



Beneficial Microorganisms in Green Synthesis of Nanoparticles and Potential Applications

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ABSTRACT

Recently, green synthesis in nanotechnology has gotten considerable attention because of its economic importance, as well as providing a clean, eco-friendly, effectual, facile, and non-toxic route to nanoparticle (NP) synthesis. The utilization of various microorganisms especially beneficial microorganisms in NP synthesis presents a sustainable and eco-friendly alternative to conventional synthesis methods, aligning with the principles of green synthesis. In this regard, beneficial microorganisms used in fermented foods as starter cultures, such as *Lactobacillus acidophilus*, *Lactiplantibacillus plantarum*, *Limosilactobacillus*

fermentum, *Secundilactobacillus kimchicus*, *Saccharomyces boulardii*, and *S. cerevisiae* have been utilized for the synthesis of Ag, Se, ZnO, Pd, Sb₂O₃, and TiO₂ NPs. These synthesized NPs have a high potential for use in drug delivery systems, agriculture, and the food industry as antimicrobial, antioxidant, and anticancer agents. Hence, further research is necessary on NP synthesis, novel sources for NP synthesis, and applications in various fields by considering its advantages and disadvantages. This review highlights the green synthesis of NPs, NPs synthesized by beneficial microorganisms, as well as the potential applications of NPs.

Keywords: Microbial synthesis, Fermented foods, Nanomaterials, Antimicrobial, Anticancer

1. Introduction

Nanotechnology is known as the reduction of atomic, molecular, and macromolecular matter to nanometer-scale molecules and is a highly prospective field of study in the twenty-first century, with applications spanning biotechnology, chemistry, medicine, and material sciences (Balakrishnan et al. 2017; Ahmed et al. 2024). Improving nanotechnology has synthesized various nanoparticles (NPs) and nanostructured materials. The particles with at least one dimension in 1-100 nm are described as NP, whose characteristics significantly change depending on the particle sizes (Pacheco-Blandino et al. 2012; Guleria et al. 2020). NPs feature inherent qualities that enable them to demonstrate reactivity and effectively bind, absorb, and transport a range of chemicals, such as DNA, RNA, small-molecule medications, proteins, and probes, with a significant level of efficiency (Elmer & White 2018). These NPs are very beneficial in various sectors such as materials and manufacturing, agriculture, food, medical, pharmaceutical, and environmental, as well as medical applications, ranging from diagnostics to cancer treatment (Guleria et al. 2020; Mitchell et al. 2021).

NPs consist of a wide variety of elements such as aluminum (Al), cerium (Ce), copper (Cu), gold (Au), iron (Fe), manganese (Mn), platinum (Pt), silver (Ag), titanium (Ti), thallium (Tl) and zinc (Zn). Both bottom-up and top-down methodologies in NP synthesis are used (Singh et al. 2021). The bottom-up approach is a very effective method, wherein NPs occur by assembling smaller molecular components. In contrast, NP synthesis by the top-down approach involves utilizing a diverse range of techniques such as electro-explosion, milling, etching, lithographic processes, and laser ablation (Fu et al. 2018). These approaches are important in producing nanomaterials with excellent stability, and their morphology can change depending on desired applications (Jamkhande et al. 2019). Nonetheless, NPs produced can be highly toxic for humans and the environment, and high-cost equipment is also required for some physicochemical synthesis approaches, which increases their production cost and restricts their large-scale commercial production (Siaw et al. 2020). In this regard, the importance of green synthesis techniques has been increasing for the last decades. The major pathways of green synthesis techniques are schematized in Figure 1. The green synthesis of NPs has attracted great attention because of the biocompatibility, cost-effectiveness, low toxicity, and eco-friendly nature of the process and NP products (Mariotti et al. 2020). Also, there is a wide range of green synthesis methods for NPs, plant extracts, or the microorganisms are generally reacted with a metallic salt and then the biological reduction is performed to convert the metal to NPs. The obtained NPs are readily suitable for utilization after proper characterization (Mittal et al. 2013).

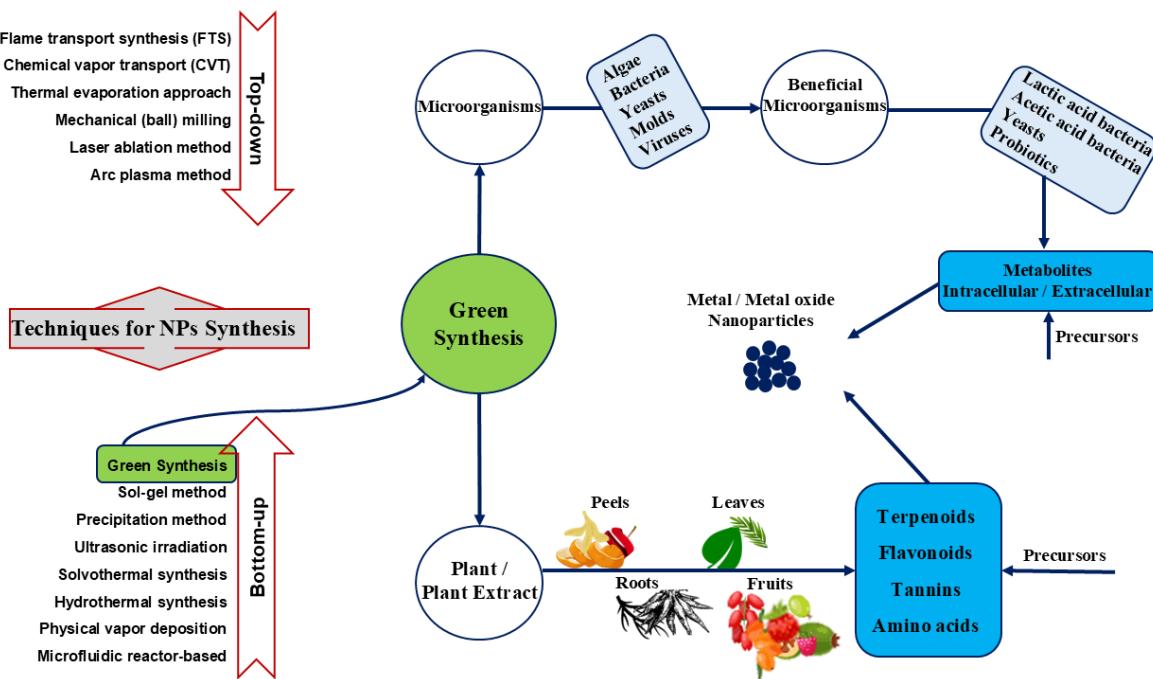


Figure 1- Schematic diagram for the green synthesis of nanoparticles

On the other hand, beneficial microorganisms, including probiotics, are frequently utilized in fermentations associated with foods/food applications and nutraceutical industries. These microorganisms with a positive effect on health are generally lactic acid bacteria (LAB), yeast, and acetic acid bacteria (AAB) (Hazards et al. 2017; Harandi et al. 2021; De Bellis & Rizzello 2024). The exceptional health benefits associated with AAB, LAB, and yeasts position them as highly promising biological scaffolds for the synthesis and targeted delivery of NPs. These microorganisms not only contribute to the sustainable and eco-friendly synthesis of NPs but also enhance their biocompatibility, stability, and therapeutic potential. By leveraging their unique metabolic pathways and bioactive properties, these microorganisms offer a versatile platform for optimizing NP formulations, improving drug delivery efficiency, and minimizing potential toxicity in various biomedical applications (Famararzi et al. 2020; Majumder et al. 2022; Abd Qasim & Yaqaqob 2023; Mohammed et al. 2023). Various LAB and yeast species commonly found in foods have been used for NP (such as Ag and Zn) synthesis (Sásková et al. 2016; Abdulradha & Alhadrawi 2023). Generally, NPs produced by beneficial microorganisms are used for various purposes, such as antimicrobial and anticancer agents, in agriculture, drug delivery systems, and the food industry (Gomez-Zavaglia et al. 2022; Shanmugam et al. 2023). In this review article, the green synthesis of NPs, microbial synthesis, the usage of beneficial microorganisms for NP synthesis, and their potential applications are highlighted.

2. Green Synthesis of Nanoparticles

The present approach for NP synthesis is characterized by the absence of hazardous chemicals, elevated temperatures, high pressure, and providing that ensures both human health and environmental safety (Mariotti et al. 2020). The utilization of economically viable and ecologically sustainable substances, including botanical extracts, microorganisms (bacteria, fungi, microalgae, and viruses), organic polymers, and proteins, has solidified this methodology as a widely favored way for the synthesis of NPs (Taha 2022).

2.1. Nanoparticles from plants or plant extracts

Different parts of plants and vegetables, including peels, extracts, and shells, can be utilized in various applications. The active natural chemical compound obtained from these parts by removing their tissue with a solvent is a plant extract, which is also rich in terms of proteins, soluble fibers, bioflavonoids, and insoluble fibers (Doan et al. 2020; Priya et al. 2023). Plants have been accepted as a more reliable approach to the synthesis of nanomaterial (Figure 2) as they avoid the usage of toxic substances formation, are abundant and environment-friendly, and allow better control over NPs' morphology. Hence, the functional substances such as alkaloids, amino acids, carbonyl, ketones, phenolics, polysaccharides, proteins, vitamins, and tannins found in plants are used for NP synthesis as a reductant source by reducing metal ions into atoms (Vijayaraghavan & Ashokkumar 2017; Kumar & Rajeshkumar 2018). In this scope, NP synthesis has been utilized in plant extracts such as *Aloe vera*, *Camellia sinensis*, *Cinnamomum zeylanicum* leaf, *Jatropha curcas*, *Mangifera indica* leaf, *Mangosteen* leaf, *Murraya koenigii* leaf, gooseberry, mushroom, lemon, pear, papaya, and tansy. NPs prepared using the extracts ensure the advantage in long-term storage conditions due to the non-aggregation of NPs (Jayaprakash et al. 2017; Bhardwaj et al. 2020). Also, tea and coffee

extracts were exploited to synthesize stable NP such as Pd and Ag NPs (Size ranges 20-60 nm). Accordingly, these approaches might be employed for NP synthesis of other noble metals (Nadagouda & Varma 2008).

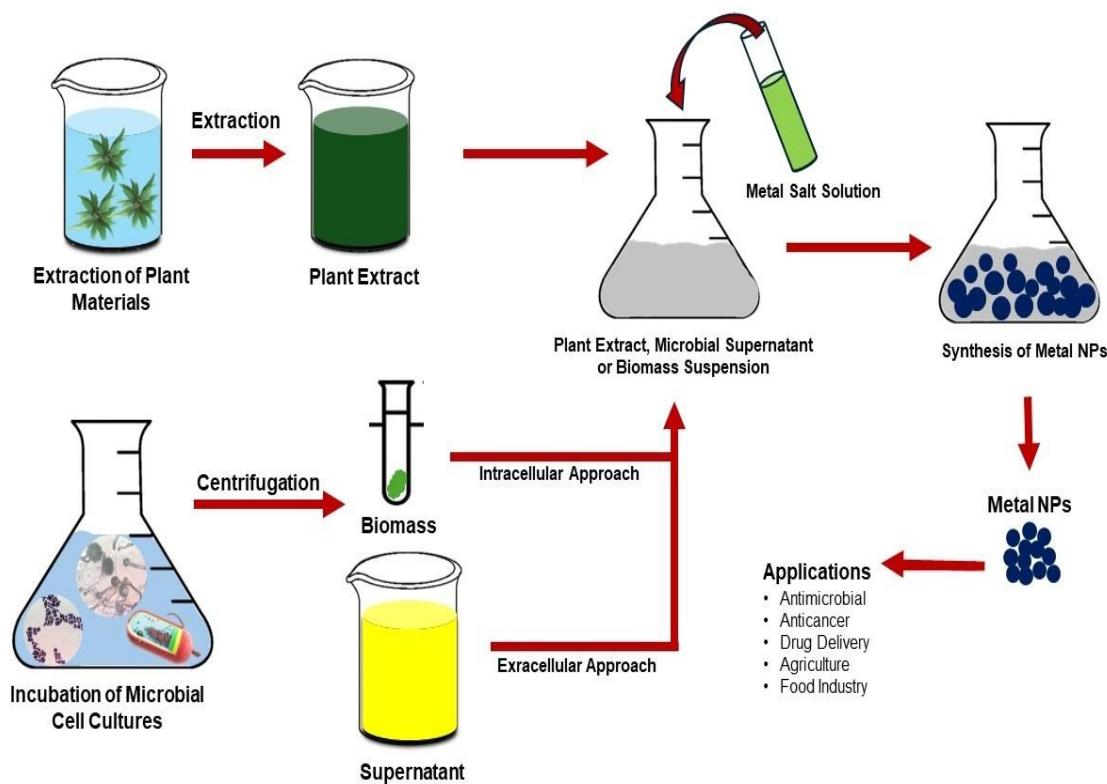


Figure 2- Schematic illustration for plant and microbial synthesis of nanoparticles

On the other hand, various plant sources were explored for the synthesis of metal NPs that had different characteristic properties. For example, in a study, spherical Ag NPs with sizes of 45-110 nm were synthesized using leaf extracts from Congolese plant species (*Brillantaisia patula*, *Crossopteryx febrifuga*, and *Senna siamea*). The study revealed that Ag NPs showed antimicrobial activity against *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa* (Kambale et al. 2020). In another study, Jayaprakash et al. (2017) produced Ag NPs from the natural fruit extract of *Tamarindus indica*. The plant extract acted as both a reducing and a capping mediator for Ag NPs synthesis. The morphology of NPs was a face-centered cubic shape and 6-8 nm sizes. Besides, they possess good antibacterial action. In a different study, Abbas et al. (2020) showed that plant-mediated NP synthesis could be used in wastewater treatment applications and for this purpose, Ag NPs and Au NPs obtained from *Aloe barbadensis*, *Azadirachta indica*, and *Coriandrum sativum* were fabricated. Table 1 compiles various plant sources for biogenic NP synthesis and their characteristics.

Plants provide advantages for natural product synthesis owing to their inherent safety, widespread availability, and diverse biomolecules or metabolites, which aid in stabilizing and reducing NPs (El-Seedi et al. 2019). Plant-derived nanotherapeutic medications have become a potential asset in contemporary cancer therapy. Over the past few years, substantial research has been dedicated to producing metallic NPs with potential anticancer properties. This exploration has spanned both laboratory experiments and studies conducted within living organisms (Andleeb et al. 2021). The anticancer properties of bioactive compounds obtained from various plants and/or plant extracts have been investigated (Majumder et al. 2019). It has been detected that the functional groups attached to NPs directly or indirectly improve anticancer effectiveness, reduce toxicity, and increase bioavailability/absorption (Pei et al. 2020). In a study conducted by In & Nieva (2015) examined the efficacy and toxicity of various NP-based chemotherapeutic treatments. It was detected that chemotherapy using NPs has substantially enhanced the management of non-small cell lung cancer compared to conventional chemotherapy. Also, Dutta et al. (2020) synthesized and stabilized orange peel-based NPs to induce apoptosis in hepatic cancer cell lines, while Andleeb et al. (2021) found that the effectiveness of various NPs in addressing cancer differs due to variations like the biological material used in their manufacturing process.

Table 1- Nanoparticle synthesis from various plant sources and their characteristics

<i>Plant sources</i>	<i>NP/Size/Shape</i>	<i>Characteristics</i>	<i>References</i>
<i>Ananas comosus</i> leaves	Ag NPs, 40-150 nm with hexagonal or spherical shape	Antimicrobial activity	Anis et al. 2023
<i>Azadirachta indica</i>	Ag NPs, 19.27-22.15 nm with spherical shape	Antiinflammatory, antidiabetic activity	Chi et al. 2022
<i>Azadirachta indica</i>	Ag NPs, 41-60 nm with spherical shape	Biolarvicidal activity	Poopathi et al. 2015
<i>Boerhavia diffusa</i>	Cu NPs, 2-10 nm with irregular spherical shape	Antibacterial activity	Vaghela et al. 2018
<i>Catharanthus roseus</i>	Pd NPs, 38 nm with spherical shape	Catalytic activity in dye degradation	Kalaiselvi et al. 2015
<i>Citrus medica</i>	Cu NPs, 10-60 nm	Antimicrobial activity	Shende et al. 2015
<i>Curcumin</i>	Au NPs, 45.10 nm with spherical shape	Cancer chemoperathy	Amini et al. 2023
<i>Datura stramonium</i>	Au NPs, 75.10-156.50 nm with spherical shape	Antifungal, antioxidant, anticoagulant, thrombolytic activity	Oladipo et al. 2020
<i>Datura stramonium</i>	MgO NPs, 0.17 nm with rod-like shape	Antibacterial activity	Saka et al. 2022
<i>Echinochloa stagnina</i>	Ag NPs, 30 nm with spherical shape	Antibacterial, cytotoxic, larvacidal activity	Shehabeldine et al. 2021
<i>Garcinia xanthochymus</i>	ZnO NPs, 20-30 nm with spherical shape	Antioxidant and photocatalytic activity	Nethravathi et al. 2015
<i>Ocimum sanctum</i>	TiO ₂ NPs 75-123 nm with spherical and polygonal	Healing efficacy in diabetic wounds	Ahmad et al. 2022
<i>Ocimum sanctum</i>	CeO ₂ NPs, 34 nm with spherical shape	Photocatalytic and antibacterial activity	Bakkiyaraj et al. 2021
Oil palm	Ag NPs, 18-20 nm with spherical	Antimicrobial	Torres et al. 2021
<i>Phyllanthus emblica</i>	Ag NPs, 19.8-92.8 with spherical shape	Antibacterial and biofilm inhibition activity	Masum et al. 2019
<i>Phyllanthus emblica</i>	Au NPs, 5-60 nm with circular, triangular, and polygonal shape	Potential anticancer activity	Wang et al. 2021
<i>Phyllanthus emblica</i>	Ag NPs, 4-5 nm with spherical shape	Biogenic reducing agent, antioxidant and antibacterial activity	Suvandee et al. 2022
<i>Piper nigrum</i>	Ag NPs, 15-38 nm with spherical shape	Antibacterial, cytotoxicity	Kanniah et al. 2021
<i>Sambucus ebulus</i>	Ag NPs, 35-50 nm with spherical shape	Catalytic, antibacterial, anticancer	Hashemi et al. 2022
Sugar cane leaves	Ag NPs, 16.90 nm with cubic shape	Ammonia hydrogen peroxide detection	Srikhao et al. 2021
<i>Tamarindus indica</i>	Ag NPs, 20-52 nm with spherical shape	Anticancer activity	Gomathi et al. 2020
<i>Tamarindus indica</i>	Cu NPs, 88 nm with spherical shape	Catalytic and antibacterial activity	Ashok et al. 2020
<i>Tamarindus indica</i>	Carbon nanotubes, 18-65 nm with microstructure sheets with thickness	Supercapacitor	Thirumal et al. 2021
<i>Tinospora cordifoli</i>	Se NPs, 100-200 nm with spherical shape	Antioxidant and Antiproliferative activity	Puri & Patil 2022
<i>Tinospora cordifoli</i>	SnO ₂ NPs, 6.90-28.40 nm with non-uniform spherical shape	Photocatalytic activity for rhodamine dye degradation	Fatimah et al. 2022

2.2. Nanoparticle synthesis by microorganisms

Microorganisms are present all over the world, such as in plants, soil, water, and air. However, certain microorganisms, which can cause infections that are harmful to humans, while some microorganisms are significant in maintaining human health and are known as beneficial microorganisms. Microorganisms such as bacteria, fungi, viruses, and microalgae have been used for the green synthesis of NPs like plant-based sources (Shah et al. 2015; Gahlawat & Choudhury 2019; Jeevanandam et al. 2022). The microbial synthesis of NPs possesses several significant advantages, and they have defined morphology, size, and chemical composition, can scale up, are adaptable to various environmental conditions, and are also easy to cultivate and handle the microbial cells (Grasso et al. 2020; Jacob et al. 2021). It has also been reported that microorganisms such as *Pseudomonas aeruginosa*, *P. deceptionensis*, *Bacillus licheniformis*, *Pyrobaculum islandicum*, *Pyrococcus furiosus*, *Bacillus* sp., *Fusarium oxysporum*, *Lactobacillus* sp., *Saccharomyces cerevisiae*, and *Shewanella oneidensis* can be used for the green synthesis of NPs such as Au, Ag, Cu, Fe, Mn, Ni, Ti, and Zn (Shamaila et al. 2016; Sanaeimehr et al. 2018). However, the properties (such as size, morphology, and characteristics) of the NPs could be changed depending on the conditions of the synthesis process (temperature, pH, incubation period, etc.) (Jeevanandam et al. 2022).

The microbial synthesis of NPs is generally performed by the internalization of metal ions, which settle on the cell surface (described as the extracellular approach) or form inside the microbial cells (described as the intracellular approach). The reduction reaction of metal ions on the cell surface or inside the microbial cells may be catalyzed by the microbial enzymes, and so metallic NPs form (Salunke et al. 2016; Barabadi et al. 2017; Grasso et al. 2020). These synthesis methods of NPs described as the extracellular and intracellular approaches mainly include oxidoreductase enzymes like NADPH-dependent sulfite reductase and NADH-dependent nitrate reductase and factor cellular transporters. The extracellular approach ensues exterior to the microbial cell after utilizing various systems and these are (1) the microbial biomass, (2) the supernatant of microbial cultures, and (3) cell-free extracts (Al-Khattaf 2021) (Figure 2). The intracellular process to synthesize NPs involves supplementary phases to isolate the synthesized and gathered NPs from bacterial cells or cell biomass and, so is less favored (Annamalai et al. 2021). For example, in the first approach, Ag NPs fabricated from microbial supernatant can surround constituents of the media, in this way their colloidal dispersion, characterization, and retrieval could be impeded. Despite that, this method verifies the complete elimination of bacterial biomass and the organic media constituents, however, the incubation of bacteria in distilled water for a long period may be inactivated biomolecules, especially enzymes (Annamalai et al. 2021). For NP synthesis, it is suggested as an efficient method that well-grown microorganisms are washed, centrifuged, and then sonicated in ice. In this regard, it is stated that cell-free extract or supernatant is used as a highly concentrated and suitable source for organic molecules and biomolecules such as reductive enzymes (Gholami-Shabani et al. 2015; Wadhwani et al. 2018). With this technique, the eradication of organic media constituents and bacterial biomass by frequent washing is provided to protect biomolecules against proteolytic degradation and denaturation. In this way, it can be stated that the usage of organic biomolecules is more suitable to synthesize the NPs (Badoei-dalfard et al. 2019). In this scope, many enzymes such as alpha-amylase, keratinase, laccase, nitrate reductase, protease, and xylanase, have been used to synthesize Ag NPs and these NPs demonstrated important properties, such as antiradical, anticoagulant, larvicidal, and thrombolytic activities (Lateef et al. 2015a, 2105b; Adelere & Lateef 2016; Elegbede et al. 2018a, 2018b).

