



## *Echium italicum* Plant Extracts Have Wound Healing Potential in Human Dermal Fibroblast (HDF) Cell

Derya Akduman<sup>1</sup>, Rahmiye Akduman<sup>1</sup>, Merve Şimşek Geyik<sup>1</sup>, Mesut Akyüz<sup>1\*</sup>

<sup>1</sup>Department of Molecular Biology and Genetics, Erzurum Technical University, Erzurum, Turkey.

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### Abstract

Plants have an important place in the development of drugs used in the treatment of diseases, as in many areas. The flower and leaf parts of the *Echium italicum* plant used in the study have been used in many studies, but no information has been found in the literature about the use of the root part in increasing cell viability and wound healing treatment. In this study, the effects of *Echium italicum*, one of the *Echium* species naturally growing in our country, on cell viability and wound healing were investigated *in vitro*. Extracts prepared with different polarity solvents from the root and aerial parts of the plant were applied to the human dermal fibroblast (HDF) cell line to evaluate their effect on cell viability and wound healing potential. Cell viability was determined with the CVDK-8 method, and the wound healing activity of the plant was determined with the scratch assay. In summary, the effects of the *Echium italicum* root extract on wound healing were examined, and the medicinal properties of the plant suggested in traditional medicine were scientifically evaluated.

Keywords: *Echium italicum*, cell viability, wound healing, HDF cell

### Introduction

*Echium italicum* is a plant species belonging to the Boraginaceae family, native to the Mediterranean region. It grows naturally in the Aegean and Mediterranean regions of Turkey but can sometimes be found in other Mediterranean countries and Southern Europe. This plant grown in temperate climates, especially in calcareous soils and sunny areas. The bio

logical characteristics of *Echium italicum* include an annual or biennial life cycle. Its stem grows upright and usually reaches a height of 30 to 60 cm. Its leaves are arranged in rows, long and narrow, and have hairy edges. The flowers are collected in long panicles and are mostly blue, purple or pink. The flowering period usually coincides with the spring and summer months (1).

Research into the medicinal potential of the *Echium italicum* plant has focused particularly on the flower and leaf parts of the plant. These parts have been

\*Correspondence: Mesut Akyüz  
Department of Molecular Biology and Genetics, Erzurum Technical University, Erzurum, Turkey.  
E-mail: mesut.akyuz@erzurum.edu.tr, Tel: 444 5 388 (2198)



associated with antioxidant, anti-inflammatory and wound-healing properties. For example, extracts of the plant may be effective in combating free radicals due to their antioxidant capacity and may alleviate inflammatory conditions due to their anti-inflammatory properties. Additionally, many studies have been conducted on the positive effects of *Echium italicum* flower and leaf extracts on skin cells and their wound-healing potential (2).

Plant roots are a vital structure that anchors the plant to the soil and helps it absorb nutrients from the soil. Roots also enable the plant to take in and store water, helping the plant survive dry periods. Roots can also produce hormones that regulate plant growth and development (3). From a medicinal point of view, many plant roots have long been used in traditional medicine. Roots generally contain the richest bioactive components of the plant and due to these properties, they have various positive effects on health. Root extracts have biological effects such as antioxidant, anti-inflammatory, antibacterial and antiviral. These can contribute to the development of drugs used in the treatment of various diseases (4).

Plant extracts are concentrated extracts obtained from various parts of plants (e.g. leaves, flowers, roots) and contain many bioactive compounds. These bioactive compounds are generally known as phytochemicals and play a role in the defense mechanisms of plants, growth regulation, and protection against environmental stresses. Plant extracts have long been used in traditional medicine and are considered an important source in the drug development process in modern medicine (5). The cellular effects of plant extracts vary depending on the bioactive compounds they contain. Various bioactive compounds, such as antioxidants, anti-inflammatories, anticancer agents, and antimicrobial compounds, exert various effects at the cellular level. For example, antioxidants protect cells from the harmful effects of free radicals by reducing cellular oxidative stress. Anti-inflammatories,

on the other hand, alleviate inflammatory conditions by reducing inflammation. Anticancer agents can prevent the growth and spread of cancer cells, while antimicrobial compounds inhibit the growth of bacteria, viruses, and fungi (6).

