

DETERMINATION OF SOME PARAMETERS WHICH AFFECT THE ACCURACY AND PRECISION IN UV-VIS SPECTROPHOTOMETRY

UV-GÖRÜNÜR ALAN SPEKTROFOTOMETRİSİ'NDE DOĞRULUK VE KESİNLİĞİ ETKİLİYEN BAZI DEĞİŞKENLERİN TAYİNİ

Cem YÜCESOY

Ankara University, Faculty of Pharmacy, Department of Analytical Chemistry
06100 Ankara-TÜRKİYE

ABSTRACT

In this study, quality control tests on UV-Vis spectrophotometers, cells and solvents used in UV-Vis spectrophotometry were made according to the instructions of pharmacopoeia and reference books. For this purpose, baseline flatness, stray light, resolution, wavelength and absorbance accuracy of UV-Vis spectrophotometers were tested. It was found that many parameters tested deviate from acceptable limits. Likely, many of the cell-pairs and solvents of different brands tested didn't meet the requirements. The results confirmed that the maintenance of the instruments, quality of cells and solvents used in the UV-Vis spectrophotometry are prerequisite to achieve accurate and precise results.

Key Words : *UV-Vis spectrophotometry, accuracy and precision, determination of parameters.*

ÖZET

Bu çalışmada, görünür-alan spektrofotometrisi'nde kullanılan cihaz, hücre ve çözücüler farmakope ve kaynak kitaplar tarafından öngörülen kalite kontrol testlerinden geçirildiler. Bu amaçla, UV-görünür alan spektrofotometrelerinde zemin absorbansı, kaçak ışın, ayırma gücü, dalga boyu ve absorbans doğruluğu kontrol edildi. İncelenen parametrelerin çoğunun kabul edilen sınırların dışında sapma gösterdiği görüldü. Aynı şekilde, incelenen hücre çiftleri ve çözücülerin çoğunun da istenen şartları yerine getirmediği görüldü. Sonuçlar, UV-görünür

alan spektrofotometrisi'nde doğru ve kesin sonuçlar elde etmek için kullanılan cihazın bakımlı olmasının, hücre ve çözücü kalitesinin önemli olduğunu teyit etti.

Anahtar Kelimeler : *UV-görünür alan spektrofotometrisi, doğruluk ve kesinlik, değişkenlerin tayini.*

INTRODUCTION

The first UV-Vis spectrophotometers came into general use in 1940's. Since then they have become the most important analytical instrument in many pharmaceutical, clinical and environmental laboratories. Because the UV-Vis spectra of substances are highly suitable for quantitative assays and for many substances they are useful as additional means of identification. Since the technique has become so commonplace, it is assumed that every scientist knows how to 'run' an absorption spectrum. However, a proper training in the technique, maintenance of the spectrophotometer, quality of the cells and solvents used is essential to achieve reproducible and reliable results. This paper investigates the importance of these factors referring to quality control tests required from pharmacopoeias and other reference books.

MATERIALS AND METHODS

Apparatus

Five UV-Vis recording spectrophotometers (A1, A2, B1, B2 and C) were tested, which were produced from two different instrument manufacturers. Different models of the one manufacturer were coded as A and B (two of each were tested) and the one and only model of the other manufacturer is coded as C.

Seven matched quartz cell-pairs (path length = 10 mm) produced from two different manufacturers were tested.

Reagents

Analytical grade potassium chloride (KCl), sodium nitrite (NaNO₂), toluene, hexane, potassium dichromate (K₂Cr₂O₇) and sulphuric acid (H₂SO₄) were purchased from Merck and Fisher. Solvents tested were ethanol 96 %, ethanol absolute, methanol absolute and

cyclohexane. They were products of eight different manufacturers, four of which were international origin.

Solutions

- 1- Solution A : 50 ± 0.5 mg $K_2Cr_2O_7$ (dried for 1 hour at 110 °C) is dissolved and made up to 1000 ml with 0.005 M H_2SO_4 .
- 2- Solution B : 100 ± 1 mg $K_2Cr_2O_7$ (dried for 1 hour at 110 °C) is dissolved and made up to 1000 ml with 0.005 M H_2SO_4 .
- 3- 1.2 % KCl solution : 1.200 g KCl is dissolved and made up to 100 ml with water.
- 4- 5 % $NaNO_2$ solution : 5.000 g $NaNO_2$ is dissolved and made up to 100 ml with water.
- 5- 0.02 % toluene (v/v) in hexane solution : 2.0 ml toluene is dissolved and made up to 100 ml with hexane. 1.0 ml of this solution is diluted to 100 ml with hexane.