Bacteria are one of the prokaryotes, and they are unicellular organisms with a simple structure. Green synthesis of several metallic ions to produce different shapes and sizes of NPs has been attained via different kinds of bacteria strains such as *Lactobacillus*, *Bacillus*, *Pseudomonas*, *Streptomyces*, *Klebsiella*, *Enterobacter*, *Escherichia*, *Aeromonas*, *Brevibacterium*, *Corynebacterium*, *Desulfovibrio*, *Plectonemaboryanum*, *Rhodobacter*, *Rhodococcus*, *Rhodopseudomonas*, *Shewanella*, and *Weissella* (Singh et al. 2016b). Generally, pH, salt concentration, and temperature can largely affect the characteristics of bacterial NP synthesis. For example, Saifuddin et al. (2009) demonstrate the synthesis of relatively smaller Ag NPs (5-50 nm) by *B. subtilis* at 37 °C and pH 7.5, which suggests that a higher incubation temperature might facilitate a more rapid reduction of Ag ions, leading to smaller NP formation. On the other hand, Das et al. (2014) observed larger size range (42-94 nm) at lower temperature of 25 °C, indicating that the rate of NP synthesis and the resulting NP characteristics can be strongly influenced by lower temperatures. These findings emphasize the need for a careful optimization of synthesis parameters to achieve desired NP sizes and properties for specific applications. Additionally, Sinha & Khare (2011) reported that mercury (Hg) NPs intracellularly synthesized by *Enterobacter* sp. were uniform in size (2-5 nm), spherical, and monodispersed in low amounts at pH 8. Also, it has been demonstrated that *Enterobacter* sp. can be used in the decontamination of the environment polluted with toxic Hg (Sinha & Khare 2011). When compared to other microbial systems, such as those used for the synthesis of Ag or Au NPs, the unique ability of *Enterobacter* sp. to synthesize Hg NPs in a controlled manner demonstrates the organism's specialized adaptation for dealing with heavy metal contamination. While other bacteria, like *Bacillus* sp., have been shown to synthesize larger and more variable NPs (e.g., Ag NPs with sizes ranging from 5-50 nm, with some studies reporting sizes up to 94 nm) (Saifuddin et al. 2009; Das et al. 2014), *Enterobacter* sp. excels in producing smaller, uniform particles (Sinha & Khare 2011). This variability in NP size may influence their applicability in specific fields, as uniform and smaller NPs are often preferred for certain biomedical and environmental applications. These findings underscore that different bacterial species possess distinct NP synthesis capabilities, influenced by their unique metabolic pathways and environmental adaptations. Such diversity suggests that tailoring the choice of microbial species for NP synthesis can optimize the properties of the NPs for specific applications, enhancing their efficacy and functionality. Different NPs synthesized from a wide range of bacteria, and their characteristics (size, morphology, and/or characteristic properties) were summarized in Table 2.

Table 2- Nanoparticle synthesis by bacteria and their characteristics

Microorganisms	Sources	Method	NP/Size/Shape	Characteristics	References
<i>Halomonas elongate</i> IBRC-M 10214	-*	Extracellular	CuO NPs, 57-79 nm with rectangular shape	Antimicrobial activity against <i>Escherichia coli</i> ATCC 25922, and <i>Staphylococcus aureus</i> ATCC 43300	Rad et al. 2018
<i>Bacillus cereus</i> HMH1	Soil	Extracellular	FeO NPs, 18.8-28.3 nm with spherical shape	Anticancer effects against the MCF-7 (breast cancer) and 3T3 (mouse fibroblast) cell lines	Fatemi et al. 2018
<i>Bacillus subtilis</i> ZBP4	Soil	Extracellular	ZnO NPs, 22-59 nm with spherical or pseudo-spherical shape	Antimicrobial activity against <i>Bacillus cereus</i> , <i>Escherichia coli</i> O157:H7, <i>Escherichia coli</i> Type 1, <i>Listeria monocytogenes</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonella Typhimurium</i> , and <i>Staphylococcus aureus</i>	Hamk et al. 2023
<i>Escherichia coli</i> E-30, <i>Klebsiella pneumoniae</i> K-6	Stool	Extracellular	CdS NPs, from <i>E. coli</i> , 3.2-44.9 nm with spherical shape <i>K. pneumoniae</i> , 8.5-44.9 nm with spherical shape	Antimicrobial activity against <i>Aspergillus fumigatus</i> , <i>Geotrichum candidum</i> , <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> and <i>Escherichia coli</i>	Abd Elsalam et al. 2018
<i>Idiomarina</i> sp. PR58-8	Soil	Intracellular	Lead (IV) Sulfide (PbS ₂), with a tetragonal crystal lattice, a crystallite domain size of 2.38 nm, spherical shape, with an average size 6 nm	Noncytotoxic, potential for <i>in situ</i> bioimaging applications	Srivastava & Kowshik 2017
<i>Escherichia coli</i> sa2, <i>Exiguobacterium aurantiacum</i> sa3, and <i>Brevundimonas diminuta</i> sa4	Soil	Extracellular	Ag NPs, 5-50 nm with spherical shape	Antimicrobial activity against <i>Bacillus subtilis</i> , <i>Bacillus cereus</i> , methicillin-resistant <i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> , <i>Klebsiella pneumoniae</i> , <i>Escherichia coli</i> , <i>Proteus</i> , <i>Salmonella typhi</i> , and <i>Enterobacter vermicularis</i>	Saeed et al. 2020
<i>Alcaligenes faecalis</i> GH3	Poultry soil	Intracellular	Ag NPs, 32-49 nm, spherical shape	Antibacterial activity against <i>Bacillus cereus</i> , <i>Bacillus subtilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , antioxidant activity	Badoei-dalfard et al. 2019
<i>Bacillus endophyticus</i> SCU-L	Saline soil	Extracellular	Ag NPs, 5-35 nm with spherical shape	Antibacterial activity against <i>Candida albicans</i> , <i>Escherichia coli</i> , <i>Salmonella typhi</i> and <i>Staphylococcus aureus</i>	Gan et al. 2018
<i>Bacillus cereus</i> SZT1	Soil	Extracellular	Ag NPs, 18-39 nm with spherical shape	Antibacterial activity <i>in vitro</i> and <i>in vivo</i> against rice bacterial pathogen <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> , no toxicity to healthy rice plants	Ahmed et al. 2020
<i>Streptococcus griseoplanus</i> SAI-25	Rhizosphere of rice	Extracellular	Ag NPs, 19.5-20.9 nm with spherical shape	Antifungal activity against charcoal rot pathogen <i>Macromomina phaseolina</i>	Vijayabharathi et al. 2018
<i>Shewanella loihica</i> PV-4	-	Intracellular/Extracellular	Au, Pd, and Pt NPs, 2-7 nm, spherical	Catalytic activity on the degradation of methyl orange dye	Ahmed et al. 2018
<i>Pseudomonas poae</i> CO	Garlic plants	Extracellular	Ag NPs, 19.8-44.9 nm with spherical shape	Antifungal activity against <i>Fusarium graminearum</i> , inhibitory effect on spore germination, germ tube growth, mycotoxin production, and the damage of cell membrane of <i>Fusarium graminearum</i>	Ibrahim et al. 2020
<i>Micrococcus yunanensis</i> J2	Sarches mine soil	Extracellular	Au NPs, 15-55 nm, spherical shape	Antibacterial activity against <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> , <i>Micrococcus luteus</i> , <i>Pseudomonas aeruginosa</i> , <i>Klebsiella pneumoniae</i> , and cytotoxicity effect on six cancer cell lines of U87, HT1080, PC12, Caco2, MCF7, and A549	Jafari et al. 2018
<i>Listeria monocytogenes</i> J0161, <i>Bacillus subtilis</i> ATCC 11774 and <i>Streptomyces anulatus</i> MTCC 2528	-	Extracellular	Ag NPs, <i>L. monocytogenes</i> , 62.07 nm with rod and 29.64 nm with spherical shape, <i>B. subtilis</i> , 143.60 nm with hexagonal, 76.38 nm with spherical shape, <i>S. anulatus</i> , 42.44 nm with rod, 37.96 nm with hexagonal, 28.99 nm with triangular and 11.53 nm with spherical shape	Antifungal activity against the entomopathogenic fungus <i>Chrysosporium keratinophilum</i> MTCC 2828, the larvae, pupae and adults of <i>Anopheles stephensi</i> and <i>Culex quinquefasciatus</i> were susceptible to Ag NPs	Soni & Prakash 2015
<i>Weissella oryzae</i> DC6	Mountain ginseng	Intracellular	Ag NPs, 10-30 nm with spherical shape	Antimicrobial activity against clinical pathogens including <i>Vibrio parahaemolyticus</i> , <i>Bacillus cereus</i> , <i>Bacillus anthracis</i> , <i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , and <i>Candida albicans</i>	Singh et al. 2016a

*-: not described.

In NP synthesis with the intracellular approach by mold and yeasts, metal ions internalized within the fungal mycelia are reduced to less toxic forms, whereas the extracellular approach involves bioactive metabolites and enzymatic secretions facilitating NP formation (Molnár et al. 2018; Zhao et al. 2018; Rajeshkumar & Sivapriya 2020). Compared to bacteria, molds and yeasts exhibit distinct advantages in NP biosynthesis, including the production of diverse bioactive metabolites, improved production, and higher aggregation efficiency (Castro-Longoria et al. 2011; Alghuthaymi et al. 2015). However, bacterial systems offer superior genetic manipulability, faster growth rates, and more precise control over NP morphology, which are critical factors in biomedical applications (Jeevanandam et al. 2016; Shamaila et al. 2016). Furthermore, different yeast strains vary in their NP biosynthesis efficiency. For instance, yeast strains such as *Candida glabrata* and *Schizosaccharomyces pombe* have been reported the intracellular synthesis of cadmium sulfide (CdS), Ag, Se, Ti, and Au NPs, with enhanced biocompatibility and stability (Moghaddam et al. 2015; Feng et al. 2017). Conversely, certain yeast species predominantly support extracellular NP formation, which facilitates large-scale synthesis and industrial applicability. Also, NPs had various morphology (at 2-16 nm sizes with rounded, spherical, or oval shapes) were obtained by using different molds and yeasts such as *Alternata alternate*, *Aspergillus oryzae*, *Colletotrichum* sp., *Fusarium oxysporum*, *Verticillium luteoalbum*, *Trichothecium* sp., *Trichoderma viride*, *Candida glabrata*, *C. albicans*, *Rhodotorula glutinis*, and *R. mucilaginosa* (Jeevanandam et al. 2016; Ananthi et al. 2018; Cunha et al. 2018; Jalal et al. 2018; Dhabalia et al. 2020). NPs synthesized by different kinds of fungi and their various characteristics such as size, morphology, and/or characteristic properties were summarized in Table 3.

Table 3- Nanoparticle synthesis by fungi and their characteristics

Sources	Method	NP/Size/Shape	Characteristics	References
Mold				
<i>Trichoderma longibrachiatum</i>	Cucumber	Extracellular	Ag NPs, 5-25 nm with spherical shape	Antifungal against phyto-pathogenic fungi: <i>Alternaria alternate</i> , <i>Fusarium oxysporum</i> , <i>Fusarium verticillioides</i> , <i>Fusarium moniliforme</i> , <i>Aspergillus flavus</i> , <i>Aspergillus heteromorphus</i> , <i>Penicillium glabrum</i> , <i>Penicillium brevicompactum</i> , <i>Pyricularia grisea</i> , and <i>Helminthosporium oryzae</i>
<i>Penicillium oxalicum</i>	-*	Extracellular	Ag NPs, 60-80 nm with spherical shape	Antibacterial activity against <i>Staphylococcus aureus</i> , <i>Shigella dysenteriae</i> , and <i>Salmonella typhi</i>
<i>Aspergillus niger</i>	-	Extracellular	Ag NPs, 25.5-543.3 nm with spherical shape	Antifungal effect on <i>Aspergillus niger</i> , and <i>Aspergillus flavus</i>
<i>Cladosporium perangustum</i> DfI-LAKPAG05	Parasitic plant <i>Dendrophthe falcata</i> grown on mango plant	Extracellular	Ag NPs, 30-40 nm with spherical shape	Antioxidant activity, anticancer activity on MCF-7 (human breast adenocarcinoma cells) cancer cell line and nano-toxicology activity on human blood RBCs and green gram seedling growth of <i>Vigna radiata</i>
<i>Cladosporium</i> sp. MK138585	<i>Commiphora wightii</i>	Extracellular	Au NPs, 5-10 nm with spherical shape	Photocatalytic activity for the degradation of Rhodamine-B (Rh-B) and Methylene Blue (MB) under sunlight irradiation, non-toxic on normal cell lines (3T3-L1), <i>in vitro</i> cytotoxicity against MCF-7 cancer cell line and <i>in vivo</i> inhibitive effect on tumor growth in ascitic tumor model
<i>Fusarium oxysporum</i>	-	Extracellular	Pt NPs, 25 nm with face-centered cubic shape	Antioxidant and antibacterial activity against <i>Escherichia coli</i>
<i>Fusarium oxysporum</i> UTMC-5026	-	Extracellular	Au NPs, 22-30 nm with spherical and hexagonal shapes	Antibacterial activity against <i>Bacillus cereus</i> PTCC-1247, <i>Escherichia coli</i> PTCC-1270, <i>Pseudomonas aeruginosa</i> PTCC-1310, <i>Staphylococcus aureus</i> PTCC-1179
<i>Penicillium chrysogenum</i>	-	Extracellular	Pt NPs, 5-40 nm with spherical shape	Cytotoxicity activity toward myoblast C2C12 carcinoma cells via reactive oxygen species (ROS)-mediated nucleus NF- κ B and caspases activation
Yeasts				
<i>Yarrowia lipolytica</i> DSM 3286	-	Intracellular	Ag NPs, 12.4-24 nm with spherical shape	Antibacterial activity against <i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Proteus vulgaris</i> , <i>Streptococcus pyogenes</i> , <i>Pseudomonas aeruginosa</i>
<i>Candida albicans</i> C86		Extracellular	Ag NPs, 30-70 nm with spherical shape	Antifungal activity against <i>Candida</i> strain C403
<i>C. albicans</i>	Patients at Al-Elweya children's	Extracellular	Ag NPs, 40.19 nm with spherical shape	Antioxidant activity, cytotoxic action against HT-29 cancer cells, non-toxic against normal cell line
<i>Candida glabrata</i>	Oropharyngeal mucosa of patients	Extracellular	Ag NPs, 2-15 nm with spherical and oval shapes	Antimicrobial activity against <i>Escherichia coli</i> , <i>Salmonella typhimurium</i> , <i>Klebsiella pneumoniae</i> , <i>Shigella flexneri</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , <i>Candida albicans</i> , <i>Candida dubliniensis</i> , <i>Candida parapsilosis</i> , <i>Candida tropicalis</i> , <i>Candida krusei</i> , and <i>Candida glabrata</i>
<i>Rhodotorula glutinis</i> and <i>Rhodotorula mucilaginosa</i>	Soil	Intracellular	Ag NPs, <i>R. glutinis</i> , 15.45 nm, <i>R. mucilaginosa</i> , 13.70 nm with rounded shapes	Antifungal activity against <i>Candida parapsilosis</i> , catalytic capacity in the degradation of 4-nitrophenol and methylene blue and cytotoxicity activity on HK-2 (human kidney cells)
<i>Filobasidium stepposum</i>	Soil	Intracellular	Cu-In-S (in the presence of copper, indium, and sulfur) NPs, 3-5 nm with triangular shape	Photostability, showing potential as photosensitizers in solar cells

*:- not described

Algae are autotrophic organisms, which can easily develop with minimal environmental conditions. They act as a "nano factory" for NP synthesis because of their secondary metabolites and many physiologically active substances that serve as capping agents during the synthesis (Fawcett et al. 2017). Previously, da Silva Ferreira et al. (2017) used dry biomass of *Chlorella*

vulgaris for NP synthesis, AgCl-NPs (ranging from 10 to 80 nm and spherical shape) from *C. vulgaris* had antimicrobial activity against *Staphylococcus aureus* and *Klebsiella pneumonia*. In the other studies, Bhuyar et al. (2020) synthesized Ag NPs ranging between 25-60 nm sizes using the marine microalga *Padina* sp., and Ag NPs showed antimicrobial activity against *P. aeruginosa* and *S. aureus*, while Fatima et al. (2020) synthesized Ag NPs with a size range of 60-70 nm and spherical shape using the red algae *Portieria hornemannii* and these NPs showed antimicrobial activity against the fish pathogens *Vibrio harveyii*, *V. parahaemolyticus*, *V. alginolyticus*, and *V. anguillarum*. Besides, various microalgae strains such as *Cystoseira trinodis*, *Sargassum tenerimum*, *Sargassum ilicifolium* and *Turbinaria conoides* have been reported to synthesize Au, CuO, and AlO NPs (Ramakrishna et al. 2016; Gu et al. 2018; Koopi & Buazar 2018). However, microalgae-based NP synthesis often faces challenges related to extended synthesis durations and less precise control over particle size distribution when compared to bacterial systems. This is primarily due to the complex composition of microalgae biomolecules, which can lead to variability in NP characteristics. In contrast, bacterial-based synthesis can offer more controlled environments, resulting in more uniform NP sizes (Chugh et al. 2021; Kulkarni et al. 2023). NPs synthesized by microalgae and their characteristic properties were given in Table 4.

Table 4- Nanoparticle synthesis by microalgae and their characteristics

Microorganisms	Sources	Method	NP/Size/Shape	Characteristics	References
<i>Amphiroa rigida</i>	Coastal area of Kanyakumari District	Extracellular	Ag NPs, 20-30 nm with spherical shape	Antibacterial activity against <i>Staphylococcus aureus</i> and <i>Pseudomonas aeruginosa</i> , cytotoxicity on breast cancer cells (MCF-7) and larvicidal efficiency against the 3rd and 4th instar larvae of <i>Aedes aegypti</i>	Gopu et al. 2021
<i>Chlorella ellipoidea</i>	Domestic sewage water of Silchar town (Assam, India)	Intracellular	Ag NPs, 220.8 nm with mostly spherical shape	Photocatalysts for degradation of water-soluble pollutants, methylene blue and methyl orange dye and antibacterial activity against <i>Staphylococcus aureus</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> and <i>Pseudomonas aeruginosa</i>	Borah et al. 2020
<i>Cystoseira baccata</i>	lower intertidal rocky shore in the NW coast of Spain	Extracellular	Au NPs, 8.40 nm with spherical shape	The cytotoxic activity against two of the human colon cancer cell lines, Caco-2 and HT-29, and on the healthy cell line PCS-201-010 CRC	González-Ballesteros et al. 2017
<i>Spirulina platensis</i>	Freshwater canals near Shebin El-Kom, Menoufia Government, Egypt	Intracellular	Au NPs, 15.60-77.13 nm with octahedral, pentagonal and triangular shape	Antiviral activity against HSV-1 on Vero cells	El-Sheekh et al. 2022
<i>Oscillatoria</i> sp.	Freshwater canals near Shebin El-Kom, Menoufia Government, Egypt	Intracellular	AgO NPs, 14.42 to 48.97 nm with spherical shape	Antiviral activity against HSV-1 on Vero cells	El-Sheekh et al. 2022
<i>Spirulina platensis</i>	-	Intracellular	Pd NPs, 10-20 nm with spherical shape	Adsorbent for lead removal from aqueous solution	Sayadi et al. 2018
<i>Synechococcus moorigangae</i> InaCC M208	Jakarta Bay, Indonesia	Intracellular	Au NPs, 2.5 to 19.9 nm with spherical shape	Antimicrobial activity against <i>Staphylococcus aureus</i> B4, <i>Escherichia coli</i> B5, <i>Klebsiella pneumoniae</i> 1617, <i>Pseudomonas aeruginosa</i> N90PS, and <i>Escherichia coli</i> 8654, antioxidant activity	Purbani et al. 2024
<i>Phaeodactylum tricornutum</i>	-	Extracellular	Ti NPs NPs 50-200 nm, Spherical shape	Antimicrobial activity against <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> , <i>in vitro</i> cytotoxicity against human alveolar adenocarcinoma (A549), human prostate adenocarcinoma (PC-3), human mammary gland adenocarcinoma (MDA-MB) cancer cell lines and human neonatal fibroblasts (CCD34LU) non-cancerous cell lines	Caliskan et al. 2022

*-: not described.

Viruses are also non-cellular parasites (nanosized) that can infect host cells (including animals, bacteria, fungi, humans, and plants) by delivering their genetic materials. The general sizes of most viral capsids are between 20 and 500 nm in diameter and viruses are considered natural NPs because of their sizes (Soto & Ratna 2010; Jeevanandam et al. 2019). Viruses, on the other hand, serve as naturally occurring nanoscale structures with well-defined morphological and structural properties, making them promising candidates for nanotechnology applications (Jeevanandam et al. 2019). Their inherent self-assembling capabilities and biocompatibility have facilitated their utilization in biomedical applications such as gene and drug delivery, vaccines/immunotherapeutics, and theranostics and imaging (Ghosh et al. 2021). Plant viruses exist morphologically in various

forms, generally in the form of rigid rods (helical) and roughly spherical (icosahedral). Frequently used icosahedral viruses in nanotechnology are the Brome mosaic virus, cowpea chlorotic mottle virus, and cowpea mosaic virus. In contrast, the most frequently utilized rod-shaped virus is the tobacco mosaic virus (Wen & Steinmetz 2016; Zhang et al. 2018). However, the major limitation of virus-based NP synthesis is their potential immunogenicity, which may pose biosafety concerns for clinical applications (Nooraei et al. 2021).

On the other hand, all these microbial-based NPs with antimicrobial, anticancer, antioxidant, and catalytic properties have been used in various fields. Notably, Au NPs synthesized using *Penicillium aculeatum* have demonstrated promising scolicidal agent against *Echinococcus granulosus*, indicating potential applications in the treatment of cystic hydatid disease (Barabadi et al. 2017). Similarly, Ag NPs obtained from *Cryptococcus laurentii* had anticancer activity against breast cancer cell lines, and the reason that the stimulation of apoptosis, sustainability, and endocytic action of tumor cell lines was influenced via synthesized Ag NPs (Ortega et al. 2015). Another study conducted by Pugazhendhi et al. (2018) also stated the potential of microbial-based NPs, where they synthesized Ag NPs by marine red alga, *Gelidium amansii* and determined that these NPs can show antifouling characteristics at an important level due to their bactericidal activities against biofilm-forming pathogens. Additionally, hybrid approaches involving different bacterial consortia have shown enhanced NP synthesis efficiency and bioactivity. In this regard, Ag NPs synthesized using a mixture of *Lactobacillus* sp. isolated from raw milk and *Bacillus* sp. isolated from soil exhibited superior antimicrobial efficacy against *S. aureus* and *P. aeruginosa* compared to NPs synthesized by either bacterium alone, highlighting the potential advantages of synergistic microbial systems (Al-Asbahi et al. 2024).

Generally, different types of microorganism species such as *Bacillus*, *Halomonas*, *Escherichia*, *Klebsiella*, *Alcaligenes*, *Pseudomonas*, *Weissella*, *Penicillium*, *Aspergillus*, *Cladosporium*, *Candida*, *Rhodotorula*, *Amphiroa*, and *Spirulina* have been successfully used to NP synthesis such as Ag, Au, Pd, CdS, ZnO, FeO NPs with antimicrobial, anticancer, antioxidant, and catalytic activity. Although these studies were performed, more research about using microbial synthesis of NPs in various diagnostics would provide a more feasible perspective in the future, especially *in vivo* studies.