The cellular effects and therapeutic potential of plant extracts have been the focus of many studies investigating their use in the pharmaceutical industry. Such studies help us understand how plant extracts act at the cellular level and identify potential therapeutic agents (7).

Human dermal fibroblast (HDF) cells are the most common cell type found in the skin and have a variety of functions. Fibroblasts play important roles in maintaining the structural integrity of skin tissue, promoting wound healing, and contributing to tissue regeneration. During the wound healing process, fibroblasts synthesize collagen and other extracellular matrix components that promote wound healing (8).

Wound healing is generally described as a three-phase process: inflammation, regeneration (proliferation), and remodeling. First, inflammation occurs at the wound site and clears damaged tissue and microorganisms. Second, during the regeneration phase, fibroblasts synthesize extracellular matrix components such as collagen and elastin in the wound site and stimulate tissue regeneration by forming new vessels. Finally, during the remodeling phase, the structural integrity of the tissue is restored, and excess matrix material is cleared (9).

The role of fibroblasts in wound healing is mediated by cell migration, proliferation, and production of extracellular matrix. These processes allow cells to come together to close the wound and repair damaged tissue. The effectiveness of fibroblasts in wound healing depends on several factors, including growth factors, cytokines, and extracellular matrix components (10).

Studies on HDF cells can contribute to our understanding of the molecular and cellular

mechanisms of wound healing and the development of new therapeutic methods targeting these processes. Therefore, studies on HDF cells and wound healing play an important role in clinical applications such as the treatment of skin diseases and wound healing (11). This study adopts a scientific approach to understand the potential health effects of *Echium italicum* root parts. The focus of the study was to evaluate the effects of *Echium italicum* root extracts on cell viability and wound healing *in vitro*.

## Material and Method

### Plant Collection and Storage Conditions:

*Echium italicum* plant was freshly collected from Mustafakemalpaşa district of Bursa province and brought to the laboratory via cargo. The drying process of the roots was carried out at low temperature and in a dark environment to preserve the biological activity of the plant. The solvents used in obtaining the plant extracts were stored in storage cabinets at room temperature (25°C).

**Preparation of Extracts:** Three different extracts were obtained from the dried roots using polarity solvents such as acetone, methanol and H<sub>2</sub>O<sub>2</sub>. For the extraction process, 20 grams of plant material was weighed using a precision balance and 100 ml of acetone, methanol and H<sub>2</sub>O<sub>2</sub> were added separately, mixed until a homogeneous mixture was obtained and filtered. The obtained extracts were concentrated using an evaporator at 55°C, 65°C, 95°C for acetone, methanol and H<sub>2</sub>O<sub>2</sub> respectively. The extracts were air dried and calculated in mg/g. They were stored at -20 °C for further studies (12).

**Concentration Calculation:** 20 mg of the extracts were taken from the sand and dissolved in 200 µl DMSO (Dimethyl sulfoxide) and subjected to sonification. After the concentrations determined from these extracts prepared as stocks (0,10 mg/mL, 0,25 mg/mL, 0,50 mg/mL, 1 mg/mL) were prepared, a

homogeneous structure was obtained by passing through a 0.45 mm filter.

**Cell Culture:** Human dermal fibroblast (HDF) cells were propagated in DMEM medium (10% FBS, 1X L-Glutamine and 1X Penicillin/Streptomycin) (EcoTech Biotechnology, Türkiye) and then seeded into T25 flasks with 5 ml medium and incubated in a 5% CO<sub>2</sub> oven at 37°C. Cells were dissociated using trypsin every 2-3 days and cultured for 4 weeks. All cell culture studies were performed in a sterile laminar air flow cabinet. Appropriate conditions were provided for cell proliferation and passaging was performed when the cell density reached 60-80% confluency.

**Cell Viability Assay:** HDF cells were counted under a microscope and distributed equally in a 96-well plate so that 1.5x10<sup>3</sup> cells would be seeded in each well, and then the plates were kept in the incubator for 24 hours. At the end of the incubation, plant extracts at different concentrations (0,10 mg/mL, 0,25 mg/mL, 0,50 mg/mL, 1 mg/mL) and negative (DMEM) and positive (10% DMSO) controls were applied to the cells prepared for the experiment and kept in the incubator for 24 hours. Three replicates were run for each experimental group to ensure repeatability. Cell viability was determined using the CVDK-8 kit (EcoTech Biotechnology, Türkiye). 50 µl (medium and 10% kit) of the CVDK-8 kit solution was applied to each well of the 96-well plate where the concentration application was made. Cells were incubated in the dark at 37°C. Measurements were taken every half hour on the Biotech Epoch device (13).