Control of the Instrumental Baseline (Baseline flatness):

It should be ensured that there are no cells in the cell holders. The spectrum between 200-800 nm are recorded. Absorbance settings : ± 0.010 A.

Control of the Cell Baseline :

The sample and reference cells are filled with water and placed in the cell holder. The spectrum between 200-800 nm are recorded. Absorbance settings : ± 0.050 A.

If there are considerable differences from the instrumental baseline, the cells are changed and checked whether the deviation is reverse in sign.

Control of the Stray Light:

The absorbance of 1.2 % KCl solution is measured at 200 nm against water.

The transmittance of 5 % $NaNO_2$ solution is measured at 340 nm against water.

Control of the Resolution :

The spectrum of 0.02% toluene in hexane solution is recorded between 200-300 nm. The ratio of the absorbance at 269 nm to that at 266 nm is calculated.

Control of Wavelengths :

The spectrum of solution A is recorded between 200-400 nm using 0.005 M H₂SO₄ as the reference. The absorption maxima and minima are compared with the wavelengths given in Table 2.

Control of Absorbance :

The absorbance values of solution A and B are measured at 235, 257, 313 and 350 nm for the instrument tested and compared with the data given in Table 3. The measurements should be made with 10 mm pathlength cells with the temperature controlled within the range 15 - 25 °C using 0.005 M H₂SO₄ as the reference.

Control of the Precision :

The absorbance of solution A is measured 10 times at 235, 257, 313 and 350 nm using two different techniques.

a) Common technique : The sample-cell is removed from the cell holder, emptied, filled and placed again in the cell holder.

b) Proposed technique : The cells are placed in the cell holder. The filling and emptying process of the sample-cell is made on site by means of Pasteur-pipette (Table 3).

RESULTS AND DISCUSSION

The instrumental baseline is a useful check of the condition of the whole system. Even if measurements are made at a single wavelength, it is worth of scanning on each side of that wavelength to ensure that the baseline is not changing rapidly (1). In this study five spectrophotometers of different models and age were tested. By all of them, the deviation of the baseline was out of the acceptable limits before baseline correction was made (Table 1a). As an example, the baseline of the instrument coded as B1 was shown in Figure 1. The results show the importance of baseline correction before actual sample-readings were made.

Table 1: a) Instrumental Baseline, b) Stray light and c) Resolution of spectrophotometers tested.

a) INSTRUMENTAL BASELINE (Deviation from zero-Absorbance)					
ALLOWED DEVIATION	OBSERVED DEVIATION BY SPECTROPHOTOMETERS				
	A1	A2	B1	B2	C
$\pm 0.002 A *$	0.003	0.003	0.004	0.005	0.003
Comment	Failed	Failed	Failed	Failed	Failed
b) STRAY LIGHT at 200 nm (as Absorbance) and 340 nm (as % Transmittance)					
VALUES REQUIRED	VALUES OBSERVED BY SPECTROPHOTOMETERS				
	A1	A2	B1	B2	C
$A_{200} > 2.000$	1.822	1.855	3.173	3.155	2.033
Comment	Failed	Failed	Passed	Passed	Passed
$T_{340} < 0.1\%$	0.48	0.41	0.06	0.06	0.09
Comment	Failed	Failed	Passed	Passed	Passed
c) RESOLUTION (as A_{269} / A_{266})					
REQUIRED RATIO	RATIOS OBSERVED BY SPECTROPHOTOMETERS				
	A1	A2	B1	B2	C
1.5^{**}	1.21	1.23	1.30	1.31	1.24
Comment	Failed	Failed	Failed	Failed	Failed

* Specified from instrument manufacturers. ** BP 1990.

The light, which reaches the detector by-passing the sample is called stray light. This problem, met near the wavelength limits of an instrument and at the crossover point between UV and Visible lamps, reduces the apparent absorption and causes asymmetrical peaks (2). The stray light of the instruments were checked at two wavelengths. The absorbance of 1.2% KCl at 200 nm should be more than 2.000 and the transmittance of 5 % NaNO_2 at 340 nm should be

less than 0.1 % (2). Two relatively older instruments, coded as A1 and A2 had more stray light than allowed (Table 1 b).