2.3. Nanoparticle synthesis by beneficial microorganisms

Beneficial microorganisms, which positively affect human health, include various species such as lactic acid bacteria (LAB), acetic acid bacteria (AAB), and yeasts that play a role in the production of fermented foods, especially probiotics (Hill et al. 2014; De Bellis & Rizzello 2024). The investigation of the beneficial microorganisms as green synthesis is important for nanobiotechnological promises due to their nontoxic and/or environmentally friendly properties. For this scope, NPs synthesized by beneficial microorganisms can exhibit diverse therapeutic applications, such as antimicrobial, antioxidant, catalytic, anticancer, and cytoprotective activities (Ghosh et al. 2022).

LABs are Gram-positive and generally accepted as probiotics, they have thick cell walls consisting of glycosides, peptides, polysaccharides, and proteins. These structures are known as bioreduction sites and attract metal ions because of their negative electric potential, this property facilitates NP synthesis and serves as a protective mechanism against metal toxicity (Król et al. 2018; Mohd Yusof et al. 2019). Various NPs, including Ag, Se, Au, and ZnO, have been synthesized using LAB strains (Nair & Pradeep 2002; Bandeira et al. 2020; Spyridopoulou et al. 2021). For example, Ag NPs synthesized by *Lactiplantibacillus plantarum* HDL-03 had crystalline and metallic properties, with the particle sizes of 36.63 nm, and antimicrobial activity against *B. subtilis*, *M. luteus*, *E. coli*, *S. aureus*, *S. enterica*, and *S. paratyphi*. The results showed that Ag NPs synthesized by LAB can be used as antimicrobial agents at safe biological concentrations (Zang et al. 2024). Similarly, *Lactobacillus acidophilus* HN23 was used to synthesize Se NPs, yielding particles ranging from 60 to 300 nm. Compared to Se, these Se NPs exhibited significantly lower toxicity and demonstrated therapeutic potential in mitigating lipid accumulation and enhancing mitochondrial membrane potential in WRL68 human hepatic cells, indicating their potential application in treating nonalcoholic fatty liver disease (Lei et al. 2024). Additionally, *Lacticaseibacillus casei* ATCC 393 assisted synthesis of Se NPs exhibited *in vitro* antiproliferative activity, induced apoptosis, and increased ROS levels in BALB/c mice's CT26 syngeneic colorectal cancer cell models (Spyridopoulou et al. 2021). On the other hand, ZnO NPs synthesized using probiotic bacteria (*Limosilactobacillus fermentum*) exhibited an average size between 100 and 120 nm, with pronounced antimicrobial activity, especially against *Vibrio harveyi* (Shanmugam et al. 2023). Different NPs synthesized by LAB, and their characteristic properties such as size, morphology, and/or various activity were summarized in Table 5.

AAB are Gram-negative α -Proteobacteria, which are obligate aerobes. They are found commonly in environments such as fruits, flowers, rotten fruits or flowers, and so on they are well known for their ability to oxidize sugars and alcohols into organic acids, which has been utilized in the production of fermented foods such as vinegar, Kombucha, lambic beer, and water kefir, all of which possess various health benefits (Buddhika et al. 2021; Hata et al. 2023). Additionally, AAB can produce exopolysaccharides that received significant attention in designing hydrogel materials with applications in tissue engineering for their viscoelasticity, swelling, morphology, and thermal stability, these exopolysaccharides commonly referred to as bacterial cellulose, have also been investigated in NP synthesis (Qi et al. 2020; Gupte et al. 2021). A notable example includes a study utilizing *Glucanoacetobacter kombuchae* to synthesize Ag NPs. Results showed that NPs were 20 nm with spherical shapes and demonstrated potent antioxidant and antimicrobial activity against *B. cereus*, *E. coli*, *S. aureus*, *S. sonnei*, *S. typhi*, and *V.*

cholerae. Besides, the anticancer effect of NPs detected on HEPG2 hepatoblastoma, MCF-7 breast, and MDA-MB 468 triple-negative breast cancer cell lines (Majumder et al. 2022). Similarly, *Acetobacter xylinum* NCIM 2526 produced Au NPs with excellent catalytic activity in the sodium borohydride-mediated reduction of 4-nitro phenol and methylene blue, demonstrating the potential of AAB-derived NPs in catalytic applications (Ahmed et al. 2014).

Table 5- Nanoparticle synthesis by lactic acid bacteria and their characteristics

Microorganisms	Sources	Method	NP/Size/Shape	Characteristics	References
<i>Bifidobacterium bifidum</i>	Patients with diarrhea who had known clinical symptoms	Extracellular	TiO ₂ NPs, 81 nm with spherical or oval	Antimicrobial activity against <i>Pseudomonas aeruginosa</i> , <i>Acinetobacter baumanii</i> , <i>Klebsiella pneumoniae</i> , <i>Escherichia coli</i> and <i>Salmonella typhi</i>	Ibrahem et al. 2020
<i>Lactobacillus kimchicus</i> (<i>Secundilactobacillus kimchicus</i>) DCY51	Korean kimchi	Intracellular	Au NPs, 5-30 nm with spherical shape	Antioxidant activity, <i>in vitro</i> low toxicity in murine macrophage (RAW264.7) and human colon cancer cell lines (HT29)	Markus et al. 2016
<i>Lactobacillus</i> sp.	Raw milk	Intracellular	Ag NPs, 11-22.8 nm, spherical shape	Antimicrobial activity against <i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> and non-hemolysis effect	Al-Asbahi et al. 2024
<i>Lactobacillus sporogenes</i>	-	Intracellular	ZnO NPs, 145.70 nm with hexagonal shape	Antimicrobial activity against <i>Staphylococcus aureus</i>	Mishra et al. 2013
<i>Lactiplantibacillus plantarum</i> VITES07	Curd sample (fermented milk product)	Intracellular	ZnO NPs, 7-19 nm with roughly spherical shape	Potential non-toxic	Selvarajan & Mohanasrinivasan 2013
<i>Lactobacillus</i> sp.	-	Extracellular	TiO ₂ NPs, 50-100 nm, spherical shape	Antibacterial activity against <i>Staphylococcus aureus</i> and <i>Escherichia coli</i> , less haemolytic activity compared to the antibiotic ampicillin	Ahmad et al. 2014
<i>Lactobacillus acidophilus</i>	Fermented milk products	Intracellular	Ag NPs, 19-25 nm with spherical shape	Antimicrobial activity against <i>Escherichia coli</i> ATCC25,922, <i>Pseudomonas aeruginosa</i> ATCC 27,853, and <i>Salmonella enterica</i> ATTC 14,028, Gram-positive: <i>Staphylococcus aureus</i> ATCC 29,213 and <i>Bacillus subtilis</i> ATCC 6051, and fungus <i>Candida albicans</i> ATCC 10,231 and cytotoxic effect on A549 (Lung), Caco (Colon), and HepG2 (Liver) cancer cell lines	Mohammed et al. 2023
<i>Lactobacillus</i> sp.	Fermented food product	Extracellular	Fe ₂ O ₃ NPs, 31.34 nm with spherical shape	Antibacterial activity on <i>Pseudomonas aeruginosa</i>	Abd Qasim & Yaaqoob 2023
<i>Lactobacillus</i> sp. strain LCM5	Brined cucumbers	Extracellular	Ag NPs, 3-35 nm with spherical shape	Antimicrobial activity against <i>Aspergillus ochraceus</i> , <i>Aspergillus flavus</i> , <i>Penicillium expansum</i> , and <i>Chromobacterium violaceum</i>	Matei et al. 2020
<i>Lactobacillus acidophilus</i>	Mother dairy's curd	Extracellular	Se NPs, 2-15 nm with spherical shape	Antibacterial activity on <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , <i>Bacillus subtilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Klebsiella pneumoniae</i> , cytotoxicity activity against HEK-293 (Human Embryonic Kidney) cell lines	Alam et al. 2020
<i>Lactobacillus</i> sp.	Cow milk	Intracellular	ZnO NPs, 32 nm with spherical shape	Antimicrobial activity against <i>Clostridium difficile</i> , <i>Clostridium perfringens</i> , <i>Escherichia coli</i> , <i>Salmonella typhi</i> , <i>Candida albicans</i> , <i>Aspergillus flavus</i> , and biocompatibility activity against human colon cancer (HT-29) cell lines	Suba et al. 2021
<i>Lactiplantibacillus plantarum</i>	-	Extracellular	Ag NPs, 40-50 nm with spherical shape	Antioxidant activity, antimicrobial activity against <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , and <i>Enterococcus faecalis</i>	Prema et al. 2022
<i>Lactiplantibacillus plantarum</i> BSe	Citrus	Intracellular	Se NPs, 100 nm with round shape	Antimicrobial activity against <i>Escherichia coli</i> GDMCC 1.118 and <i>Staphylococcus aureus</i> GDMCC 1.2442 and cytotoxicity on 293T cell line of human renal epithelial cells	Zhong et al. 2024

*-: not described.

Yeasts are widely used as starter culture in various food industries, such as winemaking, bakery, brewing, and dairy production, because of their desirable technological and sensory properties. The most frequently utilized yeasts in food products and supplements include *Saccharomyces cerevisiae* and *S. boulardii*, the latter being recognized as a probiotic yeast (Chan & Liu 2022). Beyond their traditional applications, yeasts have recently been investigated as potential alternative protein sources due to their optimal amino acid composition, and their ability to be cultivated using industrial waste products, making them an eco-friendly protein source (Jach et al. 2022). On the other hand, yeasts also provide several advantages in NP synthesis, including ease of cultivation under laboratory conditions, rapid growth rates, and the ability to utilize simple nutrients, making them highly suitable for scalable and environmentally sustainable NP production (Skalickova et al. 2017). For this purpose, in a study reported by Abdulradha & Alhadrawi (2023), Zn NPs were synthesized using the probiotic *S. boulardii*, yielding spherical NPs with an average size of 24.61 nm. These Zn NPs showed antimicrobial activity against *Burkholderia cepacia*, a pathogen associated with diabetic foot ulcers. Furthermore, Pt NPs synthesized by *S. boulardii* had important anticancer activity against A-431 epidermoid squamous carcinoma and MCF-7 breast cancer cell lines (Borse et al. 2015). NPs synthesized by beneficial yeast, which are commonly used as starter cultures in the food industry, and their characteristic properties are compiled in Table 6.

Table 6- Nanoparticle synthesis by beneficial yeasts and their characteristics

Microorganisms	Sources	Method	NP/Size/Shape	Characteristics	References
<i>Saccharomyces cerevisiae</i>	Baker's yeast	Extracellular	Ag NPs, 10-60 nm with spherical shape	Antimicrobial effect against <i>Cryptococcus gastricus</i> , <i>Esherichia coli</i> , <i>Trichophyton rubrum</i> , <i>Shigella flexneri</i> , <i>Fusarium oxysporum</i>	Sowbarnika et al. 2018
<i>Saccharomyces cerevisiae</i>	Baker's yeast	Intracellular	Se NPs, 30-100 nm, spherical shape	Antioxidant activity	Faramarzi et al. 2020
<i>Pichia kudriavzevii</i> HA-NY2 and <i>Saccharomyces uvarum</i> HA-NY3	Egyptian sweet fruits	Extracellular	Ag NPs, <i>P. kudriavzevii</i> , 20.66 nm with cubic regular shape, <i>S. uvarum</i> , 12.4 nm with round shape	Antimicrobial effect against <i>Bacillus subtilis</i> ATCC6633, <i>Staphylococcus aureus</i> ATCC29213, <i>Pseudomonas aeruginosa</i> ATCC27953, <i>Candida tropicalis</i> ATCC750, and <i>Fusarium oxysporum</i> NRC21, inhibiting of paw edema by oral administration, anticancer activity on HCT-116 (Colon cell line) and PC3 (Prostate cell line), no ulcerogenic effects	Ammar et al. 2021
<i>Saccharomyces cerevisiae</i>	Dry yeast	Intracellular	Pd NPs, 32 nm, hexagonal shape	Photocatalytic activity on degradation of the azo dye direct blue 71	Sriramulu & Sumathi 2018
<i>Saccharomyces cerevisiae</i>	Baker's yeast	Intracellular	TiO ₂ NPs, 36-12.0 nm with spherical shape and rough surfaces	Antibacterial activity against <i>Pseudomonas aeruginosa</i> ATCC 27853, <i>Staphylococcus aureus</i> ATCC 25923, <i>Escherichia coli</i> ATCC 25922, <i>Candida albicans</i> ATCC 10231, Methicillin resistant <i>Staphylococcus aureus</i> (MRSA) and <i>Acinetobacter baumannii</i> and photocatalytic activity into methylene blue dye	Peiris et al. 2018
<i>Saccharomyces cerevisiae</i>	Baker's yeast	Intracellular	ZnO NPs, 13.0-20.0 nm with spherical shape	Antimicrobial activity against <i>Staphylococcus aureus</i> and <i>Escherichia coli</i> , photocatalytic efficiency on degradation of Eriochrome Black T	El-Khawaga et al. 2023
<i>Saccharomyces cerevisiae</i> PTCC 5269	-*	Extracellular	Ag NPs, 29.07 nm with spherical shape	The antibacterial effect on <i>Listeria monocytogenes</i> ATCC 7644 and <i>Streptococcus marcescens</i> PTCC 1111, antioxidant activity, thrombolytic potential to lyse the blood clots, the ability of alpha-amylase inhibition	Barabadi et al. 2024
<i>Saccharomyces cerevisiae</i>	-	Extracellular	Ag NPs, 5-30 nm with spherical to oval shapes	Antibacterial properties against <i>Escherichia coli</i> (KCCM 11234) and <i>Staphylococcus aureus</i> (KCCM 11335) and enhancing seed germination, seedling vigor, and disease resistance for <i>Sorghum jowar</i> and <i>Zea mays</i> seeds	Kim et al. 2024

*-: not described

Considering the diverse applications and synthesis mechanisms of LAB, AAB, and yeasts, each microbial system exhibits distinct advantages in NP biosynthesis. LAB-mediated synthesis is characterized by the production of biofunctional NPs with antimicrobial and therapeutic properties, whereas NPs synthesized by AAB offer enhanced catalytic activity and potential in drug delivery systems. Yeasts, with their rapid growth and high biomass yield, provide a scalable platform for NP synthesis, particularly for biomedical applications. Despite these advantages, differences in NP morphology, particle size distribution, and synthesis efficiency must be considered when selecting a microbial system for NP production.

3. Potential Applications of Nanoparticles

Green synthesis technology is a promising interdisciplinary field that provides novel synthetic materials with broad application prospects in the fields of biology, agriculture, food medicine, and structural materials (Osman et al. 2024). There have been several researches on the utilization of microorganisms for NP synthesis, and microbial NPs are used for anticancer, antimicrobial, anti-biofouling, biosensor, catalysis, medical diagnosis, and mosquito larvicidal purposes (Salunke et al. 2016). Also, NPs are commonly employed in combination with traditional therapies or active substances to augment cellular permeability and effectiveness, while simultaneously reducing the negative effects and morbidity linked to advanced cancer treatment (Ma et al. 2020). Potential application fields of microbially synthesized NPs are presented in Figure 3.

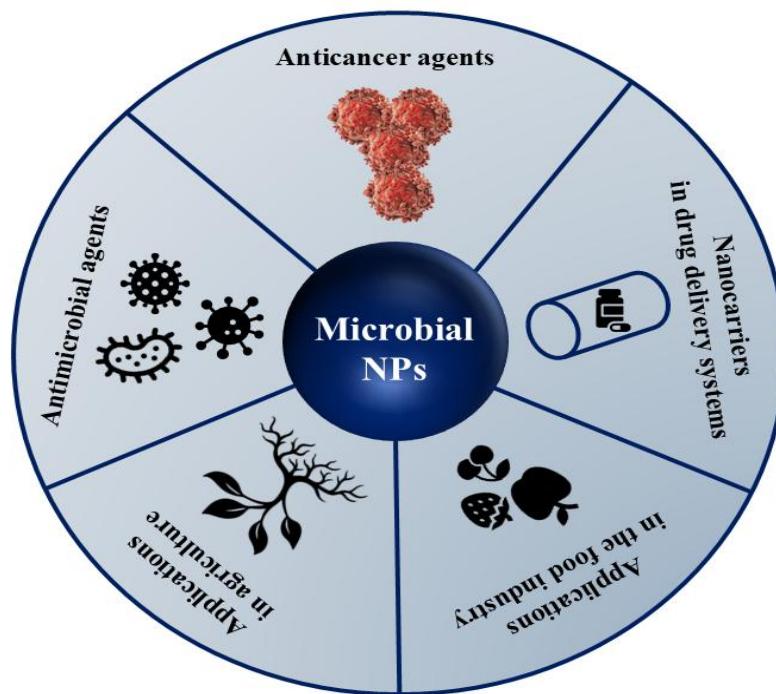


Figure 3- Schematic for the potential applications of microbial nanoparticles

3.1. NPs as antimicrobial agents

The occurrence of resistant strains because of the continuous utilization of antibiotics has dramatically increased infectious diseases. In this context, NPs have an important potential in terms of antimicrobial agents. Some physicochemical characteristics of NPs underlying their antimicrobial activities include their size, surface morphology, charge, and crystal structure (Wang et al. 2017). NPs' small size is a main advantage in achieving strong antimicrobial actions and effectively combating intracellular bacteria since it facilitates the penetration of NPs through bacterial cell walls into the bacteria (Shaikh et al. 2019). The antimicrobial effect mechanism of NPs includes the destruction of the cell wall, causing the leakage of cellular components including proteins, enzymes, DNA, and metabolites into the surrounding environment, ultimately leading to the breakdown of cellular structure (Gomaa 2017; Yuan et al. 2017; Ravichandran et al. 2018; Jalal et al. 2019). The schematic representation of the antimicrobial mechanisms of NPs is given in Figure 4. Firstly, NPs adhere to the cell surface to the plasma membrane and later on transport within the cell where they interact with DNA, so both inhibiting replication and changing the respiratory chain (Slavin et al. 2017). Several studies have investigated the antimicrobial activity of NPs synthesized by microorganisms. For example, Gan et al. (2018) reported that *B. endophyticus*, isolated from soil, extracellularly synthesized Ag NPs with spherical morphology and size range of 5-35 nm, exhibiting significant antibacterial activity with inhibition zones of 15 mm against *C. albicans*, 13 mm against *E. coli*, 11 mm against *S. typhi*, and 19 mm against *S. aureus*. Similarly, *Lactobacillus* sp. (isolated from raw milk) and *Bacillus* sp. (isolated from soil) were utilized for the biosynthesis of Ag NPs. The intracellularly synthesized Ag NPs exhibited spherical morphology with sizes ranging from 4.65 to 22.8 nm. Antibacterial activity analysis demonstrated that Ag NPs derived from *Lactobacillus* sp. exhibited inhibition zones of 12 mm against *S. aureus* and 11 mm against *P. aeruginosa*,

whereas those synthesized by *Bacillus* sp. showed inhibition zones of 11 mm and 10 mm, respectively (Al-Asbahi et al. 2024). Additionally, Ibrahem et al. (2020) reported that *B. bifidum*, isolated from patients with diarrhea exhibiting known clinical symptoms, extracellularly synthesized TiO₂ NPs with spherical or oval morphology and an average size of 81 nm. These TiO₂ NPs demonstrated antimicrobial activity at concentrations of 16 mg/mL and 32 mg/mL against *E. coli* and *S. typhi*, respectively.

The other antimicrobial mechanism of NPs is the occurrence of pits leading to structural damage in the cell membrane and death (Figure 4). After NPs enter the cytoplasm of microorganisms, they induce the generation of ROS, primarily through electron transfer interactions and oxidative stress. These ROS, including superoxide anions (O₂⁻), hydroxyl radicals (•OH), and hydrogen peroxide (H₂O₂), cause oxidative damage to essential biomolecules such as DNA, lipids, and proteins, ultimately leading to cellular toxicity and microbial death (Abo-zeid & Williams 2020). In this scope, for example, MgO NPs have shown a broad range of toxicity against various microorganisms such as *E. coli*, *S. aureus*, *Aspergillus niger*, and *Penicillium oxalicum* (Cai et al. 2017; Sierra-Fernandez et al. 2017). Also, NPs such as Ag, Cu, TiO₂, and ZnO are evaluated as antimicrobial agents because of the formation of ROS (e.g., hydrogen peroxide, hydroxide, or superoxide anion), which can damage cellular structure (Hemeg 2017).

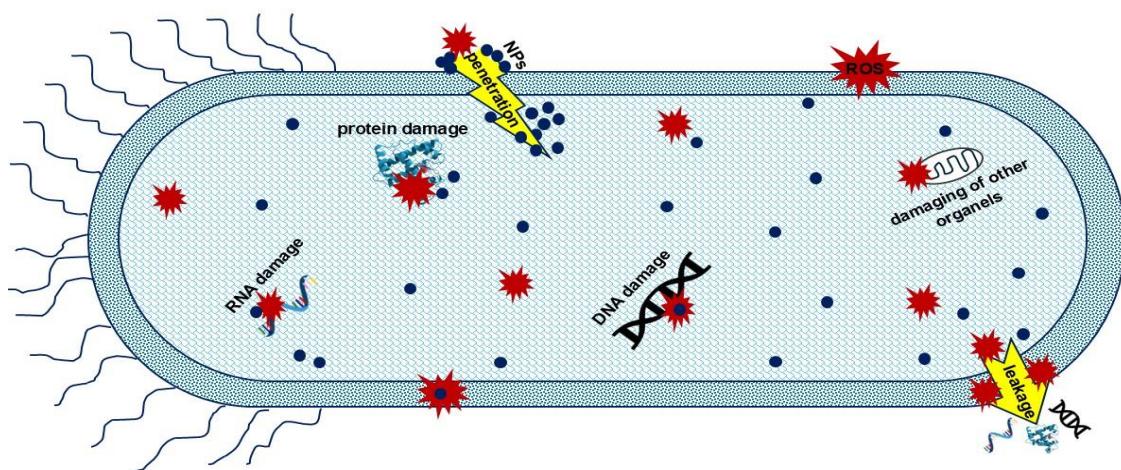


Figure 4- Schematic illustration for the inhibitory mechanisms of NPs on a microbial cell

3.2. NPs as anticancer agents

Cancer has been one of the prominent causes of mortality around the globe in recent years. It includes an assemblage of illnesses, characterized by indefinite growth and changed or no apoptosis. Generally, surgery, chemotherapy, and radiotherapy, which have undue side effects are used as methods of cancer treatment (Gegechkori et al. 2017). However, these methods are inadequate for the treatment and hence, incidences of various cancers such as blood, breast, colon, and lung are increasing each day. Hence, it is crucial to explore alternative solutions to address these challenges. According to numerous researchers, nanomedicine applications can be used in rapid tumor detection and targeted drug delivery and treatment (Bajpai et al. 2018b; Wang et al. 2020; Bloise et al. 2021; Ikram et al. 2021; Mortezaee et al. 2021; Razak et al. 2021). NPs are very comparable in size to most biological structures and compounds and thus have functional properties such as *in vivo* and *in vitro* cancer research (Oroojalian et al. 2021). Recent studies have highlighted the potential of NPs synthesized by microorganisms in cancer treatment. For example, *L. kimchicus* (*Secundilactobacillus kimchicus*) DCY51 isolated from Korean kimchi, has been utilized for the intracellular synthesis of Au NPs ranging from 5 to 30 nm in size, exhibiting spherical shape. These Au NPs have demonstrated significant antioxidant activity, as well as low toxicity in both murine macrophage (RAW264.7) and HT29 colon cancer cell lines, making them potential candidates for anticancer applications (Markus et al. 2016). Similarly, *L. acidophilus*, commonly found in fermented milk products, has been employed to synthesize Ag NPs with size range of 19 to 25 nm. The Ag NPs, also spherical in shape, displayed cytotoxic effects on several cancer cell lines, including A549 lung, Caco colon, and HepG2 liver cancer cell lines, suggesting their potential as anticancer agents (Mohammed et al. 2023). Additionally, *Lactobacillus* sp., isolated from cow milk, was used for the intracellular synthesis of ZnO NPs with spherical shape and size of approximately 32 nm. These ZnO NPs exhibited biocompatibility in HT-29 colon cancer cell lines, further supporting their role in cancer therapy (Suba et al. 2021). Moreover, yeasts such as *P. kudriavzevii* HA-NY2 and *S. uvarum* HA-NY3, isolated from Egyptian sweet fruits, have been used for the extracellular biosynthesis of Ag NPs. The Ag NPs synthesized by *P. kudriavzevii* were approximately 20.66 nm in size with cubic regular shape, whereas those synthesized by *S. uvarum* had round shape and were approximately 12.4 nm in size. These NPs exhibited potent anticancer activity against HCT-116 colon and PC3 prostate cancer cell lines (Ammar et al. 2021).