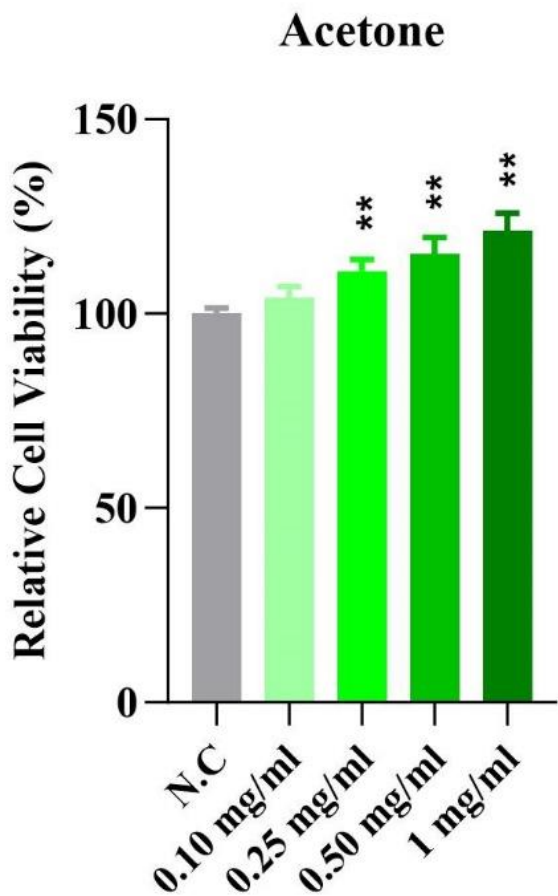
**Wound Healing Test (Scratch Assay):** To evaluate the effect of the plant extract on wound healing, HDF cells were counted and 75x10<sup>4</sup> cells were seeded into each well of a 6-well plate and kept in an incubator for 24 hours to ensure monolayer formation. Scratches were created on the cell surface attached to the ground using a 200 µl sterile pipette tip in a regular and standard manner. After 24 hours, wound closure

formations were observed and recorded with a Leica inverted microscope (14).

**Statistical Analysis:** The obtained results were visualized using GraphPad Prism 10 and ImageJ programs. Statistical analysis for all work packages was performed using two-sided Student’s t-test with log transformed data, and  $p < 0.05$  values were considered statistically significant.

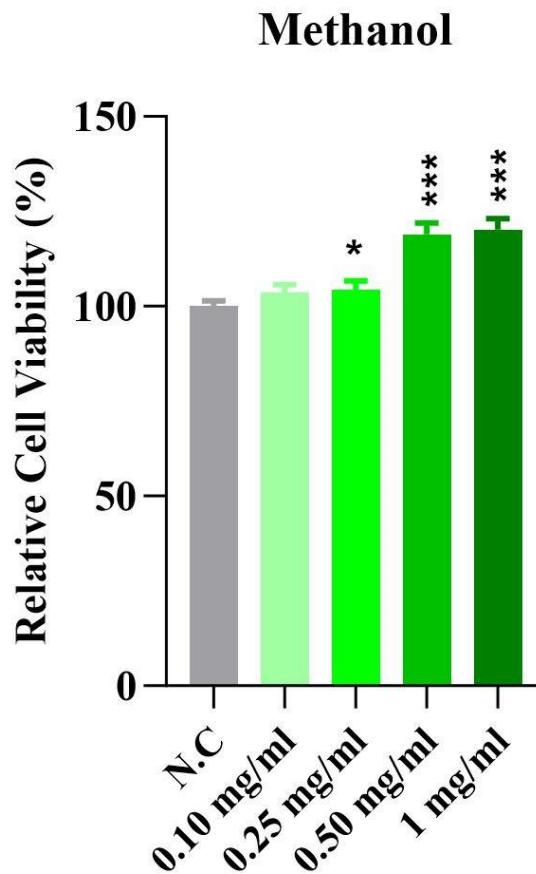
**Results**

**Cell Viability Result:** After incubating the acetone extract applied at various concentrations for 24 hours, the 0,10 mg/ml concentration of *Echium italicum* had no significant effect on cell viability, while other concentrations increased cell viability (Figure 1,  $p < 0.05$ ).