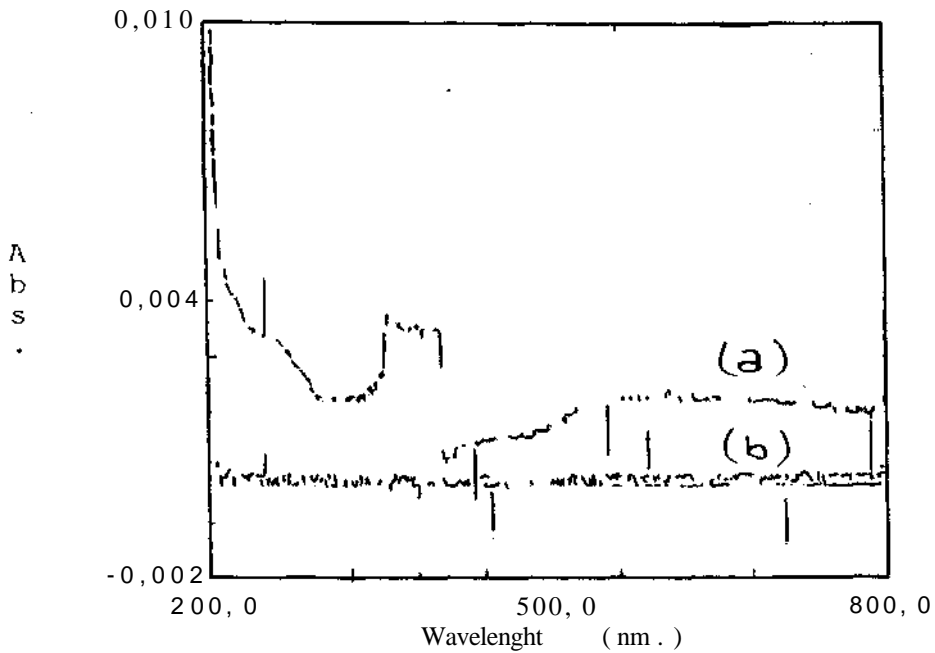


Figure 1. Instrumental baseline of spectrophotometer B1(a) before and (b) after correction (suppressed spectrum) was made.

However the effect of stray light on measured absorbance is dependent on the type of sample being measured. Consequently, this check serves to monitor the deterioration of the instrument only (1).

The resolution of the spectrophotometers is important in qualitative analysis only (3-5). To find the resolution, the ratio of the absorbance at the maximum at 269 nm to that at the minimum at 266 nm is calculated. The pharmacopoeias considered refer to the numerical value stated in the particular monograph. As an exception, BP 1990

requires that this ratio should be more than 1.5 (4). None of instruments tested met this requirement (Table 1c).

The accuracy of the wavelengths of the instruments were investigated using solution A, which was actually prepared to check the absorbance accuracy. Because wavelength standards such as holmium oxide solution (6) couldn't be obtained. $K_2Cr_2O_7$ absorbs approximately between 200-450 nm and have two maxima and minima at 235, 257, 313 and 350 nm. Accepted deviation for wavelength-readings in the UV region is ± 1 nm. With one exception, all the instruments tested, passed this test, although it cannot be assumed that the wavelengths in the visible range were also correct (Table 2).

Table 2. Wavelength Accuracy of the spectrophotometers tested.

WAVELENGTH REQUIRED	OBSERVED BY SPECTROPHOTOMETERS				
	A1	A2	B1	B2	C
235 \pm 1 (Comment)	234.6 (P)	235.0 (P)	234.0 (P)	235.3 (P)	235.2 (P)
257 + 1 (Comment)	256.6 (P)	257.0 (P)	257.0 (P)	257.5 (P)	257.0 (P)
313 + 1 (Comment)	313.2 (P)	313.3 (P)	312.0 (P)	314.0 (P)	313.1 (P)
350 \pm 1 (Comment)	348.8 Failed	349.2 (P)	349.6 (P)	350.0 (P)	350.2 (P)

* (P) = Passed the control.