Also, it has been believed that NPs serve as tools that facilitate molecular-level interaction, and traverse biological barriers but do not affect the functioning of healthy cells. As well as their usage as a drug carrier, NPs can have an anticancer effect on their own. For instance, Au NPs promoted apoptosis in pancreatic cancer cells and reversed epithelial-mesenchymal transition, and in this way, gemcitabine-resistant cells became sensible to the drug (Qiu et al. 2018; Huai et al. 2019; Zhang et al. 2019; Tomşa et al. 2021). On the other hand, the toxicity of Au NPs can be changed depending on size itself, the larger particles with 4.8-12 nm sizes may be less toxic against healthy cells while maintaining significant toxicity against tumor cells, whereas smaller NPs with 1-2 nm sizes are toxic to both healthy and tumor cells (Liu et al. 2021; Tomşa et al. 2021). Also, Au NPs have been utilized in photothermal and photodynamic therapies for various cancer types (Liu et al. 2021). Additionally, Ag NPs can induce pro-apoptotic effects by promoting ROS production through electron transfer reactions and redox cycling processes. These mechanisms lead to oxidative stress, resulting in increased caspase-3 activity and the upregulation of the p53 protein, which collectively contribute to apoptosis (Lee & Lee 2019; Halawani et al. 2020). For example, it has been stated that the cytotoxicity of Ag NPs against MCF-7 breast cancer cells resulted from the apoptosis induction by activation of the pro-apoptotic Bax protein and the inhibition of the anti-apoptotic Bcl-2 protein, also the release of cytochrome c from mitochondria (Kulandaivelu & Gothandam 2016).

Beyond their direct cytotoxic effects, NPs can also modulate the immune response, offering additional mechanisms for cancer therapy. Microbial NPs could be loaded with immunostimulatory agents, such as cytokines, Toll-like receptor agonists, or checkpoint inhibitors, and delivered to the tumor site. These NPs can stimulate antigen-presenting cells, leading to enhanced T-cell activation and a more robust adaptive immune response against tumor cells (Chohan et al. 2023; Zeng et al. 2023; Gholami et al. 2024). In terms of cytotoxicity, Au NPs have been shown to enter human cervical cancer cells via endocytosis, where they elevate intracellular ROS, disrupt matrix metalloproteinase activity, and induce apoptosis by causing cell cycle arrest at the S-phase (Khatua et al. 2020). While these mechanisms contribute to anticancer efficacy, the potential off-target effects on healthy cells due to ROS elevation and DNA damage warrant further investigation. In addition to Au NPs, Ag NPs have been reported to induce oxidative stress and mitochondrial dysfunction, ultimately triggering apoptosis in cancer cells. Their cytotoxicity is primarily associated with the disruption of cellular membranes and increased production of ROS (Xu et al. 2023). Interestingly, the microbial NPs also demonstrated significant cytotoxicity against MCF-7 breast cancer and A549 lung carcinoma cell lines, further supporting their potential as alternative anticancer agents (Fahmy et al. 2020). However, before microbial NPs can be fully integrated into clinical practice, extensive *in vivo* studies are necessary to evaluate the long-term immunogenicity, biodistribution, and potential off-target effects (Gavas et al. 2021).

Nanoparticles synthesized from microorganisms offer a promising, environmentally friendly alternative to traditional synthetic methods. They can not only exhibit anticancer effects but also modulate immune responses, further enhancing their potential in cancer therapy.

3.3. NPs as nanocarriers in drug delivery systems

Generally, the new drug delivery systems are designed to increase therapeutic effects, but these systems should provide safe, accurate, and controlled drug delivery at the target sites. NPs are extensively used as drug delivery carriers for various therapeutic agents (nucleic acids, antibodies, chemotherapeutic drugs, etc.) (Thanki et al. 2013). Target-specific nanocarriers must traverse blood-tissue barriers to reach the target cells. These carriers enter the target cells by endocytosis and transcytosis pathways and interact with cytoplasmic constituents (Fadeel & Garcia-Bennett 2010). NP drug carriers can also pass through the blood-brain barrier and junctions of tight epithelial cells in the skin, which provides drug delivery to the desired target site. Besides, these nanocarriers can increase the biodistribution of therapeutic agents due to their high surface area-to-volume ratio and reduce toxicity at the target site (Wolfram & Ferrari 2019). For this scope, NPs like Ag, Au, Cu, Pt, TiO₂, and ZnO NPs possess improved tunable optical properties and can be used as nanocarriers. Besides, their surface can be easily functionalized to conjugate targeting agents and active biomolecules through covalent bonding, Hydrogen bonding and electrostatic interactions, and multiple drugs can be easily added to improve therapeutic efficacy (Thanki et al. 2013).

On the other hand, certain types of NPs, such as lipid-based NPs (e.g., liposomes), polymeric NPs, and micelles, increase the solubility of hydrophobic compounds, making them suitable for parenteral administration. Additionally, these NPs enhance the stability of therapeutic agents, such as oligonucleotides, peptides, and other biomolecules (Emerich & Thanos 2006). Besides, adhesion and other interfacial interactions between nanocarrier and biomembrane are necessary, especially for intracellular drug delivery (Vedernykova et al. 2018). Therefore, it is stated that drug delivery by NPs could also decrease the dosage concentration of anticancer drugs with higher specificity/efficacy and lower toxicity, like that in melanoma treatment (Cassano et al. 2021). For instance, it has been reported that ZnO NPs synthesized by *Rhodococcus pyridinivorans* loaded with anthraquinone demonstrated cytotoxicity toward colon cancer cells (HT-29 colon carcinoma) depending on the dose-dependent manner used (Kundu et al. 2014). Moreover, it was stated that NPs could show features to target tumor vessels and microenvironments and enhance the efficacy of antiangiogenic drugs (Alsaab et al. 2021). On the other hand, the study by Liu et al. (2020) demonstrated the intracellular synthesis of Au NPs by *Gluconacetobacter liquefaciens* kh-1, which served as carriers for the peptide CopA3 and ginsenoside compound K, enhancing their therapeutic efficacy. These biogenic NPs facilitated targeted drug delivery to lipopolysaccharide-activated macrophages, ensuring controlled intracellular release without significant cytotoxicity. This innovative approach showed the potential of microbial synthesized NPs as biocompatible and effective drug delivery systems in

inflammation-related disorders. Additionally, the study by Pradeepa et al. (2016) introduced a novel antibiotic delivery system using Au NPs synthesized via bacterial exopolysaccharide derived from *L. plantarum*. The biogenic Au NPs were functionalized with various antibiotics, including levofloxacin, cefotaxime, ceftriaxone, and ciprofloxacin, to enhance their therapeutic efficacy against multidrug-resistant bacterial strains. Conjugation of antibiotics to Au NPs significantly improved their stability, bioavailability, and controlled release, ensuring a prolonged antibacterial effect while reducing the minimum inhibitory concentration and minimum bactericidal concentration of the drugs. Mechanistically, the Au NP-antibiotic conjugates effectively penetrated bacterial cells, overcoming resistance mechanisms such as efflux pumps and enzymatic degradation. Transmission electron microscopy analysis revealed that these NP-drug complexes disrupted bacterial membranes, leading to cytoplasmic leakage and eventual cell death. This targeted delivery system not only enhanced the antibacterial activity of conventional antibiotics but also minimized potential cytotoxic effects by reducing the required dosage. These studies not only demonstrate the therapeutic potential of NP-based drug delivery systems but also highlight the crucial role of microorganisms in the synthesis of biocompatible NPs. In both cases, *G. liquefaciens* and *L. plantarum* serve as key microbial agents, enabling the intracellular synthesis of Au NPs that have different characteristics and are functionalized for specific therapeutic applications. The use of these microorganisms ensures a sustainable, eco-friendly synthesis process and enhances the efficiency of the drug delivery system, as their metabolic pathways provide a natural method for producing NPs with tailored properties suitable for targeted drug delivery.

3.4. NPs applications in agriculture

Nanotechnology in agriculture is accepted as one of the most appropriate methods to enhance crop yield and sustain the world's continuously growing population. NPs have significant applications such as nano-fertilizers, nano-pesticides, and crop protection for the regulated dispensing of agrochemicals (de Oliveira et al. 2014; Grillo et al. 2016; Qureshi et al. 2018). The applications of NPs in agriculture as fertilizers are related to their enhanced characterization such as absorption, adhesion effects, responsiveness, and surface properties (Qureshi et al. 2018). Nanofertilizers are micro- or macro-nutrient fertilizers with a particle size of <100 nm and can be used to increase yields. They are also responsible for supplying types of nutrients (one or more) to growing plants, promoting their growth, and increasing production (Chhipa 2017). Nanofertilizers are used in two different types. Firstly, they can provide nutrients to plants to enhance their development and yield. Secondly, they are used as nutrient carriers like nanocarriers used in drug delivery systems, and they help not only in the transport and release of nutrients but also as a nutrient source (Liu & Lal 2015). However, foliar spray or irrigation can be delivered above or below ground to encourage plant growth and yield, or NPs can be added to seeds or primed (Mittal et al. 2020). The efficiency of NPs can change depending on various factors such as the plant type, as well as NP type, size, concentration, stability, chemical composition, and transformation rate after biological interaction (Prasad et al. 2017; Chen 2018).

NPs synthesized by microorganisms such as *B. subtilis*, *Paenibacillus elgii*, and *P. fluorescens* can be used as nano-biofertilizers. Since one liter of nano-biofertilizers can be applied to several hectares of crops, their requirements are so (relatively minimal in comparison with other fertilizers), making them costs-affordable. In addition to carbon-based nanomaterials, nano-clay minerals, and zeolites can also be used as fertilizers (Guo et al. 2018; Bisinoti et al. 2019). Many crops such as tomatoes, chickpeas, spinach, and maize have benefited from the use of different natural nanofertilizers such as ZnO, SiO₂, Fe slag powder, and TiO₂ NPs (Sivarethnamohan & Sujatha 2021). Additionally, the studies conducted for the potential application in food and agriculture fields of NPs are also given in Table 7.

Table 7- Potential applications in the agriculture and food fields of NPs

Applications	NPs Type	Synthesis Sources	Properties	References
Agriculture - Fertilizers	Se NPs, 50-500 nm	Extracellular synthesis by <i>L. acidophilus</i> , <i>L. casei</i> , <i>Bifidobacterium</i> sp.	Plant disease enhancer	Eszenyi et al. 2011
Agriculture- Fertilizer and Pathogen Control	Ag NPs, 12-27 nm	Extracellular synthesis by <i>L. casei</i>	Plant-growth stimulator, antimicrobial effect	Singh et al. 2015
Agriculture - Pathogen Control	Ag NPs, 20-50 nm	Extracellular synthesis by <i>Achromobacter</i> sp., <i>Pseudomonas</i> sp.	Antifungal activity against <i>Fusarium oxysporum</i> in chickpeas	Kaur et al. 2018
Agriculture - Pathogen Control	ZnO NPs, 2-83 nm	Extracellular synthesis by <i>Trichoderma harzianum</i>	Antifungal agent against soil pathogens <i>Macrophomina phaseolina</i> , <i>Rhizoctonia solani</i> , and <i>Fusarium</i> sp.	Zaki et al. 2021
Agriculture - Pathogen Control	CuO NPs, 31-45 nm	Extracellular synthesis by <i>Trichoderma harzianum</i>	Antifungal effects against <i>Alternaria brassicicola</i> (the foliar pathogen)	Gaba et al. 2022
Agriculture - Pathogen Control	ZnO NPs, 32 nm	Extracellular synthesis by <i>B. subtilis</i>	Control growth of agricultural pathogens including <i>Alternaria solani</i> , <i>Colletotrichum capsici</i> , <i>F. oxysporum</i> , <i>Sclerotium rolfsii</i>	Nargund et al. 2022
Food Industry - Antimicrobial Packaging	Ag NPs	Cellulose and collagen	Prevent the growth of <i>E. coli</i> and <i>S. aureus</i> in sausage casings	Fedotova et al. 2010
Food Industry – Packaging	Ag NPs, 300 nm	Chitosan	Prevent the growth of <i>E. coli</i> O157: H7, <i>L. monocytogenes</i> , <i>S. aureus</i> , and <i>S. typhimurium</i> in food packaging	Rhim et al. 2013
Food Industry - Packaging	Ag NPs embedded in carboxymethylcellulose film	-*	Prevent the growth of <i>E. coli</i> and <i>E. faecalis</i> in food packaging	Siqueira et al. 2014
Food Industry - Food Preservation	Ag NPs, ZnO NPs, 20 nm	-	Antibacterial biodegradable films based on polylactide and polybutylene adipate terephthalate (as matrix) loaded with ZnO or Ag NPs in active packaging to preserve the fresh noodles	Yana et al. 2024

*:- not described.

3.5. NP applications in the food industry

The Food and Agriculture Organization of the United Nations highlights that over 1.3 billion metric tons of consumable food is trashed annually due to inadequate storage and transport facilities, poor post-harvest techniques, and market/consumer waste. Preventing food wastage is critical to mitigate the food crisis exacerbated by environmental challenges and growing populations (Nile et al. 2020). The primary causes of food wastage are microbial contaminations and/or spoilages, which compromise food quality and safety and increase the risk of foodborne diseases (Sperber 2009). Nanotechnology presents various solutions across the food supply chain, including production, processing, storage, and distribution. It increases food safety through nanosensors, which detect foodborne pathogens, enhance packaging with antimicrobial properties, and extend shelf life (Sekhon 2010; Nile et al. 2020). Besides, nanotechnology helps in toxin detection, color formation, and flavor production (Seklon et al. 2010). Moreover, smart nanotechnology systems provide efficient localization, sensing, and remote control of food items. Also, nanobased delivery systems can improve the nutritional value of food (Bajpai et al. 2018a). For example, carbon nanotubes serve as potent antimicrobial agents by causing cellular damage or death in *E. coli* through direct contact (Kang et al. 2007), nano-biosensors can be also used for detecting carcinogenic pathogens in food production (Nile et al. 2020). Besides, SiO₂ (E551) and TiO₂ (E171) were allowed as food additives in bulk quantities to produce healthier foods (EFSA 2009), while Ag NPs were used for the production of fortified Jambu juice to increase their content of essential vitamins and minerals (CFS 2017). Moreover, Ag NPs were utilized for sterilization, quality control, and bioavailability, supporting the immune system and defense for natural healing in supplemented functional drinks (CFS 2017). NPs are also utilized as carriers to deliver enzymes, antibrowning agents, antioxidants, flavors, etc. to develop the shelf life even if the package is opened (Cha & Chinnan 2004; Weiss et al. 2006). Fe, ZnO, C, MgO, TiO₂, and SiO₂ NPs are used as antimicrobial agents and in some conditions as food components and/or ingredients (He et al. 2019). Besides, NPs were used in various food applications, as food colorants, anticaking, hygroscopic, and drying agents, as a whitener in dairy products, and as acting antibacterial agents, absorbing and decomposing ethylene in fruit and vegetables (Mohammad et al. 2022).

On the other side, NPs are widely used in packaging systems since they show antioxidant and antimicrobial activities against the microorganisms found on the surface of food and/or food packaging materials (Wang et al. 2022). NPs (namely Ag, CuO, Cu, Fe, MgO, Pd, TiO₂, and ZnO) with antimicrobial properties that could be adhered by various interactions (such as covalent, electrostatic, and hydrogen bonding) are used to develop advanced packaging systems that possess antimicrobial activities (Morris et al. 2017). On the other hand, for nanocomposite production, polymers such as ethylene–vinyl acetate copolymer, polyolefins, nylons, polyethylene terephthalate, polystyrene, and polyamides can be used because polymer matrices control the release of active components and regulate the function of nanocomposites (Nie et al. 2020). The studies conducted for the potential application in food and agriculture fields of nanoparticles are given in Table 7.

As a result, the application of NPs is vast in the food industry, spanning from ingredients to packaging, and product analysis. Despite its potential, the interaction of NPs with food systems can cause health concerns for humans and animals. Nanoformulated products can be toxic to plants and animals, yet no standard regulatory laws exist for their use in the food and agriculture sectors. Thus, effective guidelines and policies should be legislated as crucial for the safe use of NPs in the food industry (Nie et al. 2020). In the European Union, “Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR)” conducts risk assessments for nanotechnology. EU regulations highlight that nanotechnology-based food ingredients should undergo safety evaluations before authorization for consumption (Tinkle et al. 2014; Nie et al. 2020).

4. Challenges and future prospects

NP synthesis through green methods, especially by using beneficial microorganisms, has garnered significant attention because of its eco-friendly nature and potential applications in various fields such as medicine, agriculture, food, and environmental remediation. However, this area of research is not without its challenges, and addressing these is significant for future advancements and applications.

The reproducibility and scalability of NP synthesis using microorganisms pose important challenges. Variability in biological systems can lead to inconsistent NP characteristics, affecting their performance and application. Besides, scaling up from laboratory to industrial production while maintaining the NP's quality and characteristics is complex and requires standardization of processes (Gomez-Zavaglia et al. 2022). In this regard, genetic engineering could play a vital role in producing microorganisms with enhanced reproducibility, thus overcoming some of the scalability challenges.

On the other hand, controlling the shape, size, and crystallinity of NPs synthesized by microorganisms is also challenging. Contrary to chemical synthesis, where conditions can be finely tuned, biological synthesis is influenced by many factors such as the microorganism's metabolic state, growth conditions, and the presence of other biomolecules. To address this, advanced bioreactors with precise control over parameters like pH, temperature, and nutrient availability could enhance NP consistency and quality. Achieving uniformity in NP properties is important for their usages, particularly in fields, such as drug delivery and food (Gomez-Zavaglia et al. 2022; Shanmugam et al. 2023; Osman et al. 2024).

The other one, understanding the exact mechanisms by which microorganisms synthesize NPs is still incomplete. Different microorganisms use varied biochemical pathways for NP synthesis, and these pathways can be both intracellular and extracellular. Comprehensive knowledge of these mechanisms is important for optimizing and controlling the synthesis process (Shanmugam et al. 2023). Future work should focus on identifying and controlling these pathways to improve NP synthesis. The type of microorganisms can also affect the characteristics of nanoparticles synthesized by green technology. Some properties of ZnO NPs synthesized by different beneficial microorganisms (*Lactobacillus* sp., *L. plantarum* VITES07, and *Saccharomyces cerevisiae*) are presented and compared in Figure 5. While the shape of all ZnO NPs synthesized by different beneficial microorganisms was observed as spherical by SEM analyses, the sizes of nanoparticles were measured as 32 nm, 7-19 nm, and 13-20 nm for *Lactobacillus* sp., *L. plantarum* VITES07 and *S. cerevisiae*, respectively. The average crystalline sizes of ZnO NPs were also calculated by XRD peaks using Scherrer's equation as 7 nm, 18.6 nm, and 13.58 nm, respectively. FTIR results showed the ZnO stretching vibrations patterns were correlated with 528 cm⁻¹, 641 cm⁻¹, and 570.2 cm⁻¹ for NPs synthesized by *L. plantarum* VITES07, *Lactobacillus* sp. and *S. cerevisiae*, respectively. Ensuring the biocompatibility and non-toxicity of biosynthesized NPs is vital, especially for medical and food applications. The presence of residual biomolecules from the synthesis process can impact the toxicity profile of NPs. Detailed toxicological studies are required to ensure safety and efficacy, which can be time-consuming and resource-intensive (Kışla et al. 2023).

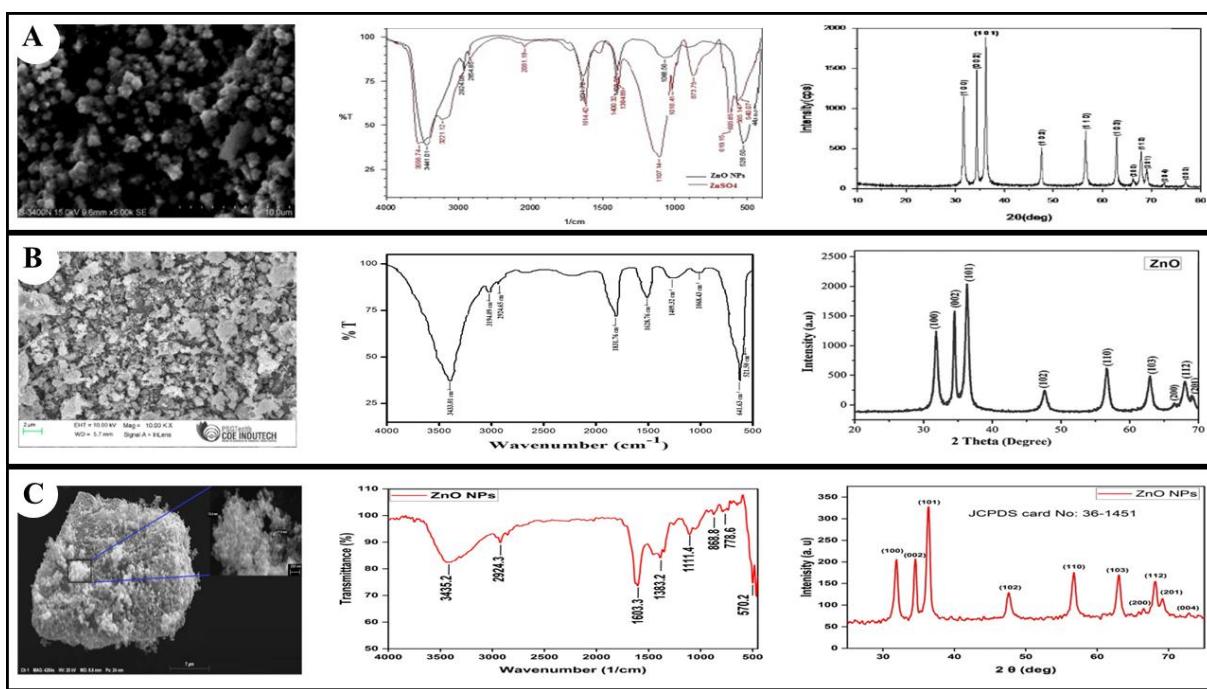


Figure 5- Comparison of the properties of ZnO NPs synthesized by different beneficial microorganisms including *Lactiplantibacillus plantarum* VITES07 (A) (Selvarajan et al. 2013), *Lactobacillus* sp. (B) (Suba et al. 2021), and *Saccharomyces cerevisiae* (C) (El-Khawaga et al. 2023)

While green synthesis is environmentally friendly, its economic viability is a concern factor. The costs associated with cultivating microorganisms, optimizing growth conditions, and harvesting NPs can be high. Besides, the extraction and purification processes should be efficient to make the overall process cost-effective compared to conventional methods (Alsaiari et al. 2023; Osman et al. 2024). Nevertheless, advances in genetic engineering offer promising solutions to improve the efficiency and control of NP synthesis. By manipulating the genetic pathways involved in metal ion reduction and NP formation, microorganisms can be engineered to synthesize NPs with desired characteristics. This approach could increase the reproducibility and scalability of biosynthesis processes. Also, detailed studies on optimizing the growth conditions of microorganisms can lead to more consistent and controllable NP synthesis. Factors such as pH, temperature, nutrient availability, and the presence of specific inducers need to be finely tuned to maximize NP yield and quality. Developing bioreactors with precise control over these parameters could facilitate large-scale production. Combining biological and chemical synthesis methods could overcome some of the limitations of pure biological synthesis (Alsaiari et al. 2023).