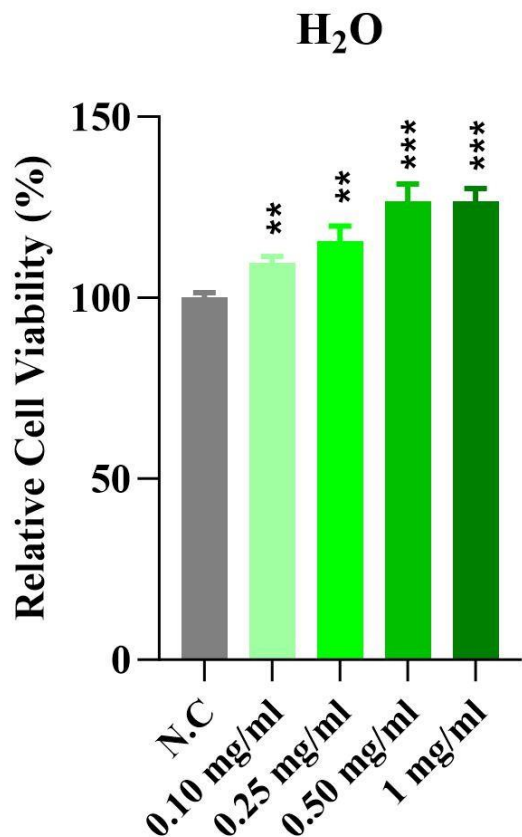
**Figure 1.** Effects of various concentrations of acetone extraction on HDF cells.

In the examination of the effects of the methanol extract of *Echium italicum* applied to HDF cells for 24 hours on cell viability, it was determined that the dose of 0,10 mg/ml did not have a significant effect, while other doses significantly increased cell viability (Figure 2,  $p < 0.05$ ).

