



Antimicrobial activities of chitosan-based edible films produced by adding different macrofungi extracts and plants essential oils

Kağan VERYER¹, Özge SÜFER², Merve BÜLBÜL³, Mukaddes GÖKYERMEZ⁴, Fuat BOZOK⁵

^{1,5}Osmaniye Korkut Ata University, Engineering and Natural Sciences Faculty, Department of Biology, Osmaniye, Türkiye

²Osmaniye Korkut Ata University, Engineering and Natural Sciences Faculty, Department of Food Engineering, Osmaniye, Türkiye

^{3,4}Osmaniye Korkut Ata University, Postgraduate Education Institute, Osmaniye, Türkiye

*fbozok@osmaniye.edu.tr, ¹kaganveryer@osmaniye.edu.tr, ²ozgesufer@osmaniye.edu.tr,

³merve.bulbul.ea@gmail.com, ⁴drkyrmz@hotmail.com

Received : 02.09.2024

Accepted : 20.10.2024

Online : 15.02.2025

Farklı makromantar ekstraktları ve bitki uçucu yağları eklenerek üretilen kitosan bazlı yenilebilir filmlerin antimikrobiyal aktiviteleri

Abstract: This research aims to reveal the antimicrobial activity of chitosan-based edible films containing different mushroom extracts and plant essential oils. In this study, edible films were produced using the essential oils of *Satureja cuneifolia* Ten., *Mentha longifolia* (L.) Hudson subsp. *typhoides* (Brig.) Harley var. *typhoides* (L.) Hudson and extracts of *Amanita caesarea* (Scop.) Pers. and *Boletus reticulatus* Schaeff. collected from different localities of Osmaniye province. The antimicrobial activities of these films were investigated on *Escherichia coli* by using the Kirby-Bauer disk diffusion test. At the end of the research, it was determined that the edible film obtained by adding *S. cuneifolia* Ten. essential oil (3%) had the highest antimicrobial activity. And also, it could be said that all the edible films produced had antimicrobial activity.

Key words: Antibacterial activity, edible film, volatile, mushroom

Özet: Bu araştırma, farklı mantar ekstraktı ve bitki uçucu yağı içeren kitosan temelli yenilebilir filmlerin antimikrobiyal aktivitesini açığa çıkarmayı amaçlamaktadır. Bu çalışmada, Osmaniye'nin farklı lokalitelerinden toplanan *Satureja cuneifolia* Ten., *Mentha longifolia* (L.) Hudson subsp. *typhoides* (Brig.) Harley var. *typhoides* (L.) Hudson'un uçucu yağları ve *Amanita caesarea* (Scop.) Pers. ve *Boletus reticulatus* Schaeff. ekstraktları kullanılarak yenilebilir filmler üretilmiştir. Üretilen bu filmlerin *Escherichia coli* üzerindeki antimikrobiyal aktiviteleri Kirby-Bauer disk difüzyon testi kullanılarak araştırılmıştır. Araştırma sonunda, *S. cuneifolia* Ten.'in uçucu yağının (%3) ilave edilmesiyle üretilen yenilebilir filmin en yüksek antimikrobiyal aktiviteye sahip olduğu belirlenmiştir. Bununla birlikte, üretilen tüm yenilebilir filmlerin antimikrobiyal aktiviteye sahip olduğu söylenebilir.

Anahtar Kelimeler: Antibakteriyel aktivite, yenilebilir film, aroma, mantar

Citation: Veryer K, Süfer Ö, Bülbül M, Gökyermez M, Bozok F (2025). Antimicrobial activities of chitosan-based edible films produced by adding different macrofungi extracts and plants essential oils. *Anatolian Journal of Botany* 9(1): 1-5.