Post-synthesis functionalization and surface modification of NPs can also improve their biocompatibility, stability, and functionality. Techniques such as coating NPs with peptides or biopolymers etc. can improve their dispersibility in different media and target-specific applications. This is particularly important for medical applications where targeted drug delivery is significant (Gavas et al. 2021; Kişi et al. 2023). Conducting application-specific research to tailor NPs for particular uses can accelerate their adoption in various fields. For example, in agriculture, NPs can be designed for slow-release fertilizers or pesticides, while in medicine, they can be used for specific drug delivery systems or imaging agents (Liu et al. 2021; Sivarethnamohan & Sujatha 2021). So, collaborative research across disciplines can lead to innovative solutions and practical applications of synthesized NPs.

A comprehensive assessment of the environmental impact of NP synthesis and application is necessary. Despite green synthesis being eco-friendly compared to traditional methods, the long-term effects of NPs in the environment need to be understood. For this scope, in addition to the increase in the synthesis of NPs with different types, shapes, and characteristics, the scientific community has increasingly focused on examining their environmental impact, as well as their effects on living things (Martínez et al. 2021). Multiple pathways exist for NPs to enter the environment, these sources can be categorized into point sources (such as factories, landfills, and wastewater treatment plants) and non-point sources (materials containing NPs). Besides, NPs are directly introduced into the environment through applications such as remediation technologies, causing their aggregation and subsequent infiltration into soil, groundwater, and sediments. This has resulted in a novel research domain described as "Nanoecotoxicology" (Bimová et al. 2021). The intrinsic toxicity of NPs can be related to their propensity to reach and accumulate in air, soil, and water, with their effect directly proportional to their degree of accumulation. The risks of NPs are affected by various processes that impact their release mechanisms (Martínez et al. 2021). The principal mechanism underlying the potential toxicity of NPs includes their property to induce peroxidation of membrane lipids and oxidative stress, which subsequently causes the generation of ROS and resultant damage to cellular structures (Czyżowska & Barbasz 2022). While the application of NPs offers numerous advantages across various industries, it is imperative to consider and address their

potential negative effects on the environment. Hence, toxicology studies of NP synthesized on negative effects not only on humans but also on the environment should be thoroughly studied. The interaction of these NPs with crucial nutrients, especially for plants is also a compelling area of research. So, a detailed assessment of all factors such as application method, exposure time, and environmental factors, as well as NP's shape, size, and concentration is required (Bundschuh et al. 2016; Selmani et al. 2020).

As a result, while the green synthesis of NPs using beneficial microorganisms holds great promise, several challenges must be addressed to fully realize its potential. Future research should focus on improving reproducibility, scalability, and understanding the underlying mechanisms of NP synthesis, alongside advancements in genetic engineering, hybrid synthesis, and environmental safety. Collaborative efforts between microbiologists, engineers, chemists, and material scientists will be crucial to overcome these challenges and enable the widespread application of green-synthesized NPs.

5. Conclusions

The green synthesis of NPs offers a sustainable and environmentally friendly alternative to traditional chemical and physical methods. Beneficial microorganisms have also attracted special interest in the green synthesis of NPs due to their non-pathogenic properties of them, high production of various enzymes, as well as GRAS status that allows their use in various food and nutraceutical products. In this review, NP synthesis by beneficial microorganisms as green synthesis has been highlighted, and the potential application fields of NPs summarising their efficiency have been described. The green synthesis approach leverages the natural metabolic processes of microorganisms to produce NPs with unique properties suitable for various applications. Different NPs such as Ag, Se, Pd, Sb₂O₃, ZnO, and TiO₂ are synthesized by beneficial microorganisms, and these NPs have various characteristics in terms of size (20-300 nm), morphology (spherical or hexagonal), as well as characteristic properties such as antimicrobial, antioxidant, antiproliferative, anticancer, and antidiabetic activity, as presented in this review. Besides, microbial NPs have potential use in various fields as antimicrobial, anticancer, nanocarrier, and nanofertilizer, and can also play an important role in enhancing food quality and safety through developed processing and/or packaging, and long-term storage techniques. Despite the evident potential and diverse applications, significant challenges remain in ensuring reproducibility, controlling NP characteristics, understanding synthesis mechanisms, and ensuring economic viability. For this scope, the toxicity effect of NP synthesis and applications on living things and the environment should be comprehensively assessed. In conclusion, while microbial synthesis of NP presents several challenges, the future prospects are bright. Future research focused on application-specific development and comprehensive environmental impact assessments will be crucial in harnessing the full potential of this green synthesis method. As the field progresses, the integration of interdisciplinary research and innovative technological approaches will likely lead to significant advancements, making synthesized NP a cornerstone in various industries, especially in food, agriculture, and medical applications.

Conflicts of Interest

The author declares that there is no conflict of interest to disclose.

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References

Abbas S, Nasreen S, Haroon A & Ashraf M A (2020). Synthesis of silver and copper nanoparticles from plants and application as adsorbents for naphthalene decontamination. *Saudi Journal of Biological Sciences* 27: 1016-1023. <https://doi.org/10.1016/j.sjbs.2020.02.011>

Abd Elsalam S S, Taha R H, Tawfeik A M, El-Monem A, Mohamed O & Mahmoud H A (2018). Antimicrobial activity of bio and chemical synthesized cadmium sulfide nanoparticles. *The Egyptian Journal of Hospital Medicine* 70(9): 1494-1507

Abd Qasim M & Yaaqoob L A (2023). Evaluation of antibacterial activity of iron oxide nanoparticles synthesis by extracellular *Lactobacillus* against *Pseudomonas aeruginosa*. *Journal of Medicinal and Chemical Sciences* 6: 1100. <https://doi.org/10.26655/JMCHEMSCI.2023.5.15>

Abdulradha H A & Alhadrawi S W (2023). Biological and medical efficacy of zinc oxide nanoparticles manufactured using *Saccharomyces boulardii* against *Burkholderia* sp. isolated from diabetic foot patients. *International Journal of Advanced Multidisciplinary Research and Studies* 3(3): 703-708

Abo-zeid Y & Williams G R (2020). The potential anti-infective applications of metal oxide nanoparticles: A systematic review. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology* 12(2): e1592. <https://doi.org/10.1002/wnan.1592>

Adelere I A & Lateef A (2016). A novel approach to the green synthesis of metallic nanoparticles: the use of agro-wastes, enzymes, and pigments. *Nanotechnology Reviews* 5(6): 567-587. <https://doi.org/10.1515/ntrev-2016-0024>

Ahmad M Z, Alasiri A S, Ahmad J, Alqahtani A A, Abdullah M M, Abdel-Wahab B A, Pathak K, Saikia R, Das A, Sarma H & Alzahrani S A (2022). Green synthesis of titanium dioxide nanoparticles using *Ocimum sanctum* leaf extract: *In vitro* characterization and its healing efficacy in diabetic wounds. *Molecules* 27(22): 7712. <https://doi.org/10.3390/molecules27227712>

Ahmad R, Khatoon N & Sardar M (2014). Antibacterial effect of green synthesized TiO₂ nanoparticles. *Advanced Science Letters* 20(7-8): 1616-1620. <https://doi.org/10.1166/asl.2014.5563>

Ahmed B, Tahir M B, Sagir M & Hassan M (2024). Bio-inspired sustainable synthesis of silver nanoparticles as next generation of nanoproduct in antimicrobial and catalytic applications. *Materials Science and Engineering: B* 301: 117165. <https://doi.org/10.1016/j.mseb.2023.117165>

Ahmed E, Kalathil S, Shi L, Alharbi O & Wang P (2018). Synthesis of ultra-small platinum, palladium and gold nanoparticles by *Shewanella loihica* PV-4 electrochemically active biofilms and their enhanced catalytic activities. *Journal of Saudi Chemical Society* 22(8): 919-929. <https://doi.org/10.1016/j.jscs.2018.02.002>

Ahmed K B A, Kalla D, Uppuluri K B & Anbazhagan V (2014) Green synthesis of silver and gold nanoparticles employing levan, a biopolymer from *Acetobacter xylinum* NCIM 2526, as a reducing agent and capping agent. *Carbohydrate Polymers* 112: 539-545. <https://doi.org/10.1016/j.carbpol.2014.06.033>

Ahmed T, Shahid M, Noman M, Niazi M B K, Mahmood F, Manzoor I, Zhang Y, Li B, Yang Y & Yan Chen J (2020). Silver nanoparticles synthesized by using *Bacillus cereus* SZT1 ameliorated the damage of bacterial leaf blight pathogen in rice. *Pathogens* 9(3): 160. <https://doi.org/10.3390/pathogens9030160>

Alam H, Khatoon N, Khan M A, Husain S A, Saravanan M & Sardar M (2020). Synthesis of selenium nanoparticles using probiotic bacteria *Lactobacillus acidophilus* and their enhanced antimicrobial activity against resistant bacteria. *Journal of Cluster Science* 31: 1003-1011. <https://doi.org/10.1007/s10876-019-01705-6>

Al-Asbahi M G, Al-Ofiry B A, Saad F A, Alnehia A & Al-Gunaid M Q (2024). Silver nanoparticles biosynthesis using mixture of *Lactobacillus* sp. and *Bacillus* sp. growth and their antibacterial activity. *Scientific Reports* 14(1): 10224. <https://doi.org/10.1038/s41598-024-59936-1>

Alghuthaymi M A, Almoammar H, Rai M, Said-Galiev E & Abd-Elsalam K A (2015) Myconanoparticles: synthesis and their role in phytopathogens management. *Biotechnology & Biotechnological Equipment* 29(2): 221-236. <https://doi.org/10.1080/13102818.2015.1008194>

Al-Khattaf F S (2021). Gold and silver nanoparticles: Green synthesis, microbes, mechanism, factors, plant disease management and environmental risks. *Saudi Journal of Biological Sciences* 28(6): 3624-3631. <https://doi.org/10.1016/j.sjbs.2021.03.078>

Alsaab H O, Al-Hibs A S, Alzahrani R, Alrabighi K K, Alqathama A, Alwithenani A, Almaliki A H & Althobaiti Y S (2021). Nanomaterials for antiangiogenic therapies for cancer: a promising tool for personalized medicine. *International Journal of Molecular Sciences* 22(4): 1631. <https://doi.org/10.3390/ijms22041631>

Alsaiari N S, Alzahrani F M, Amari A, Osman H, Harharah H N, Elboughdiri N & Tahoob M A (2023). Plant and microbial approaches as green methods for the synthesis of nanomaterials: synthesis, applications, and future perspectives. *Molecules* 28(1): 463. <https://doi.org/10.3390/molecules28010463>

Amini S M, Hadighi R, Najm M, Alipour M, Hasanpour H, Vosoogh M, Vosough A, Hajizadeh M & Badirzadeh A (2023) The therapeutic effects of curcumin-coated gold nanoparticle against *Leishmania* major causative agent of zoonotic cutaneous leishmaniasis (ZCL): an *in vitro* and *in vivo* study. *Current Microbiology* 80(4): 104. <https://doi.org/10.1007/s00284-022-03172-1>

Ammar H A, El Aty A A A & El Awdan S A (2021) Extracellular myco-synthesis of nano-silver using the fermentable yeasts *Pichia kudriavzevii* HA-NY2 and *Saccharomyces uvarum* HA-NY3, and their effective biomedical applications. *Bioprocess and Biosystems Engineering* 44: 841-854. <https://doi.org/10.1007/s00449-020-02494-3>

Ananthi V, Prakash G S, Rasu K M, Gangadevi K, Boobalan T, Raja R, Anand K, Sudhakar M, Chuturgoon A & Arun A (2018) Comparison of integrated sustainable biodiesel and antibacterial nano silver production by microalgal and yeast isolates. *Journal of Photochemistry and Photobiology B: Biology* 186: 232-242. <https://doi.org/10.1016/j.jphotobiol.2018.07.021>

Andleeb A, Andleeb A, Asghar S, Zaman G, Tariq M, Mahmood A & Abbasi B H (2021). A systematic review of biosynthesized metallic nanoparticles as a promising anti-cancer-strategy. *Cancers* 13(11): 2818. <https://doi.org/10.3390/cancers13112818>

Anis S N S, Liew W C, Marsin A M, Muhamad I I, Teh S H & Khudzari A Z M (2023) Microwave-assisted green synthesis of silver nanoparticles using pineapple leaves waste. *Cleaner Engineering and Technology* 15: 100660. <https://doi.org/10.1016/j.clet.2023.100660>

Annamalai J, Ummalyma S B, Pandey A & Bhaskar T (2021) Recent trends in microbial nanoparticle synthesis and potential application in environmental technology: a comprehensive review. *Environmental Science and Pollution Research* 28(36): 49362-49382. <https://doi.org/10.1007/s11356-021-15680-x>

Arriaza-Echanes C, Campo-Giraldo J L, Valenzuela-Ibaceta F, Ramos-Zúñiga J & Pérez-Donoso J M (2024) Biosynthesis of Cu-In-S nanoparticles by a yeast isolated from Union glacier, Antarctica: A platform for enhanced quantum dot-sensitized solar cells. *Nanomaterials* 14(6): 552. <https://doi.org/10.3390/nano14060552>

Ashok B, Hariram N, Siengchin S & Rajulu A V (2020). Modification of tamarind fruit shell powder with in situ generated copper nanoparticles by single step hydrothermal method. *Journal of Bioresources and Bioproducts* 5(3): 180-185. <https://doi.org/10.1016/j.jobab.2020.07.003>

Badoei-dalfard A, Shaban M & Karami Z (2019). Characterization, antimicrobial, and antioxidant activities of silver nanoparticles synthesized by uricase from *Alcaligenes faecalis* GH3. *Biocatalysis and Agricultural Biotechnology* 20: 101257. <https://doi.org/10.1016/j.bcab.2019.101257>

Bajpai V K, Kamle M, Shukla S, Mahato D K, Chandra P, Hwang S K, Kumar P, Huh Y K & Han Y K (2018a). Prospects of using nanotechnology for food preservation, safety, and security. *Journal of Food and Drug Analysis* 26(4): 1201-1214. <https://doi.org/10.1016/j.jfda.2018.06.011>

Bajpai V K, Shukla S, Kang S M, Hwang S K, Song X, Huh Y S & Han Y K (2018b). Developments of cyanobacteria for nano-marine drugs: Relevance of nanoformulations in cancer therapies. *Marine Drugs* 16(6): 179. <https://doi.org/10.3390/md16060179>

Bakkiyaraj R, Subramanian R, Balakrishnan M & Ravichandran K (2021). Biofabrication of CeO₂ nanoparticles, characterization, photocatalytic, and biological activities. *Inorganic and Nano-Metal Chemistry* 1-9. <https://doi.org/10.1080/24701556.2021.1983841>

Balakrishnan S, Sivaji I, Kandasamy S, Duraisamy S, Kumar N S & Gurusubramanian G (2017). Biosynthesis of silver nanoparticles using *Myristica fragrans* seed (nutmeg) extract and its antibacterial activity against multidrug-resistant (MDR) *Salmonella enterica* serovar Typhi isolates. *Environmental Science and Pollution Research* 24: 14758-14769. <https://doi.org/10.1007/s11356-017-9065-7>

Bandeira M, Giovanelia M, Roesch-Ely M, Devine D M & da Silva Crespo J (2020). Green synthesis of zinc oxide nanoparticles: A review of the synthesis methodology and mechanism of formation. *Sustainable Chemistry and Pharmacy* 15: 100223. <https://doi.org/10.1016/j.scp.2020.100223>

Barabadi H, Honary S, Ali Mohammadi M, Ahmadpour E, Rahimi M T, Alizadeh A, Naghibi F & Saravanan M (2017). Green chemical synthesis of gold nanoparticles by using *Penicillium aculeatum* and their scolicidal activity against hydatid cyst protoscolices of *Echinococcus granulosus*. *Environmental Science and Pollution Research* 24: 5800-5810. <https://doi.org/10.1007/s11356-016-8291-8>

Barabadi H, Vahidi H, Arjmand M, Abdorashidi M, Jahani R, Amidi S, Hosseini O, Sadeghian-Abadi S, Jounaki K & Ashouri F (2024). Exploring the biological properties of *Saccharomyces cerevisiae*-derived silver nanoparticles: *In vitro* structural characteristics, antibacterial, biofilm inhibition and biofilm degradation, antioxidant, anticoagulant, thrombolytic, and antidiabetic performance. *Inorganic Chemistry Communications* 162: 112291. <https://doi.org/10.1016/j.inoche.2024.112291>

Bhardwaj B, Singh P, Kumar A, Kumar S & Budhwar V (2020). Eco-friendly greener synthesis of nanoparticles. *Advanced Pharmaceutical Bulletin* 10(4): 566. <https://doi.org/10.34172/apb.2020.067>

Bhuyar P, Rahim M H A, Sundararaju S, Ramaraj R, Maniam G P & Govindan N (2020). Synthesis of silver nanoparticles using marine macroalgae *Padina* sp. and its antibacterial activity towards pathogenic bacteria. *Beni-Suef University Journal of Basic and Applied Sciences* 9: 1-15. <https://doi.org/10.1186/s43088-019-0031-y>

Bimová P, Barbieríková Z, Grenčíková A, Šipoš R, Škulcová A B, Krivjanská A & Mackuľák T (2021). Environmental risk of nanomaterials and nanoparticles and EPR technique as an effective tool to study them-a review. *Environmental Science and Pollution Research* 28: 22203-22220. <https://doi.org/10.1007/s11356-021-13270-5>

Bisinoti M C, Moreira A B, Melo C A, Fregolente L G, Bento L R, dos Santos J V & Ferreira O P (2019). Application of carbon-based nanomaterials as fertilizers in soils, in do Nascimento R F, Ferreira O P, De Paula A J, Neto V O S (Eds.), *Nanomaterials Applications for Environmental Matrices*. Elsevier, Netherlands, pp. 305-333

Bloise N, Okkeh M, Restivo E, Della Pina C & Visai L (2021). Targeting the "Sweet Side" of tumor with glycan-binding molecules conjugated-nanoparticles: Implications in cancer therapy and diagnosis. *Nanomaterials* 11(2): 289. <https://doi.org/10.3390/nano11020289>

Bolbanabad E M, Ashengroh M & Darvishi F (2020). Development and evaluation of different strategies for the clean synthesis of silver nanoparticles using *Yarrowia lipolytica* and their antibacterial activity. *Process Biochemistry* 94: 319-328. <https://doi.org/10.1016/j.procbio.2020.03.024>

Borah D, Das N, Das N, Bhattacharjee A, Sarmah P, Ghosh K, Chandel M, Rout J, Pandey P, Ghosh N N & Bhattacharjee C R (2020). Alga-mediated facile green synthesis of silver nanoparticles: Photophysical, catalytic and antibacterial activity. *Applied Organometallic Chemistry* 34(5): e5597. <https://doi.org/10.1002/aoc.5597>

Borse V, Kaler A & Banerjee U C (2015). Microbial synthesis of platinum nanoparticles and evaluation of their anticancer activity. *International Journal of Emerging Trends in Electrical and Electronics* 11(2): 26-31. <https://doi.org/10.13140/RG.2.1.3132.5283>

Buddhika M A A, Perumpuli P A B N & Kaumal M N (2021). Identification and characterization of acetic acid bacteria species isolated from various sources in Sri Lanka. *Ceylon Journal of Science* 50(4): 521-532. [http://doi.org/10.4038/cjs.v50i4.7951](https://doi.org/10.4038/cjs.v50i4.7951)

Bundschuh M, Seitz F, Rosenfeldt R R & Schulz R (2016). Effects of nanoparticles in fresh waters: risks, mechanisms and interactions. *Freshwater Biology* 61(12): 2185-2196. <https://doi.org/10.1111/fwb.12701>

Cai Y, Wu D, Zhu X, Wang W, Tan F, Chen J, Qiao X & Qiu X (2017). Sol-gel preparation of Ag-doped MgO nanoparticles with high efficiency for bacterial inactivation. *Ceramics International* 43(1): 1066-1072. <https://doi.org/10.1016/j.ceramint.2016.10.041>

Caliskan G, Mutaf T, Agba H C & Elibol M (2022). Green synthesis and characterization of titanium nanoparticles using microalga, *Phaeodactylum tricornutum*. *Geomicrobiology Journal* 39(1): 83-96. <https://doi.org/10.1080/01490451.2021.2008549>

Cassano R, Cuconato M, Calviello G, Serini S & Trombino S (2021). Recent advances in nanotechnology for the treatment of melanoma. *Molecules* 26(4): 785. <https://doi.org/10.3390/molecules26040785>

Castro-Longoria E, Vilchis-Nestor A R & Avalos-Borja M (2011). Biosynthesis of silver, gold and bimetallic nanoparticles using the filamentous fungus *Neurospora crassa*. *Colloids and surfaces B: Biointerfaces* 83(1): 42-48. <https://doi.org/10.1016/j.colsurfb.2010.10.035>

CFS Center for Food Safety (2017). (www.centerforfoodsafety.org) Retrieved from http://salsa3.salsalabs.com/o/1881/p/salsa/web/common/public/content?content_item_KEY=14112%20#showJoin

Cha D S & Chinnan M S (2004). Biopolymer-based antimicrobial packaging: A review. *Critical Reviews in Food Science and Nutrition* 44(4): 223-237. <https://doi.org/10.1080/10408690490464276>