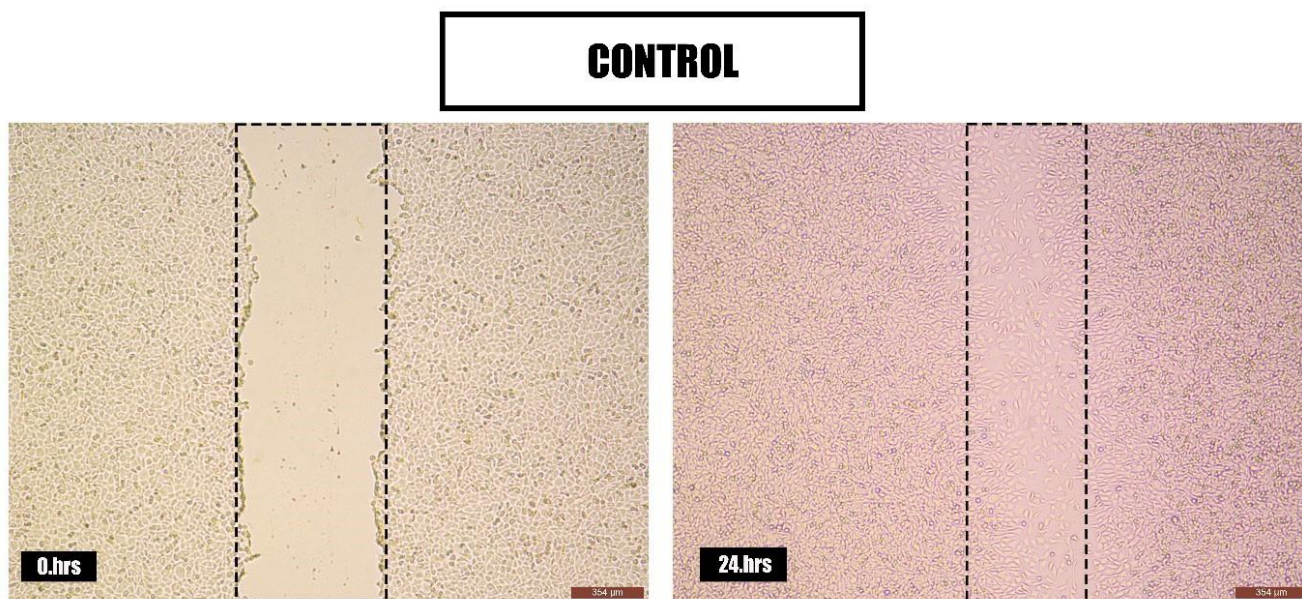


**Figure 2.** Effects of various concentrations of methanol extraction on HDF cells.

In the examination of the effects of 24-hour application of *Echium italicum* H<sub>2</sub>O extract on cell viability of HDF cells, all applied concentrations of *Echium italicum* significantly increased cell viability (Figure 3,  $p < 0.05$ ).



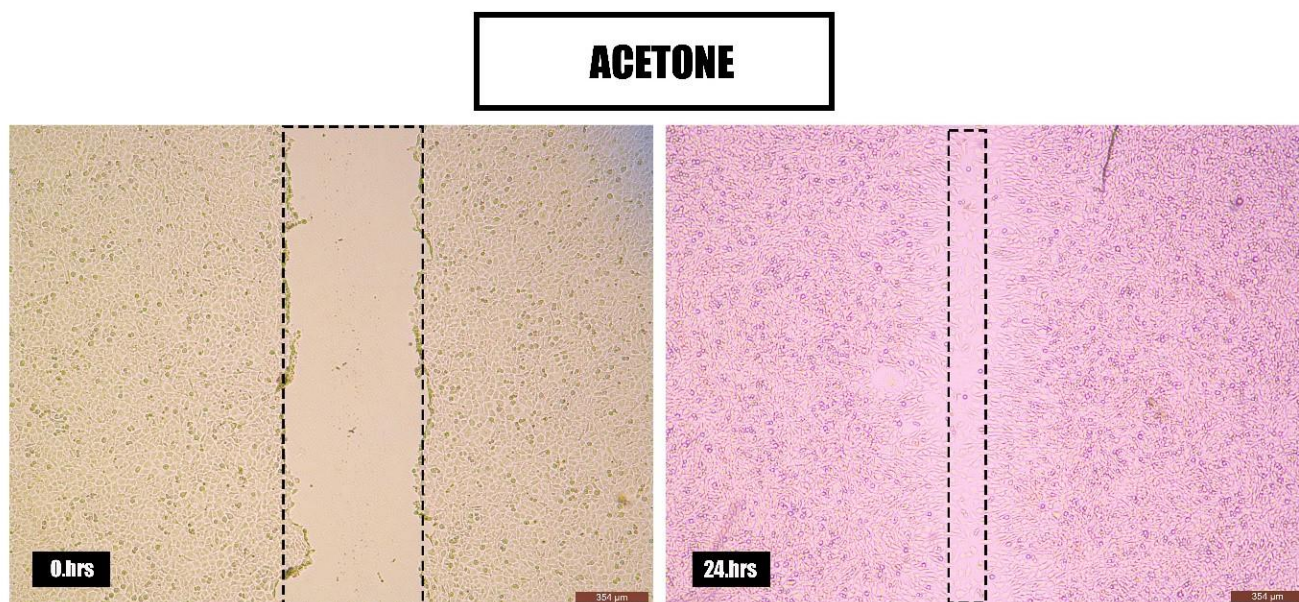
**Figure 3.** Effects of various concentrations of H<sub>2</sub>O extraction on HDF cells.