1. Introduction

The deterioration of synthetic plastic materials has been a major cause of the growing interest in edible films in recent years (Kumar et al., 2022). Plastics are among the waste elements that harm the environment when interacting with nature since they take a long time to break down in the natural world (Lakhiar et al., 2024). However, they swiftly deteriorate after being thrown away, edible coatings from consumable goods can improve food quality by extending its shelf life (Xu et al., 2005). Chitosan is one of the polymer materials that is commonly utilized in the creation of edible films (Ebrahimi et al., 2024; Singh et al., 2024). It is created by deacetylating chitin, which is found in large quantities in shrimp and crab shells (Priyadarshi et al., 2018; Campalani et al., 2024). Chitin becomes more reactive and soluble in diluted acetic and citric acids, as a result of the deacetylation process (Ma et al., 2021). A valuable and promising biomaterial, chitosan is a derivative of chitin and a helpful bioactive polymer that has gained attention recently (Safarzadeh et al., 2024). Important

characteristics of the edible polymer chitosan include its adsorption, non-toxicity, biodegradability, and biocompatibility (Amin et al., 2024). Chitosan finds application in the food business because of its unique chemical and physical characteristics (Gutiérrez, 2017). Chitosan film is a viable substitute for commercial polymers used to prolong food shelf life because it is a rigid, flexible, semi-permeable substance that rips readily (Butler vd., 1996; Kittur vd., 1998). In recent years, studies on chitosan-based edible films have started to examine their antibacterial and antioxidant properties by adding essential oils derived from aromatic and medicinal plants (Altiok et al., 2010). The biological effects of essential oils differ since they are composed of a diverse range of chemicals (Wińska et al., 2019; Parveen et al., 2024).

Numerous experts have found that non-toxic biocompatible chitosan has antibacterial and antioxidant properties in earlier investigations (Begin and Calsteren, 1999; Xie et al., 2001; Rabea et al., 2003; Yen et al., 2008; Goy et al., 2009; Raafat and Sahl, 2009; Kong et al., 2010; Siripatrawan and

Harte, 2010; Leceta et al., 2013; Ke et al., 2021). Several investigations have been carried out to examine the antibacterial and antioxidant activities of mushroom extracts (Mau et al., 2002; Yang et al., 2002; Cheung et al., 2003; Gao et al., 2005; Gezer vd., 2006; Puttaraju et al., 2006; Barros et al., 2007; Jayakumar et al., 2009; Smolskaite et al., 2015). A few studies were conducted previously on edible films produced by adding essential oil and mushroom extract (Kumar et al., 2021; John, 2022; Kaya et al., 2022). However, there is no sufficient study in the literature on chitosan-based films containing the extracts of *Boletus reticulatus* Schaeff. and *Amanita caesarea* (Scop.) Pers. and the essential oils of *Satureja cuneifolia* Ten., and *Mentha longifolia* (L.) Hudson subsp. *typhoides* (Brig.) Harley var. *typhoides* (L.) Hudson. This study aims to investigate the antimicrobial activities of chitosan-based edible films obtained by adding the extracts of *A. caesarea* (Scop.) Pers., and *B. reticulatus* Schaeff. macrofungi and the essential oils of *S. cuneifolia* Ten., and *M. longifolia* (L.) Hudson subsp. *typhoides* (Brig.) Harley var. *typhoides* (L.) Hudson. plants collected from Osmaniye province.

2. Materials and Method

2.1. Sampling, extraction, production and characterization of edible films

The localities of plant and fungal species are given in Table 1. Edible film production steps and physical and chemical analyses of the produced films were mentioned in the studies conducted by Bülbül et al., (2023) and Gökyermez et al., (2023). Plant and fungal species were collected and identified by the last author.

2.2. Antimicrobial activity

In the study, *Escherichia coli* was used to determine the antimicrobial activities of the edible films. The Kirby-Bauer disc diffusion assay was used to determine the antibacterial activities of the produced edible films. According to the assay, the commercial Müller-Hinton Agar (MHA, Merck) medium was autoclaved at 121°C. After the processing, the obtained hot liquid (70 °C) medium was poured into sterile petri plates of 9 cm diameter. After the cooling of medium, The bacterial strain was spread on the surface of the rigid MHA plate with a swab and pre-incubated for 10 minutes. The inoculated samples were incubated at 25 °C for 15 minutes and then at 37°C for 24 hours. The resulting inhibition zones were measured with a ruler in mm (Sedefoğlu et al., 2023). Assays were performed in three replicates.