Chan M Z A & Liu S Q (2022). Fortifying foods with symbiotic and postbiotic preparations of the probiotic yeast, *Saccharomyces boulardii*. *Current Opinion in Food Science* 43: 216-224. <https://doi.org/10.1016/j.cofs.2021.12.009>

Chen H (2018). Metal based nanoparticles in agricultural system: Behavior, transport, and interaction with plants. *Chemical Speciation & Bioavailability* 30: 123-134. <https://doi.org/10.1080/09542299.2018.1520050>

Chhipa H (2017). Nanofertilizers and nanopesticides for agriculture. *Environmental Chemistry Letters* 15: 15-22. <https://doi.org/10.1007/s10311-016-0600-4>

Chi N T L, Narayanan M, Chinnathambi A, Govindasamy C, Subramani B, Brindhadevi K, Pimpimon T & Pikulkaew S (2022). Fabrication, characterization, anti-inflammatory, and anti-diabetic activity of silver nanoparticles synthesized from *Azadirachta indica* kernel aqueous extract. *Environmental Research* 208: 112684. <https://doi.org/10.1016/j.envres.2022.112684>

Chohan K L, Siegler E L & Kenderian S S (2023). CAR-T cell therapy: the efficacy and toxicity balance. *Current Hematologic Malignancy Reports* 18(2), 9-18. <https://doi.org/10.1007/s11899-023-00687-7>

Chugh D, Viswamalya V S & Das B (2021). Green synthesis of silver nanoparticles with algae and the importance of capping agents in the process. *Journal of Genetic Engineering and Biotechnology*, 19(1), 126. <https://doi.org/10.1186/s43141-021-00228-w>

Cunha F A, Cunha M D C, da Frotta S M, Mallmann E J, Freire T M, Costa L S, Paula A J, Menezes E A & Fechine P B (2018). Biogenic synthesis of multifunctional silver nanoparticles from *Rhodotorula glutinis* and *Rhodotorula mucilaginosa*: Antifungal, catalytic and cytotoxicity activities. *World Journal of Microbiology and Biotechnology* 34: 1-15. <https://doi.org/10.1007/s11274-018-2514-8>

Czyżowska A & Barbasz A (2022). A review: zinc oxide nanoparticles-friends or enemies? *International Journal of Environmental Health Research* 32(4): 885-901. <https://doi.org/10.1080/09603123.2020.1805415>

da Silva Ferreira V, ConzFerreira M E, Lima L M T, Frasés S, de Souza W & Sant'Anna C (2017). Green production of microalgae-based silver chloride nanoparticles with antimicrobial activity against pathogenic bacteria. *Enzyme and Microbial Technology* 97: 114-121. <https://doi.org/10.1016/j.enzmictec.2016.10.018>

Das V L, Thomas R, Varghese R T, Soniya E V, Mathew J & Radhakrishnan E K (2014). Extracellular synthesis of silver nanoparticles by the *Bacillus* strain CS 11 isolated from industrialized area. *3 Biotech* 4: 121-126. <https://doi.org/10.1007/s13205-013-0130-8>

De Bellis P & Rizzello C G (2024). Advances in the use of beneficial microorganisms to improve nutritional and functional properties of fermented foods. *Foods* 13(1): 155. <https://doi.org/10.3390/foods13010155>

de Oliveira J L, Campos E V R, Bakshi M, Abhilash P C & Fraceto L F (2014). Application of nanotechnology for the encapsulation of botanical insecticides for sustainable agriculture: prospects and promises. *Biotechnology Advances* 32(8): 1550-1561. <https://doi.org/10.1016/j.biotechadv.2014.10.010>

Dhabalia D, Ukkund S J, Syed U T, Uddin W & Kabir M A (2020). Antifungal activity of biosynthesized silver nanoparticles from *Candida albicans* on the strain lacking the CNP41 gene. *Materials Research Express* 7(12): 125401. <https://doi.org/10.1088/2053-1591/abcc83>

Doan V D, Huynh B A, Nguyen T D, Cao X T, Nguyen V C, Nguyen T L H, Nguyen H T & Le V T (2020). Biosynthesis of silver and gold nanoparticles using aqueous extract of *Codonopsis pilosula* roots for antibacterial and catalytic applications. *Journal of Nanomaterials* 8492016. <https://doi.org/10.1155/2020/8492016>

Dutta T, Chattopadhyay A P, Ghosh N N, Khatua S, Acharya K, Kundu S, Mitra D & Das M (2020). Biogenic silver nanoparticle synthesis and stabilization for apoptotic activity; insights from experimental and theoretical studies. *Chemical Papers* 74: 4089-4101. <https://doi.org/10.1007/s11696-020-01216-z>

EFSA (European Food Safety Authority) (2009). The potential risks arising from nanoscience and nanotechnologies on food and feed safety. *EFSA Journal* 7(3): 958. <https://doi.org/10.2903/j.efsa.2009.958>

Elamawi R M, Al-Harbi R E & Hendi A A (2018). Biosynthesis and characterization of silver nanoparticles using *Trichoderma longibrachiatum* and their effect on phytopathogenic fungi. *Egyptian Journal of Biological Pest Control* 28(1): 1-11. <https://doi.org/10.1186/s41938-018-0028-1>

Elegbede J A, Lateef A, Azeez M A, Asafa T B, Yekeen T A, Oladipo I C, Adebayo E A, Beukes L S & Gueguim-Kana E B (2018a). Fungal xylanases-mediated synthesis of silver nanoparticles for catalytic and biomedical applications. *IET Nanobiotechnology* 12(6): 857-863. <https://doi.org/10.1049/iet-nbt.2017.0299>

Elegbede J A, Lateef A, Azeez M A, Asafa T B, Yekeen T A, Oladipo I C, Aina D A, Beukes L S & Gueguim-Kana E B (2018b). Biofabrication of gold nanoparticles using xylanases through valorization of corncobs by *Aspergillus niger* and *Trichoderma longibrachiatum*: antimicrobial, antioxidant, anticoagulant and thrombolytic activities. *Waste Biomass Valorization* 1-11. <https://doi.org/10.1007/s12649-018-0540-2>

El-Khawaga A M, Elsayed M A, Gobara M, Suliman A A, Hashem A H, Zaher A A, Mohsen M & Salem S S (2023). Green synthesized ZnO nanoparticles by *Saccharomyces cerevisiae* and their antibacterial activity and photocatalytic degradation. *Biomass Conversion and Biorefinery* 1-12. <https://doi.org/10.1007/s13399-023-04827-0>

Elmer W & White J C (2018). The future of nanotechnology in plant pathology. *Annual Review of Phytopathology* 56: 111-133. <https://doi.org/10.1146/annurev-phyto-080417-050108>

El-Seedi H R, El-Shabasy R M, Khalifa S A, Saeed A, Shah A, Shah R & Guo W (2019). Metal nanoparticles fabricated by green chemistry using natural extracts: Biosynthesis, mechanisms, and applications. *RSC Advances* 9(42): 24539-24559. <https://doi.org/10.1039/C9RA02225B>

El-Sheekh M M, Shabaan M T, Hassan L & Morsi H H (2022). Antiviral activity of algae biosynthesized silver and gold nanoparticles against Herpes Simplex (HSV-1) virus in vitro using cell-line culture technique. *International Journal of Environmental Health Research* 32(3): 616-627. <https://doi.org/10.1080/09603123.2020.1789946>

Emerich D F & Thanos C G (2006). The pinpoint promise of nanoparticle-based drug delivery and molecular diagnosis. *Biomolecular Engineering* 23(4): 171-184. <https://doi.org/10.1016/j.bioeng.2006.05.026>

Eszenyi P, Sztrik A, Babka B & Prokisch J (2011). Elemental, nano-sized (100-500 nm) selenium production by probiotic lactic acid bacteria. *International Journal of Bioscience, Biochemistry and Bioinformatics* 1(2), 148.

Fadeel B & Garcia-Bennett A E (2010). Better safe than sorry: Understanding the toxicological properties of inorganic nanoparticles manufactured for biomedical applications. *Advanced Drug Delivery Reviews* 62(3): 362-374. <https://doi.org/10.1016/j.addr.2009.11.008>

Fahmy S A, Preis E, Bakowsky U & Azzazy H M E S (2020). Platinum nanoparticles: green synthesis and biomedical applications. *Molecules*, 25(21), 4981. <https://doi.org/10.3390/molecules25214981>

Faramarzi S, Anzabi Y & Jafarizadeh-Malmiri H (2020). Nanobiotechnology approach in intracellular selenium nanoparticle synthesis using *Saccharomyces cerevisiae*—fabrication and characterization. *Archives of Microbiology* 202(5): 1203-1209. <https://doi.org/10.1007/s00203-020-01831-0>

Fatemi M, Mollania N, Momeni-Moghaddam M & Sadeghifar F (2018). Extracellular biosynthesis of magnetic iron oxide nanoparticles by *Bacillus cereus* strain HMH1: Characterization and *in vitro* cytotoxicity analysis on MCF-7 and 3T3 cell lines. *Journal of Biotechnology* 270: 1-11. <https://doi.org/10.1016/j.jbiotec.2018.01.021>

Fatima R, Priya M, Indurthi L, Radhakrishnan V & Sudhakaran R (2020). Biosynthesis of silver nanoparticles using red algae *Portieria hornemannii* and its antibacterial activity against fish pathogens. *Microbial Pathogenesis* 138: 103780. <https://doi.org/10.1016/j.micpath.2019.103780>

Fatimah I, Purwiandono G, Jauhari M H, Maharani A A A P, Sagadevan S, Oh W C & Doong R. A (2022). Synthesis and control of the morphology of SnO₂ nanoparticles via various concentrations of *Tinospora cordifolia* stem extract and reduction methods. *Arabian Journal of Chemistry* 15(4): 103738. <https://doi.org/10.1016/j.arabjc.2022.103738>

Fawcett D, Verduin J J, Shah M, Sharma S B & Poinern G E J (2017). A review of current research into the biogenic synthesis of metal and metal oxide nanoparticles via marine algae and seagrasses. *Journal of Nanoscience* 2017(1): 8013850. <https://doi.org/10.1155/2017/8013850>

Fedotova A V, Snejhko A G, Sdobnikova O A, Samoilova L G, Smurova T A, Revina A A & Khailova E B (2010). Packaging materials manufactured from natural polymers modified with silver nanoparticles. *International Polymer Science and Technology* 37(10): 59-64. <https://doi.org/10.1177/0307174X1003701010>

Feng H, Liu S Y, Huang X B, Ren R, Zhou Y, Song C P & Qian D H (2017). Green biosynthesis of CdS nanoparticles using yeast cells for fluorescence detection of nucleic acids and electrochemical detection of hydrogen peroxide. *International Journal of Electrochemical Science* 12(1): 618-628. <https://doi.org/10.20964/2017.01.57>

Feroze N, Arshad B, Younas M, Afzidi M I, Saqib S & Ayaz A (2020). Fungal mediated synthesis of silver nanoparticles and evaluation of antibacterial activity. *Microscopy Research and Technique* 83(1): 72-80. <https://doi.org/10.1002/jemt.23390>

Fu X, Cai J, Zhang X, Li W D, Ge H & Hu Y (2018). Top-down fabrication of shape-controlled, monodisperse nanoparticles for biomedical applications. *Advanced Drug Delivery Reviews* 132: 169-187. <https://doi.org/10.1016/j.addr.2018.07.006>

Gaba S, Varma A, Prasad R & Goel A (2022). Exploring the impact of bioformulated copper oxide nanoparticles on cytomorphology of *Alternaria brassicicola*. *Current Microbiology* 79(8): 244. <https://doi.org/10.1007/s00284-022-02927-0>

Gahlawat G & Choudhury A R (2019). A review on the biosynthesis of metal and metal salt nanoparticles by microbes. *RSC Advances* 9(23): 12944-12967. <https://doi.org/10.1039/C8RA10483B>

Gan L, Zhang S, Zhang Y, He S & Tian Y (2018). Biosynthesis, characterization and antimicrobial activity of silver nanoparticles by a halotolerant *Bacillus endophyticus* SCU-L. *Preparative Biochemistry and Biotechnology* 48(7): 582-588. <https://doi.org/10.1080/10826068.2018.1476880>

Gavas S, Quazi S & Karpinski T M (2021). Nanoparticles for cancer therapy: Current progress and challenges. *Nanoscale Research Letters* 16(1): 173. <https://doi.org/10.1186/s11671-021-03628-6>

Gegechkori N, Haines L & Lin J J (2017). Long-term and latent side effects of specific cancer types. *Medical Clinics* 101(6): 1053-1073. <https://doi.org/10.1016/j.mcna.2017.06.003>

Gholami A, Mohkam M, Soleimanian S, Sadraeian M & Lauto A (2024). Bacterial nanotechnology as a paradigm in targeted cancer therapeutic delivery and immunotherapy. *Microsystems & Nanoengineering*, 10(1), 113. <https://doi.org/10.1038/s41378-024-00743-z>

Gholami-Shabani M, Shams-Ghahfarokhic M, Gholami-Shabanid Z, Akbarzadeh A, Riazi G, Ajdarif S, Amanig A & Razzaghi-Abyaneh M (2015). Enzymatic synthesis of gold nanoparticles using sulfite reductase purified from *Escherichia coli*: a green eco-friendly approach. *Process Biochemistry* 50: 1076-1085. <https://doi.org/10.1016/j.procbio.2015.04.004>

Ghosh S, Ahmad R, Zeyaullah M & Khare S K (2021). Microbial nano-factories: Synthesis and biomedical applications. *Frontiers in Chemistry* 9: 626834. <https://doi.org/10.3389/fchem.2021.626834>

Ghosh S, Sarkar B, Kaushik A & Mostafavi E (2022). Nanobiotechnological prospects of probiotic microflora: Synthesis, mechanism, and applications. *Science of the Total Environment* 838: 156212. <https://doi.org/10.1016/j.scitotenv.2022.156212>

Gomaa E Z (2017). Silver nanoparticles as an antimicrobial agent: A case study on *Staphylococcus aureus* and *Escherichia coli* as models for Gram-positive and Gram-negative bacteria. *The Journal of General and Applied Microbiology* 63(1): 36-43. <https://doi.org/10.2323/jgam.2016.07.004>

Gomathi A C, Rajarathinam S X, Sadiq A M & Rajeshkumar S (2020). Anticancer activity of silver nanoparticles synthesized using aqueous fruit shell extract of *Tamarindus indica* on MCF-7 human breast cancer cell line. *Journal of Drug Delivery Science and Technology* 55: 101376. <https://doi.org/10.1016/j.jddst.2019.101376>

Gomez-Zavaglia A, Cassani L, Hebert E M & Gerbino E (2022). Green synthesis, characterization and applications of iron and zinc nanoparticles by probiotics. *Food Research International* 155: 111097. <https://doi.org/10.1016/j.foodres.2022.111097>

González-Ballesteros N, Prado-López S, Rodríguez-González J B, Lastra M & Rodríguez-Argüelles M (2017). Green synthesis of gold nanoparticles using brown algae *Cystoseira baccata*: Its activity in colon cancer cells. *Colloids and Surfaces B: Biointerfaces* 153: 190-198. <https://doi.org/10.1016/j.colsurfb.2017.02.020>

Gopu M, Kumar P, Selvankumar T, Senthilkumar B, Sudhakar C, Govarthanan M, Kumar R S & Selvam K (2021). Green biomimetic silver nanoparticles utilizing the red algae *Amphiroa rigida* and its potent antibacterial, cytotoxicity and larvicidal efficiency. *Bioprocess and Biosystems Engineering* 44: 217-223. <https://doi.org/10.1007/s00449-020-02426-1>

Govindappa M, Lavanya M, Aishwarya P, Pai K, Lunked P, Hemashekhar B, Arpitha B M, Ramachandra Y L & Raghavendra V B (2020). Synthesis and characterization of endophytic fungi, *Cladosporium perangustum* mediated silver nanoparticles and their antioxidant, anticancer and nano-toxicological study. *BioNanoScience* 10: 928-941. <https://doi.org/10.1007/s12668-020-00719-z>

Grasso G, Zane D & Dragone R (2020). Microbial nanotechnology: Challenges and prospects for green biocatalytic synthesis of nanoscale materials for sensoristic and biomedical applications. *Nanomaterials* 10: 11. <https://doi.org/10.3390/nano10010011>

Grillo R, Abhilash P C & Fraceto L F (2016). Nanotechnology applied to bio-encapsulation of pesticides. *Journal of Nanoscience and Nanotechnology* 16(1): 1231-1234. <https://doi.org/10.1166/jnn.2016.12332>

Gu H, Chen X, Chen F, Zhou X & Parsaei Z (2018). Ultrasound-assisted biosynthesis of CuO-NPs using brown alga *Cystoseira trinodis*: Characterization, photocatalytic AOP, DPPH scavenging and antibacterial investigations. *Ultrasonics Sonochemistry* 41: 109-119. <https://doi.org/10.1016/j.ultsonch.2017.09.006>

Guleria A, Neogy S, Raorane B S & Adhikari S (2020). Room temperature ionic liquid assisted rapid synthesis of amorphous Se nanoparticles: Their prolonged stabilization and antioxidant studies. *Materials Chemistry and Physics* 253: 123369. <https://doi.org/10.1016/j.matchemphys.2020.123369>

Guo H, White J C, Wang Z & Xing B (2018). Nano-enabled fertilizers to control the release and use efficiency of nutrients. *Current Opinion in Environmental Science & Health* 6: 77-83. <https://doi.org/10.1016/j.coesh.2018.07.009>

Gupta K & Chundawat T S (2019). Bio-inspired synthesis of platinum nanoparticles from fungus *Fusarium oxysporum*: its characteristics, potential antimicrobial, antioxidant and photocatalytic activities. *Materials Research Express* 6(10): 1050d6. <https://doi.org/10.1088/2053-1591/ab4219>

Gupte Y, Kulkarni A, Raut B, Sarkar P, Choudhury R, Chawande A, Kumar G R K, Bhadra B., Satapathy A, Das G, Vishnupriya B & Dasgupta S (2021). Characterization of nanocellulose production by strains of *Komagataeibacter* sp. isolated from organic waste and Kombucha. *Carbohydrate Polymers* 266: 118176. <https://doi.org/10.1016/j.carbpol.2021.118176>

Gursoy N (2020). Fungus-mediated synthesis of silver nanoparticles (AgNP) and inhibitory effect on *Aspergillus* spp. in combination with antifungal agent. *Cumhuriyet Science Journal* 41(1): 311-318. <https://doi.org/10.17766/csj.653627>

Halawani E M, Hassan A M & Gad El-Rab S M (2020). Nanoformulation of biogenic cefotaxime-conjugated-silver nanoparticles for enhanced antibacterial efficacy against multidrug-resistant bacteria and anticancer studies. *International Journal of Nanomedicine* 1889-1901. <https://doi.org/10.2147/IJN.S236182>

Hamk M, Akçay F A & Avcı A (2023). Green synthesis of zinc oxide nanoparticles using *Bacillus subtilis* ZBP4 and their antibacterial potential against foodborne pathogens. *Preparative Biochemistry & Biotechnology* 53(3): 255-264. <https://doi.org/10.1080/10826068.2022.2076243>

Harandi F N, Khorasani A C, Shojaosadati S A & Hashemi-Najafabadi S (2021). Living *Lactobacillus*-ZnO nanoparticles hybrids as antimicrobial and antibiofilm coatings for wound dressing application. *Materials Science and Engineering: C* 130: 112457. <https://doi.org/10.1016/j.msec.2021.112457>

Hashemi Z, Mizwari Z M, Mohammadi-Aghdam S, Mortazavi-Derazkola S & Ebrahimpour M A (2022). Sustainable green synthesis of silver nanoparticles using Sambucus ebulus phenolic extract (AgNPs@ SEE): Optimization and assessment of photocatalytic degradation of methyl orange and their *in vitro* antibacterial and anticancer activity. *Arabian Journal of Chemistry* 15(1): 103525. <https://doi.org/10.1016/j.arabjc.2021.103525>

Hata N N Y, Surek M, Sartori D, Vassoler Serrato R & Aparecida Spinosa W (2023). Role of acetic acid bacteria in food and beverages. *Food Technology and Biotechnology* 61(1): 85-103. <https://doi.org/10.17113/ftb.61.01.23.7811>

Hazards E P O B, Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Girones R, Herman L, Koutsoumaniss K, Lindqvist R, Norrung B, Robertson L, Ru G, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Kuile B T & Lindqvist R (2017). Scientific opinion on the update of the list of QPS recommended biological agents intentionally added to food or feed as notified to EFSA. *EFSA Journal* 15: e04664. <https://doi.org/10.2903/j.efsa.2017.4664>

He X, Deng H & Hwang H M (2019). The current application of nanotechnology in food and agriculture. *Journal of Food and Drug Analysis* 27(1): 1-21. <https://doi.org/10.1016/j.jfda.2018.12.002>

Hemeg H A (2017). Nanomaterials for alternative antibacterial therapy. *International Journal of Nanomedicine* 8211-8225. <https://doi.org/10.2147/IJN.S132163>

Hikmet R A & Hussein N N (2021). Mycosynthesis of silver nanoparticles by *Candida albicans* yeast and its biological applications. *Archives of Razi Institute* 76(4): 857-869. <https://doi.org/10.22092/ARI.2021.355935.1741>

Hill C, Guarner F, Reid G, Gibson G R, Merenstein D J, Pot B, Morelli L, Canani R B, Flint H J, Salminen S, Calder P C & Sanders M E (2014). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature Reviews Gastroenterology & Hepatology* 11: 506-514. <https://doi.org/10.1038/nrgastro.2014.66>

Huai Y, Zhang Y, Xiong X, Das S, Bhattacharya R & Mukherjee P (2019). Gold nanoparticles sensitize pancreatic cancer cells to gemcitabine. *Cell Stress* 3(8): 267. <https://doi.org/10.15698/cst2019.08.195>

Ibrahem K H, Ali F A & Sorchee S M A (2020). Biosynthesis and characterization with antimicrobial activity of TiO₂ nanoparticles using probiotic *Bifidobacterium bifidum*. *Cellular and Molecular Biology* 66(7): 111-117. <https://doi.org/10.14715/cmb/2020.66.7.17>

Ibrahim E, Zhang M, Zhang Y, Hossain A, Qiu W, Chen Y, Wang Y, Wu W, Sun G & Li B (2020). Green-synthesis of silver nanoparticles using endophytic bacteria isolated from garlic and its antifungal activity against wheat *Fusarium* head blight pathogen *Fusarium graminearum*. *Nanomaterials* 10(2): 219. <https://doi.org/10.3390/nano10020219>

Ikram M, Javed B, Raja N I & Mashwani Z U R (2021). Biomedical potential of plant-based selenium nanoparticles: a comprehensive review on therapeutic and mechanistic aspects. *International Journal of Nanomedicine* 249-268. <https://doi.org/10.2147/IJN.S295053>

In G K & Nieva J (2015). Emerging chemotherapy agents in lung cancer: nanoparticles therapeutics for non-small cell lung cancer. *Translational Cancer Research* 4(4). <https://doi.org/10.3978/j.issn.2218-676X.2015.08.05>

Jach M E, Serefko A, Ziaja M & Kieliszek M (2022). Yeast protein as an easily accessible food source. *Metabolites* 12(1): 63. <https://doi.org/10.3390/metabo12010063>