** Spectrum of solution A is recorded between 200 - 400 nm.

The accuracy of the absorbance of the instruments is checked with two solutions of $K_2Cr_2O_7$ of known concentration (Solution A and B). The second column of Table 3 shows the accepted range of absorbances being measured by the instruments at 235, 257, 313 and 350 nm. The tolerances represent acceptable limits based on the uncertainties of the literature values, the temperature coefficient of the molar absorptivities of $K_2Cr_2O_7$ over the temperature range maintained in the method and the degree of human error normally expected in an analytical

laboratory (1). The absorbance values of the older instrument was totally out of acceptable limits. The deviations observed correspond to about 4 % error. Other instruments had relative lower absorbance deviations and this at some wavelengths, the reason of which may be optical, electronic or mechanical origin (1).

It is assumed that the filling and emptying of the cells and their relocation in the cell holder is a serious source of error in uv-vis spectrophotometry. To evaluate the eventual effect of cell relocation on accuracy and precision of absorbance readings, the results of the common technique (T1) and proposed technique (T2) were compared (1). For this work, solution A was used only.

In the common technique, sample cell was removed after each measurement from the cell holder, emptied, filled with the same solution and relocated in the cell holder for following absorbance measurement. In the proposed technique, sample cell was held in the cell holder during measurements. The emptying and filling process was made with a Pasteur pipette. The average of 10 measurements and the relative standard deviation (RSD) for both techniques were calculated. According to the results, there was no remarkable difference between the means and RSDs (Column A1, A2, B1, B2 and C of Table 3). Conclusively, the proposed technique might be useful for spectrophotometers with relative older technology.

Table 3 : Absorbance Accuracy and Precision (by *technique I and II) of the spectrophotometers tested.

SOLN.	λ (nm) and ABS. REQUIRED at λ	T*	ABS. OBSERVED BY SPECTROPHOTOMETERS				
			A1	A2	B1	B2	C
A	235 \Rightarrow 0.626 \pm 0.009	I	0.602 \pm 0.001 F	0.615 \pm 0.001 F	0.632 \pm 0.000	0.619 \pm 0.001	0.622 \pm 0.001
A	235 \Rightarrow 0.626 \pm 0.009	II	0.602 \pm 0.001 F	0.615 \pm 0.001 F	0.631 \pm 0.001	0.620 \pm 0.000	0.623 \pm 0.001
B	235 \Rightarrow 1.251 \pm 0.019	I	1.215 F	1.228 F	1.260	1.241	1.233
A	257 \Rightarrow 0.727 \pm 0.007	I	0.698 \pm 0.000 F	0.717 \pm 0.001 F	0.729 \pm 0.000	0.720 \pm 0.001 F	0.723 \pm 0.000
A	257 \Rightarrow 0.727 \pm 0.007	II	0.698 \pm 0.000 F	0.718 \pm 0.000 F	0.728 \pm 0.001	0.719 \pm 0.001	0.722 \pm 0.000
B	257 \Rightarrow 1.454 \pm 0.015	I	1.412 F	1.429 F	1.461	1.437 F	1.452
A	313 \Rightarrow 0.244 \pm 0.004	I	0.234 \pm 0.001 F	0.239 \pm 0.000 F	0.249 \pm 0.000 F	0.242 \pm 0.000	0.240 \pm 0.000
A	313 \Rightarrow 0.24 \pm 0.004	II	0.234 \pm 0.000 F	0.240 \pm 0.000	0.248 \pm 0.001	0.241 \pm 0.000	0.239 \pm 0.001 F
B	313 \Rightarrow 0.48 \pm 0.007	I	0.465 F	0.483	0.499 F	0.478 F	0.485
A	350 \Rightarrow 0.536 \pm 0.005	I	0.515 \pm 0.001 F	0.533 \pm 0.000	0.538 \pm 0.000	0.529 \pm 0.001	0.527 \pm 0.000 F
A	350 \Rightarrow 0.536 \pm 0.005	II	0.514 \pm 0.000 F	0.532 \pm 0.001	0.537 \pm 0.001	0.528 \pm 0.000	0.527 \pm 0.001 F
B	350 \Rightarrow 1.071 \pm 0.011	I	1.026 F	1.063	1.072	1.050	1.065

* Technique I (TI) = The cells are emptied and filled outside and relocated in the cell holder.

Technique II (TII) = The cells are emptied and filled inside the cell holder.

F (=Failed) show the absorbance values, which are outside the required range.