2.3. Statistical analysis

The results were evaluated using the one-way analysis of variance Duncan test (95% confidence interval) with the help of the Statistical Package for the Social Sciences

(SPSS) program (IBM Statistics, USA) (Version 18.0). In addition, the effects of the variables on the outputs were determined by two-way analysis of variance.

3. Results and Discussion

The research by Bülbül et al. (2023) and Gökyermez et al. (2023) displayed the edible films' physical (thickness, moisture content, water solubility, opacity, UV-Vis, FTIR, and SEM) and chemical analysis (total phenolic compound content, DPPH activity) results. However, *E. coli* was used to examine the films' antibacterial properties in the present study. As seen in Figure 1, it was determined that antimicrobial activity increased with the increase in the amount of mushroom extracts and essential oils added to the edible films. It was determined that the edible film obtained by adding *S. cuneifolia* Ten. essential oil (3%) had the highest antimicrobial activity. And also, it could be said that all the edible films produced had antimicrobial activity.

In several previous studies, chitosan-based edible films were produced using different samples, and their various activities (antioxidant and antimicrobial activities, etc.) were investigated (Yuan et al., 2016; Hromiš et al., 2017; Kaya et al., 2018; Xu et al., 2021; Sarfraz et al., 2024). It was determined that these produced films had antioxidant and antimicrobial activities.

Gómez-Estaca et al. (2009) produced a chitosan-based film utilizing clove oil and used this film to preserve raw sliced salmon and investigated its antibacterial properties against the bacterial strains (*L. acidophilus*, *P. fluorescens*, *L. innocua* and *E. coli*.) At the end of the study, a decrease in total bacterial growth was observed after 11 days of storage, and therefore it was stated that, the produced films are suitable for use as an active packaging substance applied to fish products. In the study conducted by Sánchez-González et al. (2009), chitosan-based edible films including different concentrations of *Melaleuca alternifolia* (Maiden & Betche) Cheel essential oil, were obtained and the water vapor permeability, mechanical, and optical features of dried films were investigated to evaluate the effect of incorporating essential oil into these films. In addition, composite films were photographed by utilizing a scanning electron microscope (SEM) and their antibacterial features were tested against two bacterial strains (*Listeria monocytogenes* and *Penicillium italicum*). It was found that, chitosan-based films have a significant antimicrobial activity. Handayasari et al (2019) produced chitosan-based films containing garlic oil and nitrite-added gelatin-chitosan. They also investigated the mechanical properties and antibacterial effect of the films. At the end of the study, the tensile strength of the film increased gradually with the increasing amount of chitosan, but the elongation at break decreased. Furthermore, with the addition of the oil, the antibacterial activity of the films increased significantly. In

Table 1. Localities of plant and fungal species

Samples	Localities	GPS coordinates	Collection Date
<i>Mentha longifolia</i> subsp. <i>typhoides</i> var. <i>typhoides</i>	Bahçe, Yukarı Kardere village, Mandal deresi place	37° 13'K, 36° 37'D, 878 m,	10.07.2021
<i>Satureja cuneifolia</i>	Düziçi, Baskonuş high plateau, Odunluk place	37° 21'K, 36°30'D, 1681 m,	07.09.2021
<i>Amanita caesarea</i>	Osmaniye, Zorkun high plateau, Karıncalı place	36°57'K, 36°20'D, 1324 m	08.10.2021
<i>Boletus reticulatus</i>	Osmaniye, Zorkun high plateau, Karıncalı place	36°57'K, 36°19'D, 1240 m	08.10.2021