Jacob J M, Ravindran R, Narayanan M, Samuel S M, Pugazhendhi A & Kumar G (2021). Microalgae: A prospective low cost green alternative for nanoparticle synthesis. *Current Opinion in Environmental Science & Health*, 20: 100163. <https://doi.org/10.1016/j.coesh.2019.12.005>

Jafari M, Rokhbakhsh-Zamin F, Shakibaie M, Moshafi M H, Ameri A, Rahimi H R & Forootanfar H (2018). Cytotoxic and antibacterial activities of biologically synthesized gold nanoparticles assisted by *Micrococcus yunnanensis* strain J2. *Biocatalysis and Agricultural Biotechnology* 15: 245-253. <https://doi.org/10.1016/j.bcab.2018.06.014>

Jalal M, Ansari M A, Alzohairy M A, Ali S G, Khan H M, Almatroudi A & Raees K (2018). Biosynthesis of silver nanoparticles from oropharyngeal *Candida glabrata* isolates and their antimicrobial activity against clinical strains of bacteria and fungi. *Nanomaterials* 8(8): 586. <https://doi.org/10.3390/nano8080586>

Jalal M, Ansari M A, Alzohairy M A, Ali S G, Khan H M, Almatroudi A & Siddiqui M I (2019). Anticandidal activity of biosynthesized silver nanoparticles: Effect on growth, cell morphology, and key virulence attributes of *Candida* species. *International Journal of Nanomedicine* 4667-4679. <https://doi.org/10.2147/IJN.S210449>

Jamkhande P G, Ghule N W, Bamer A H & Kalaskar M G (2019). Metal nanoparticles synthesis: An overview on methods of preparation, advantages and disadvantages, and applications. *Journal of Drug Delivery Science and Technology* 53: 101174. <https://doi.org/10.1016/j.jddst.2019.101174>

Jayaprakash N, Vijaya J J, Kaviyarasu K, Kombaiah K, Kennedy L J, Ramalingam R J, Munusamy M A & Al-Lohedan H A (2017). Green synthesis of Ag nanoparticles using Tamarind fruit extract for the antibacterial studies. *Journal of Photochemistry and Photobiology B: Biology* 169: 178-185. <https://doi.org/10.1016/j.jphotobiol.2017.03.013>

Jeevanandam J, Chan Y S & Danquah M K (2016). Biosynthesis of metal and metal oxide nanoparticles. *ChemBioEng Reviews* 3(2): 55-67. <https://doi.org/10.1002/cben.201500018>

Jeevanandam J, Pal K & Danquah M K (2019). Virus-like nanoparticles as a novel delivery tool in gene therapy. *Biochimie* 157: 38-47. <https://doi.org/10.1016/j.biochi.2018.11.001>

Jeevanandam J, Ling J K U, Barhoum A, San Chan Y & Danquah M K (2022). Bionanomaterials: Definitions, sources, types, properties, toxicity, and regulations, in Barhoum A, Jeevanandam J, Danquah M K (Eds.), *Fundamentals of bionanomaterials*. Elsevier, Netherlands, pp. 1-29

Kalaiselvi A, Roopan S M, Madhumitha G, Ramalingam C & Elango G (2015). Synthesis and characterization of palladium nanoparticles using *Catharanthus roseus* leaf extract and its application in the photo-catalytic degradation. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* 135: 116-119. <https://doi.org/10.1016/j.saa.2014.07.010>

Kambale E K, Nkanga C I, Mutonkole B P I, Bapolisi A M, Tassa D O, Liesse J M I, Krause R W M & Memvanga P B (2020). Green synthesis of antimicrobial silver nanoparticles using aqueous leaf extracts from three Congolese plant species (*Brillantaisia patula*, *Crossopteryx febrifuga* and *Senna siamea*). *Heliyon* 6(8). <https://doi.org/10.1016/j.heliyon.2020.e04493>

Kang S, Pinault M, Pfefferle L D & Elimelech M (2007). Single-walled carbon nanotubes exhibit strong antimicrobial activity. *Langmuir* 23(17): 8670-8673. <https://doi.org/10.1021/la701067r>

Kanniah P, Chelliah P, Thangapandi J R, Gnanadhas G, Mahendran V & Robert M (2021). Green synthesis of antibacterial and cytotoxic silver nanoparticles by *Piper nigrum* seed extract and development of antibacterial silver based chitosan nanocomposite. *International Journal of Biological Macromolecules* 189: 18-33. <https://doi.org/10.1016/j.ijbiomac.2021.08.056>

Kaur P, Thakur R, Duhan J S & Chaudhury A (2018). Management of wilt disease of chickpea *in vivo* by silver nanoparticles biosynthesized by rhizospheric microflora of chickpea (*Cicer arietinum*). *Journal of Chemical Technology & Biotechnology* 93(11): 3233-3243. <https://doi.org/10.1002/jctb.5680>

Khatua A, Prasad A, Priyadarshini E, Patel A K, Naik A, Saravanan M, Barabadi H, Ghosh I, Paul B, Paulraj R & Meena R (2020). Emerging antineoplastic plant-based gold nanoparticle synthesis: a mechanistic exploration of their anticancer activity toward cervical cancer cells. *Journal of Cluster Science*, 31, 1329-1340. <https://doi.org/10.1007/s10876-019-01742-1>

Kim D Y, Kim M, Sung J S, Koduru J R, Nile S H, Syed A, Bahkali A H, Seth C S & Ghodake G S (2024). Extracellular synthesis of silver nanoparticle using yeast extracts: Antibacterial and seed priming applications. *Applied Microbiology and Biotechnology* 108(1): 150. <https://doi.org/10.1007/s00253-023-12920-7>

Kışla D, Gökmen G G, Evrendilek G A, Akan T, Vlčko T, Kulawik P, Jambrak A R, & Ozogul F (2023). Recent developments in antimicrobial surface coatings: Various deposition techniques with nanosized particles, their application and environmental concerns. *Trends in Food Science & Technology* 135: 144-172. <https://doi.org/10.1016/j.tifs.2023.03.019>

Koopi H & Buazar F (2018). A novel one-pot biosynthesis of pure alpha aluminum oxide nanoparticles using the macroalgae *Sargassum ilicifolium*: a green marine approach. *Ceramics International* 44(8): 8940-8945. <https://doi.org/10.1016/j.ceramint.2018.02.091>

Król A, Railean-Plugaru V, Pomastowski P, Złoch M & Buszewski B (2018). Mechanism study of intracellular zinc oxide nanocomposites formation. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 553: 349-358. <https://doi.org/10.1016/j.colsurfa.2018.05.069>

Kulandaivelu B & Gothandam K M (2016). Cytotoxic effect on cancerous cell lines by biologically synthesized silver nanoparticles. *Brazilian Archives of Biology and Technology* 59: e16150529. <https://doi.org/10.1590/1678-4324-2016150529>

Kulkarni D, Sherkar R, Shirasathe C, Sonwane R, Varpe N, Shelke S, More M P, Pardeshi S R, Dhaneshwar G, Junnuthula V & Dyawanapelly S (2023). Biofabrication of nanoparticles: sources, synthesis, and biomedical applications. *Frontiers in Bioengineering and Biotechnology*, 11, 1159193. <https://doi.org/10.3389/fbioe.2023.1159193>

Kumar S V & Rajeshkumar S (2018). Plant-based synthesis of nanoparticles and their impact. *Nanomaterials in Plants, algae, and Microorganisms* 1: 33-57. <https://doi.org/10.1016/B978-0-12-811487-2.00002-5>

Kundu D, Hazra C, Chatterjee A, Chaudhari A & Mishra S (2014). Extracellular biosynthesis of zinc oxide nanoparticles using *Rhodococcus pyridinivorans* NT2: multifunctional textile finishing, biosafety evaluation and in vitro drug delivery in colon carcinoma. *Journal of Photochemistry and Photobiology B: Biology* 140: 194-204. <https://doi.org/10.1016/j.jphotobiol.2014.08.001>

Lateef A, Adelere I A & Gueguim-Kana E B (2015a). *Bacillus safensis* LAU 13: a new source of keratinase and its multi-functional biocatalytic applications. *Biotechnology & Biotechnological Equipment* 29(1): 54-63. <https://doi.org/10.1080/13102818.2014.986360>

Lateef A, Adelere I A, Gueguim-Kana E B, Asafa T B & Beukes L S (2015b). Green synthesis of silver nanoparticles using keratinase obtained from a strain of *Bacillus safensis* LAU 13. *International Nano Letters* 5: 29-35. <https://doi.org/10.1007/s40089-014-0133-4>

Lee B & Lee D G (2019). Synergistic antibacterial activity of gold nanoparticles caused by apoptosis-like death. *Journal of Applied Microbiology* 127(3): 701-712. <https://doi.org/10.1111/jam.14357>

Lei X, Peng Y, Li Y, Chen Q, Shen Z, Yin W, Lemiasheuski V, Xu S & He J (2024). Effects of selenium nanoparticles produced by *Lactobacillus acidophilus* HN23 on lipid deposition in WRL68 cells. *Bioorganic Chemistry* 107165. <https://doi.org/10.1016/j.bioorg.2024.107165>

Liu R & Lal R (2015). Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions. *Science of the Total Environment* 514: 131-139. <https://doi.org/10.1016/j.scitotenv.2015.01.104>

Liu X Y, Wang J Q, Ashby Jr C R, Zeng L, Fan Y F & Chen Z S (2021). Gold nanoparticles: Synthesis, physiochemical properties and therapeutic applications in cancer. *Drug Discovery Today* 26(5): 1284-1292. <https://doi.org/10.1016/j.drudis.2021.01.030>

Liu Y, Perumalsamy H, Kang C H, Kim S H, Hwang J S, Koh S C, Yi T H & Kim Y J (2020). Intracellular synthesis of gold nanoparticles by *Gluconacetobacter liquefaciens* for delivery of peptide CopA3 and ginsenoside and anti-inflammatory effect on lipopolysaccharide-activated macrophages. *Artificial Cells, Nanomedicine, and Biotechnology* 48(1), 777-788. <https://doi.org/10.1080/21691401.2020.1748639>

Ma D, Han T, Karimian M, Abbasi N, Ghaneialvar H & Zangeneh A (2020). Immobilized Ag NPs on chitosan-biguanidine coated magnetic nanoparticles for synthesis of propargylamines and treatment of human lung cancer. *International Journal of Biological Macromolecules* 165: 767-775. <https://doi.org/10.1016/j.ijbiomac.2020.09.193>

Majumder J, Pal K, Chakraborty W, Karmakar P & Gachhui R (2022). *Glucanoacetobacter kombuchae* (RG3T), a novel bacteria for AgNPs biosynthesis: Characterization and comprehensive evaluation of bioactivity. *Materials Today Communications* 33: 104410. <https://doi.org/10.1016/j.mtcomm.2022.104410>

Majumder R, Das C K & Mandal M (2019). Lead bioactive compounds of *Aloe vera* as potential anticancer agent. *Pharmacological Research* 148: 104416. <https://doi.org/10.1016/j.phrs.2019.104416>

Mariotti N, Bonomo M, Fagiolari L, Barbero N, Gerbaldi C, Bella F & Barolo C (2020). Recent advances in eco-friendly and cost-effective materials towards sustainable dye-sensitized solar cells. *Green Chemistry* 22(21): 7168-7218. <https://doi.org/10.1039/D0GC01148G>

Markus J, Mathiyalagan R, Kim Y J, Abbai R, Singh P, Ahn S, Perez Z E J, Hurt J & Yang D C (2016). Intracellular synthesis of gold nanoparticles with antioxidant activity by probiotic *Lactobacillus kimchicus* DCY51T isolated from Korean kimchi. *Enzyme and Microbial Technology* 95: 85-93. <https://doi.org/10.1016/j.enzmictec.2016.08.018>

Martínez G, Merinero M, Pérez-Aranda M, Pérez-Soriano E M, Ortiz T, Villamor E, Begines B & Alcudia A (2021). Environmental impact of nanoparticles' application as an emerging technology: A review. *Materials* 14(1): 166. <https://doi.org/10.3390/ma14010166>

Masum M M I, Siddiqa M M, Ali K A, Zhang Y, Abdallah Y, Ibrahim E, Qiu W, Yan C & Li B (2019). Biogenic synthesis of silver nanoparticles using *Phyllanthus emblica* fruit extract and its inhibitory action against the pathogen *Acidovorax oryzae* strain RS-2 of rice bacterial brown stripe. *Frontiers in Microbiology* 10: 820. <https://doi.org/10.3389/fmicb.2019.00820>

Matei A, Matei S, Matei G M, Cogălniceanu G & Cornea C P (2020). Biosynthesis of silver nanoparticles mediated by culture filtrate of lactic acid bacteria, characterization and antifungal activity. *The EuroBiotech Journal* 4(2): 97-103. <https://doi.org/10.2478/ebtj-2020-0011>

Mishra M, Paliwal J S, Singh S K, Selvarajan E, Subathradevi C & Mohanasrinivasan V (2013). Studies on the inhibitory activity of biologically synthesized and characterized zinc oxide nanoparticles using *Lactobacillus sporogens* against *Staphylococcus aureus*. *Journal of Pure Applied Microbiology* 7(2): 1263-1268

Mitchell M J, Billingsley M M, Haley R M, Wechsler M E, Peppas N A & Langer R (2021). Engineering precision nanoparticles for drug delivery. *Nature Reviews Drug Discovery* 20(2): 101-124. <https://doi.org/10.1038/s41573-020-0090-8>

Mittal A K, Chisti Y & Banerjee U C (2013). Synthesis of metallic nanoparticles using plant extracts. *Biotechnology Advances* 31(2): 346-356. <https://doi.org/10.1016/j.biotechadv.2013.01.003>

Mittal D, Kaur G, Singh P, Yadav K & Ali S A (2020). Nanoparticle-based sustainable agriculture and food science: Recent advances and future outlook. *Frontiers in Nanotechnology* 2: 579954. <https://doi.org/10.3389/fnano.2020.579954>

Moghaddam A B, Namvar F, Moniri M, Md Tahir P, Azizi S & Mohamad R (2015). Nanoparticles biosynthesized by fungi and yeast: a review of their preparation, properties, and medical applications. *Molecules* 20(9): 16540-16565. <https://doi.org/10.3390/molecules200916540>

Mohammad Z H, Ahmad F, Ibrahim S A & Zaidi S (2022). Application of nanotechnology in different aspects of the food industry. *Discover Food* 2(1): 12. <https://doi.org/10.1007/s44187-022-00013-9>

Mohammed A A, Hegazy A E & Salah A (2023). Novelty of synergistic and cytotoxicity activities of silver nanoparticles produced by *Lactobacillus acidophilus*. *Applied Nanoscience* 13(1): 633-640. <https://doi.org/10.1007/s13204-021-01878-5>

Mohd Yusof H, Mohamad R, Zaidan U H & Abdul Rahman N A (2019). Microbial synthesis of zinc oxide nanoparticles and their potential application as an antimicrobial agent and a feed supplement in animal industry: a review. *Journal of Animal Science and Biotechnology* 10: 1-22. <https://doi.org/10.1186/s40104-019-0368-z>

Molnár Z, Bódai V, Szakacs G, Erdélyi B, Fogarassy Z, Sáfrán G, Varga T, Konya Z, Toth-Szeles E, Szucs R & Lagzi I (2018). Green synthesis of gold nanoparticles by thermophilic filamentous fungi. *Scientific Reports* 8(1): 3943. <https://doi.org/10.1038/s41598-018-22112-3>

Morris M A, Padmanabhan S C, Cruz-Romero M C, Cummins E & Kerry J P (2017). Development of active, nanoparticle, antimicrobial technologies for muscle-based packaging applications. *Meat Science* 132: 163-178. <https://doi.org/10.1016/j.meatsci.2017.04.234>

Mortezaee K, Narmani A, Salehi M, Bagheri H, Farhood B, Hagh-Aminjan H & Najafi M (2021). Synergic effects of nanoparticles-mediated hyperthermia in radiotherapy/chemotherapy of cancer. *Life Sciences* 269: 119020. <https://doi.org/10.1016/j.lfs.2021.119020>

Munawer U, Raghavendra V B, Ningaraju S, Krishna K L, Ghosh A R, Melappa G & Pugazhendhi A (2020). Biofabrication of gold nanoparticles mediated by the endophytic *Cladosporium* species: Photodegradation, *in vitro* anticancer activity and *in vivo* antitumor studies. *International Journal of Pharmaceutics* 588: 119729. <https://doi.org/10.1016/j.ijpharm.2020.119729>

Nadagouda M N & Varma R S (2008). Green synthesis of silver and palladium nanoparticles at room temperature using coffee and tea extract. *Green Chemistry* 10(8): 859-862. <https://doi.org/10.1039/B804703K>

Naimi-Shamel N, Pourali P & Dolatabadi S (2019). Green synthesis of gold nanoparticles using *Fusarium oxysporum* and antibacterial activity of its tetracycline conjugant. *Journal de Mycologie Medicale* 29(1): 7-13. <https://doi.org/10.1016/j.mycmed.2019.01.005>

Nair B & Pradeep T (2002). Coalescence of nanoclusters and formation of submicron crystallites assisted by *Lactobacillus* strains. *Crystal Growth & Design* 2(4): 293-298. <https://doi.org/10.1021/cg0255164>

Nargund V B, Patil R R & Vanti G L (2022). *Bacillus* sp. extract used to fabricate ZnO nanoparticles for their antagonist effect against phytopathogens. *Biometals* 35(6): 1255-1269. <https://doi.org/10.1007/s10534-022-00440-2>

Nethravathi P C, Shruthi G S, Suresh D, Nagabhushana H & Sharma S C (2015). *Garcinia xanthochymus* mediated green synthesis of ZnO nanoparticles: photoluminescence, photocatalytic and antioxidant activity studies. *Ceramics International* 41(7): 8680-8687. <https://doi.org/10.1016/j.ceramint.2015.03.084>

Nile S H, Baskar V, Selvaraj D, Nile A, Xiao J & Kai G (2020). Nanotechnologies in food science: applications, recent trends, and future perspectives. *Nano-micro Letters* 12: 1-34. <https://doi.org/10.1007/s40820-020-0383-9>

Nooraei S, Bahrulolum H, Hoseini Z S, Katalani C, Hajizade A, Easton A J & Ahmadian G (2021). Virus-like particles: preparation, immunogenicity and their roles as nanovaccines and drug nanocarriers. *Journal of Nanobiotechnology*, 19, 1-27. <https://doi.org/10.1186/s12951-021-00806-7>

Oladipo I C, Lateef A, Azeez M A, Asafa T B, Yeeken T A, Ogunsona S B, Irshad H M & Abbas S H (2020). Characterization and biomedical application of phytosynthesized gold nanoparticles from *Datura stramonium* seed extract. *IOP Conference Series: Materials Science and Engineering*, IOP Publishing 805(1): 012021

Oroojalian F, Beygi M, Baradaran B, Mokhtarzadeh A & Shahbazi M A (2021). Immune cell Membrane-Coated biomimetic nanoparticles for targeted cancer therapy. *Small* 17(12): 2006484. <https://doi.org/10.1002/smll.202006484>

Ortega F G, Fernández-Baldo M A, Fernandez J G, Serrano M J, Sanz M I, Diaz-Mochon J J, Lorente J A & Raba J (2015). Study of antitumor activity in breast cell lines using silver nanoparticles produced by yeast. *International Journal of Nanomedicine* 2021-2031. <https://doi.org/10.2147/IJN.S75835>

Osman A I, Zhang Y, Farghali M, Rashwan A K, Eltaweil A S, Abd El-Monaem E M, Mohamed I M A, Badr M M, Ihara I, Rooney D W & Yap P S (2024). Synthesis of green nanoparticles for energy, biomedical, environmental, agricultural, and food applications: A review. *Environmental Chemistry Letters* 22(2): 841-887. <https://doi.org/10.1007/s10311-023-01682-3>

Pacheco-Blandino I, Vanner R & Buzea C (2012). Toxicity of nanoparticles, in F. Pacheco-Torgal F, Jalali S, Fucic A (Eds.), *Toxicity of Building Materials*. Woodhead Publishing, Sawston, United Kingdom, pp. 427-475

Pei X, Zhu Z, Gan Z, Chen J, Zhang X, Cheng X & Wang J (2020). PEGylated nano-graphene oxide as a nanocarrier for delivering mixed anticancer drugs to improve anticancer activity. *Scientific Reports* 10(1): 2717. <https://doi.org/10.1038/s41598-020-59624-w>

Peiris M M K, Guansekera T D C P, Jayaweera P M & Fernando S S N (2018). TiO₂ nanoparticles from Baker's yeast: a potent antimicrobial. *Journal of Microbiology and Biotechnology* 28(10): 1664-1670. <https://doi.org/10.4014/jmb.1807.07005>

Poopathi S, De Britto L J, Praba V L, Mani C & Praveen M (2015). Synthesis of silver nanoparticles from *Azadirachta indica*—a most effective method for mosquito control. *Environmental Science and Pollution Research* 22: 2956-2963. <https://doi.org/10.1007/s11356-014-3560-x>

Pradeepa Vidya S M, Mutalik S, Bhat K U, Huilgol P & Avadhani K (2016). Preparation of gold nanoparticles by novel bacterial exopolysaccharide for antibiotic delivery. *Life Sciences* 153, 171-179. <https://doi.org/10.1016/j.lfs.2016.04.022>

Prasad R, Bhattacharyya A & Nguyen Q D (2017). Nanotechnology in sustainable agriculture: recent developments, challenges, and perspectives. *Frontiers in Microbiology* 8: 1014. <https://doi.org/10.3389/fmicb.2017.01014>

Prema P, Ranjani S S, Kumar K R, Veeramanikandan V, Mathiyazhagan N, Nguyen V H & Balaji P (2022). Microbial synthesis of silver nanoparticles using *Lactobacillus plantarum* for antioxidant, antibacterial activities. *Inorganic Chemistry Communications* 136: 109139. <https://doi.org/10.1016/j.inoche.2021.109139>

Priya A S, Afzal O, Khalid M, Ahmad M F, Upadhyay A, Kumar S, Garg A, Ramzan M, Hussain A, Altamimi M A, Altamimi A S A, Webster T J & Khanam A (2023). Biogenic nanoparticles from waste fruit peels: Synthesis, applications, challenges and future perspectives. *International Journal of Pharmaceutics* 643: 123223. <https://doi.org/10.1016/j.ijpharm.2023.123223>

Pugazhendhi A, Prabakar D, Jacob J M, Karuppusamy I & Saratale R G (2018). Synthesis and characterization of silver nanoparticles using *Gelidium amansii* and its antimicrobial property against various pathogenic bacteria. *Microbial Pathogenesis* 114: 41-45. <https://doi.org/10.1016/j.micpath.2017.11.013>

Purbani D C, Pratiwi R D, Muttaqien S E, Nantapong N, Warsito M F, Fikri M N, Ruth F, Gustini N, Syahputra G, Padri M, Noerdjito D R, Nurkanto A & Afani H (2024). Eco-friendly synthesis of gold nanoparticles by marine microalgae *Synechococcus moorigangae*: Characterization, antimicrobial, and antioxidant properties. *Kuwait Journal of Science* 51(2): 100194. <https://doi.org/10.1016/j.kjs.2024.100194>