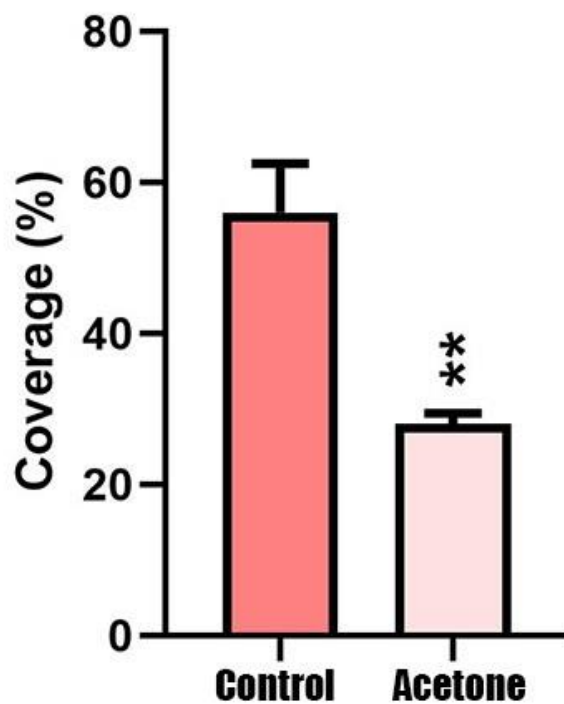
**Figure 4.** Control group 0-24th hour surface coverage image

**Wound Healing (Scratch Assay) Result:** The effects of the obtained plant extract on wound healing were evaluated with the wound healing test. The lowest concentration of acetone extract (0,10 mg/ml) was selected and given to the cells. At the end of 24 hours of incubation, the wound closure status was observed under a microscope and photographed.

In the analysis of cell wound area closure (scratch test), while no significant increase in wound area closure was observed in the control group, it was determined that the wound area closed significantly in the group to which *Echium italicum* acetone extract was applied (Figures4,5,6).



**Figure 5.** Acetone extraction group 0-24th hour surface coverage image



**Figure 6.** Comparison of wound opening area in acetone extraction application compared to control in HDF cell line.

### Discussion

*Echium italicum* is a plant belonging to the Boraginaceae family and is widely found in the Mediterranean region. The root parts of this plant have

been used for various purposes in traditional medicine (15). In this study, the medicinal properties of the *Echium italicum* plant recommended by traditional medicine were evaluated and the traditional use of *Echium Sp.* was brought to a rational basis. In addition,

the potential effects of the root extract of the *Echium italicum* plant on wound healing were examined. In the literature, the red roots of the *Echium italicum* plant are used especially in the treatment of burns (16). Based on these data, the effects of the plant on cell viability and wound healing using solvents not used in the literature were examined by following scientific methodology. The study and the results obtained revealed important findings that *Echium italicum* root extract can positively affect the wound healing process. First, the effect of the plant extract on cellular proliferation was evaluated and it was observed that it increased the proliferation ability of HDF cells when applied at different concentrations (Figure 1, 2, 3). This suggests that the plant extract can promote the regeneration of fibroblast cells. In addition, its effects on cellular migration, which plays an important role in the wound healing process, were examined. Scratch assay experiments showed that *Echium italicum* root extract increased the migration potential of HDF cells (Figure 4, 5, 6). In conclusion, this study reveals the positive effects of *Echium italicum* plant root extract on wound healing.

Studies show that phytochemical components such as naphthoquinones and flavonoids support the wound healing process mainly due to their antimicrobial properties that promote cell proliferation and granulation tissue and are responsible for wound contraction (17). It is considered that the high naphthoquinone content in *Echium italicum* roots may be responsible for the wound healing activity, probably by promoting cell proliferation and migration, and can form thick granulation tissue and re-epithelialization of

wounds, but these results need to be supported by further studies.


## Conclusions

The main purposes of analyzing crude plant extracts are to isolate bioactive agents either for direct use as drugs or to identify bioactive compounds that can be used as precursors for the preparation of semi-synthetic drugs. Numerous new wound healing drugs have been discovered from natural products in the past and new ones are being developed continuously. These natural products can play an important role in wound treatment by working together with traditional drugs, thus increasing their efficacy or reducing their toxicity (18,19). The results of our study indicate that *Echium italicum* plant extract has promising wound healing activities *in vitro*. It is expected that more comprehensive studies will be conducted in the future to obtain therapeutic implications of these results and to add new drugs to existing ones.