Control of cell baseline should be performed every time that clean cells are put into the instrument and at least once per day. If the cells and the solvent they contain are identical, the cell baseline should coincide with the instrumental baseline. A difference of ± 0.005 cm in the cell-pathlength is tolerable, which again causes such a difference in the absorbance-measurements (3-5). The baselines of cell-pairs tested were recorded between 200-800 nm using water as solvent. Absorbance difference of four cell-pairs inspected was out of the limits, which can be explained with the contamination that the cleaning procedure has failed to remove or fine scratches that were not visible, when the cells are filled (Table 4). Figure 2 shows the absorbance deviation of cell baseline for matched cell-pair No.3 from acceptable limits.

BP 1980 requires that ethanol 96 %, ethanol absolute, methanol absolute and cyclohexane should have an absorbance, measured in a 1-cm cell at 240 nm with reference to water, not exceeding 0.100 A (7). This statement was discarded in the latest edition of British Pharmacopoeia. To find whether it is important or not, nine

Table 4 : Cell Baseline of matched quartz cell-pairs tested.

No.	\pm ABSORBANCE DEVIATION	COMMENT	PATHLENGTH	BRAND*
1	0.000	Passed	10 mm	B1
2	0.004	Passed	10 mm	B1
3	0.040	Failed	10 mm	B2
4	0.010	Failed	10 mm	B2
5	0.003	Passed	10 mm	B2
6	0.008	Failed	10 mm	B2
7	0.006	Failed	10 mm	B2

* Cells of 2 different brands were measured.

Spectrum between 200 - 800 nm were recorded.

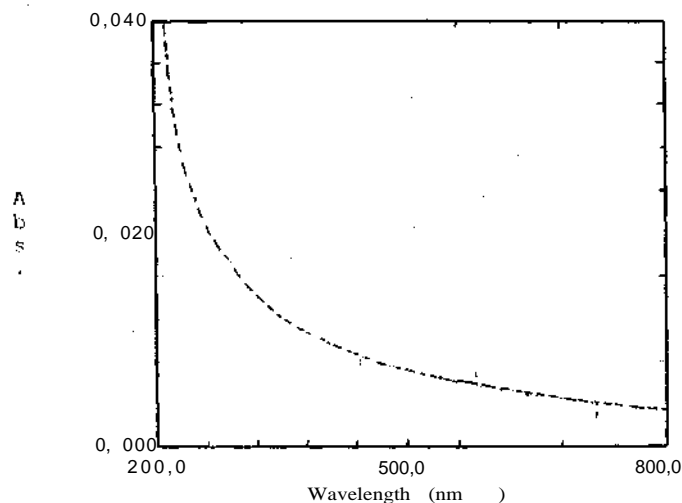


Figure 2. The cell baseline of the matched cell-pair No. 3

solvents of different commercial origin were tested. As all the solvents of international manufacturers passed this exam, 3 out of 4 Turkish products failed, showing the importance of solvent purity in the uv-spectrophotometry (Table 5).

Table 5 : Absorbance of some solvents of different commercial origin at 240 nm

SOLVENTS	ABSORBANCE		COMMENT
	OBSERVED	REQUIRED	
Ethanol 96 % (TP-1) ^a	0.085	< 0.100	Passed
Ethanol Abs. (TP-2)	0.151	< 0.100	Failed
Methanol Abs (IP-3) ^b	0.030	< 0.100	Passed
Methanol Abs (IP-4)	0.026	< 0.100	Passed
Methanol Abs (TP-5)	1.852	< 0.100	Failed
Methanol Abs (TP-6)	4.000	< 0.100	Failed
Cyclohexane (IP-3)	0.037	< 0.100	Passed
Cyclohexane (IP-7)	0.051	< 0.100	Passed

TP-1 = Turkish product, **IP-3** = Imported product. Following numbers (1-7) represent different manufacturers.

The results of the study show that achievement of accurate and precise results in uv-vis spectrophotometry depend on the maintenance of the spectrophotometer, quality of the cell-pairs and solvents chosen for the assay. It can not be expected that spectrophotometers in use reach the performance of a brandnew one. Therefore, it is more realistic to set personal tolerances based on the level of performances that is required from the instrument. But as soon as the regular checks show that the performance has fallen outside these tolerances, action must be taken.

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