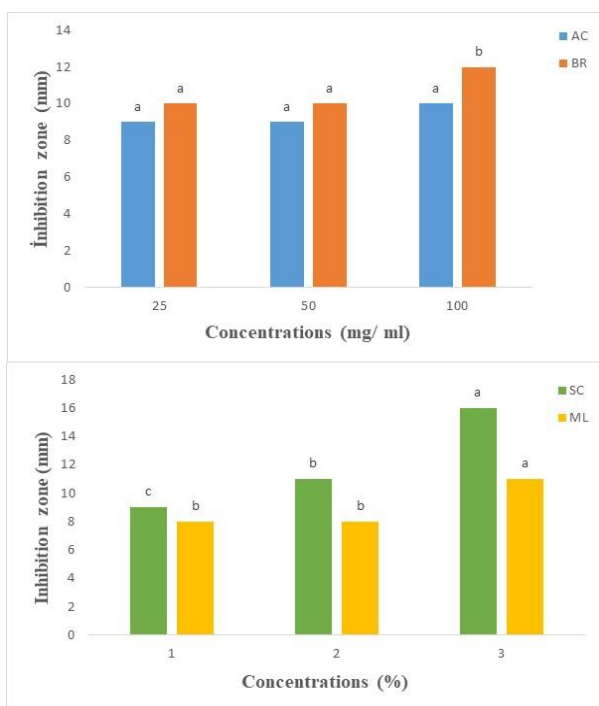


Figure 1. Antimicrobial activities of chitosan-based edible films (AC: *Amanita caesarea*, BR: *Boletus reticulatus*, SC: *Satureja cunefolia*, ML: *Mentha longifolia* subsp. *typhoides* var. *typhoides*). Mean with different letter in columns is significant at $p < 0.05$.

the study by Şimşek et al. (2020), essential oils of *Eucalyptus globulus* Labill., *Schinus molle* L., and *Santolina chamaecyparissus* L. gathered in Osmaniye (province of Türkiye) were extracted by using a Clevenger apparatus. Carboxy-methyl cellulose films were obtained by utilizing different concentrations of the oils to investigate the physical, chemical, and antibacterial effects of these films. The conclusion of the study revealed that films with added essential oil exhibited significant changes in their physicochemical properties and antimicrobial effects compared to the control groups. In another study, the chitosan-based films with the included water-based extract of *Tricholoma terreum* (Schaeff.) P. Kumm. gathered from the İnönü plateau in Sakarya province of

Türkiye, were produced by Koç et al (2020). The phenolic content, optical transparency, thermal analysis, thermogravimetric analysis, functional groups by FTIR, water contact angles, and solubility in water, along with antioxidant, antimicrobial, and anti-quorum sensing activities were tested to determine the potential of the films. As a result of this study, the phenolic content of 0.1 g of fungi extract included $2659.82 \mu\text{g g}^{-1}$ of p-coumaric acid and $2126.69 \mu\text{g g}^{-1}$ of gallic acid. Moreover, the researchers found that, the chitosan-based films including fungi extract have a thickness of $190 \mu\text{m}$ and a light brown color. Furthermore, the films have optical transparency ranging from 17% to 53% and water contact angles of 76.06 degrees. Lastly, the elongation of break lengths of the films was determined to increase by 111.1%, when compared to the control group. In terms of antioxidant activity, the films had more than double the effect of the control group. Similarly, the study showed that the films had remarkable antimicrobial activity compared to the control group as well. In the research conducted by Savin et al. (2020), chitosan-based films with added extract of *Ganoderma lucidum* (Curtis) P. Karst., which was obtained from Romania medica SRL laboratories exhibited substantial effects of antioxidant, cytotoxic, and antimicrobial. It was revealed that, the films had more effective antimicrobial activity against Gram-positive bacterial strains than against Gram-negative bacterial strains and that in terms of antioxidant tests such as ABTS and DPPH, the films showed a moderate level of activity.

Conflict of Interest

The authors have declared no conflict of interest.

Authors' Contributions

The authors contributed equally.

Acknowledgements

Thanks to the Scientific Research Projects Unit of Osmaniye Korkut Ata University (project numbers: OKÜBAP-2021-PT3-004 and OKÜBAP-2021-PT3-006) for the invaluable support.