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## ORCID:

Derya AKDUMAN  0009-0008-8534-0608

Rahmiye AKDUMAN  0009-0005-6510-9482

Merve ŞİMŞEK GEYİK  0000-0002-4088-183X

Mesut AKYÜZ  0000-0001-8161-2479

## References

- Mifsud, S. (2002). *Echium italicum* (Pale Bugloss): MaltaWildPlants.com-the online Flora of the Maltese Islands.
- Uysal, İ., Mohammed, F. S., Şabik, A. E., Kına, E., & Sevindik, M. (2021). Antioxidant and Oxidant status of medicinal plant *Echium italicum* collected from different regions. Turkish Journal of Agriculture-Food Science and Technology, 9(10), 1902-1904.
- Jin, J., Zhang, Y., Xu, Y., & Zhang, Y. (2020). Antioxidant properties and reported ethnomedicinal use of the genus *Echium* (Boraginaceae). Antioxidants, 9(8), 722.
- Zhang, L., Ravipati, A. S., Koyyalamudi, S. R., Jeong, S. C., Reddy, N., Smith, P. T., & Wu, M. J. (2011). Antioxidant and anti-inflammatory activities of selected medicinal plants containing

- phenolic and flavonoid compounds. Journal of agricultural and food chemistry, 59(23), 12361-12367.
5. Balunas, M. J., & Kinghorn, A. D. (2005). Drug discovery from medicinal plants. Life sciences, 78(5), 431-441.
  6. Gibbons, S. (2005). Plants as a source of bacterial resistance modulators and anti-infective agents. Phytochemistry reviews, 4, 63-78.
  7. Alamholo, M. (2020). Antiradical and antibacterial activity of *Echium altissimum* extracts on human infective bacteria and chemical composition analysis. Microbiology, Metabolites and Biotechnology, 3(1), 19-27.
  8. Martin, P. (1997). Wound healing aiming for perfect skin regeneration. Science, 276(5309), 75-81.
  9. Diegelmann, R. F., & Evans, M. C. (2004). Wound healing: an overview of acute, fibrotic and delayed healing. Frontiers in Bioscience, 9, 283-289. doi:10.2741/1184
  10. Rodrigues, M., Kosaric, N., Bonham, C. A., & Gurtner, G. C. (2019). Wound healing: a cellular perspective. Physiological reviews, 99(1), 665-706.
  11. Abd Haghighi, M. J., Azadbakht, M., Habibi, E., Vahedi, L., & Hosseinimehr, S. J. (2024). Phytochemical profile, antioxidant, and wound healing activities of *Echium amoenum* (Boraginaceae). Natural Product Research, 1-7.
  12. Gami, B., & Parabia, F. (2011). Screening of methanol & acetone extract for antimicrobial activity of some medicinal plants species of Indian folklore. Int J Res Pharm Sci, 2(1), 69-75.
  13. Zhou, N., et al. (2022). Deferasirox shows inhibition activity against cervical cancer *in vitro* and *in vivo*. Gynecol Oncol 166(1): 126-137.
  14. Ozel Capik, Fatma Sanli, Ali Kurt, Onur Ceylan, Ilknur Suer, Murat Kaya, Michael Ittmann & Omer Faruk Karatas (2021). CASC11 promotes aggressiveness of prostate cancer cells through miR-145/IGF1R axis. Prostate Cancer and Prostatic Diseases volume 24, pages 891–902.
  15. Jin, J., Boersch, M., Nagarajan, A., Davey, A. K., & Zunk, M. (2020). Antioxidant properties and reported ethnomedicinal use of the genus *Echium* (Boraginaceae). Antioxidants, 9(8), 722.
  16. Pazyar, N., Yaghoobi, R., Rafiee, E., Mehrabian, A., & Feily, A. (2014). Skin wound healing and phytomedicine: a review. Skin pharmacology and physiology, 27(6), 303-310.
  17. Papageorgiou, V. P., Assimopoulou, A. N., Couladouros, E. A., Hepworth, D., & Nicolaou, K. C. (1999). The chemistry and biology of alkannin, shikonin, and related naphthazarin natural products. Angewandte Chemie International Edition, 38(3), 270-301.
  18. Maver T, Maver U, Kleinschek K S, Smrke D M, and Kreft S. A review of herbal medicines in wound healing. International Journal of Dermatology 2015, 54, 740–751
  19. Ghuman S., Ncube B., Finniea J.F., McGawa L.J., Njoya E M., R.M. Coopoosamyc R M., Stadena J V. Antioxidant, anti-inflammatory and wound healing properties of medicinal plant extracts used to treat wounds and dermatological disorders. South African Journal of Botany 126 (2019) 232–240.