 2020;25(5):1020.