References

- Altıok D, Altıok E, Tihminlioglu F (2010). Physical, antibacterial and antioxidant properties of chitosan films incorporated with thyme oil for potential wound healing applications. *Journal of Materials Science: Materials in Medicine* 21: 2227-2236.
- Amin T, Naik HR, Hussain SZ, Wani SM (2024). *Polysaccharide Based Films for Food Packaging: Fundamentals, Properties and Applications*. Singapore: Springer Verlag.
- Barros L, Calhelha RC, Vaz JA, Ferreira IC, Baptista P, Estevinho LM (2007). Antimicrobial activity and bioactive compounds of Portuguese wild edible mushrooms methanolic extracts. *European Food Research and Technology* 225: 151-156.
- Bégin A, Van Calsteren MR (1999). Antimicrobial films produced from chitosan. *International Journal of Biological Macromolecules* 26(1): 63-67.
- Butler BL, Vergano PJ, Testin RF, Bunn JM Wiles JL (1996). Mechanical and barrier properties of edible chitosan films as affected by composition and storage. *Journal of Food Science* 61(5): 953-956.
- Bülbül M, Süfer Ö, Bozok F (2023). Bazı Aromatik Bitkilerin Uçucu Yağlarını İçeren Yenilebilir Film Üretimi. In Talaz O (ed.) *Fen Bilimleri ve Matematik Alanında Gelişmeler*. Ankara: Platanus Publishing. pp: 99-128.
- Campalani C, Bertuol I, Bersani C, Calmanti R, Filonenko S, Rodríguez-Padrón D, Perosa A (2024). Green extraction of chitin from hard spider crab shells. *Carbohydrate Polymers* 345: 122565.
- Cheung LM, Cheung PC, Ooi VE (2003). Antioxidant activity and total phenolics of edible mushroom extracts. *Food Chemistry* 81(2): 249-255.
- Ebrahimi SS, Hamzeh Y, Ashori A, Roohani M, Marlin N, Spigno G (2024). Ozone-activated lignocellulose films blended with chitosan for edible film production. *International Journal of Biological Macromolecules* 270: 132285.