Puri A & Patil S (2022). *Tinospora cordifolia* stem extract-mediated green synthesis of selenium nanoparticles and its biological applications. *Pharmacognosy Research* 14(3). <https://doi.org/10.5530/pj.2019.11.x>

Qi X, Su T, Zhang M, Tong X, Pan W, Zeng Q & Shen J (2020). Sustainable, flexible and biocompatible hydrogels derived from microbial polysaccharides with tailorable structures for tissue engineering. *Carbohydrate Polymers* 237: 116160. <https://doi.org/10.1016/j.carbpol.2020.116160>

Qiu W, Chen R, Chen X, Zhang H, Song L, Cui W, Zhang J, Ye D, Zhang Y & Wang Z (2018). Oridonin-loaded and GPC1-targeted gold nanoparticles for multimodal imaging and therapy in pancreatic cancer. *International Journal of Nanomedicine* 6809-6827. <https://doi.org/10.2147/IJN.S177993>

Qureshi A, Singh D K & Dwivedi S (2018). Nano-fertilizers: a novel way for enhancing nutrient use efficiency and crop productivity. *International Journal of Current Microbiology and Applied Sciences* 7(2): 3325-3335. <https://doi.org/10.20546/ijcmas.2018.702.398>

Rad M, Taran M & Alavi M (2018). Effect of incubation time, CuSO₄ and glucose concentrations on biosynthesis of copper oxide (CuO) nanoparticles with rectangular shape and antibacterial activity: Taguchi method approach. *Nano Biomedicine and Engineering* 10(1): 25-33. <https://doi.org/10.5101/nbe.v10i1.p25-33>

Rajeshkumar S & Sivapriya D (2020). Fungus-mediated nanoparticles: characterization and biomedical advances, in Shukla A, (Ed.), *Nanoparticles in Medicine*. Springer, Singapore, pp. 185-199

Ramakrishna M, Rajesh Babu D, Gengan R M, Chandra S & Nageswara Rao G (2016). Green synthesis of gold nanoparticles using marine algae and evaluation of their catalytic activity. *Journal of Nanostructure in Chemistry* 6: 1-13. <https://doi.org/10.1007/s40097-015-0173-y>

Ravichandran A, Subramanian P, Manoharan V, Muthu T, Periyannan R, Thangapandi M, Ponnuchamy K, Pandi B & Marimuthu P N (2018). Phyto-mediated synthesis of silver nanoparticles using fucoidan isolated from *Spatoglossum asperum* and assessment of antibacterial activities. *Journal of Photochemistry and Photobiology B: Biology* 185: 117-125. <https://doi.org/10.1016/j.jphotobiol.2018.05.031>

Razak A, Mohd Gazzali S A, Fisol A, Abdulbaqi F A M, Parumasivam I, Mohtar T & Wahab H A (2021). Advances in nanocarriers for effective delivery of docetaxel in the treatment of lung cancer: an overview. *Cancers* 13(3): 400. <https://doi.org/10.3390/cancers13030400>

Rhim J W, Park H M & Ha C S (2013). Bio-nanocomposites for food packaging applications. *Progress in Polymer Science* 38(10-11): 1629-1652. <https://doi.org/10.1016/j.progpolymsci.2013.05.008>

Saeed S, Iqbal A & Ashraf M A (2020). Bacterial-mediated synthesis of silver nanoparticles and their significant effect against pathogens. *Environmental Science and Pollution Research* 27(30): 37347-37356. <https://doi.org/10.1007/s11356-020-07610-0>

Saifuddin N, Wong C W & Yasumira A N (2009). Rapid biosynthesis of silver nanoparticles using culture supernatant of bacteria with microwave irradiation. *Journal of Chemistry* 6(1): 61-70. <https://doi.org/10.1155/2009/734264>

Saka A, Jule L T, Gudata L, Gindaba A, Abdisa S S, Nagaprasad N & Ramaswamy K (2022). Green synthesis of *Datura stramonium* (Asaangira) leaves infusion for antibacterial activity through magnesium oxide (MgO) nanoparticles. *Advances in Materials Science and Engineering* 2022(1): 7134991. <https://doi.org/10.1155/2022/7134991>

Salunke B K, Sawant S S, Lee S I & Kim B S (2016). Microorganisms as efficient biosystem for the synthesis of metal nanoparticles: current scenario and future possibilities. *World Journal of Microbiology and Biotechnology* 32: 1-16. <https://doi.org/10.1007/s11274-016-2044-1>

Sanaeimehr Z, Javadi I & Namvar F (2018). Antiangiogenic and antiapoptotic effects of green-synthesized zinc oxide nanoparticles using *Sargassum muticum* algae extraction. *Cancer Nanotechnology* 9: 1-16. <https://doi.org/10.1186/s12645-018-0037-5>

Sásková D, Pořízka J, Kulich P, Španová A, Diviš P & Rittich B (2016). Silver nanoparticles production with probiotic bacteria. *Materials Science Forum* 851: 32-36

Sayadi M H, Salmani N, Heidari A & Rezaei M R (2018). Bio-synthesis of palladium nanoparticle using *Spirulina platensis* alga extract and its application as adsorbent. *Surfaces and Interfaces* 10: 136-143. <https://doi.org/10.1016/j.surfin.2018.01.002>

Sekhon B S (2010). Food nanotechnology—an overview. *Nanotechnology, Science and Applications* 1-15. <https://doi.org/10.2147/nsa.s12187498>

Selmani A, Ulm L, Kasemets K, Kurvet I, Erceg I, Barbir R, Pem B, Santini P, Marion I D, Včnkošek T, Krivohlavek A, Sikiric M D, Kahru A & Vrček I V (2020). Stability and toxicity of differently coated selenium nanoparticles under model environmental exposure settings. *Chemosphere* 250: 126265. <https://doi.org/10.1016/j.chemosphere.2020.126265>

Selvarajan E & Mohanasrinivasan V J M L (2013). Biosynthesis and characterization of ZnO nanoparticles using *Lactobacillus plantarum* VITES07. *Materials Letters* 112: 180-182. <https://doi.org/10.1016/j.matlet.2013.09.020>

Shah M, Fawcett D, Sharma S, Tripathy S K & Poinern G E J (2015). Green synthesis of metallic nanoparticles via biological entities. *Materials* 8(11): 7278-7308. <https://doi.org/10.3390/ma8115377>

Shaikh S, Nazam N, Rizvi S M D, Ahmad K, Baig M H, Lee E J & Choi I (2019). Mechanistic insights into the antimicrobial actions of metallic nanoparticles and their implications for multidrug resistance. *International Journal of Molecular Sciences* 20(10): 2468. <https://doi.org/10.3390/ijms20102468>

Shamaila S, Sajjad A K L, Farooqi S A, Jabeen N, Majeed S & Farooq I (2016). Advancements in nanoparticle fabrication by hazard free eco-friendly green routes. *Applied Materials Today* 5: 150-199. <https://doi.org/10.1016/j.apmt.2016.09.009>

Shanmugam R, Munusamy T, Jayakodi S, Al-Ghanim K A, Nicoletti M, Sachivkina N & Govindarajan M (2023). Probiotic-bacteria (*Lactobacillus fermentum*)-wrapped zinc oxide nanoparticles: Biosynthesis, characterization, and antibacterial activity. *Fermentation* 9(5): 413. <https://doi.org/10.3390/fermentation9050413>

Shehabeldine A M, Elbahnaawy M A & Hasaballah A I (2021). Green phytosynthesis of silver nanoparticles using *Echinochloa stagnina* extract with reference to their antibacterial, cytotoxic, and larvicidal activities. *BioNanoScience* 11: 526-538. <https://doi.org/10.1007/s12668-021-00846-1>

Shende S, Ingle A P, Gade A & Rai M (2015). Green synthesis of copper nanoparticles by *Citrus medica* Linn.(Idilimbu) juice and its antimicrobial activity. *World Journal of Microbiology and Biotechnology* 31: 865-873. <https://doi.org/10.1007/s11274-015-1840-3>

Siaw Y M, Jeevanandam J, Hii Y S & Chan Y S (2020). Photo-irradiation coupled biosynthesis of magnesium oxide nanoparticles for antibacterial application. *Naunyn-Schmiedeberg's Archives of Pharmacology* 393(12): 2253-2264. <https://doi.org/10.1007/s00210-020-01934-x>

Sierra-Fernandez A, De la Rosa-García S C, Gomez-Villalba L S, Gómez-Cornelio S, Rabanal M E, Fort R & Quintana P (2017). Synthesis, photocatalytic, and antifungal properties of MgO, ZnO and Zn/Mg oxide nanoparticles for the protection of calcareous stone heritage. *ACS Applied Materials & Interfaces* 9(29): 24873-24886. <https://doi.org/10.1021/acsami.7b06130>

Singh K R, Nayak V, Singh J, Singh A K & Singh R P (2021). Potentialities of bioinspired metal and metal oxide nanoparticles in biomedical sciences. *RSC Advances* 11(40): 24722-24746. <https://doi.org/10.1039/DRA04273D>

Singh P, Kim Y J, Wang C, Mathiyalagan R & Yang D C (2016a). *Weissella oryzae* DC6-facilitated green synthesis of silver nanoparticles and their antimicrobial potential. *Artificial Cells, Nanomedicine, and Biotechnology* 44(6): 1569-1575. <https://doi.org/10.3109/21691401.2015.1064937>

Singh P, Kim Y J, Zhang D & Yang D C (2016b). Biological synthesis of nanoparticles from plants and microorganisms. *Trends in Biotechnology*, 34(7): 588-599. <https://doi.org/10.1016/j.tibtech.2016.02.006>

Singh S, Singh B K, Yadav S M & Gupta A K (2015). Applications of nanotechnology in agricultural and their role in disease management. *Research Journal of Nanoscience and Nanotechnology* 5(1), 1-5.

Sinha A & Khare S K (2011). Mercury bioaccumulation and simultaneous nanoparticle synthesis by *Enterobacter* sp. cells. *Bioresource Technology* 102(5): 4281-4284. <https://doi.org/10.1016/j.biortech.2010.12.040>

Siqueira M C, Coelho G F, de Moura M R, Bresolin J D, Hubinger S Z, Marconcini J M & Mattoso L H (2014). Evaluation of antimicrobial activity of silver nanoparticles for carboxymethylcellulose film applications in food packaging. *Journal of Nanoscience and Nanotechnology* 14(7): 5512-5517. <https://doi.org/10.1166/jnn.2014.8991>

Sivarethnamohan R & Sujatha S (2021). Unlocking the potentials of using nanotechnology to stabilize agriculture and food production. *Third International Conference on Material Science, Smart Structures and Applications: (ICMSS 2020)*, AIP Publishing 2327: 1

Skalickova S, Baron M & Sochor J (2017). Nanoparticles biosynthesized by yeast: a review of their application. *Kvasny Prumysl* 63(6): 290-292. <https://doi.org/10.18832/kp201727>

Slavin Y N, Asnis J, Híñfeli U O & Bach H (2017). Metal nanoparticles: understanding the mechanisms behind antibacterial activity. *Journal of Nanobiotechnology* 15: 1-20. <https://doi.org/10.1186/s12951-017-0308-z>

Soni N & Prakash S (2015). Antimicrobial and mosquitocidal activity of microbial synthesized silver nanoparticles. *Parasitology Research* 114: 1023-1030. <https://doi.org/10.1007/s00436-014-4268-z>

Soto C M & Ratna B R (2010). Virus hybrids as nanomaterials for biotechnology. *Current Opinion in Biotechnology* 21(4): 426-438. <https://doi.org/10.1016/j.copbio.2010.07.004>

Sowbarnika R, Anhuradha S & Preetha B (2018). Enhanced antimicrobial effect of yeast mediated silver nanoparticles synthesized from baker's yeast. *International Journal of Nanoscience and Nanotechnology* 14(1): 33-42

Sperber W H (2009). Introduction to the microbiological spoilage of foods and beverages, in Sperber W, Doyle M (Eds.), Compendium of the Microbiological Spoilage of Foods and Beverages. Food Microbiology and Food Safety. Springer, German, pp. 1-40

Spiridopoulou K, Tryfonopoulou E, Aindelis G, Ypsilantis P, Sarafidis C, Kalogirou O & Chlichlia K (2021). Biogenic selenium nanoparticles produced by *Lactobacillus casei* ATCC 393 inhibit colon cancer cell growth *in vitro* and *in vivo*. *Nanoscale Advances* 3(9): 2516-2528. <https://doi.org/10.1039/D0NA00984A>

Srikhao N, Kasemsiri P, Lorwanishpaisarn N & Okhawilai M (2021). Green synthesis of silver nanoparticles using sugarcane leaves extract for colorimetric detection of ammonia and hydrogen peroxide. *Research on Chemical Intermediates* 47: 1269-1283. <https://doi.org/10.1007/s11164-020-04354-x>

Sriramulu M & Sumathi S (2018). Biosynthesis of palladium nanoparticles using *Saccharomyces cerevisiae* extract and its photocatalytic degradation behaviour. *Advances in Natural Sciences: Nanoscience and Nanotechnology* 9(2): 025018. <https://doi.org/10.1088/2043-6254/aac506>

Srivastava P & Kowshik M (2017). Fluorescent lead (IV) sulfide nanoparticles synthesized by *Idiomarina* sp. strain PR58-8 for bioimaging applications. *Applied and Environmental Microbiology* 83(7): e03091-16. <https://doi.org/10.1128/AEM.03091-16>

Suba S, Vijayakumar S, Vidhya E, Punitha V N & Nilavukkarasi M (2021). Microbial mediated synthesis of ZnO nanoparticles derived from *Lactobacillus* spp: Characterizations, antimicrobial and biocompatibility efficiencies. *Sensors International* 2: 100104. <https://doi.org/10.1016/j.sintl.2021.100104>

Subramaniyan S A, Sheet S, Vinothkannan M, Yoo D J, Lee Y S, Belal S A & Shim K S (2018). One-pot facile synthesis of Pt nanoparticles using cultural filtrate of microgravity simulated grown *P. chrysogenum* and their activity on bacteria and cancer cells. *Journal of Nanoscience and Nanotechnology* 18(5): 3110-3125. <https://doi.org/10.1166/jnn.2018.14661>

Suvandee W, Teeranachaideekul V, Jeenduang N, Nooeaid P, Makarases A, Chuenchom L & Dechtrirat D (2022). One-pot and green preparation of *Phyllanthus emblica* extract/silver nanoparticles/polyvinylpyrrolidone spray-on dressing. *Polymers* 14(11): 2205. <https://doi.org/10.3390/polym14112205>

Taha R H (2022). Green synthesis of silver and gold nanoparticles and their potential applications as therapeutics in cancer therapy; a review. *Inorganic Chemistry Communications* 143: 109610. <https://doi.org/10.1016/j.inoche.2022.109610>

Thanki K, Gangwal R P, Sangamwar A T & Jain S (2013). Oral delivery of anticancer drugs: challenges and opportunities. *Journal of Controlled Release* 170(1): 15-40. <https://doi.org/10.1016/j.jconrel.2013.04.020>

Thirumal V, Dhamodharan K, Yuvakkumar R, Ravi G, Saravanakumar B, Thambidurai M & Velauthapillai D (2021). Cleaner production of tamarind fruit Shell into bio-mass derived porous 3D-activated carbon nanosheets by CVD technique for supercapacitor applications. *Chemosphere* 282: 131033. <https://doi.org/10.1016/j.chemosphere.2021.131033>

Tinkle S, McNeil S E, Mühlebach S, Bawa R, Borchard G, Barenholz Y, Tamarkin L & Desai N (2014). Nanomedicines: addressing the scientific and regulatory gap. *Annals of the New York Academy of Sciences* 1313(1): 35-56. <https://doi.org/10.1111/nyas.12403>

Tomşa A M, Răchişan A L, Aldea A A & Ciumărnean L (2021). Perspectives of gold nanoparticles and their applications in pancreatic cancer. *Experimental and Therapeutic Medicine* 21(3): 1-1. <https://doi.org/10.3892/etm.2021.9689>

Torres L A Z, Woiciechowski A L, de Andrade Tanobe V O, Zandoná Filho A, de Freitas R A, Noseda M D, Szameitat E S, Faulds C, Coutinho P, Bertrand E & Soccol C R (2021). Lignin from oil palm empty fruit bunches: Characterization, biological activities and application in green synthesis of silver nanoparticles. *International Journal of Biological Macromolecules* 167: 1499-1507. <https://doi.org/10.1016/j.ijbiomac.2020.11.104>

Vaghela H, Shah R & Parmar K A (2018). Plant mixture mediated biogenic copper nanoparticles: antibacterial assay. *Current Nanomaterials* 3(2): 86-94. <https://doi.org/10.2174/2405461503666180803152152>

Vedernykova I O, Koval A A, Antonenko O V, Chan T M, Shpychak O S & Marchenko M V (2018). Synthesis, technology and analysis of nanoparticles of barium hexaferrite for creation of magnetically controlled drug delivery systems. *Journal of Pharmaceutical Science and Research* 10: 2122

Vijayabharathi R, Sathy A & Gopalakrishnan S (2018). Extracellular biosynthesis of silver nanoparticles using *Streptomyces griseoplanus* SAI-25 and its antifungal activity against *Macrophomina phaseolina*, the charcoal rot pathogen of sorghum. *Biocatalysis and Agricultural Biotechnology* 14: 166-171. <https://doi.org/10.1016/j.bcab.2018.03.006>

Vijayaraghavan K & Ashokkumar T (2017). Plant-mediated biosynthesis of metallic nanoparticles: A review of literature, factors affecting synthesis, characterization techniques and applications. *Journal of Environmental Chemical Engineering* 5(5): 4866-4883. <https://doi.org/10.1016/j.jece.2017.09.026>

Wadhwani S A, Shedbalkar U U, Singh R & Chopade B A (2018). Biosynthesis of gold and selenium nanoparticles by purified protein from *Acinetobacter* sp. SW 30. *Enzyme and Microbial Technology* 111: 81-86. <https://doi.org/10.1016/j.enzmictec.2017.10.007>

Wang C, Gong C, Qin Y, Hu Y, Jiao A, Jin Z, Qiu C & Wang J (2022). Bioactive and functional biodegradable packaging films reinforced with nanoparticles. *Journal of Food Engineering* 312: 110752. <https://doi.org/10.1016/j.jfoodeng.2021.110752>

Wang L, Hu C & Shao L (2017). The antimicrobial activity of nanoparticles: present situation and prospects for the future. *International Journal of Nanomedicine* 12: 1227-1249. <https://doi.org/10.2147/IJN.S121956>

Wang R, Xu X, Puja A M, Perumalsamy H, Balusamy S R, Kim H & Kim Y J (2021). Gold nanoparticles prepared with *Phyllanthus emblica* fruit extract and *Bifidobacterium animalis* subsp. *lactis* can induce apoptosis via mitochondrial impairment with inhibition of autophagy in the human gastric carcinoma cell line AGS. *Nanomaterials* 11(5): 1260. <https://doi.org/10.3390/nano11051260>

Wang Y, Xu J, Shi L & Yang H (2020). Recent advances in the antilung cancer activity of biosynthesized gold nanoparticles. *Journal of Cellular Physiology* 235(12): 8951-8957. <https://doi.org/10.1002/jcp.29789>

Weiss J, Takhistov P & McClements D J (2006). Functional materials in food nanotechnology. *Journal of Food Science* 71(9): 107-116. <https://doi.org/10.1111/j.1750-3841.2006.00195.x>

Wen A M & Steinmetz N F (2016). Design of virus-based nanomaterials for medicine, biotechnology, and energy. *Chemical Society Reviews* 45(15): 4074-4126. <https://doi.org/10.1039/C5CS00287G>

Wolfram J & Ferrari M (2019). Clinical cancer nanomedicine. *Nano today* 25: 85-98. <https://doi.org/10.1016/j.nantod.2019.02.005>

Xu J, Yıldıztekin M, Han D, Keskin C, Baran A, Baran M F, Eftekhari A, Ava C A, Kandemir S İ, Cebe D B, Dağ B, Beileri A & Khalilov R (2023). Biosynthesis, characterization, and investigation of antimicrobial and cytotoxic activities of silver nanoparticles using *Solanum tuberosum* peel aqueous extract. *Heliyon*, 9(8). <https://doi.org/10.1016/j.heliyon.2023.e19061>

Yana L, Yutong C & Qinghui H (2024). Antibacterial films based on polylactide and polybutylene adipate terephthalate loaded with zinc oxide or silver nanoparticles: Characterization and application in fresh noodles packaging. *Journal of Food Engineering* 367: 111889. <https://doi.org/10.1016/j.jfoodeng.2023.111889>

Yuan Y G, Peng Q L & Gurunathan S (2017). Effects of silver nanoparticles on multiple drug-resistant strains of *Staphylococcus aureus* and *Pseudomonas aeruginosa* from mastitis-infected goats: an alternative approach for antimicrobial therapy. *International Journal of Molecular Sciences* 18(3): 569. <https://doi.org/10.3390/ijms18030569>

Zaki S A, Ouf S A, Albarakaty F M, Habeb M M, Aly A A & Abd-Elsalam K A (2021). *Trichoderma harzianum*-mediated ZnO nanoparticles: A green tool for controlling soil-borne pathogens in cotton. *Journal of Fungi* 7(11), 952. <https://doi.org/10.3390/jof7110952>

Zang W, Cao H, Ge J & Zhao D (2024). Structures, physical properties and antibacterial activity of silver nanoparticles of *Lactiplantibacillus plantarum* exopolysaccharide. *International Journal of Biological Macromolecules* 263: 130083. <https://doi.org/10.1016/j.ijbiomac.2024.130083>

Zeng Q, Liu Z, Niu T, He C, Qu Y & Qian Z (2023). Application of nanotechnology in CAR-T-cell immunotherapy. *Chinese Chemical Letters*, 34(3), 107747. <https://doi.org/10.1016/j.cclet.2022.107747>

Zhang Y, Dong Y, Zhou J, Li X & Wang F (2018). Application of plant viruses as a biotemplate for nanomaterial fabrication. *Molecules* 23(9): 2311. <https://doi.org/10.3390/molecules23092311>

Zhang F, Han X, Hu Y, Wang S, Liu S, Pan X, Wang H, Ma J, Wang W, Li S, Wu Q, Shen H, Yu X, Yuan Q & Liu H (2019). Interventional photothermal therapy enhanced brachytherapy: A new strategy to fight deep pancreatic cancer. *Advanced Science* 6(5): 1801507. <https://doi.org/10.1002/advs.201801507>

Zhao X, Zhou L, Riaz Rajoka M S, Yan L, Jiang C, Shao D, Zhu J, Shi J, Huang Q, Yang H & Jin M (2018). Fungal silver nanoparticles: synthesis, application and challenges. *Critical Reviews in Biotechnology* 38(6): 817-835. <https://doi.org/10.1080/07388551.2017.1414141>

Zhong B, Xu W, Xie H & Wu Z (2024). Biosynthesis and characterization of selenium nanoparticles by Se-tolerant *Lactiplantibacillus plantarum*. *Food Bioscience* 59: 104061. <https://doi.org/10.1016/j.fbio.2024.104061>



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