- Gao Y, Tang W, Gao HE, Chan E, Lan J, Li X, Zhou S (2005). Antimicrobial activity of the medicinal mushroom *Ganoderma*. *Food Reviews International* 21(2): 211-229.
- Gezer K, Duru ME, Kivrak I, Turkoglu A, Mercan N, Turkoglu H, Gulcan S (2006). Free-radical scavenging capacity and antimicrobial activity of wild edible mushroom from Turkey. *African Journal of Biotechnology* 5(20): 1924-1928.
- Gómez-Estaca J, López-Lacey A, Gómez-Guillén MC, López-Caballero ME, Montero P (2009). Antimicrobial activity of composite edible films based on fish gelatin and chitosan incorporated with clove essential oil. *Journal of Aquatic Food Product Technology* 18(1-2): 46-52.
- Goy RC, Britto DD, Assis OB (2009). A review of the antimicrobial activity of chitosan. *Polímeros* 19: 241-247.
- Gökyermez M, Süfer Ö, Bozok F (2023). Edible film production containing *Amanita caesarea* and *Boletus reticulatus* extracts. In: Akpınar A (edt.), *Research on Mathematics and Science- III*. Özgür Publications. pp. 1-26.
- Gutiérrez, TJ, Tomy J (2017). Chitosan applications for the food industry. In: Ahmed S, Ikram S (eds), *Chitosan: Derivatives, Composites and Applications*. pp. 183-232.
- Handayasari F, Suyatma NE, Nurjanah S (2019). Physicochemical and antibacterial analysis of gelatin–chitosan edible film with the addition of nitrite and garlic essential oil by response surface methodology. *Journal of Food Processing and Preservation* 43(12): e14265.
- Hromiš NM, Lazić VL, Popović SZ, Šuput DZ, Bulut SN (2017). Antioxidative activity of chitosan and chitosan-based biopolymer film. *Food and Feed Research* 44(2): 91-100.
- Jayakumar T, Thomas PA, Geraldine P (2009). *In-vitro* antioxidant activities of an ethanolic extract of the oyster mushroom, *Pleurotus ostreatus*. *Innovative Food Science & Emerging Technologies* 10(2): 228-234.
- John S (2022). Fungal Biopolymers as an alternative construction material. In: John S (edt.) *Fungal Biopolymers and Biocomposites, Prospects and Avenues* Singapore: Springer Nature. pp. 169-188.
- Kaya E, Kahyaoglu LN, Sumnu G (2022). Development of curcumin incorporated composite films based on chitin and glucan complexes extracted from *Agaricus bisporus* for active packaging of chicken breast meat. *International Journal of Biological Macromolecules* 221: 536-546.
- Kaya M, Ravikumar P, Ilk S, Mujtaba M, Akyuz L, Labidi J, Erkul SK (2018). Production and characterization of chitosan based edible films from *Berberis crataegina's* fruit extract and seed oil. *Innovative Food Science & Emerging Technologies* 45: 287-297.
- Ke CL, Deng FS, Chuang CY, Lin CH (2021). Antimicrobial actions and applications of chitosan. *Polymers* 13(6): 904.
- Kittur FS, Kumar KR, Tharanathan RN (1998). Functional packaging properties of chitosan films. *Zeitschrift für Lebensmitteluntersuchung und-Forschung A* 206: 44-47.
- Koc B, Akyuz L, Cakmak YS, Sargin I, Salaberria AM, Labidi J, Kaya M (2020). Production and characterization of chitosan-fungal extract films. *Food Bioscience* 35: 10054.
- Kong M, Chen XG, Xing K, Park HJ (2010). Antimicrobial properties of chitosan and mode of action: a state of the art review. *International Journal of Food Microbiology* 144(1): 51-63.
- Kumar A, Hasan M, Mangaraj S, Pravitha M, Verma DK, Srivastav PP (2022). Trends in edible packaging films and its prospective future in food: a review. *Applied Food Research* 2(1): 100118.
- Kumar H, Bhardwaj K, Sharma R, Nepovimova E, Cruz-Martins N, Dhanjal DS, Kuča K (2021). Potential usage of edible mushrooms and their residues to retrieve valuable supplies for industrial applications. *Journal of Fungi* 7(6): 427.
- Lakhiar IA, Yan H, Zhang J, Wang G, Deng S, Bao R, Wang X. (2024). Plastic pollution in agriculture as a threat to food security, the ecosystem, and the environment: An overview. *Agronomy* 14(3): 548.
- Leceta I, Guerrero P, Cabezudo S, de la Caba K (2013). Environmental assessment of chitosan-based films. *Journal of Cleaner Production* 41: 312-318.
- Ma Q, Gao X, Bi X, Xia M, Han Q, Peng M, Wang M (2021). Combination of steam explosion and ionic liquid pretreatments for efficient utilization of fungal chitin from citric acid fermentation residue. *Biomass and Bioenergy* 145: 105967.
- Mau JL, Lin HC, Chen CC (2002). Antioxidant properties of several medicinal mushrooms. *Journal of Agricultural and Food Chemistry* 50(21): 6072-6077.
- Parveen A, Abbas MG, Keefover-Ring K, Binyameen M, Mozūraitis R, Azeem M (2024). Chemical Composition of Essential Oils from Natural Populations of *Artemisia scoparia* collected at different altitudes: Antibacterial, mosquito repellent, and larvicidal effects. *Molecules* 29(6): 1359.
- Priyadarshi, R., Kumar, B., Negi, YS. (2018). Chitosan film incorporated with citric acid and glycerol as an active packaging material for extension of green chilli shelf life. *Carbohydrate polymers* 195: 329-338.
- Puttaraju NG, Venkateshaiah SU, Dharmesh SM, Urs SMN, Somasundaram R (2006). Antioxidant activity of indigenous edible mushrooms. *Journal of Agricultural and Food Chemistry* 54(26): 9764-9772.
- Raafat D, Sahl HG (2009). Chitosan and its antimicrobial potential—a critical literature survey. *Microbial Biotechnology* 2(2): 186-201.
- Rabea EI, Badawy MET, Stevens CV, Smagghe G, Steurbaut W (2003). Chitosan as antimicrobial agent: applications and mode of action. *Biomacromolecules* 4(6): 1457-1465.

- Safarzadeh S, Mozafari MR, Naghib SM (2024). Chitosan-incorporated bioceramic-based nanomaterials for localized release of therapeutics and bone regeneration: An overview of recent advances and progresses. *Current Organic Chemistry* 28(15): 1190-1214.
- Sánchez-González L, Vargas M, González-Martínez C, Chiralt A, Cháfer M (2019). Characterization of edible films based on hydroxypropylmethylcellulose and tea tree essential oil. *Food Hydrocolloids* 23(8): 2102-2109.
- Sarfraz MH, Hayat S, Siddique MH, Aslam B, Ashraf, A, Saqalein M, Muzammil S (2024). Chitosan based coatings and films: A perspective on antimicrobial, antioxidant, and intelligent food packaging. *Progress in Organic Coatings* 188: 108235.
- Savin S, Craciunescu O, Oancea A, Ilie D, Ciucan T, Antohi LS, Oancea F (2020). Antioxidant, cytotoxic and antimicrobial activity of chitosan preparations extracted from *Ganoderma lucidum* mushroom. *Chemistry & Biodiversity* 17(7): e2000175.
- Sedefoglu N, Er S, Veryer K, Zalaoglu Y, Bozok F (2023). Green synthesized CuO nanoparticles using macrofungi extracts: Characterization, nanofertilizer and antibacterial effects. *Materials Chemistry and Physics* 309: 128393.
- Simsek M, Eke B, Demir H (2020). Characterization of carboxymethyl cellulose-based antimicrobial films incorporated with plant essential oils. *International Journal of Biological Macromolecules* 163: 2172-2179.
- Singh M, Saroj R, Kaur D (2024). Optimized chitosan edible coating for guava and its characterization. *Measurement*. *Food* 14: 100145.
- Siripatrawan U, Harte BR (2010). Physical properties and antioxidant activity of an active film from chitosan incorporated with green tea extract. *Food Hydrocolloids* 24(8): 770-775.
- Smolskaitė L, Venskutonis PR, Talou T (2015). Comprehensive evaluation of antioxidant and antimicrobial properties of different mushroom species. *LWT-Food Science and Technology* 60(1): 462-471.
- Wińska K, Mączka W, Łyczko J, Grabarczyk M, Czubaszek A, Szumny A (2019). Essential oils as antimicrobial agents-myth or real alternative?. *Molecules* 24(11): 2130.
- Xie Y, Xie Z, Zhou N (2001). The study on anti-*Helicobacter pylori* mechanisms of chitosans *in vitro*. *Chinese Journal of Digestion* 24(11): 655-658.
- Xu D, Chen T, Liu Y (2021). The physical properties, antioxidant and antimicrobial activity of chitosan–gelatin edible films incorporated with the extract from hop plant. *Polymer Bulletin* 78: 3607-3624.
- Xu YX, Kim KM, Hanna MA, Nag D (2005). Chitosan–starch composite film: preparation and characterization. *Industrial crops and Products* 21(2): 185-192.
- Yang JH, Lin, HC, Mau JL (2002). Antioxidant properties of several commercial mushrooms. *Food Chemistry* 77(2): 229-235.
- Yen MT, Yang JH, Mau JL (2008). Antioxidant properties of chitosan from crab shells. *Carbohydrate Polymers* 74(4): 840-844.
- Yuan G, Chen X, Li D (2016). Chitosan films and coatings containing essential oils: The antioxidant and antimicrobial activity, and application in food systems. *Food Research International* 89: 117-128.